



**Indole: An evolutionarily conserved influencer of behavior
across kingdoms**

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Indole: An evolutionarily conserved influencer of behavior across kingdoms

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Keywords: behavioral cue; behavioral signal; inter-kingdom interactions; competition; mutualism

Abbreviations: QS, quorum-sensing; Trp, tryptophan.

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3 **Abstract.** Indole is a key environmental cue that is used by many organisms. Based
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5 on its biochemistry, we suggest indole is used so universally, and by such different
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7 organisms, because it derives from the metabolism of tryptophan, a resource essential
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9 for many species yet rare in nature. These properties make it a valuable,
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11 environmental cue for resources almost universally important for promoting fitness.
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13 We then describe how indole is used to coordinate actions within organisms, to
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15 influence the behavior of conspecifics and can even be used to change the behavior of
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17 species that belong to other kingdoms. Drawing on the evolutionary framework that
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19 has been developed for understanding animal communication, we show how this is
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21 diversely achieved by indole acting as a cue, a manipulative signal, and an honest
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23 signal, as well as how indole can be used synergistically to amplify information
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25 conveyed by other molecules. Clarifying these distinct functions of indole identifies
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27 patterns that transcend different kingdoms of organisms.
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Introduction

With the advent of high-throughput sequencing and other molecular techniques, researchers are now able to peer into the microscopic world and determine the ecological and evolutionary interactions of single cell organisms in more detail than ever before, yielding new insights into the way in which microbial cells interact with each other and with other organisms such as plants and animals. Recent reviews have highlighted that microbes, and particularly bacteria, are adept at influencing the behavior of animals [1-3]. Furthermore, researchers from across a multitude of disciplines have discovered a number of molecules produced by microbes that mediate changes in animal behavior. However, one molecule in particular, indole, seems to be ubiquitous in nature across the organismal scale from microbes and plants to invertebrates and vertebrates [4,5] (Table 1). The goal of this review is to describe the diverse ways in which indole mediates interactions between organisms, and to map the extraordinary natural history that has recently been uncovered onto long-established evolutionary concepts from the study of animal communication [6]. The purpose is to understand more about the evolution and function of indole in the natural world, and to identify gaps in understanding that might be profitably filled by future research.

The initial portion of the review covers the biochemistry of indole, to identify special properties that might explain its ubiquity in mediating interactions among organisms. We then review the types of interactions between organisms that are regulated by indole, and attempt to classify them using existing concepts from the theory of animal communication. To make sense of what follows, we therefore begin with a brief primer in the terminology and concepts from evolutionary communication theory. This lays the foundation for our understanding of the function of indole in

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3 mediating interactions within and among kingdoms.
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7 **Setting the stage- a signaling primer**

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10 Evolutionary theory of animal communication makes important distinctions between
11 the different ways that animals can collect information from one another, and their
12 wider world. The animal collecting information is often referred to as the **receiver**. In
13 the simplest case, receivers can use the behavior of others as a source of information.
14 For example, bees (Hymenoptera: Apidae) are able to gain information about the
15 nectar content of flowers on a plant simply by the presence of other bees – the
16 presence of conspecifics is a “cue” to the potential availability of a resource.
17 Importantly, the **cue** conveys useful information to the receiver but it has not evolved
18 specifically for that purpose. Honeybees, *Apis* sp., (Hymenoptera: Apidae) also have
19 famously sophisticated ways of actively transmitting information about the location of
20 nectar-producing flowers to one another through the waggle dance. In this case, the
21 location of a resource is being “signaled” and the individual imparting that
22 information is known as the **signaler**. The key distinction between a **signal** and a cue
23 is that the signal has evolved for the purpose of imparting information, because the
24 signaler benefits in some way as a result of the receiver acting on the information
25 conveyed.
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45 An animal will only respond to a cue if to do so is of benefit to that receiver –
46 otherwise it will be selected to ignore it. An animal that responds to a signal could
47 benefit too, if the information sent by the signaler is accurate and useful to the
48 receiver – in other words, if it is an ‘**honest signal**’. The conditions that enforce the
49 evolution of honest signaling are still the subject of some debate [6], but one
50 suggestion is that honesty evolves when the nature of the signal is intimately
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3 connected to the information it conveys, making it harder to fake. The depth of a
4 toad's croak is tightly associated with its body size, for example, and so accurately
5 conveys the competitive ability of the sender [7].
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10 However, signals need not be honest, and signalers can potentially use their
11 signals to manipulate others [8]. A **manipulative signal** is one, which brings a fitness
12 benefit to the signaler, but at some fitness cost to the receiver. Any receiver that is
13 routinely manipulated in this way is then placed under intense selection to ignore the
14 manipulative signal, because it will instantly gain fitness as a result. Nevertheless,
15 some receivers remain vulnerable to manipulation by signalers. They may be caught
16 in a **sensory trap**, for example. This could happen when it is sometimes – though not
17 always - adaptive for receivers to respond to a particular signal, and the receiver
18 cannot distinguish the contexts when it should and should not respond. Manipulative
19 signalers can further exploit receiver uncertainty here by sending signals that **mimic**
20 the credible signal to which the signaler is attuned. This is how cuckoo nestlings
21 succeed in manipulating their host parents into feeding them, for example [7].
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36 Finally, whether or not a signal is honest, it is under selection to be salient and
37 detectable by the receiver. Signals therefore often comprise multiple elements [9].
38 More than one element might convey the same information, and this **redundancy** of
39 information might ensure that the message gets across even if it is partially degraded
40 during transmission [10]. Other elements might convey no information at all, but
41 serve simply to **amplify** information conveyed by other elements of the display [11].
42 Or each element of the display might convey different information, transmitting
43 **multiple messages** simultaneously to the receiver [10]. The different parts of a
44 nestling's begging display probably serve each of these functions, for example
45 [12,13].
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Indole biochemistry and production

We now return our attention specifically to indole and apply these concepts to understanding its many functions in the natural world. Since its initial discovery in the mid-twentieth century, a tremendous amount has been determined about indole's chemistry, relevance in ecology, and more recently its inter-kingdom interactions. A key point is that indole is a by-product from the metabolism of tryptophan (L-tryptophan (Trp; α -amino- β -3-indolepropionic acid)), a large neutral amino acid containing an aromatic ring [14]. Trp is a relatively rare but nutritionally essential (indispensable) amino acid for mammals and it is generally more abundant in animal- than in plant-source foods. Importantly, Trp can be degraded by free-living bacteria, as well as plants and bacterial flora in animals to yield indole or indole-based compounds. In monogastric animals (e.g. pigs and rats), ~15% of dietary Trp is degraded by intestinal bacteria, while in animal cells, three pathways are responsible for degrading Trp in a highly cell- and tissue-specific manner: the kynurenine, serotonin, and transamination pathways [15]; however, these paths do not produce indole from tryptophan, so animals acquire indole from the bacteria which colonize them.

