

Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- | | | |
|-------------------------------------|-------------------------------------|--|
| <input type="checkbox"/> | <input checked="" type="checkbox"/> | The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> | A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> | The statistical test(s) used AND whether they are one- or two-sided
<i>Only common tests should be described solely by name; describe more complex techniques in the Methods section.</i> |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> | A description of all covariates tested |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> | A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> | A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals) |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> | For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
<i>Give P values as exact values whenever suitable.</i> |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> | For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> | For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> | Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated |

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection TECAN SparkControl, TCGA, GTEX, Ella Automated ELISA software,

Data analysis ImageJ, QuantStudio Design & Analysis Software, FastQC, samtools, Cutadapt, Trim Galore!, grape-nf pipeline, wrapping STAR, RSEM, R (DESeq2 package), Reactome website, NetworkAnalyst 3.0, FlowJo V.10, Adobe Photoshop, MS Excel, IMARIS 10.1, GraphPad Prism 7

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our [policy](#)

Provide your data availability statement here.

Research involving human participants, their data, or biological material

Policy information about studies with [human participants or human data](#). See also policy information about [sex, gender \(identity/presentation\), and sexual orientation](#) and [race, ethnicity and racism](#).

Reporting on sex and gender

Use the terms *sex* (biological attribute) and *gender* (shaped by social and cultural circumstances) carefully in order to avoid confusing both terms. Indicate if findings apply to only one sex or gender; describe whether sex and gender were considered in study design; whether sex and/or gender was determined based on self-reporting or assigned and methods used. Provide in the source data disaggregated sex and gender data, where this information has been collected, and if consent has been obtained for sharing of individual-level data; provide overall numbers in this Reporting Summary. Please state if this information has not been collected. Report sex- and gender-based analyses where performed, justify reasons for lack of sex- and gender-based analysis.

Reporting on race, ethnicity, or other socially relevant groupings

Please specify the socially constructed or socially relevant categorization variable(s) used in your manuscript and explain why they were used. Please note that such variables should not be used as proxies for other socially constructed/relevant variables (for example, race or ethnicity should not be used as a proxy for socioeconomic status). Provide clear definitions of the relevant terms used, how they were provided (by the participants/respondents, the researchers, or third parties), and the method(s) used to classify people into the different categories (e.g. self-report, census or administrative data, social media data, etc.) Please provide details about how you controlled for confounding variables in your analyses.

Population characteristics

Describe the covariate-relevant population characteristics of the human research participants (e.g. age, genotypic information, past and current diagnosis and treatment categories). If you filled out the behavioural & social sciences study design questions and have nothing to add here, write "See above."

Recruitment

Describe how participants were recruited. Outline any potential self-selection bias or other biases that may be present and how these are likely to impact results.

Ethics oversight

Identify the organization(s) that approved the study protocol.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

☒ Life sciences ☐ Behavioural & social sciences ☐ Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://www.nature.com/documents/nr-reporting-summary-flat.pdf)

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	Sample sizes were not predetermined. Cell numbers were chosen according to protocols provided by the kit suppliers.
Data exclusions	-
Replication	At least three technical replicates were employed for experiments. For sensitive assays (cytokine secretion, immune cell related assays) at least two biological replicates using different donors were generated. The samples sent for RNA sequencing were generated in biological triplicates to rule out small cell culture related variations.
Randomization	-
Blinding	-

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

n/a	Involved in the study
<input type="checkbox"/>	<input checked="" type="checkbox"/> Antibodies
<input type="checkbox"/>	<input checked="" type="checkbox"/> Eukaryotic cell lines
<input checked="" type="checkbox"/>	<input type="checkbox"/> Palaeontology and archaeology
<input checked="" type="checkbox"/>	<input type="checkbox"/> Animals and other organisms
<input checked="" type="checkbox"/>	<input type="checkbox"/> Clinical data
<input checked="" type="checkbox"/>	<input type="checkbox"/> Dual use research of concern
<input checked="" type="checkbox"/>	<input type="checkbox"/> Plants

Methods

n/a	Involved in the study
<input checked="" type="checkbox"/>	<input type="checkbox"/> ChIP-seq
<input type="checkbox"/>	<input checked="" type="checkbox"/> Flow cytometry
<input checked="" type="checkbox"/>	<input type="checkbox"/> MRI-based neuroimaging

