

2 The cascade to pathogenicity in 3 autoantibody-mediated CNS diseases

4 Federico Montini,^{1,2,†} Elinor Wing,^{3,†} Max Herman,⁴ Sean J. Pittock,^{1,2} Sebastian Lopez,^{2,4}
5 Eoin P. Flanagan^{1,2} and Sarosh R. Irani^{2,3,4,5}

6 **†These authors contributed equally to this work.**

7 **Abstract**

8 The discovery of pathogenic neuroglial-surface directed autoantibodies (NGSAb) has
9 fundamentally transformed clinical neurology, by enabling molecular-level diagnoses in
10 potentially treatable, yet previously unrecognised, diseases. Annual descriptions of novel
11 CNS-targeting antibodies create a continuous stream of new conditions in which to
12 evaluate distinct phenotypes, specific tumour associations and immunotherapy
13 responses.

14 Alongside this clinical growth, increasing basic knowledge has highlighted origins and
15 mechanisms underlying disease causation, most comprehensively interrogated in the well-
16 established autoantibody-mediated conditions of autoimmune encephalitis (AE),
17 neuromyelitis optica spectrum disorder (NMOSD) and myelin oligodendrocyte glycoprotein
18 antibody-associated disease (MOGAD). The corresponding most common ‘big six’
19 autoantigens are LGI1, the NMDA receptor, CASPR2, IgLON5, in forms of AE, AQP4 and
20 MOG. Each of these autoantigens associates with a homogenous set of basic clinical
21 features, across age, sex, tumour associations and ethnicities, coupled with partly
22 distinctive profiles of triggers and predispositions, paradigms of immune tolerance escape
23 in the periphery, how cells and autoantibodies gain access to the CNS and discrete
24 mechanisms by which the CNS autoantibodies induce neuroglial dysfunction. These
25 observations lead us to reconstruct a proposed chronological series of events as the
26 “cascade to pathogenicity”, which together culminate in a rare CNS disease. By extension,
27 we hypothesize elucidating the underlying biology of each condition will present differing
28 precision medicine approaches to optimize patient care. Despite distinctions, there are
29 also clinical and biological overlaps between these diseases, collectively creating
30 opportunities to compare and contrast their individual features.

1 Here, in each condition, we review current knowledge regarding the similarities and
2 differences between the triggering events, underlying immunological processes and
3 pathogenic mechanisms of autoantibodies. In some instances, we identify scientific clues
4 which drive hypothetical pathways of pathogenesis and, for others, highlight striking
5 observations which aim to generate hypothesis-driven next steps.

6 Our aim is to construct a model across the major autoantibody-mediated CNS diseases to
7 highlight distinct components of cascades to pathogenicity which may offer targeted
8 therapeutic approaches to improve patient outcomes, and identify key areas and questions
9 for future research.

10

11 **Author affiliations:**

12 1 Department of Neurology, Mayo Clinic, Rochester, MN, 55905, USA

13 2 Center for MS and Autoimmune Neurology, Mayo Clinic, Rochester, MN 55905, USA

14 3 Department of Neuroscience, Mayo Clinic, Jacksonville, FL, 32224, USA

15 4 Department of Neurology, Mayo Clinic, Jacksonville, FL, 32224, USA

16 5 Oxford Autoimmune Neurology Group, Nuffield Department of Clinical Neurosciences,
17 University of Oxford, Oxford, OX3 9DU, UK

18

19 Correspondence to: Professor Sarosh R. Irani

20 Departments of Neurology and Neurosciences, 4500 San Pablo Road, Mangurian Building,
21 5th Floor, Jacksonville, FL 32224, USA

22 E-mail: irani.sarosh@mayo.edu

23

24 **Running title:** Pathogenicity of CNS autoantibodies

25 **Keywords:** autoimmune; encephalitis; neuromyelitis optica; MOGAD; autoantibodies

26

1 Introduction

2 Autoimmune neurology is one of the most dynamic and rapidly advancing fields in
3 contemporary medicine. Over the past two decades, the discovery of multiple
4 autoantibodies directed against autoantigens with epitopes expressed on the surface of
5 neurons and glia has transformed our understanding of neuroinflammatory diseases,
6 shifting the paradigm from syndromic descriptions to molecularly-precise autoantibody-
7 mediated diagnoses.^{1,2} These neuroglial surface-directed autoantibody (NGSAb) mediated
8 diseases also carry major pragmatic importance for our patients as they are typically
9 immunotherapy-responsive, and should be considered “not to miss” conditions.³⁻⁵

