

**NO[•]/RUNX3/Kynurenine Metabolic Signaling Enhances Disease Aggressiveness
in Pancreatic Cancer**

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List of abbreviations:

NO[•]: Nitric Oxide
RUNX3: RUNX Family Transcription Factor 3
PDAC: Pancreatic ductal adenocarcinoma
NOS2: nitric oxide synthase 2 (inducible nitric oxide synthase)
IDO1: indoleamine-2,3-dioxygenase 1
IDO2: indoleamine-2,3-dioxygenase 2
TDO: Tryptophan dioxygenase
AHR: aryl hydrocarbon receptor
NFE2L2: Nuclear Factor, Erythroid 2 Like 2
NRF2: NFE2-Related Factor 2
SERPINB2: Serpin Family B Member 2
IL1b: interleukin 1, beta
IL6: interleukin 6
IL8: interleukin 8
UHPLC: ultra-high performance liquid chromatography
MS: mass spectrometry
SPER/NO: Spermine NONOate, N-[4-[1-(3-Aminopropyl)-2-hydroxy-2-nitrosohydrazino]butyl]-1,3-propanediamine
TCA: Trichloroacetic acid
ChIP: Chromatin immunoprecipitation
CH223191: 1-Methyl-N-[2-methyl-4-[2-(2-methylphenyl)diazenyl]phenyl]-1H-pyrazole-5-carboxamide
STR: short tandem repeat
qRT-PCR: quantitative real-time PCR
DAB: diaminobenzene

Novelty and Impact

Pancreatic cancer is a lethal malignancy with no effective treatment. Metabolic reprogramming is a key biological event supporting pancreatic cancer progression. However, the regulatory mechanisms of metabolic adaptations in pancreatic cancer is not adequately defined. This study illustrates an interactive role of inflammatory signaling and metabolic reprogramming by identifying a novel NO[•]/RUNX3/Kynurenine metabolic axis and its clinical relevance in disease aggressiveness, which may be potentially useful in designing novel approaches to improve patient outcome.

Abstract

Pancreatic ductal adenocarcinoma (PDAC) is a lethal malignancy and is refractory to available treatments. Delineating the regulatory mechanisms of metabolic reprogramming, a key event in pancreatic cancer progression, may identify candidate targets with potential therapeutic significance. We hypothesized that inflammatory signaling pathways regulate metabolic adaptations in pancreatic cancer. Metabolic profiling of tumors from PDAC patients with a high- (>median, N=31) and low-NOS2 (inducible nitric oxide synthase) (<median, N=32) mRNA expression was performed. Differentially abundant metabolites were analyzed and linked with patient survival. The functional role of the prognostically significant metabolite and the mechanism of its regulation by NOS2/NO[•] (nitric oxide)-mediated signaling pathway was elucidated. The level of kynurenine, a tryptophan metabolite, was associated with high NOS2 expression, and a higher level of kynurenine predicted poor survival in patients (N=63, *P* = 0.01). Gene expression analysis in PDAC tumors (N=63) showed a positive correlation between the expression of NOS2 and the tryptophan/kynurenine pathway genes, including indoleamine-2,3-dioxygenase 1 (IDO1) and several aryl hydrocarbon receptor (AHR)-target genes including *NFE2L2* (*NRF2*), *SERPINB2*, *IL1b*, *IL6* and *IL8*, which are implicated in pancreatic cancer. Consistently, treatment of pancreatic cancer cell lines with NO[•] donor induced IDO1, kynurenine production, and the expression of AHR-target genes. Furthermore, kynurenine treatment enhanced spheroid growth and invasive potential of pancreatic cancer cell lines. Mechanistically, NO[•]-induced IDO1/Kynurenine/AHR signaling was mediated by RUNX3 transcription factor.

Our findings identified a novel NO[•]/RUNX3/Kynurenine metabolic axis, which enhances disease aggressiveness in pancreatic cancer and may have potential translational significance in improving disease outcome.

Introduction

Pancreatic cancer is a lethal malignancy and remains the third leading cause of death due to cancer with an estimated 56,770 new cases and 45,750 deaths in 2019 alone in the United States ¹. Late diagnosis with locally advanced or metastatic disease and a largely ineffective response to available treatments contribute to the dismal patient outcome. Therefore, delineating the key biological pathways involved in pancreatic cancer progression and disease aggressiveness is essential to identify novel targets, achieve effective treatment and improve patient survival in this lethal malignancy. Recent advances in pancreatic cancer research have improved our understanding of pancreatic tumor biology and continues to provide rationale for developing potential treatment strategies. However, an effective therapeutic approach is still lacking and therefore novel clinically relevant targets and strategies are needed to improve patient outcome.

Metabolic reprogramming is one of the key events in pancreatic tumor biology which helps cancer cells to survive and grow under harsh microenvironment with depleted nutrient and oxygen level ²⁻⁴. Several of the metabolic adaptations including increase in glucose uptake, enhanced glycolysis, increased flux of glycolysis intermediates into anabolic pathways,

canonical and non-canonical glutamine metabolism, and activation of recycling and scavenging processes support an increased demand for energy and biosynthesis by tumor cells and also maintain an optimal redox balance to support growth and progression ³. Although critically important, metabolic adaptations are highly complex and context dependent dynamic events, and the mechanisms of their regulation are not yet completely understood. Therefore, understanding the regulation of metabolic adaptations in tumor may identify critical metabolic switch, which may be used as a candidate target to impede cancer growth and progression.

Inflammatory and immune mediators are implicated in the development and progression of pancreatic cancer. Many of these inflammatory mediators are produced by both inflammatory as well as tumor cells and are involved in mediating key inflammatory signaling pathways responsible for conferring survival and growth advantage ⁵. Interestingly, chronic inflammation may result in metabolic dysfunction, and the inflammatory/immune signaling and metabolic regulations are described as highly integrated events ⁶. Furthermore, inflammatory and metabolic processes, associated with obesity, cooperatively enhance tumorigenesis and chemoresistance ^{7,8}. In a recent study, inhibitor of kappa kinase subunit-epsilon (IKKe), which plays a central role in innate immunity, is reported to reprogram glucose metabolism and enhance pancreatic tumor growth in mouse model ⁹. Additionally, an increased expression of IKKe associated with poorer survival in patients with PDAC ¹⁰. However, the role of inflammatory signaling pathways in the regulation of metabolic adaptations is not clear in pancreatic cancer.

Nitric oxide (NO[•]) is an inflammatory and immune mediator and is generated during the conversion of arginine to citrulline, a reaction that is catalyzed by nitric oxide synthases (NOS), which is categorized as constitutive NOS [NOS1(neuronal NOS) and NOS3 (endothelial NOS)] and inducible NOS (NOS2) ¹¹⁻¹³. NO[•] plays an important role in several critical physiological processes including vasodilation, neurotransmission, immune regulation and host defense ¹². However, a high level of NO[•] production for a prolonged period may contribute to tumorigenesis. NOS2 is primarily responsible for a high and sustained level of NO[•] production during chronic inflammation and cancer. We have recently described that a higher NOS2 expression in tumors is associated with poor survival in resected PDAC patients and NOS2/NO[•] signaling enhances tumor progression and disease aggressiveness ^{14, 15}. Moreover, NOS2 is described as a potential mediator of immune suppression in PDAC ¹⁶. NO[•] is a modulator of bioenergetics pathways and may play a role in metabolic reprogramming in tumor cells for sustained growth and progression under low nutrient and hypoxic microenvironment ¹⁷. Consistently, a higher expression of NO[•] is reported to enhance glycolysis and lactate production in breast cancer ¹⁸. Furthermore, NO[•] is shown as a positive regulator of the Warburg effect in ovarian cancer ¹⁹. However, the role of NO[•]-mediated signaling in tumor metabolic reprogramming in pancreatic cancer is previously not described. In this study we tested the hypothesis that NOS2/NO[•]-signaling pathway regulate metabolic adaptations in PDAC. Our

findings identified a NO[•]/RUNX3/kynurenine metabolic signaling that enhances pancreatic cancer progression and disease aggressiveness.