The biochemistry of indole provides some clues to explain its ubiquity in mediating interactions among organisms in nature. First, as previously mentioned, it can be readily produced by plants, by bacteria in animals and by free-living microbes simply through the metabolism of tryptophan. This means that in principle any of these organisms can produce indole. Second, tryptophan is a rare and valuable resource, and the production of indole, its metabolite, provides useful information about its potential location. In other words, the rarity yet importance of tryptophan makes indole a valuable cue for diverse organisms. Accordingly, cells are able to detect and respond to Trp metabolites such as indole and can quickly change their

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3 patterns of gene expression as a result. This is well illustrated by detailed analyses of
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5 the action of indole (and other Trp metabolites) in animal cells (**Figure 1**). Here Trp
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7 metabolites are natural ligands and activators of the aryl hydrocarbon receptor (AhR;
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9 also known as dioxin receptor) [16], which is a cytosolic ligand-activated
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11 transcription factor. AhR is normally present in a dormant state but upon ligand
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13 binding, AhR undergoes a conformational change leading to the exposure of a nuclear
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15 localization signal. Thereafter, the ligand-activated AhR translocates into the nucleus,
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17 dissociates from the complex, and forms a heterodimer with the closely related Arnt
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19 protein in the nucleus. This in turn enhances expression of target genes.
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23 A final key point is that indole closely resembles human and plant hormones
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25 such as serotonin and indole-3-acetic acid, respectively. This has also led to
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27 speculation that indole is the archetype for cell hormones [17]. It might also explain
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29 how indole can mediate interactions among kingdoms, ranging from bacteria
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31 stimulating seed germination in orchids [18], microalga [19] and diatom division [20]
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33 to fungi causing wilt in chickpeas [21], and increasing chlorophyll content and root
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35 growth in rice [22]. In short, the biochemistry of indole explains why it is a valuable
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37 cue for many diverse organisms and shows how cells are organized to detect and
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39 respond quickly to fluctuations in indole concentrations. However, as we now show,
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41 the function of indole has moved beyond a simple cue in many contexts and now
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43 plays a key role in regulating complex intra- and inter-specific interactions.
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50 **Indole as an honest intraspecific signal**

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52 The clearest evidence that indole functions as a signal comes from analyses of its role
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54 in mediating quorum-sensing (QS): the ability of microbial cells to measure
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56 population size and modulate their activities accordingly (**Figure 2**) [23]. QS systems
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3 are important for multicellular bacterial behavior such as sporulation,
4 bioluminescence, and virulence factor production [24-26]. QS is used for sensing the
5 same species, but it can also be used to sense populations of other bacteria [27],
6 sometimes known as “eaves-dropping”. In the former case, indole is most likely being
7 used as a signal, but in the latter it is more likely a cue (because the consensus is that
8 it is unlikely that one bacterial species evolved a QS signal specifically to
9 communicate with another bacterial species [23]).

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19 The *Escherichia coli* volatile metabolic product indole is emerging as a signal
20 that is important in QS interactions. Indole is produced by at least 27 different
21 bacterial genera that produce tryptophanase (TnaA) [17], the enzyme that converts
22 tryptophan into indole. Indole was first discovered as a signal in *E. coli* in which it
23 activates *gabT* and *astD* [28]. Using enterohemorrhagic *E. coli* (EHEC) and *E. coli* K-
24 12, it was then shown that indole is a QS signal [29] since it satisfies the four criteria
25 for compounds to be called cell-to-cell signals [30]: (i) the putative signal must be
26 produced during a specific stage (indole is produced primarily in the stationary-phase
27 [28]), (ii) the putative signal must accumulate extra-cellularly and be recognized by a
28 specific receptor (indole is a known extracellular signal [28,31] that is exported by
29 AcrEF [32] and is imported by Mtr [33] although it may pass through the membrane
30 at a slower rate [34]), (iii) the putative signal must accumulate and generate a
31 concerted response (indole has been shown to delay cell division [35]), and (iv) the
32 putative signal must elicit a response that extends beyond the physiological changes
33 required to metabolize or detoxify the signal (indole has been shown to control
34 biofilms [17] and cell division [35,36] which are not related to indole metabolism). *E.*
35 *coli* appears to have at least two QS systems when it lives in the mammalian
36 gastrointestinal tract. At low temperatures, indole is the primary signal, while
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3 autoinducer 2 (AI-2) fills this role at higher temperatures [37].
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7 **Indole as a manipulative interspecific bacterial signal**

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10 Indole also mediates interactions with other species of bacterial cells. It reduces the
11 pathogenicity of cells that do not synthesize it [38-40] and influences the biofilm
12 formation of other cells [17]. For example, indole reduces the virulence of
13 *Pseudomonas aeruginosa* in guinea pigs (**Figure 3**) by repressing the *mexGHI-opmD*
14 multidrug efflux pump and the genes involved in the synthesis of pyocyanin (*phz*
15 operon), 2-heptyl-3-hydroxy-4(1*H*)-quinolone (PQS) signal (*pqs* operon), pyochelin
16 (*pch* operon) and pyoverdine (*pvd* operon) which results in reduced levels of
17 pyocyanin, rhamnolipid, PQS and pyoverdine [38]. Each of these effects on other
18 bacterial species is likely to be to the advantage of the signaler but to the detriment of
19 the receiver. In this context, therefore, indole probably represents a form of coercion
20 between species, mediated by manipulative signaling.
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37 **Indole as an honest inter-kingdom bacterial signal with animals**