10 NGSAb-mediated diseases encompass a diverse spectrum affecting multiple levels of the
11 nervous system, from cortex, through subcortical white and grey matter, brainstem,
12 cerebellum, optic nerve, spinal cord and peripheral nerve, resulting in varied clinical
13 phenotypes. While traditional disease classifications often pivot on such anatomical
14 localizations, experiential patterns and clinical courses, the molecular revolution
15 exemplified by NGSAb-mediated diseases offers an opportunity to classify these
16 conditions via their fundamental biological basis.⁶ This molecular-led evolution has already
17 commenced by strictly clinically distinguishing aquaporin-4-antibody associated
18 neuromyelitis optica spectrum disorder (AQP4⁺NMOSD) from myelin oligodendrocyte
19 glycoprotein antibody disease (MOGAD).⁷ Here, we aim to also distinguish between the
20 varied and clinically divergent forms of autoimmune encephalitis (AE) and incorporate a
21 molecular viewpoint, dissecting triggers and predisposing factors, through key drivers of
22 the aberrant immune system, to mechanisms by which end effector autoantibodies modify
23 neuroglial function and determine patient outcomes (Fig. 1).

24 Central to a molecular understanding is recognition that these disorders arise from the
25 breakdown of immune tolerance, typically against a single autoantigen. Hence, we propose
26 that B and T cells which recognize this single autoantigen are both sufficient and necessary
27 for pathogenesis of NGSAb-mediated disorders, placing the role of the pivotal autoantigen
28 ‘centre-stage’.^{2,8,9} Their ultimate generation of NGS-reactive autoantibodies leads to
29 diseases, occurring at around 0.2 to 3 per million per year. Our working hypothesis is that
30 such rare and non-inherited diseases likely require a series of multiple more common
31 events to culminate in causation: their “cascade to pathogenicity”.¹⁰

32 In this *Review*, we aim to apply current knowledge of triggers, immunology and
33 autoantibody-autoantigen interactions to propose a cascade to pathogenicity.
34 Characterizing and interrogating this cascade aims to provide biological insights and
35 multiple therapeutic targets to catalyse the development of molecularly informed precision
36 medicine approaches in NGSAb-mediated diseases. By focusing on the commonest ‘big

1 six' NGSAb targets (Table 1), leucine-rich glioma inactivated 1 (LGI1), the N-methyl D-
2 aspartate receptor (NMDAR), contactin-associated protein like 2 (CASPR2),
3 immunoglobulin-like cell adhesion molecule 5 (IgLON5), AQP4 and MOG,^{11,12} those most
4 commonly encountered in autoimmune neurological practice, we highlight similarities and
5 differences between the cascades in each condition. Additionally, as the field of NGSAb-
6 mediated diseases is relatively nascent, we selectively incorporate other autoimmune
7 conditions to illustrate concepts of likely pertinence to NGSAb-mediated disorders.

9 The longitudinal cascade to pathogenicity

10 Consistent with a proposed cascade, we construct a timeline which begins with genetic
11 predispositions, including HLA associations and other emerging variants, that likely alter
12 the thresholds or nature of immune tolerance, permitting the escape of NGS-reactive
13 lymphocytes (Fig.1). Triggering events, such as infectious agents, NGS-autoantigen
14 expressing tumours and immune checkpoint inhibitor therapies, further augment
15 probabilities of breaking immune tolerance and priming of the immune system against
16 these autoantigens. Tolerance breaks, in both central and peripheral B and T cell
17 checkpoints, likely leads to the generation of NGS-reactive memory B cells and antibody-
18 secreting cells which gain access to the CNS to secrete pathogenic antibodies, where they
19 have immediate proximity to the extracellular domains of their molecular CNS targets
20 (Fig.2).¹³⁻¹⁵ Here, the fundamental qualities of the pathogenic antibodies and the nature and
21 site of the target autoantigen interact to manifest varied mechanisms of dysfunction at
22 molecular, circuit and systems levels, ultimately resulting in characteristic clinical
23 phenotypes and disease trajectories (Fig.3).