Materials and Methods

Human samples

Fresh-frozen human primary tumor tissues from resected PDAC patients were collected at the University of Maryland Medical System (UMMS) at Baltimore, MD, through NCI-UMD resource contract and University Medical Center Göttingen, Germany. Tumor histopathology was classified by Board certified pathologists and the patient's characteristics are described in the Supplementary Table S1. NCI-Office of the Human Subject Research Protection (OHSRP), NIH, Bethesda, MD, approved the use of these clinical samples in our study (OHSRP Exempt # 4678)

Metabolic profiling and data analysis of PDAC

Metabolic profiling of 33 tumor samples from PDAC patients was performed at Metabolon Inc. by using standard protocol as described earlier³⁹⁻⁴¹. To describe briefly, Metabolon untargeted metabolic platform uses two separate ultra-high performance liquid chromatography/tandem mass spectrometry (UHPLC/MS/MS) injections and one gas chromatography/mass spectrometry injection for each sample. The data from this sample set was combined with our earlier metabolic profiling data sets (N=33 and N=31)⁴¹ Metabolic profiling of these sample sets was also

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conducted at Metabolon Inc. Only patients with stage I and II disease with follow up data were included in further analysis (N=63). We used a total of 235 common compounds of known identity to query all three datasets and normalized the merged data by assigning the median of each compound level to equal to 1 for each dataset termed “block normalization”. Missing data were imputed with the minimum observed value for each compound in each dataset. The combined dataset was log2 transformed and served for further data analysis (Supplementary Table S2).

Cell lines and culture condition

Human pancreatic cancer cell lines Panc-1 (RRID: CVCL_0480), CFPAC-1 (RRID: CVCL_1119), MIA PaCa-2 (RRID: CVCL_0428), AsPC-1 (RRID: CVCL_0152) and Capan-1 (RRID: CVCL_0237) were purchased from the ATCC and were authenticated by short tandem repeat (STR) analysis. All cell lines were mycoplasma-free. Cells were grown in RPMI 1640 medium, supplemented with 10% FBS and 1% penicillin–streptomycin in a humidified incubator containing 5% CO₂ at 37°C. All reagents for cell culture were purchased from Gibco/Thermo Fisher Scientific, Grand Island, NY.

NO[•] treatment, RNA isolation and quantitative real-time PCR (qRT-PCR)

Cells were plated in 24-well plates at 30-50% confluency the day before treatment. On the following day medium was replenished with fresh medium supplemented with 0.5mM nitric oxide donor drug Spermine/NONOate (SPER/NO) (Cayman Chemical, Ann Arbor, MI).

Following 48 hours of treatment, cells were collected for total RNA isolation. Total RNA was extracted from frozen tumor samples and cultured cells with standard TRIzol protocol (Invitrogen/Thermo Fisher Scientific, Grand Island, NY). First strand cDNA was synthesized with High Capacity cDNA Reverse Transcription Kit (Applied biosystems, Thermo Fisher Scientific, Grand Island, NY) from total RNA and used for quantitative real-time PCR using Taqman probes (Applied Biosystems/ Thermo Fisher Scientific, Grand Island, NY) for corresponding target genes: *IDO1* (Hs00984148_m1), *RUNX3* (Hs01091094_m1), *IDO2* (Hs01589373_m1), *TDO2* (Hs00194611_m1), *AHR* (Hs00907314_m1), *NRF2/NFE2L2* (Hs00975961_g1), *IL1b* (Hs01555410_m1), *IL6* (Hs00174131_m1), *IL8* (Hs00174103_m1), *SERPINB2* (Hs01010736_m1), *RUNX3* (Hs01091094_m1).

Measurement of kynurenine and tryptophan in cell culture medium

Cells were cultured in RPMI medium supplemented with 0.5 mM nitric oxide donor SPER/NO for 48 hours, then culture medium was collected and centrifuged at 12,000 rpm for 10 min. Supernatant was used for metabolite measurement. In a 500 μ l microcentrifuge tube, 20 μ l of samples or standards were mixed with 20 μ l of Isotopic Standard (IS), and 40 μ l of trichloroacetic acid (TCA, 0.6 N). The supernatant was transferred to a HPLC vial for analysis. LC/MS/MS was performed with a Shimadzu 20AC-XR system followed by a TSQ Quantiva triple quadrupole mass spectrometer (Thermo Fisher Scientific, Grand Island, NY) operating in selected reaction monitoring mode with positive electrospray ionization at NCI-Frederic Mass

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Spectrometry core facility. The target peaks were detected using the following m/z precursor/product ions: kynurenine (209/192); d6-kynurenine (215/198); tryptophan (205/188), d5-tryptophan (210/192). Kynurenine and tryptophan in the samples were determined using the Thermo Xcalibur software. Calibration curves constructed by plotting the peak area ratios vs. standard concentration were fitted by linear regressions ($R^2 > 0.99$). The peak area ratios were calculated by dividing the peak areas of kynurenine or tryptophan by the peak area of their respective isotopic standards. Analytical standards of tryptophan and kynurenine were obtained from Sigma-Aldrich, St. Louis, MO. The isotopic standards (IS) d5-Trp and d6-Kyn were obtained from Cambridge Isotope Laboratories, Tewksbury, MA. Stock solutions of the tryptophan metabolite standards and the IS were prepared in 50% acetonitrile. The calibration standards Kyn (0.08-5 μ M) and Trp (0.8-50 μ M) were prepared by diluting the stocks with water. The IS solution containing d6-Kyn (2 μ M) and d5-Trp (10 μ M) was prepared in water. Kynurenine and tryptophan concentration in the cell culture medium was measured before and after SPER/NO treatment. The level of kynurenine is presented as a ratio of kynurenine production to the tryptophan consumption.

Immunohistochemistry

5 μ m thick paraffin-embedded tumor sections were incubated with mouse monoclonal anti-NOS2 (BD Transduction Laboratories, Franklin Lakes, NJ) and rabbit monoclonal anti-IDO-1 antibody (Cell Signaling, Danvers, MA). Signals were amplified using biotinylated IgG,

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followed by horseradish peroxidase-conjugated avidin-biotin complex (Vectastain ABC Kit, Vector Lab, Burlingame, CA) and diaminobenzene (DAB) as the chromogen (Dako Envision System, Dako, Carpinter, CA). Immunostained slides were blindly evaluated by a board-certified pathologist to assign the intensity as well as prevalence score as described elsewhere ⁴². A score of 0-3, representing negative, weak, moderate or strong expression was assigned for intensity and prevalence was given a score of 0-4, representing <10%, 10-30%, >30-50%, >50-80% and >80% cells showing either NOS2 or IDO1 expression. Then the overall IHC score was obtained by adding the intensity and prevalence score.

Cell migration and invasion assay

Migration and invasion assays were performed with 24-well BD Falcon Cell Culture Insert and Corning Bio Coat Matrigel Invasion Chamber (BD Biosciences, Bedford, MA) using manufacturer's protocol. Briefly, 10^5 cells in 500 μ l serum-free DMEM were loaded into each upper insert, and 750 μ l of appropriate medium with 10% FBS were added to the lower chamber. Kynurenine at 0, 10, 50 and 100 μ M dose was supplemented in both upper and lower chambers.