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39 Remarkably, indole also mediates interactions between *E. coli* and the mammalian
40 host in which it resides since it is one of the first compounds made by commensal
41 bacteria in the mammalian gastrointestinal tract and has been shown to be beneficial
42 by tightening gut epithelial cell junctions, thereby preventing invasion by pathogens
43 [41,42]. Here we can consider indole to be an honest signal because the exclusion of
44 pathogenic microbes is to the benefit of both the mammalian host and the commensal
45 gut bacteria. Indole serves a similar defensive function in other animals that also have
46 intimately associated microbiomes. For example, the indole derivative indole-3-
47 carboxaldehyde produced by microbes associated with the frog *Smilisca phaeota* [43]
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3 or the red-backed salamander, *Plethodon cinereus* [44], has been shown to repel or
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5 inhibit infection by fungal pathogens. These examples are slightly different to the
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7 honest signaling outlined above: they involve indole acting as an apparently
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9 manipulative signal to pathogenic fungi, to the detriment of their fitness, but to the
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11 benefit of the commensal microbes and their amphibian hosts.
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15 Honest inter-kingdom signaling also occurs between microbes and animals in
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17 the context of dispersal. Here selection has resulted in close relationships between the
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19 one seeking the ride and the one providing it, and indole has been shown to play an
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21 important role in mediating the provision of this service. For example, the fetid
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23 fungus, *Lysurus mokusin*, relies upon the dispersal of its spores via fecal deposition of
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25 mycophagous insects. Insects are attracted to the scent of the fungus, of which indole
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27 is a key constituent, and feed upon the fungus. Consumed spores then pass through
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29 the insect alimentary tract, enhancing their ability to germinate and increasing their
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31 dispersal range [45]. Bioassays using a synthetic mixture of the characterized scent,
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33 which included about 7.5% indole (95-99% pure), found that the odor of the fungus is
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35 attractive to the earwig, *Anisolabis maritima* (Bonelli) (Dermaptera: Anisolabididae)
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37 as well as flies belonging to 10 different genera from five families (Sarcophagidae,
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39 Calliphoridae, Muscidae, Sepsidae and Drosophilidae) [45]. Here indole is an
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41 important part of the honest signaling system that mediates dispersal.
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46 However, not all instances of dispersal mediated by indole are so obviously
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48 part of an honest signaling system. For example, the Hippelates eye gnat (genus
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50 *Tricimba* (Lioy)) (Diptera: Chloropidae)) feeds on the mucous and sebaceous
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52 secretions around the eye of vertebrates and is capable of spreading microbial
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54 organisms that cause diseases, such as conjunctivitis (pink eye), anaplasmosis, and
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56 bovine mastitis in the vertebrate host, and which themselves produce indole during
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3 infections. Hwang et al. [46] found that gnats were especially attracted to odors that
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5 included indole or skatole – presumably because this environmental cue potentially
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7 guides them to a profitable feeding location. However, it is unknown whether gnats
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9 are more attracted to feeding locations infested with microbes, and unclear that these
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11 microbes produce indole for the purpose of attracting insects. Therefore we cannot
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13 conclude that indole is an honest inter-kingdom signal in this example.
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16 The same problem exists for understanding dispersal of microbes mediated by
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18 the house fly *Musca domestica* L. (Diptera: Muscidae) and *E. coli*. In a study of
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20 chemical attractants to house flies, indole was determined to be a primary attractant
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22 [47]. When indole was compared with the closely related skatole or an indole-skatole
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24 mixture, it was determined that house flies respond more specifically to indole than
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26 skatole or the combination [47,48]. Indole is produced by *E. coli*, which is associated
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28 with vertebrate feces [49], which is an ephemeral resource, which lasts a few days or
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30 less depending on conditions [50]. As such, house flies must locate this resource
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32 quickly in order to maximize their use of it. Therefore, house flies utilize indole,
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34 which is a signature of feces and present in high concentrations, as a means to locate
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36 and colonize these resources. Adult flies attracted to the waste are then contaminated
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38 with *E. coli* and disperse it from this ephemeral resource to new locations [51].
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40 Furthermore, adults that develop as larvae feeding on the manure are also
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42 contaminated with the bacteria and can disperse it into the surrounding areas [52],
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44 often resulting in the contamination of resources consumed by vertebrate hosts [53].
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46 Nevertheless, it is unlikely that house flies and *E. coli* have evolved an honest indole
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48 signaling system to mediate *E. coli* dispersal. We should more conservatively
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50 conclude that flies are drawn to indole-rich resources because indole is a cue that
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52 conveys information about the value of the resource to the fly. Until further evidence
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3 is produced to indicate otherwise, the dispersal of *E. coli* is an incidental part of this
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5 process.
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8 Similar processes of cue-mediated incidental dispersal probably also occur
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10 within the vertebrate carrion system where indole serves as a mediator of fly (Diptera)
11 and beetle (Coleoptera) (**Figure 4**) attraction and utilization of associated resources.
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13 blow flies (Diptera: Calliphoridae) are attracted by indole to decomposing remains as
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15 a means to locate mates and provide resources to resulting offspring [54-57].
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17 Research prior to Liu et al. [54] also determined that inhibiting behavioral responses
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19 by bacteria associated with carrion [58], specifically swarming by *Proteus mirabilis*,
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21 which is regulated by a quorum sensing pathway, resulted in reduced blow fly
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23 attraction and oviposition [59]. Furthermore, responses by flies to these bacteria were
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25 regulated by sex, age, and adult nutrition history [60]. As with the house fly example,
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27 resulting contaminated adults [61] disperse into the surrounding environment
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29 allowing for microbial colonization of other resources. Likewise, mosquitoes
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31 (Diptera: Culicidae) also utilize indole as a means to locate hosts for blood-meals [62]
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33 or oviposition sites [63]. However, any microbial dispersal that also ensues is likely to
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35 be a secondary part of their search for food and egg-laying sites.
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43 **Indole as an honest inter-kingdom bacterial signal with plants**

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45 Just as indole mediates interactions between microbes and animals, it similarly
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47 mediates interactions between bacteria and plants. Soil microbes are integral to plant
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49 health and influence root architecture [64]. In fact, over 80% of land plants are able to
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51 establish mutualistic interactions with soil microbes [65]. Signaling between soil
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53 microbes and plants, with indole serving as the medium through which this interaction
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55 occurs, has been well documented. Indole produced by microbes often stimulates
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3 plant growth directly [66] and in a dose-dependent manner, such as with the
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5 rhizobacterium *Proteus vulgaris*, whose production of indole accelerates cabbage and
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7 cress growth [67,68]. In many instances, the associated microbes release mineral
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9 nutrients to the plant and in return the plant releases carbon that is then utilized by the
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11 microbes [65].
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14 Indole stimulates plant growth through the interplay of the auxin, cytokinin
15
16 and brassinosteroid hormonal pathways [67]. When produced by soil bacteria, it is
17
18 specifically able to promote early lateral root development by modulating the plant
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20 secondary root network via interference with the auxin-signaling machinery, an
21
22 essential local signal for lateral root growth [66,69]. Indole-3-acetic acid (IAA) is
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24 among the most common natural auxin growth regulators found in plants and exerts a
25
26 positive influence on root growth and length, thus increasing the total root surface
27
28 area [70]. However, as mentioned above, a dose-dependent, plant-growth-promoting
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30 property exists and long-term exposure to high concentrations of indole can have
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32 negative effects on growth and development, as demonstrated with the rockcress,
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34 *Arabidopsis thaliana* (L.) [69]. The bacterium *Pseudomonas putida* and the fungus
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36 *Trichoderma atrovirid* both produce IAA, which can increase the weight of the shoots
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38 and roots of tomato plant seedlings, but these microbes were also determined to have
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40 the capacity to reduce the deleterious effect of excess IAA by microbial degradation
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42 [71]. Therefore, microbes may help plants in two ways: by stimulating growth
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44 through production of an indole derivative and by helping to degrade harmful
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46 excesses in concentrations of these indole-related compounds. Whether these
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48 compounds are functioning as cues or signals in this context remains to be formally
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50 determined. However, it is conceivable that indole is an honest signal in this context
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52 because the microbes potentially benefit by closely regulating plant growth to
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3 optimize the levels of carbon the plant then releases back to them.
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7 **Indole signaling between plant cells**