Antibodies

Antibodies used	anti-CD25 AlexaFluor488 (BC96, BioLegend, UK), anti-CD69 AlexaFluor647 (FN50, BioLegend, UK), anti-Her2-CD3 BiTE antibody (BSAB-002, Creative Biolabs, USA), Anti-E-Cadherin (mouse, Proteintech, 60335-1-Ig), Anti-Integrin b1 (rabbit, Proteintech, 82729-3-RR) Anti-Integrin a1b1 (rabbit, Proteintech, 22146-1-AP), Anti-Integrin a2b1 (rabbit, Proteintech, 30703-1-AP), Anti-Integrin a5b1 (mouse, Proteintech, 68561-1-Ig), Anti-Integrin a6b1 (mouse, Proteintech, 66906-1-Ig), Anti-Calnexin (rabbit, Proteintech, 10427-2-AP), IRDye® 680LT Goat anti-Rabbit IgG Secondary Antibody (LI-COR Biosciences, 926-68021), IRDye® 800CW Goat anti-Rabbit IgG Secondary Antibody (LI-COR Biosciences, 926-32211), IRDye® 800CW Goat anti-Mouse IgG Secondary Antibody (LI-COR Biosciences, 926-32210)
Validation	Antibodies were used according to the suppliers provided recommendations. All antibodies were verified by the supplier for the methods employed in this study.

Eukaryotic cell lines

Policy information about [cell lines and Sex and Gender in Research](#)

Cell line source(s)	The immortalized cell lines Panc-1, MCF-7, Capan-1, Capan-2 and BT-747 are all derived from human. The primary T cells were isolated from human blood, no data on the sex of the donors was provided.
Authentication	Cell lines were verified by the supplier and not further authenticated.
Mycoplasma contamination	Cell lines were tested negative by the supplier, no further testing was performed.
Commonly misidentified lines (See ICLAC register)	-

Plants

Seed stocks	<i>Report on the source of all seed stocks or other plant material used. If applicable, state the seed stock centre and catalogue number. If plant specimens were collected from the field, describe the collection location, date and sampling procedures.</i>
Novel plant genotypes	<i>Describe the methods by which all novel plant genotypes were produced. This includes those generated by transgenic approaches, gene editing, chemical/radiation-based mutagenesis and hybridization. For transgenic lines, describe the transformation method, the number of independent lines analyzed and the generation upon which experiments were performed. For gene-edited lines, describe the editor used, the endogenous sequence targeted for editing, the targeting guide RNA sequence (if applicable) and how the editor was applied.</i>
Authentication	<i>Describe any authentication procedures for each seed stock used or novel genotype generated. Describe any experiments used to assess the effect of a mutation and, where applicable, how potential secondary effects (e.g. second site T-DNA insertions, mosaicism, off-target gene editing) were examined.</i>

Flow Cytometry

Plots

Confirm that:	
<input checked="" type="checkbox"/> The axis labels state the marker and fluorochrome used (e.g. CD4-FITC).	
<input checked="" type="checkbox"/> The axis scales are clearly visible. Include numbers along axes only for bottom left plot of group (a 'group' is an analysis of identical markers).	
<input type="checkbox"/> All plots are contour plots with outliers or pseudocolor plots.	
<input type="checkbox"/> A numerical value for number of cells or percentage (with statistics) is provided.	

Methodology

Sample preparation	T cells were isolated from human blood according to the instructions provided with the RosetteSep Human CD8+ T cell Enrichment Cocktail (STEMCELL technologies, Germany). Panc-1 cells were cultured according to the suppliers instructions.
Instrument	Attune NxT flow cytometer
Software	FlowJo V.10
Cell population abundance	-
Gating strategy	T cells: rough SSC/FSC gating was performed to discard any cell debris. FSC-A/FSC-H gating was employed to check for doublets. positive CD25 and CD69 populations were identified by using unstained and unstimulated cells as negative ctrls. Panc-1: rough SSC/FSC gating was performed to discard any cell debris. Positive Hoechst signal was used to separate the cell line from DSLBs, the cutoff point was determined via unstained ctrl. The Cyanine 5 signal was compared to a DSLB-free (and therefore Cyanine 5-free) sample.

☒ Tick this box to confirm that a figure exemplifying the gating strategy is provided in the Supplementary Information.