Spheroid formation assay

Spheroid formation assay was performed using AlgiMatrix 3D Culture System (Gibco/Thermo Fisher Scientific, Grand Island, NY). 10^5 cells were resuspended in culture medium supplemented with 0, 10, 50, and 100 μ M kynurenine or DMSO and plated into a 24-well plate

containing the 3D culture matrix. Cells were then cultured for 10 days in the presence of kynurenine, and medium was replenished every other day. Cells were then collected by dissolving the matrix and then fixed with 3.7% formaldehyde, stained with crystal violet, counted and measured with Image J software, NIH (<https://imagej.nih.gov/ij/>). More than 100 spheroids were measured for each treatment and control groups.

Chromatin immunoprecipitation (ChIP)

Chromatin immunoprecipitation (ChIP) was performed with ChIP-IT High Sensitivity® kit from Active Motif (Carlsbad, CA) following manufacture's protocol. Briefly, for each immunoprecipitation reaction, 10^7 cells were treated with or without 0.5 mM Sper/NO for 24h and then fixed with formaldehyde for 15 min. Nuclear fraction was then prepared according to manufacturer's protocol and chromatin was sheared to 500-800 bp fragments with a Bioruptor sonication device (Diagenode, Denville, NJ). 5% of total extract was saved as input and the rest was used for immunoprecipitation with RUNX3 antibody (Santa Cruz, Dallas, TX). Recovered DNA samples were used as template for qRT-PCR with the following IDO1 primers purchased from IDT DNA Technology (Coralville, IA): positive primers: 5'-GCACAGAGATGCTTTTGTGGTTT-3' and 5'-CCCCTCAGTGTCTGAAGAGTT-3'; negative primers: 5'-GCATTCAATCAAATAGCAACAAC-3' and 5'-CAGCCATGAAGCAATGGACC-3'.

Statistical Analysis

Student t-test was used to evaluate the difference in mRNA expression, metabolite abundance level, cell spheroids size, invasion and immunohistochemistry score among different experimental groups. Kaplan-Meier analysis and Log Rank test was utilized to assess the difference in survival between two groups of PDAC patients. Pearson's correlation was used to determine any correlation between the expression of genes or metabolite levels. These statistical analyses were performed using GraphPad Prism 7.0. Univariable and multivariable Cox-regression analysis was performed using STATA 15 (StataCorp LP, College Station, TX). For all the statistical analyses *P* value <0.05 was deemed statistically significant.

Data availability

All the data and other items supporting the results in the paper are included as the supplementary materials. Additionally, any data that are minimally required to replicate the outcomes of the study will be made available upon reasonable request.

Results

Kynurenine positively associated with NOS2 and predicted prognosis in PDAC patients

We compared metabolic profile of tumors with a high (>median)- and low (<median)-NOS2 gene expression in a cohort of 63 PDAC patients (Figure 1A) (patients characteristics are described in Supplementary Table S1). Seventeen metabolites with ≥ 1.5 -fold difference in their

relative level and a statistical significance of $P < 0.05$ were identified in NOS2-high versus NOS2-low tumors. Among these metabolites the level of kynurenine showed the most significant association with survival by univariable Cox-regression analysis (HR = 2.24, 95% CI, 1.18-4.24; $P = 0.01$) (Supplementary Table S3), therefore, we investigated the role of kynurenine and its potential regulation by NOS2/NO^{*} signaling in PDAC. A higher level of kynurenine was found in high-NOS2 tumors as compared to low-NOS2 tumors ($P = 0.026$), and patients with a higher level of kynurenine in tumors (upper quartile) showed poorer survival by Kaplan-Meier (Log Rank test, $P = 0.01$) (Fig 1A-C) and multivariable (HR = 2.61, 95% CI, 1.24-5.51; $P = 0.01$) Cox regression analysis (Supplementary Table S4). As kynurenine is a product of tryptophan metabolism, which is mediated by indoleamine 2,3-dioxygenase enzymes (IDO1 and IDO2) and tryptophan dioxygenase (TDO), we examined their association in PDAC tumors. Tumors with a higher (>median) IDO1 expression showed a higher kynurenine level as compared with tumors with low IDO1 expression (<median), and a positive correlation was observed between kynurenine level and IDO1 gene-expression (Supplementary Figure S1A, B). Although NOS2 positively correlated with TDO expression, no significant correlation was observed between NOS2 and IDO2 or Kynurenine and IDO2 or TDO (Supplementary Figure S1C). Taken together, these findings showed that the level of kynurenine in tumors is positively correlated with NOS2 expression and a higher kynurenine level is associated with poorer survival in PDAC patient. Furthermore, based on these findings, we hypothesized that NOS2/NO^{*} signaling may enhance kynurenine level in tumors through the upregulation of IDO1.

NO[•] enhanced kynurenine production, IDO-1 expression and IDO-1/aryl hydrocarbon receptor (AHR)-target genes

To test the hypothesis that NO[•] signaling enhances the production of kynurenine in pancreatic cancer, Panc-1, CFPac-1 and AsPC-1 pancreatic cancer cell lines were treated with 0.5 mM NO[•] donor drug Spermine/NONOate (Sper/NO), which resulted in an increased production of kynurenine as compared to control cells ($P < 0.001$) (Fig 2A). We then checked, if NO[•] affects the expression of IDO1 gene, which catalyzes the production of kynurenine and positively associated with Kynurenine in our analysis of tumors from PDAC patients, as described above (Supplementary Fig S1A, B). Sper/NO-treatment significantly enhanced the expression of IDO1 in pancreatic cancer cell lines ($p < 0.01$) (Fig 2B), suggesting that NO[•]-mediated increase in IDO1 may be responsible for the increased production of kynurenine. It is earlier described that endogenously produced kynurenine binds to aryl hydrocarbon receptor (AHR) in tumor cells leading to its translocation into the nucleus and activation of AHR-target genes, which promoted tumor cell survival and migration in human gliomas²⁰. Therefore, we examined the expression of known Kynurenine/AHR target genes following NO[•] treatment in pancreatic cancer cells. NO[•] treatment significantly increased the expression of *NFE2L2* (Nrf2), *SERPINB2*, *IL1b*, *IL6* and *IL8*, which are implicated in the development and progression of pancreatic cancer (Fig 2C). Furthermore, an AHR antagonist, CH223191, suppressed the NO[•]-induced increase in expression of these target genes. Taken together these findings revealed that NO[•] signaling enhances IDO1

expression and kynurenine production, which subsequently augmented the expression of AHR-target genes.

NOS2, IDO-1 and AHR target genes expression are correlated in human PDAC

To understand the relevance of NO[•] induced signaling in the regulation of AHR-target genes in human disease, we analyzed tumors from patients with PDAC (N = 63). Tumors with a higher NOS2 expression (>median) also expressed a high level of IDO1 as compared with tumors with a lower NOS2 expression (<median) ($P = 0.006$) and a positive correlation existed between NOS2 and IDO1 ($r = 0.43$, $P = 0.0005$) (Fig 3A, B). Consistent with these findings immunohistochemical staining showed a positive correlation between NOS2 and IDO1 protein expression ($r = 0.70$, $P = 0.002$) (Fig 3C). Additionally, high-NOS2 tumors showed an enhanced expression of AHR-target genes *NFE2L2* (Nrf2), *SERPINB2*, *IL1b*, *IL6* and *IL8* as compared to low-NOS2 tumors and a positive correlation existed between NOS2 and the above-mentioned AHR-target genes (Fig 3D, E). Furthermore, IDO1-high (>median) tumors also showed an upregulation in the expression of AHR-target genes and a positive correlation was found between IDO1 and AHR target genes *NFE2L2* (Nrf2), *SERPINB2*, *IL1b*, *IL6* and *IL8* (Fig 3F, G). These findings provide evidence of an association between NOS2, IDO1 and the regulation of AHR-target genes in human PDAC.