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10 Indole also plays an important role in regulating plant defense mechanisms. In the
11 case of rice *Oryza sativa*, *GH3-8*, an auxin-responsive gene responds to indole-3-
12 acetic acid and activates disease resistance via jasmonic acid and salicylic acid
13 signaling-independent pathways [72]. Here this variant of indole serves a signaling
14 function within the plant, to coordinate its defense mechanisms. In addition, following
15 attack by herbivores, plants can release a suite of volatile organic compounds,
16 including indole, which can induce nearby plants of the same species to enhance their
17 defensive mechanisms [73]. This phenomenon, called priming, triggers increased
18 transcription of defense-related genes thus allowing nearby plants to respond more
19 rapidly and robustly to an imminent assault [74]. For example, in a study by Erb et al.
20 [75], maize was injured and treated with African cotton leafworm, *Spodoptera*
21 *littoralis* Boisduval (Lepidoptera: Noctuidae) regurgitate. The authors found that
22 indole was produced within 45 min and peaked after about 2 h. They also determined
23 that in the presence of indole the production of volatiles and terpenoids by plants that
24 were subsequently injured was enhanced as a result of priming. The simplest
25 interpretation here is that plants are eavesdropping on indole cues produced by their
26 neighbors, to adaptively modulate their defense mechanisms against the likelihood of
27 herbivore attack. However, it is possible that indole is used as a signal in this context
28 if neighboring plants are closely related, or if collectively unrelated neighboring
29 plants can more effectively repel attack by herbivores. If these conditions are met,
30 then the indole-producing plant gains fitness benefits by producing indole, either by
31 helping to defend relatives against attack or by reducing its own future vulnerability
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7 **Indole in manipulative inter-kingdom signaling by plants**

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10 Indole production by some plants directly prevents fungal infection. For example,
11 indole produced by barley, *Hordeum vulgare* L. cv. Goseshikoku and cv. Morex can
12 reduce the likelihood of powdery mildew, *Blumeria graminis* f.sp. hordei, infection
13 [76]. Similarly, rice, Sekiguchi lesion (sl)-mutant produces indole to reduce rice blast
14 fungal infection [77], while indole and other associated compounds also reduce the
15 likelihood of infection by the fungus responsible for brassica dark leaf spot,
16 *Alternaria brassicicola* [78]. Here indole can be viewed as a manipulative signal, just
17 as it is when used by microbes carried by amphibian as a defense against fungi [43]:
18 the plant gains fitness from indole production, while the fungi lose fitness.
19 Manipulative signaling also apparently occurs between the mouse-ear, *Arabidopsis*
20 *thaliana*, and the cabbage white butterfly [79]. Here some concentrations of indole
21 produced by the plant inhibit oviposition by the butterfly, although at other doses,
22 oviposition is enhanced. The dose-dependent response to indole by the butterfly might
23 explain why manipulative signaling by the plant can persist in this context and the
24 butterfly has not evolved to ignore it: any fitness costs to the butterfly through lost
25 fecundity are potentially offset by fitness it might gain in response to other levels of
26 indole signaling.
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50 **Indole in honest inter-kingdom signaling by plants**

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52 The cocktail of chemicals released by plants in response to damage by herbivores has
53 also been implicated in inter-kingdom signaling, with indole serving a key role in this
54 function. For example, the release of indole directly attracts parasitoids, which then
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3 kill the plant's insect herbivores. An example comes from Alborn et al. [80] and
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5 involves the beet armyworm caterpillar *Spodoptera exigua* (Hübner) (Lepidoptera:
6
7 Noctuidae). This caterpillar secretes volicitin (a fatty acid derivative regurgitate N-
8
9 (17-hydroxylinolenoyl)-L-glutamine) while consuming plants, such as maize, *Zea*
10
11 *mays* (L.). Contact between volicitin and the plant elicits the release of a blend of
12
13 volatile terpenoids and indole systemically from the maize plant, not just from the
14
15 damaged maize leaves. Volicitin selectively activates the formation of free indole
16
17 [81], drawing in parasitoids which lay their eggs in the caterpillar. Here indole is an
18
19 honest signal because it enables both the plant and the parasitoid to gain fitness
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21 benefits.
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25 In other cases, indole serves a more indirect function in recruiting natural
26
27 enemies of the arthropod herbivore. The volatiles produced by plants subsequent to
28
29 herbivore feeding are complex blends of compounds resulting from three primary
30
31 biosynthetic pathways: the terpenoid, the shikimate, and the fatty acid degradation
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33 pathway [82]. Indole produced from the shikimic acid pathway can play a role in
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35 indirect defense because it facilitates the release of a different volatile signal from a
36
37 damaged plant that attracts natural enemies of the arthropod herbivore inflicting the
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39 damage [83]. Here, indole's function is merely to mediate communication within the
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41 plant, which in turn leads to the release of a second honest signal that is received by
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43 the animal.
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49 **Indole in mating displays by animals**