25 Fundamental phenotypic distinctions closely associate 26 with autoantigen specificities

27 A central clinical observation is that the autoantigen target forms a pivotal feature to help
28 demarcate fundamental clinical features (Table 1). For example, patients with LGI1-,
29 CASPR2- and IgLON5-antibodies typically present after 50 years of age, and very rarely in
30 childhood.^{16,17} Whereas patients with NMDAR are mostly far younger, often children and
31 young adults, although older onset cases are increasingly well-recognised and often have
32 tumours.¹⁸⁻²⁰ Both NMDAR- and AQP4+NMOSD show a bias to females and non-

1 Caucasians; whereas MOGAD shows a slight female predilection, rare associations with
2 tumours and no marked ethnic predispositions.²¹⁻²³ Hence, the sex ratios, tumour and
3 infectious associations of these diseases show well-established and geographically
4 reproducible skews which, again, appear to co-segregate with the autoantigen (Table 1).
5 While core symptoms of individual illnesses may be attributed to the dominant CNS
6 localisations of the autoantigen, why should the target autoantigen, the end-product of the
7 cascade to pathogenicity, be positioned to determine such a distinctive range of
8 fundamental demographic and epidemiological associations? This question led us to
9 hypothesize that the corresponding aetiological and underlying biological features of each
10 autoantigen-defined syndrome will show distinctions, a major focus of our review. Yet,
11 particularly given the ongoing and rapid evolution of knowledge in NGSAb-diseases, we
12 also identify some overlapping features between these syndromes which suggest parallel
13 pathophysiological mechanisms.

14

15 Predispositions and triggers of NGSAb-mediated 16 conditions: the presymptomatic phase

17 Genetic Predispositions

18 HLA

19 Genetics forms a canonical basis to elucidate fixed risk disease factors. HLA class II
20 associations dominate the known genetic landscape in three of the six core NGSAb-
21 mediated disorders, associated with LGI1, CASPR2 and IgLON5-antibodies (Table 1; Fig. 1).
22 In each of the three diseases, one specific HLA allele is expressed in ~75-90% of the
23 patients: HLA-DRB1*07:01 with LGI1-antibodies,²⁴⁻²⁶ HLA-DRB1*11:01 with CASPR2-
24 antibody encephalitis,^{26,27} and HLA-DQB1*05:01 in IgLON5-antibody disease.²⁸ However,
25 these HLA alleles exist at ~10-25% rates in the control population, suggesting multiple
26 other factors must contribute to clinical disease. These three conditions also share two
27 other distinctive and core features: late onset age and dominance of the rarest IgG
28 subclass in serum, IgG4, a distinctive triad likely related to fundamental T-B cell
29 interactions. Indeed, HLA Class II molecules on professional antigen presenting cells,
30 notably dendritic and B cells, traditionally present peptide antigen fragments to cognate T
31 cell receptors on autoantigen reactive CD4⁺ T cells.²⁹ By providing the key signal mediating
32 this critical interaction, HLA molecules shape the pivotal CD4⁺ T - B cell interaction to
33 initiate or suppress the subsequent immune response (Fig. 1). This interaction likely

1 operates in germinal centres – sites of clustered lymphocytes which act as ‘training
2 grounds’ for T-B interactions, mostly situated in lymph nodes - and can initiate both
3 immunoglobulin (Ig) class switch recombination (CSR), constant gene region exchanges
4 which lead to expression of a different Ig isotype or subclass, and somatic hypermutation
5 (SHM), a process by which targeted mutations are introduced into variable regions of B cell
6 receptors. These are two of the major mechanisms which focus the immune response
7 against a given antigen.³⁰ A single dominant HLA likely suggests T-B interactions are
8 markedly biased by few peptides which efficiently bind the specified HLA and that this
9 peptide-HLA complex is then recognised by, likely oligoclonal, T cells. Hence, the dominant
10 HLA associations may reflect limited variation across the CD4⁺-HLA-peptide interactions,
11 presenting a relatively stereotyped drug target for future therapeutic options.