Kynurenine promoted growth of spheroids and invasiveness in pancreatic cancer cells

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To examine the effect of kynurenine on cellular growth, Capan-1 pancreatic cancer cell spheroids were grown in 3D cultures and treated with 0, 10, 50 and 100 μ M doses of kynurenine. Treatment with kynurenine resulted in an enhanced growth of spheroids as compared to control in a dose dependent manner (Fig 4 A, B). Next, we examined if kynurenine affects the invasive ability of pancreatic cancer cells. Panc-1 and Capan-1 pancreatic cancer cell lines were treated with different doses of kynurenine, as described above, which resulted in the increased invasiveness in a dose dependent manner (Fig 4 C, D). These findings demonstrate that kynurenine enhances growth and invasiveness of pancreatic cancer cells.

NO[•]-mediated regulation of kynurenine signaling involves RUNX3 transcription factor

As described above, our findings showed that NO[•] enhances IDO1 expression and kynurenine production, which results in the increased expression of Kynurenine/AHR-target genes. To further investigate the mechanism of NO[•]-mediated regulation of kynurenine signaling, we first investigated the potential IDO1 regulation by using UCSC genome browser and the transcription factor ChIP-seq data from the ENCODE Factorbook repository ²¹. This search identified ChIP-seq peak for the binding of RUNX3 transcription factor to IDO1 promoter (Fig 5A).

Furthermore, RUNX3 positively correlated with IDO1 ($r = 0.73$, $P < 0.0001$) and kynurenine ($r = 0.32$, $P = 0.01$) in tumors from patients with PDAC (Fig 5 B, C). Additionally, high-NOS2 tumors showed a higher expression of RUNX3 as compared to low-NOS2 tumors and a positive correlation existed between NOS2 and RUNX3 ($r = 0.52$, $p < 0.0001$) (Fig 5 D, E). Based on

these findings we hypothesized that NO[•] may upregulate RUNX3 in pancreatic cancer.

Treatment of pancreatic cancer cell lines Panc-1 and MiaPaCa-2 with NO[•] donor SPER/NO enhanced the mRNA and protein expression of RUNX3 (Fig 5F, G). Furthermore, SPER/NO treatment enhanced the binding of RUNX3 to IDO1 promoter in these cell lines as determined by ChIP-PCR assay (Fig 5H). Taken together, these findings show that NO[•] enhances the expression of RUNX3 and facilitates its binding to IDO1 promoter and also provides evidence of an association between NOS2 and RUNX3, RUNX3 and IDO-1, and RUNX3 and kynurenine production in human PDAC.

Discussion

Inflammatory signaling pathways and metabolic alterations are key biological elements supporting pancreatic cancer progression and disease aggressiveness²²⁻²⁵. These events may independently and/or interactively contribute to tumorigenesis. Inflammatory mediators including cytokines, chemokines and free radicals are implicated in influencing metabolic alterations in tumorigenesis⁶. However, the role of inflammatory signaling pathways in the regulation of metabolic adaptations are not adequately described in pancreatic cancer. One of the inflammatory and immune mediators, NO[•], is implicated in tumorigenesis^{5, 15}. NOS2/NO[•] signaling enhances pancreatic cancer progression and a higher NOS2 expression is associated with poor patient survival¹⁴. In this study, we have investigated the role of NOS2/NO[•]-mediated

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signaling pathway in the regulation of metabolic alterations in human PDAC. Untargeted metabolic profiling of tumors from PDAC patients with a high- (>median) and low (<median)-NOS2 expression identified a previously undescribed NO[•]/RUNX3/kynurenine metabolic signaling, which enhanced the expression of AHR-target genes involved in tumorigenesis and predict poor patient outcome.

Kynurenine is a metabolite produced by tryptophan metabolism, which is catalyzed by indoleamine 2,3-dioxygenase-1 and 2 (IDO1/2) and tryptophan dioxygenase (TDO) enzymes in both tumor and immune cells²⁶. Kynurenine is an endogenous ligand to aryl hydrocarbon receptor (AHR)²⁰. Kynurenine binds to AHR in both autocrine and paracrine fashion, which results in the nuclear translocation of AHR and expression of AHR-target genes leading to immune tolerance and migration of tumor cells^{20, 27}. However, the role of kynurenine in pancreatic cancer growth and progression is not clear. Our findings of an association between increased level of kynurenine in tumors and worse survival in early stage resected patients suggested a potential role of kynurenine in pancreatic cancer progression and disease aggressiveness. However, this analysis has been conducted in patients from a single cohort and further analysis in additional independent cohort would strengthen this observation. Additionally, enhanced growth of pancreatic cancer cell spheroids and migration of pancreatic cancer cells following kynurenine treatment in a dose dependent manner are consistent with the hypothesis that kynurenine enhances pancreatic cancer growth and progression.

Understanding the regulation of metabolic adaptations may identify critical metabolic regulatory elements that may be potentially targeted to impede tumor progression in PDAC. As mentioned earlier, inflammatory mediators are implicated in the modulation of metabolic processes. An increased level of kynurenine in NOS2-high as compared to NOS2-low tumors in PDAC patients in our study suggested an association between NOS2/NO[•]-signaling pathway and kynurenine production. Enhanced kynurenine production in pancreatic cancer cell lines upon treatment with NO[•] donor SPER/NO is consistent with this hypothesis. It is earlier described that IDO1/2 and TDO catalyzes tryptophan metabolism and production of kynurenine. In line with these earlier studies PDAC tumors with higher IDO1 expression showed a higher level of kynurenine as compared to tumors with a low IDO1 and there existed a positive correlation between IDO1 and Kynurenine. Additionally, based on our findings showing a higher IDO1 gene expression in NOS2-high tumors as compared to NOS2-low tumors and a positive correlation between NOS2 and IDO1, we hypothesized that NOS2/NO[•] signaling may regulate IDO1 expression. Our data showing an increased IDO1 expression following treatment of pancreatic cancer cells with NO[•] are consistent with this hypothesis. Further mechanistic and functional analysis to elucidate NO[•]-induced kynurenine signaling identified NO[•]-induced RUNX3 transcription factor which regulated the expression of IDO1 through binding to its promoter. A positive correlation between RUNX3 and NOS2, RUNX3 and IDO1, and RUNX3 and kynurenine in tumors further indicated their relevance in human PDAC. RUNX3 has been

earlier defined as a critical regulator of pancreatic cancer growth and metastasis ²⁸. These findings suggested that NO[•]-induced increase in kynurenine production is mediated through RUNX3 and IDO1 in PDAC.

Kynurenine binding to AHR results in the expression of AHR-target genes which enhances malignant progression of human gliomas ²⁰. NO[•]-treatment enhanced the expression of several AHR-target genes including *NFE2L2* (Nrf2), *SERPINB2*, *IL-1b*, *IL-6* and *IL8*, which could be blocked by an AHR-antagonist CH223191 in pancreatic cancer cell lines, confirming that NO[•]-induced expression of these genes is mediated through the activation of AHR. Furthermore, a higher expression of these AHR-target genes in NOS2-high and IDO1-high tumors as compared to NOS2-low and IDO1-low tumors indicate the potential clinical relevance of NO[•]-induced expression of AHR-target genes in human PDAC. *NFE2L2* encodes for Nrf2, which is a key regulator of antioxidant machinery and favors pancreatic cancer growth and progression ²⁹. Similarly, cytokines including *IL-1b*, *IL-6* and *IL-8* are implicated in tumorigenesis and their increased expression is associated with the development, progression and therapeutic resistance in pancreatic cancer ³⁰⁻³⁵. The role of *SERPINB2*, which encodes the plasminogen activator inhibitor 2, is context dependent in cancer and both tumor suppressing and promoting functions are reported ^{36, 37}. However, a potential tumor suppressive role of *SERPINB2* is earlier described in pancreatic cancer ³⁸. Therefore, the significance of *SERPINB2* in the context of NO[•]/Kynurenine/AHR mediated signaling in pancreatic cancer need further

clarification. These findings suggested that NO[•]-mediated upregulation of kynurenine, AHR activation and expression of AHR target genes are biologically relevant to the progression of pancreatic cancer.