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51 Indole has been shown to be involved in pheromonal displays that are used for mate
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53 attraction in animals. For the most part, these are displays by females for attracting
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55 males. For example, males of the scarab beetle, *Holotrichia reynaudi* Hope
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3 (Coleoptera: Scarabaeidae) rely on a mixture of abdominal exudates, including indole,
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5 produced by the female to locate a partner [84]. However, in the dung beetle, *Kheper*
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7 *bonelli* (MacLeay) (Coleoptera: Scarabaeidae) males produce and release a
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9 proteinaceous secretion to attract females for mating. Within this proteinaceous
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11 carrier material are putative sex pheromones, among which indole was identified [85].
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13 Given that this particular species relies on dung, which typically contains high levels
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15 of *E. coli* (which produces indole as previously mentioned), it would be interesting to
16
17 determine whether indole production is truly by the insect or by the *E. coli* harbored
18
19 within the insect. In general, although indole is present in these pheromonal cocktails,
20
21 its function in luring a mate remains unclear. As is illustrated by the examples we
22
23 consider next, it might function to convey important information to a potential
24
25 partner, or it may simply amplify information conveyed by other compounds in the
26
27 pheromone. It might even play no role at all in mate attraction. More work is required
28
29 to distinguish these different possibilities.
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36 **The function of indole in complex displays: information carrier or** 37 38 **amplifier?** 39

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41 In many of the examples discussed above, indole is part of a complex cocktail of
42
43 volatiles emitted by a signaler. To understand its specific function in these contexts,
44
45 we must turn to evolutionary theory connected with complex, or multicomponent
46
47 displays (summarized above). One suggestion here is that some elements of a
48
49 complex display serve to amplify other parts of the display [11]. Indole seems to serve
50
51 exactly this function in the signaling that takes place between the gourd family of
52
53 flowering plants (Cucurbitaciae) and diabroticite rootworm beetles (Coleoptera:
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55 Chrysomelidae: Luperini). These organisms are anciently associated with one another
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3 and have likely coevolved through their associated chemical ecology [86,87]. The
4
5 Cucurbita blossom is a source of nectar and pollen for diabroticite beetles, which are
6
7 attracted by its odorous bouquet, and which includes indole as a volatile [88]. Most
8
9 Cucurbitaciae produce a secondary compound called cucurbitacin, a triterpene
10
11 hydrocarbon containing an indole structure [89]. Cucurbitacins are bitter and often
12
13 toxic semiochemicals that serve to protect the plants from attack by invertebrate and
14
15 vertebrate herbivores. Diabroticite beetles, however, use these compounds as
16
17 kairomones for locating the blossom. They are able to feed on the cucurbits and store
18
19 the bitter cucurbitacins in their blood and tissues as allomones to deter predation [87].
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23 The role of indole in attracting the beetles to the blossom is to act as an
24
25 amplifier. The diabroticite beetles, *Acalymma vitatum*, *Diabrotica u. howardi*, *D.*
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27 *virgifera virgifera*, and *D. barberi*, are only weakly to moderately attracted to indole
28
29 as a single compound [90]. However, when combined with other olfactants from
30
31 Cucurbita blossoms, indole increased olfactory responses in diabroticite beetles
32
33 synergistically by 2 to 4 fold [88,90]. Further evidence that indole amplifies the
34
35 attraction of other volatiles to these beetles comes from experiments using insect traps
36
37 for diabroticite rootworm beetles, in bean, *Phaseolus vulgaris*, and soybean, *Glycine*
38
39 *max*, fields. Traps baited with veratrole + indole + phenylacetaldehyde caught 6.5
40
41 and 3.5 times more beetles than solvent controls in soybean and common bean plots,
42
43 respectively; traps baited with 1,2,4-trimethoxybenzene + indole + trans-cinnamal-
44
45 dehyde) caught 6.7 and 3.5 times more beetles, respectively [91]. Thus indole has
46
47 evolved to be part of the Cucurbita blossom's odiferous display seemingly because it
48
49 amplifies the response by beetles to other volatiles in the bouquet.
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53 Indole might serve a similar amplifying function in the scent profile of other
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55 plant species, as demonstrated in a study by Friberg et al. [92]. Here it was found to
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3 be one of the compounds within the unique floral profiles produced by two different
4 woodstar plants, *Lithophragma bolanderi* and *L. cymbalaria* (Saxifragaceae), which
5 attract the parasitic moth, *Greya politella* (Walsingham) (Prodoxidae), for pollination.
6
7 Female moths responded most strongly to the uniquely distinctive scents from their
8 local host species and were thereby more likely to pollinate the local plants. It would
9 be interesting in future work to determine whether this divergent response to floral
10 scents has been facilitated by indole.
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18 In other floral scents, however, indole might be used by the plant as a signal to
19 manipulate insects into providing a pollination service. For example, the composition
20 of the floral scent of the sapromyiophilous, *Periploca laevigata*, was investigated
21 because of its ability to lure in the common house fly as a pollinator species [93]. The
22 most abundant compound identified in the scent disseminated from cultivated
23 sapromyiophilous was indole (39%), which attracted both male and female flies [93].
24 Presumably the flies use indole as a cue for locating oviposition or food resources and
25 the plant has evolved a manipulative signal, in which the insects are sensorily trapped
26 into visiting the plant and pollinate it in the process.
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41 **Conclusions**

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43 In this review we have attempted to explain why indole is used so ubiquitously in
44 nature, and how it functions to modulate interactions among diverse organisms. We
45 suggest that indole is used so widely, and by such different organisms, because it
46 derives from the metabolism of tryptophan, a resource that is essential for many
47 species yet rare in nature. These properties make it a valuable, environmental cue for
48 resources that are almost universally important for promoting fitness. By surveying a
49 broad literature, we find that indole is used to coordinate actions within organisms, to
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3 influence the behavior of conspecifics and can even be used to change the behavior of
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5 species that belong to other kingdoms. This is variously achieved by indole acting as a
6
7 cue, a manipulative signal, and an honest signal, as well as an amplifier for
8
9 information conveyed by other molecules. Importantly, these distinct functions of
10
11 indole transcend different kingdoms of organisms. These roles across kingdoms for
12
13 indole make it special to the extent it is widely used but not necessarily unique; for
14
15 example, the bacterial QS signal *N*-acyl-L-homoserine lactone from the opportunistic
16
17 pathogen *Pseudomonas aeruginosa* represses the mammalian innate immune system
18
19 [94]
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23 We suggest that future work could profitably build on the conclusions of this
24
25 article by using indole in interventions to manage crop pests and to control vectors of
26
27 pathogens. Existing biology reviewed here suggests that such insect species could be
28
29 surprisingly vulnerable to being manipulated in this way. Finally, we wonder whether
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31 indole could even be deployed to promote the pollination services provided by insects
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33 of economically important crop plants, since existing evidence suggests this might
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35 enhance the attractiveness of the plant to potential pollinators, possibly thereby
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37 boosting pollination rates.
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3 **Figure 1. Mechanisms for the physiological actions of indole in animals.** In
4 animals, indole can scavenge free radical species and exert anti-oxidative effects [95],
5 and can also enhance expression of xenobiotic-metabolizing enzymes (e.g.,
6 cytochrome P450) and immune response through binding to aryl hydrocarbon
7 receptors (ligand-activated transcription factors) [96]. These actions of indole result in
8 the amelioration of oxidative stress (such as UV radiation- or oxidant-induced DNA
9 damage). Through binding to the serotonin receptor and serving as an α_{1A} -
10 adrenoceptor antagonist [97], indole modulates animal behavior, the contraction of
11 smooth muscle, gut motility, and food intake [98]. By interacting with iron in heme-
12 containing oxygenases [95], indole plays a role in whole-body aerobic metabolism.
13 Indole also regulates the release of secretion of luteinizing hormone, and, therefore
14 male and female reproduction [99]. Finally, indole affects the metabolism and activity
15 of gut microbes, thereby sustaining intestinal health [100,101].
16