Taken together, our findings describe a novel NO[•]/RUNX3/kynurenine metabolic signaling pathway (Figure 6), which enhances pancreatic cancer progression and disease aggressiveness and may be useful in designing novel strategies to improve patient outcome.

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References

1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2019. *CA Cancer J Clin* 2019;69:7-34.
2. Sousa CM, Biancur DE, Wang X, Halbrook CJ, Sherman MH, Zhang L, Kremer D, Hwang RF, Witkiewicz AK, Ying H, Asara JM, Evans RM, et al. Pancreatic stellate cells support tumour metabolism through autophagic alanine secretion. *Nature* 2016;536:479-83.
3. Sousa CM, Kimmelman AC. The complex landscape of pancreatic cancer metabolism. *Carcinogenesis* 2014;35:1441-50.

- Accepted Article
4. Perera RM, Bardeesy N. Pancreatic Cancer Metabolism: Breaking It Down to Build It Back Up. *Cancer Discov* 2015;5:1247-61.
 5. Hussain SP, Hofseth LJ, Harris CC. Radical causes of cancer. *Nature Reviews Cancer* 2003;3:276-85.
 6. Hotamisligil GS. Inflammation and metabolic disorders. *Nature* 2006;444:860-7.
 7. Lashinger LM, Rossi EL, Hursting SD. Obesity and resistance to cancer chemotherapy: interacting roles of inflammation and metabolic dysregulation. *Clin Pharmacol Ther* 2014;96:458-63.
 8. Harvey AE, Lashinger LM, Hays D, Harrison LM, Lewis K, Fischer SM, Hursting SD. Calorie restriction decreases murine and human pancreatic tumor cell growth, nuclear factor-kappaB activation, and inflammation-related gene expression in an insulin-like growth factor-1-dependent manner. *PLoS One* 2014;9:e94151.
 9. Zubair H, Azim S, Srivastava SK, Ahmad A, Bhardwaj A, Khan MA, Patel GK, Arora S, Carter JE, Singh S, Singh AP. Glucose Metabolism Reprogrammed by Overexpression of IKKepsilon Promotes Pancreatic Tumor Growth. *Cancer Res* 2016;76:7254-64.
 10. Cheng A, Guo J, Henderson-Jackson E, Kim D, Malafa M, Coppola D. IkappaB Kinase epsilon expression in pancreatic ductal adenocarcinoma. *Am J Clin Pathol* 2011;136:60-6.
 11. Tamir S, Tannenbaum SR. The role of nitric oxide (NO.) in the carcinogenic process. *Biochim Biophys Acta* 1996;1288:F31-6.
 12. Bredt DS, Snyder SH. Nitric oxide: a physiologic messenger molecule. *Annual review of biochemistry* 1994;63:175-95.
 13. Geller DA, Billiar TR. Molecular biology of nitric oxide synthases. *Cancer Metastasis Rev* 1998;17:7-23.
 14. Wang J, He P, Gaida M, Yang S, Schetter AJ, Gaedcke J, Ghadimi BM, Ried T, Yfantis H, Lee D, Weiss JM, Stauffer J, et al. Inducible nitric oxide synthase enhances disease aggressiveness in pancreatic cancer. *Oncotarget* 2016;7:52993-3004.
 15. Wang J, Hussain SP. NO(*) and Pancreatic Cancer: A Complex Interaction with Therapeutic Potential. *Antioxid Redox Signal* 2017;26:1000-08.

- Accepted Article
16. Bailey P, Chang DK, Forget MA, Lucas FA, Alvarez HA, Haymaker C, Chattopadhyay C, Kim SH, Ekmekcioglu S, Grimm EA, Biankin AV, Hwu P, et al. Exploiting the neoantigen landscape for immunotherapy of pancreatic ductal adenocarcinoma. *Sci Rep* 2016;6:35848.
 17. Chang CF, Diers AR, Hogg N. Cancer cell metabolism and the modulating effects of nitric oxide. *Free Radic Biol Med* 2015;79:324-36.
 18. De Vitto H, Mendonca BS, Elseth KM, Onul A, Xue J, Vesper BJ, Gallo CV, Rumjanek FD, Paradise WA, Radosevich JA. Part III. Molecular changes induced by high nitric oxide adaptation in human breast cancer cell line BT-20 (BT-20-HNO): a switch from aerobic to anaerobic metabolism. *Tumour Biol* 2013;34:403-13.
 19. Caneba CA, Yang L, Baddour J, Curtis R, Win J, Hartig S, Marini J, Nagrath D. Nitric oxide is a positive regulator of the Warburg effect in ovarian cancer cells. *Cell Death Dis* 2014;5:e1302.
 20. Opitz CA, Litzenburger UM, Sahm F, Ott M, Tritschler I, Trump S, Schumacher T, Jestaedt L, Schrenk D, Weller M, Jugold M, Guillemin GJ, et al. An endogenous tumour-promoting ligand of the human aryl hydrocarbon receptor. *Nature* 2011;478:197-203.
 21. Wang J, Zhuang J, Iyer S, Lin X, Whitfield TW, Greven MC, Pierce BG, Dong X, Kundaje A, Cheng Y, Rando OJ, Birney E, et al. Sequence features and chromatin structure around the genomic regions bound by 119 human transcription factors. *Genome Res* 2012;22:1798-812.
 22. Ying H, Kimmelman AC, Lyssiotis CA, Hua S, Chu GC, Fletcher-Sananikone E, Locasale JW, Son J, Zhang H, Coloff JL, Yan H, Wang W, et al. Oncogenic Kras maintains pancreatic tumors through regulation of anabolic glucose metabolism. *Cell* 2012;149:656-70.
 23. Shukla SK, Purohit V, Mehla K, Gunda V, Chaika NV, Vernucci E, King RJ, Abrego J, Goode GD, Dasgupta A, Illies AL, Gebregiworgis T, et al. MUC1 and HIF-1 α Signaling Crosstalk Induces Anabolic Glucose Metabolism to Impart Gemcitabine Resistance to Pancreatic Cancer. *Cancer Cell* 2017;32:71-87 e7.
 24. Son J, Lyssiotis CA, Ying H, Wang X, Hua S, Ligorio M, Perera RM, Ferrone CR, Mullarky E, Shyh-Chang N, Kang Y, Fleming JB, et al. Glutamine supports pancreatic cancer growth through a KRAS-regulated metabolic pathway. *Nature* 2013;496:101-5.

- Accepted Article
25. Gukovsky I, Li N, Todoric J, Gukovskaya A, Karin M. Inflammation, autophagy, and obesity: common features in the pathogenesis of pancreatitis and pancreatic cancer. *Gastroenterology* 2013;144:1199-209 e4.
 26. Zhai L, Spranger S, Binder DC, Gritsina G, Lauing KL, Giles FJ, Wainwright DA. Molecular Pathways: Targeting IDO1 and Other Tryptophan Dioxygenases for Cancer Immunotherapy. *Clin Cancer Res* 2015;21:5427-33.
 27. Prendergast GC. Cancer: Why tumours eat tryptophan. *Nature* 2011;478:192-4.
 28. Whittle MC, Izeradjene K, Rani PG, Feng L, Carlson MA, DelGiorno KE, Wood LD, Goggins M, Hruban RH, Chang AE, Calses P, Thorsen SM, et al. RUNX3 Controls a Metastatic Switch in Pancreatic Ductal Adenocarcinoma. *Cell* 2015;161:1345-60.
 29. DeNicola GM, Karreth FA, Humpton TJ, Gopinathan A, Wei C, Frese K, Mangal D, Yu KH, Yeo CJ, Calhoun ES, Scrimieri F, Winter JM, et al. Oncogene-induced Nrf2 transcription promotes ROS detoxification and tumorigenesis. *Nature* 2011;475:106-9.
 30. Zhang D, Li L, Jiang H, Li Q, Wang-Gillam A, Yu J, Head R, Liu J, Ruzinova MB, Lim KH. Tumor-Stroma IL1beta-IRAK4 Feedforward Circuitry Drives Tumor Fibrosis, Chemoresistance, and Poor Prognosis in Pancreatic Cancer. *Cancer Res* 2018;78:1700-12.
 31. Biffi G, Oni TE, Spielman B, Hao Y, Elyada E, Park Y, Preall J, Tuveson DA. IL1-Induced JAK/STAT Signaling Is Antagonized by TGFbeta to Shape CAF Heterogeneity in Pancreatic Ductal Adenocarcinoma. *Cancer Discov* 2018.
 32. Zhang Y, Yan W, Collins MA, Bednar F, Rakshit S, Zetter BR, Stanger BZ, Chung I, Rhim AD, di Magliano MP. Interleukin-6 is required for pancreatic cancer progression by promoting MAPK signaling activation and oxidative stress resistance. *Cancer Res* 2013;73:6359-74.
 33. Mace TA, Shakya R, Pitarresi JR, Swanson B, McQuinn CW, Loftus S, Nordquist E, Cruz-Monserrate Z, Yu L, Young G, Zhong X, Zimmers TA, et al. IL-6 and PD-L1 antibody blockade combination therapy reduces tumour progression in murine models of pancreatic cancer. *Gut* 2018;67:320-32.
 34. Chen L, Fan J, Chen H, Meng Z, Chen Z, Wang P, Liu L. The IL-8/CXCR1 axis is associated with cancer stem cell-like properties and correlates with clinical prognosis in human pancreatic cancer cases. *Sci Rep* 2014;4:5911.

- Accepted Article
35. Matsuo Y, Ochi N, Sawai H, Yasuda A, Takahashi H, Funahashi H, Takeyama H, Tong Z, Guha S. CXCL8/IL-8 and CXCL12/SDF-1alpha co-operatively promote invasiveness and angiogenesis in pancreatic cancer. *Int J Cancer* 2009;124:853-61.
 36. Jin T, Suk Kim H, Ki Choi S, Hye Hwang E, Woo J, Suk Ryu H, Kim K, Moon A, Kyung Moon W. microRNA-200c/141 upregulates SerpinB2 to promote breast cancer cell metastasis and reduce patient survival. *Oncotarget* 2017;8:32769-82.
 37. Huang Z, Li H, Huang Q, Chen D, Han J, Wang L, Pan C, Chen W, House MG, Nephew KP, Guo Z. SERPINB2 down-regulation contributes to chemoresistance in head and neck cancer. *Mol Carcinog* 2014;53:777-86.
 38. Harris NLE, Vennin C, Conway JRW, Vine KL, Pinese M, Cowley MJ, Shearer RF, Lucas MC, Herrmann D, Allam AH, Pajic M, Morton JP, et al. SerpinB2 regulates stromal remodelling and local invasion in pancreatic cancer. *Oncogene* 2017;36:4288-98.
 39. Evans AM, DeHaven CD, Barrett T, Mitchell M, Milgram E. Integrated, nontargeted ultrahigh performance liquid chromatography/electrospray ionization tandem mass spectrometry platform for the identification and relative quantification of the small-molecule complement of biological systems. *Anal Chem* 2009;81:6656-67.
 40. Budhu A, Roessler S, Zhao X, Yu Z, Forgues M, Ji J, Karoly E, Qin LX, Ye QH, Jia HL, Fan J, Sun HC, et al. Integrated metabolite and gene expression profiles identify lipid biomarkers associated with progression of hepatocellular carcinoma and patient outcomes. *Gastroenterology* 2013;144:1066-75 e1.
 41. Zhang G, He P, Tan H, Budhu A, Gaedcke J, Ghadimi BM, Ried T, Yfantis HG, Lee DH, Maitra A, Hanna N, Alexander HR, et al. Integration of Metabolomics and Transcriptomics Revealed a Fatty Acid Network Exerting Growth Inhibitory Effects in Human Pancreatic Cancer. *Clin Cancer Res* 2013.
 42. Glynn SA, Boersma BJ, Dorsey TH, Yi M, Yfantis HG, Ridnour LA, Martin DN, Switzer CH, Hudson RS, Wink DA, Lee DH, Stephens RM, et al. Increased NOS2 predicts poor survival in estrogen receptor-negative breast cancer patients. *J Clin Invest* 2010;120:3843-54.

Figure Legends

Figure 1.

A higher level of kynurenine associated with increased NOS2 expression in tumors and predicted poorer survival in PDAC patients. **A**, NOS2 mRNA expression, as determined by qRT-PCR, in tumors from NOS2-high (>median) and -low (<median) groups of patients. **B**, Violin plot showing an increased level of kynurenine in NOS2-high as compared with NOS2-low tumors. **C**, Kaplan-Meier curve showing that a higher level of kynurenine (>75th percentile) is associated with poorer patient survival as compared with patients with low kynurenine (<75th percentile).

Figure 2.

NO[•] regulates kynurenine production and expression of AHR-target genes. **A-B**, Treatment of pancreatic cancer cell lines with 0.5 mM SPER/NO (NO[•] donor drug) enhanced kynurenine production and mRNA expression of IDO1 after 48 hrs. **C**, SPER/NO-treatment also resulted in the increased expression of AHR-target genes as compared to control. The increase in AHR-target genes expression was attenuated by CH223191, an AHR antagonist, when used in combination with SPER/NO. Each experiment was repeated at least three times.

Figure 3.

NOS2, IDO-1 and AHR-target genes are positively correlated in human PDAC. **A**, Tumors from PDAC patients with high NOS2 expression (>median) showed a higher IDO1 expression as compared with tumors with low NOS2 expression (<median). **B**, A positive correlation between NOS2 and IDO1 gene expression. **C**, Immunohistochemical staining showing an increased IDO1 protein expression in NOS2-high as compared with NOS2-low tumors. A positive correlation was found between NOS2 and IDO1 protein expression as determined by using IHC score. **D**, NOS2-high tumors showed a positive correlation with AHR-target genes: NFE2L2 (Nrf2), SERPINB2, IL1b, IL6 and IL8 as compared to NOS2-low tumors. **E**, A positive correlation was found between NOS2 and NFE2L2 (Nrf2), SERPINB2, IL1b, IL6 and IL8. **F**, Tumors from PDAC patients with high IDO1 expression (>median) showed a higher NFE2L2 (Nrf2), SERPINB2, IL1b, IL6 and IL8 expression as compared with tumors with low IDO-1 expression. **G**, A positive correlation existed between IDO1 and NFE2L2, SERPINB2, IL1b, IL6 and IL8 expression in tumors from PDAC patients.

Figure 4.

Kynurenine enhances pancreatic cancer cell spheroid growth and invasiveness. **A-B**, Kynurenine treatment of pancreatic cancer cell line Capan-1 enhanced growth of spheroids in a dose dependent manner (a minimum of 100 spheroids were measured for each treatment group using Image j, NIH, software). **C**, Kynurenine treatment enhanced invasion of pancreatic cancer cell lines. All the experiments were repeated three times.

Figure 5.