17 **Figure 2. The hypothesized function of quorum sensing.** Bacterial cells produce
18 signal molecules, which can be used as a source of information about the density of
19 cells in their environment. It has been shown that cells use this information to control
20 the expression of density dependent traits, such as protease production. On the left,
21 the diagram shows how beneficial exo-products can be easily lost, providing little or
22 no benefit to cells, the right hand side shows how exo-products are more likely to
23 benefit surrounding cells at high densities. [102].
24

25 **Figure 3. Reduction of virulence of *P. aeruginosa* in guinea pigs by 7-**
26 **hydroxyindole (7HI).** Colonization and clearance of *P. aeruginosa* PAO1 pre-treated
27 with 7HI or solvent (DMF) prior to infection of guinea pigs by aerosol with $\sim 2 \times 10^5$
28 cfu. Average of five replicates, and one standard deviation is shown (A). Real-time
29 analysis of *P. aeruginosa* PAO1 pre-treated with 7HI or solvent (DMF) in the acute
30 guinea pig infection model (representative guinea pigs are shown for each group and
31 are imaged laterally) using the Xenogen IVIS CCD camera (B). Color bar represents
32 the intensity of luminescent signal in photons/sec/cm² from low (blue) to high (red)
33 [38].
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37 **Figure 4. Arthropods commonly colonizing vertebrate carrion. (a)** Beetles (e.g.
38 *Nicrophorus vespilloides* [Coleoptera: Silphidae], photo: Tom Houslay) and **(b)** flies
39 (e.g. *Chrysomya rufifacies* and *Cochliomyia macellaria* [Diptera: Calliphoridae],
40 photo: C.C. Heo) are the primary invertebrate consumers of vertebrate carrion. Such
41 invertebrates use indole and other volatiles to locate such resources essential for mate
42 location as well as adult and larval nutrition.
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Table 1. Examples of indole and indole derivatives as potential signals or cues.

| Indole or derivative (IUPAC name if different) | Sender | | | | Receiver | | | |
|---|----------|---|--|---|----------|-----------------------------------|--|--|
| | Kingdom | Common Name | Scientific Name | Notes | Kingdom | Common Name | Scientific Name | Induced Behavior & Citations |
| | Animalia | | | | | | | |
| Indole | | Vertebrate host | Varied | Most likely host-microbe derived | Animalia | Blow fly | <i>Lucilia sericata</i> | Attraction for oviposition [54,55] |
| Indole | | Vertebrate host | Varied | Most likely host-microbe derived | Animalia | Mosquito | <i>Anopheles gambiae</i> | Attraction for blood-meal [62] |
| Indole-3-carboxaldehyde | | Frog | <i>Smilisca phaeota</i> | Most likely host-microbe derived | Fungi | Chytridiomycosis (fungal disease) | <i>Batrachochytrium dendrobatidis</i> (<i>Bd</i>) | Repellence [43] |
| Indole-3-carboxaldehyde | | Red-backed salamander | <i>Plethodon cinereus</i> | Derived from host skin symbiont, <i>Janthinobacterium lividum</i> | Fungi | Chytridiomycosis (fungal disease) | <i>Batrachochytrium dendrobatidis</i> (JEL 215 strain) | Inhibited growth and mortality [44] |
| | Plantae | | | | | | | |
| Indole-3-acetonitrile | | Arabidopsis, mouse-ear cress, thale cress | <i>Arabidopsis thaliana</i> (L.) | | Animalia | Cabbage white butterfly | <i>Pieris rapae</i> | Decreased oviposition by [79] |
| Indole-3-carbinol | | Arabidopsis, mouse-ear cress, thale cress | <i>Arabidopsis thaliana</i> (L.) | | Animalia | Cabbage white butterfly | <i>Pieris rapae</i> | Increased oviposition by [79] |
| Gramine (1-(1H-indol-3-yl)-N,N-dimethylmethanamine) | | Barley | <i>Hordeum vulgare</i> L. cv. F Union | | Animalia | Greenbug | <i>Schizaphis graminum</i> | Prevents herbivory by [103] |
| Gramine (1-(1H-indol-3-yl)-N,N-dimethylmethanamine) | | Barley | <i>Hordeum vulgare</i> L. cv. Goseshikoku and cv. Morex | | Fungi | Barley powdery mildew | <i>Blumeria graminis</i> f.sp. <i>hordei</i> | Prevents infection by [76] |
| Gramine (1-(1H-indol-3-yl)-N,N-dimethylmethanamine) | | Barley | <i>Hordeum vulgare</i> L. cv. Goseshikoku and cv. Morex | | Fungi | Wheat powdery mildew | <i>Blumeria graminis</i> f.sp. <i>tritici</i> | Prevents Infection by [76] |
| Indole-3-acetonitrile | | Rice | Sekiguchi lesion (sl)-mutant <i>Oryza sativa</i> | | Fungi | Rice blast fungus | <i>Magnaporthe grisea</i> | Prevents infection by [77] |
| Hydrolysis products of indole glucosinolate | | Arabidopsis, mouse-ear cress, thale cress | <i>Arabidopsis thaliana</i> (L.) | | Fungi | Brassica dark leave spot | <i>Alternaria brassicicola</i> | Prevent infection by [78] |
| | Fungi | | | | | | | |
| Indole acetic acid | | | <i>Fusarium tricinctum</i> RSF-4L isolated from <i>Solanum nigrum</i> | | Plantae | Rice | <i>Oryza sativa japonica</i> variety Dongjin | Enhanced chlorophyll content, root-shoot length, and biomass production [22] |
| Indole acetic acid | | | <i>Alternaria alternata</i> RSF 6L isolated from <i>Solanum nigrum</i> | | Plantae | Rice | <i>Oryza sativa japonica</i> variety Dongjin | Enhanced chlorophyll content, root-shoot length, and biomass production [22] |