NO[•]-induced regulation of kynurenine signaling is mediated by RUNX3 transcription factor. **A**, ENCODE ChIP-seq data analysis identified RUNX3 binding site on IDO1 promoter. **B-C**, RUNX3 showed a positive correlation with IDO1 gene expression and kynurenine level in tumors from PDAC patients. **D**, NOS2-high tumors expressed a higher level of RUNX3 as compared with NOS2-low tumors. **E**, RUNX3 positively correlated with NOS2 expression in human PDAC tumors. **F-G**, Treatment of pancreatic cancer cell lines with 0.5mM SPER/NO (NO[•] donor drug) enhanced RUNX3 expression as shown by qRT-PCR and immunoblotting. **H**, ChIP-qPCR showing an enhanced binding of RUNX3 to IDO1 promoter in pancreatic cancer cell lines following SPER/NO treatment.

Figure 6.

NO[•]-RUNX3-kynurenine metabolic axis: Schema showing NO[•]-RUNX3-Kynurenine axis, which enhances the expression of protumorigenic AHR-target genes in pancreatic cancer.

Supplementary Figure S1

IDO1 expression in human PDAC tumors positively correlated with level of kynurenine

A, IDO1-high (>median) tumors showed a higher abundance of kynurenine as compared to IDO1-low (>median) tumors in PDAC patients. **B**, A positive correlation was found between

IDO1 gene expression and kynurenine level in PDAC tumors. C, IDO2 didn't show any significant correlation with NOS2 or kynurenine. Additionally, TDO positively correlated with NOS2, however, TDO was not correlated with kynurenine level in these tumors from PDAC patients.

Supplementary table S1

Characteristics of PDAC patient population

Supplementary Table S2

Log2 transformed metabolomic data set (Enclosed as a separate Excel File)

Supplementary table S3

Metabolites with significantly different relative abundance in NOS2-high versus NOS2-low tumors

Supplementary Table S4

Univariable and multivariable Cox-regression analysis showing kynurenine as a candidate predictor of prognosis, independent of other clinical covariates