| | | | | | | | | |
|----|--|-----------|---|--------------------------------|--------------------------------|------------------------|--|--|
| 1 | Indole-3-acetic acid | | <i>Fusarium delphinooides</i> strain GPK | | Chickpea ICCV- 10 and L-550 | <i>Cicer arietinum</i> | Caused wilt [21] | |
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| 3 | | | | | | | | |
| 4 | | | | | | | | |
| 5 | | Bacteria | | | | | | |
| 6 | Indole | commensal | <i>Escherichia coli</i> | Common in GI tract | Animalia | Humans | <i>Homo sapiens</i> | Tightens epithelial cell junctions [41,42] |
| 7 | | | | | | | | |
| 8 | Indole | commensal | <i>Escherichia coli</i> | Common in GI tract | Bacteria | Pseudomonad | <i>Pseudomonas aeruginosa</i> | Reduces virulence [38] |
| 9 | Indole | commensal | <i>Escherichia coli</i> | Common in GI tract | Bacteria | EHEC | <i>Escherichia coli O157:H7</i> | Reduces biofilm formation [29] and chemotaxis, motility, and attachment [104] |
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| 11 | | | | | | | | |
| 12 | Indole | commensal | <i>Escherichia coli</i> | Common in GI tract | Bacteria | Pseudomonad | <i>Pseudomonas fluorescens</i> | Increases biofilm formation |
| 13 | Indole | Multiple | Gram-positive and Gram- negative taxa | Common in oviposition sites | Animalia | Mosquito | <i>Aedes aegypti</i> | Attraction for oviposition [63] |
| 14 | | | | | | | | |
| 15 | Indole-3-acetic acid (2-(1 <i>H</i> - indol-3-yl)acetic acid) | Multiple | <i>Rhizobium</i> , <i>Microbacterium</i> , <i>Sphingomonas</i> , and <i>Mycobacterium</i> | | Plantae | Orchid | <i>Dendrobium moschatum</i> | Stimulates seed germination [18] |
| 16 | | | | | | | | |
| 17 | Indole | Proteus | <i>Proteus mirabilis</i> | Associated with host | Animalia | Blow fly | <i>Lucilia sericata</i> | Attraction for food [54,56,57] |
| 18 | | | | | | | | |
| 19 | Indole | Proteus | <i>Proteus mirabilis</i> | Associated with host | Animalia | Blow fly | <i>Lucilia sericata</i> | Attraction for oviposition [54,56,57] |
| 20 | | | | | | | | |
| 21 | Indole-3-acetic acid | | <i>Azospirillum brasilens</i> and <i>A.</i> <i>lipoferum</i> | | Plantae | Microalga | <i>Chlorella vulgaris</i> | Promoting growth[19] |
| 22 | | | | | | | | |
| 23 | Indole | | <i>Escherichia coli tnaA-</i> (JW3686) | Soil-borne bacteria | | | <i>Arabidopsis thaliana</i> Columbia-O ecotype | Increased plant secondary root network [66] |
| 24 | | | | | | | | |
| 25 | Indole-3-acetic acid | | <i>Sulfitobacter</i> spp. | | Protista | Diatom | <i>Thalassiosira</i> <i>pseudonana</i> CCMP1335 | Promotes cell division [20] |
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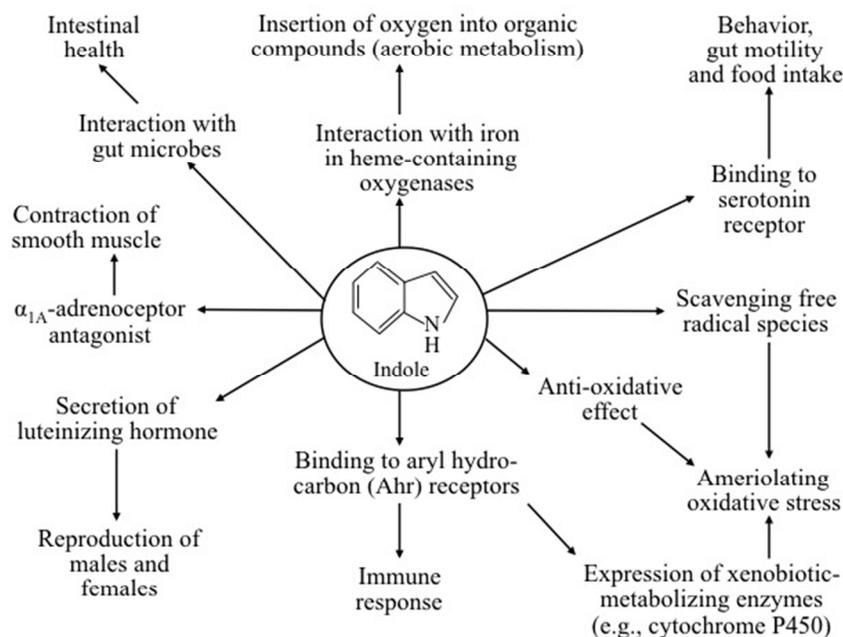
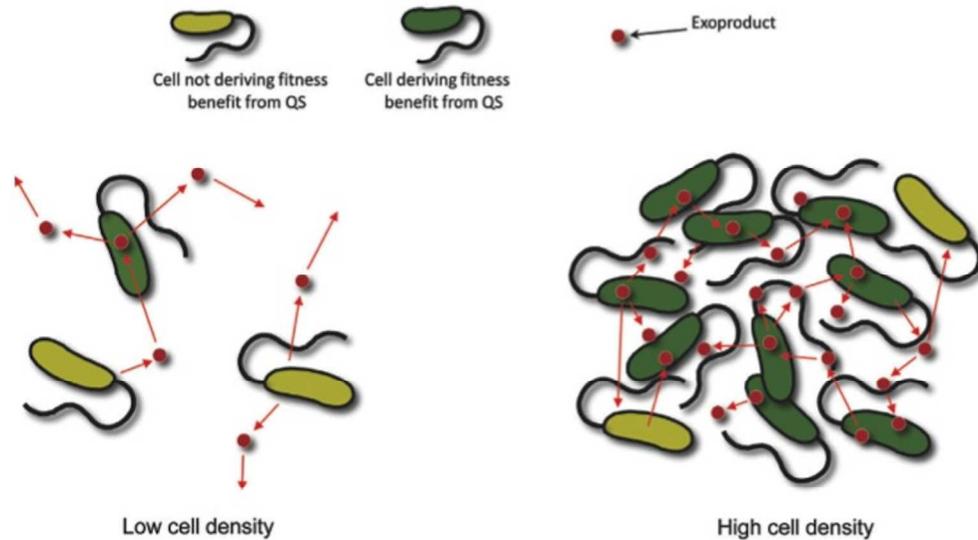


Figure 1. Mechanisms for the physiological actions of indole in animals. In animals, indole can scavenge free radical species and exert anti-oxidative effects [95], and can also enhance expression of xenobiotic-metabolizing enzymes (e.g., cytochrome P450) and immune response through binding to aryl hydrocarbon receptors (ligand-activated transcription factors) [96]. These actions of indole result in the amelioration of oxidative stress (such as UV radiation- or oxidant-induced DNA damage). Through binding to the serotonin receptor and serving as an α_{1A} -adrenoceptor antagonist [97], indole modulates animal behavior, the contraction of smooth muscle, gut motility, and food intake [98]. By interacting with iron in heme-containing oxygenases [95], indole plays a role in whole-body aerobic metabolism. Indole also regulates the release of secretion of luteinizing hormone, and, therefore male and female reproduction [99]. Finally, indole affects the metabolism and activity of gut microbes, thereby sustaining intestinal health [100,101].

254x190mm (72 x 72 DPI)



An Introduction to Behavioural Ecology, Fourth Edition. Nicholas B. Davies, John R. Krebs and Stuart A. West.
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Figure 2. The hypothesized function of quorum sensing. Bacterial cells produce signal molecules, which can be used as a source of information about the density of cells in their environment. It has been shown that cells use this information to control the expression of density dependent traits, such as protease production. On the left, the diagram shows how beneficial exo-products can be easily lost, providing little or no benefit to cells, the right hand side shows how exo-products are more likely to benefit surrounding cells at high densities. [102].

254x190mm (300 x 300 DPI)

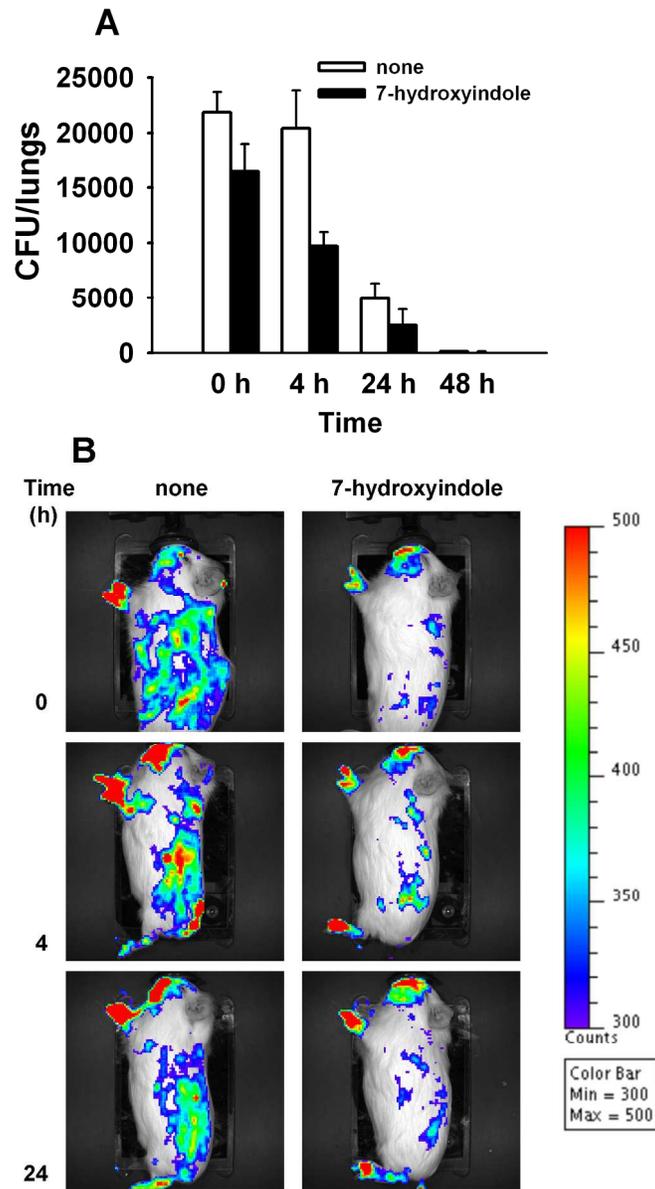


Figure 3. Reduction of virulence of *P. aeruginosa* in guinea pigs by 7-hydroxyindole (7HI). Colonization and clearance of *P. aeruginosa* PAO1 pre-treated with 7HI or solvent (DMF) prior to infection of guinea pigs by aerosol with $\sim 2 \times 10^5$ cfu. Average of five replicates, and one standard deviation is shown (A). Real-time analysis of *P. aeruginosa* PAO1 pre-treated with 7HI or solvent (DMF) in the acute guinea pig infection model (representative guinea pigs are shown for each group and are imaged laterally) using the Xenogen IVIS CCD camera (B). Color bar represents the intensity of luminescent signal in photons/sec/cm² from low (blue) to high (red) [38].

147x260mm (300 x 300 DPI)



Figure 4. Arthropods commonly colonizing vertebrate carrion. (a) Beetles (e.g. *Nicrophorus vespilloides* [Coleoptera: Silphidae], photo: Tom Houslay) and (b) flies (e.g. *Chrysomya rufifacies* and *Cochliomyia macellaria* [Diptera: Calliphoridae], photo: C.C. Heo) are the primary invertebrate consumers of vertebrate carrion. Such invertebrates use indole and other volatiles to locate such resources essential for mate location as well as adult and larval nutrition.

56x37mm (300 x 300 DPI)



Figure 4. Arthropods commonly colonizing vertebrate carrion. (a) Beetles (e.g. *Nicrophorus vespilloides* [Coleoptera: Silphidae], photo: Tom Houslay) and (b) flies (e.g. *Chrysomya rufifacies* and *Cochliomyia macellaria* [Diptera: Calliphoridae], photo: C.C. Heo) are the primary invertebrate consumers of vertebrate carrion. Such invertebrates use indole and other volatiles to locate such resources essential for mate location as well as adult and larval nutrition.

210x144mm (300 x 300 DPI)