Figure 1

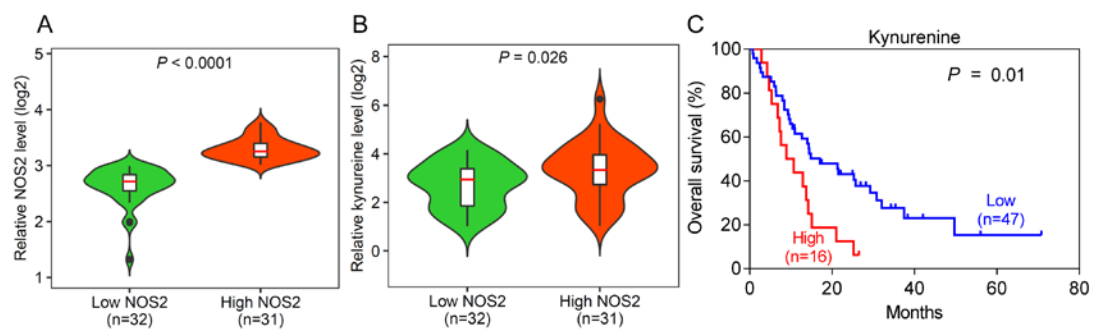


Figure 2

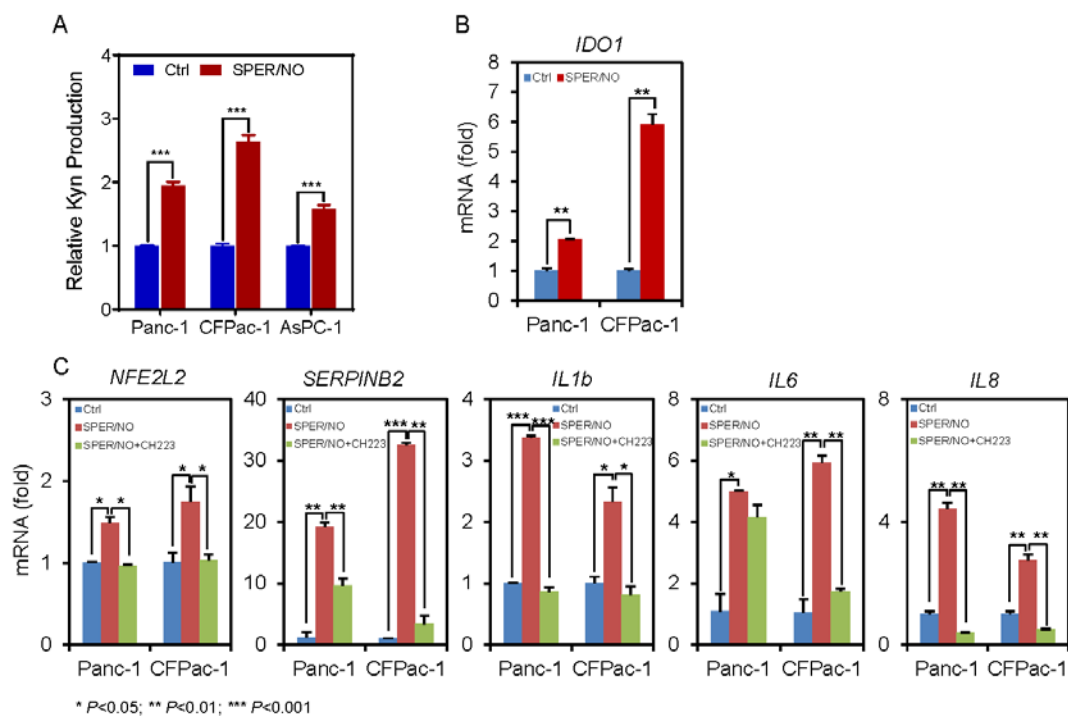


Figure 3

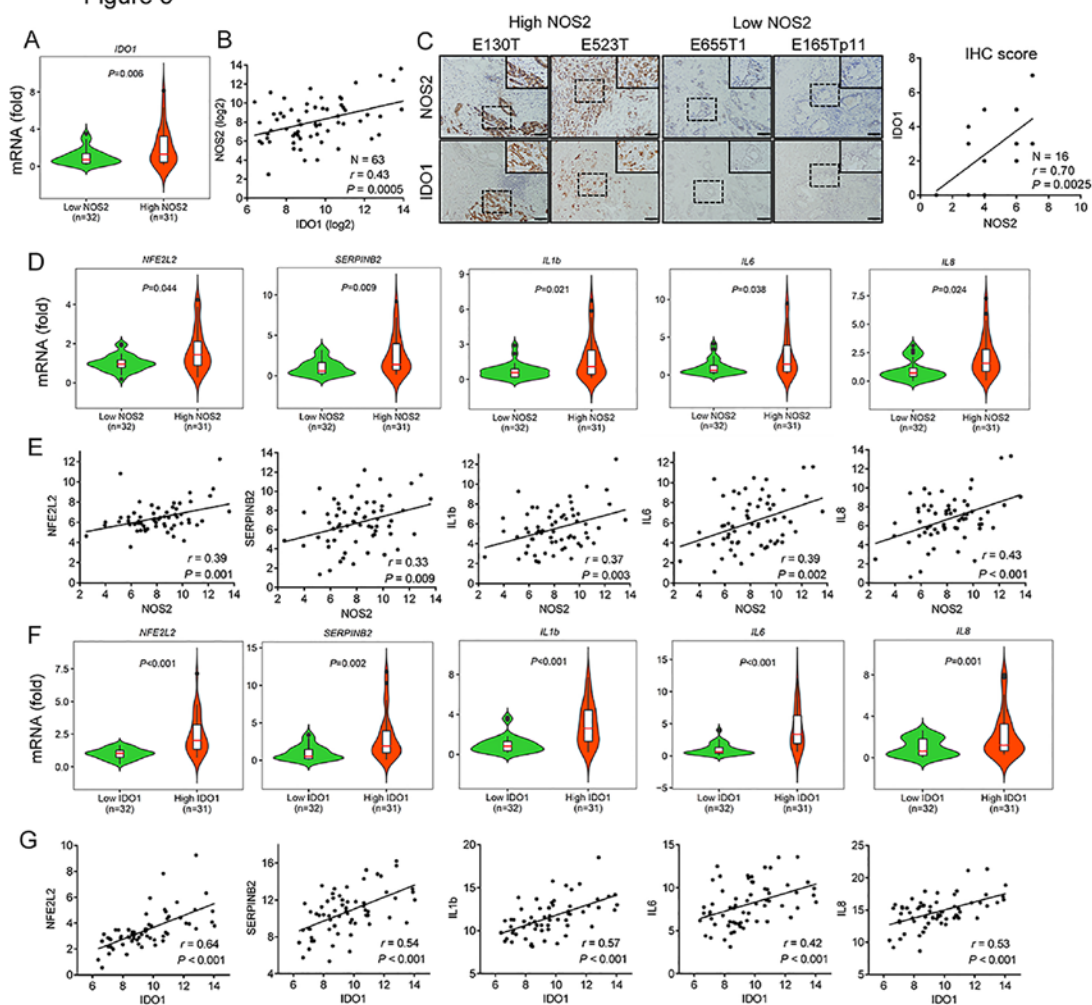


Figure 5

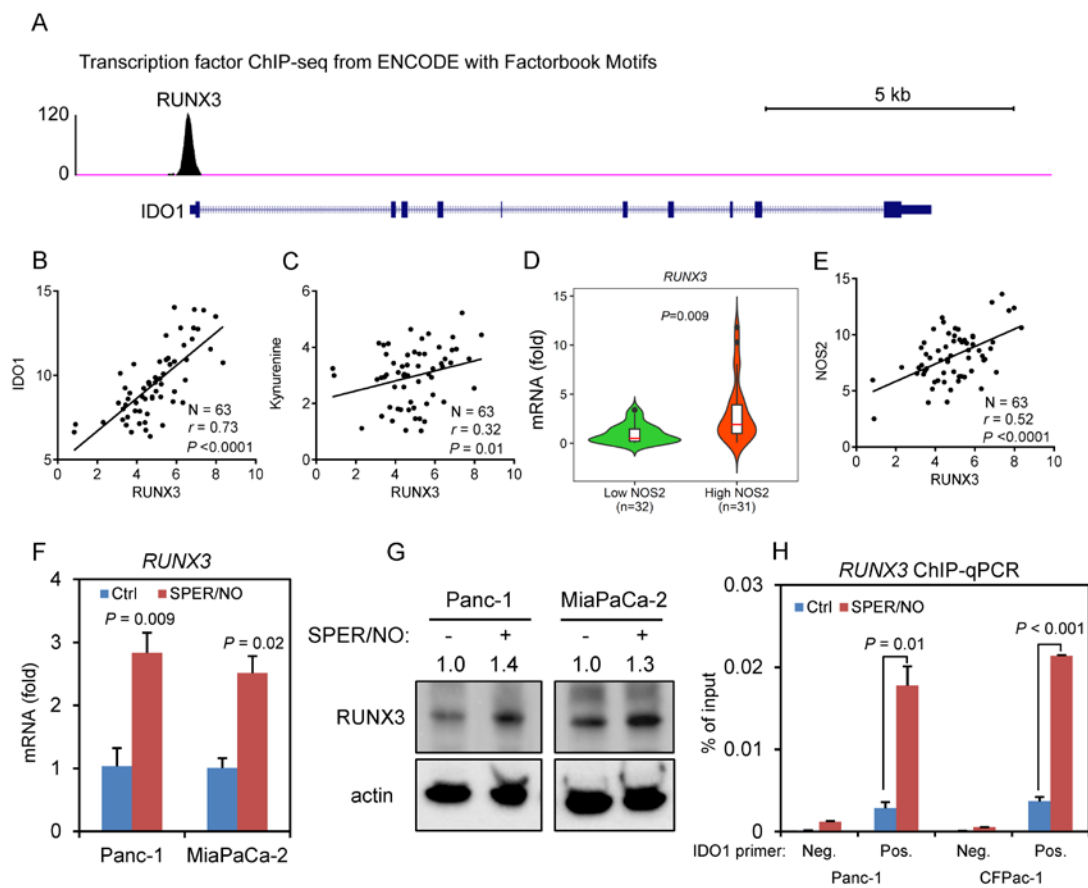
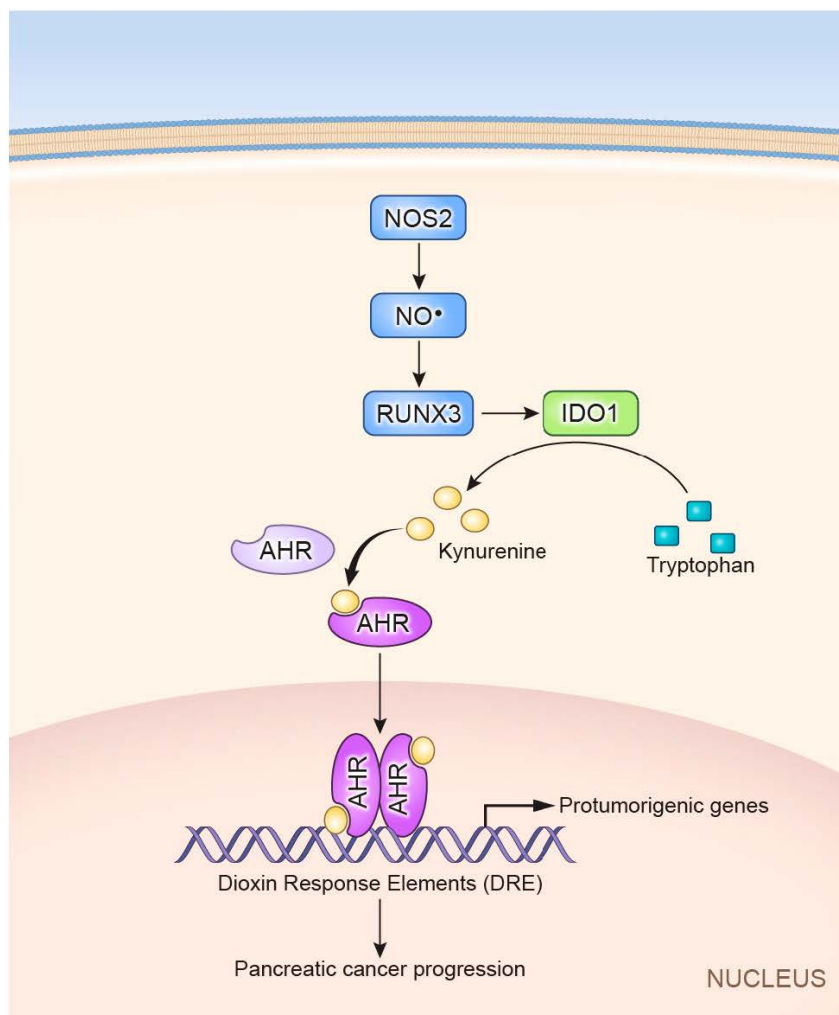


Figure 6



Metabolic reprogramming is a key biological event supporting pancreatic cancer progression but the regulatory mechanisms of metabolic adaptations in the tumors are not well defined. Here the authors focused on inflammatory signaling pathways and performed metabolic profiling of tumors with high or low inducible nitric oxide synthetase expression. They define a new metabolic axis composed of nitric oxide, the transcription factor RUNX3 and the tryptophan metabolite kynurenine that correlates with disease aggressiveness and may be useful in designing novel therapeutic approaches to improve patient outcome.

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