12

13 Outside of HLA

14 In LGI1-antibody encephalitis, polygenic risk scores attribute up to 25% of disease risk to
15 genetics.²⁴ Yet, *PTPRD* is the only non-HLA gene validated in this condition, with patients
16 very rarely reported to have mutations in *FOXP3*, the canonical transcription factor
17 regulating T regulatory cell function.^{31,32} NMDAR- and MOG-antibody diseases show weaker
18 or no HLA associations,³³⁻³⁶ perhaps suggesting greater polyclonality and promiscuity
19 within the range of potential CD4⁺-HLA-peptide complexes. However, these two conditions
20 also show stronger associations with non-HLA genes (Fig. 1). NMDAR-antibody encephalitis
21 associates with *IFIH1* and specific killer cell immunoglobulin-like receptor (KIR) genes,
22 notably KIR2DL5B*00201.^{33,34} *IFIH1* is an intracellular sensor of viral RNA that triggers
23 interferon production, and KIR genes encode proteins expressed principally by natural killer
24 (NK) cells which interact with MHC Class I molecules. Hence, rather than providing clear T
25 or B cell signals to dissect pathogenic pathways, these associations implicate innate
26 immunity in some NGSAb-diseases,⁹ supported by rare cases with mutations in *IRAK4*.²² In
27 AQP4⁺NMOSD, modest associations with HLA-DQ and DR genes have been identified, with
28 stronger signals from C4A-B complement genes: all encoded on chromosome 6.^{36,37}
29 Interestingly, of the big six NGSAb-diseases, the most marked complement deposition is in
30 AQP4⁺NMOSD tissue, linking genetic-pathological observations with the near complete
31 responsiveness of this condition to complement inhibitors.³⁸

32 In summary, genetic observations offer both confirmatory and hypothesis-modifying
33 observations which highlight fundamental pathways of innate and acquired immune
34 defects as key contributors to NGSAb-disease initiation. Yet, familial clustering of
35 neurologic autoimmunity remains rare and individual allele effect sizes are modest,
36 suggesting other major contributions to autoimmune neurology pathogenesis.

1 Exogenous triggers: infection, tumours, iatrogenic aetiologies and the 2 microbiome

3 Infection

4 Among potential environmental triggers, infectious agents can initiate inflammatory
5 responses with subsequent B and T cell proliferation.³⁹ Herpes simplex virus encephalitis
6 (HSVE) represents the best-established infectious trigger for AE, occurring in ~25% of
7 cases, most commonly with NMDAR-antibodies, and particularly in the young.⁴⁰⁻⁴³
8 Currently, Japanese encephalitis virus (JEV) is the only other consistently reported
9 infectious trigger for AE.^{44,45} It is intriguing to ask why this response is being initiated by
10 CNS, rather than peripheral, infections.⁴⁶ A plausible model suggests HSVE or JEV induce
11 neuroglial damage which exposes previously CNS-sequestered antigens. Peripheral
12 lymphocytes may never have been tolerized to CNS-restricted autoantigens. Hence these
13 autoantigen-ignorant lymphocytes, likely draining to the periphery via cervical lymph nodes
14 (CLNs), are more prone to escaping tolerance, potentially explaining the necessity for a
15 CNS infection to induce CNS autoimmunity.⁴⁷

16 The proposed model of peripheral immune priming also aligns with the several-fold higher
17 levels of NMDAR-antibodies typically observed in serum versus CSF,^{2,48} the presence of
18 serum-only NMDAR-antibodies in an additional ~20% of patients who do not develop
19 symptomatic AE after HSVE, and the higher plasma IFN gene signatures as a predictor of
20 secondary autoimmunity after HSVE.⁴⁹ It remains unclear why the NR1 subunit of the
21 NMDAR is commonly targeted in this process. Explanations may include a high frequency
22 of NR1-reactive naïve B cells, as observed with CASPR2,⁵⁰ or in limited thymic tolerance
23 against the NMDAR, a phenomenon observed for both NR1 and NR2B subunits.⁵¹
24 Additionally, it is not understood why other modes of CNS tissue injury, for example stroke,
25 traumatic brain injury or even other brain infections recognised to date,^{41,52-55} do not
26 generate secondary AE. Perhaps this suggests the degree or nature of tissue damage alone
27 is an insufficient trigger, and that a specific inflammatory milieu may be required. This
28 environment may associate with age-related immune alterations to explain the predilection
29 for AE in younger HSVE patients.⁴⁹ The higher rates of serum HSV-IgG in patients with
30 NMDAR-antibody encephalitis without symptomatic CNS infections led to the alternative
31 suggestion, that non-encephalitic HSV exposure may trigger cases of NMDAR-antibody
32 encephalitis.⁵⁶ Yet, the overall high (~25-50%) rates of HSV-IgG in healthy controls suggest
33 this exposure itself is insufficient for a rare disease to manifest, but may be a factor in
34 breaking peripheral tolerance, towards a pathogenic cascade. The high HSV-IgG, yet low
35 NMDAR-IgG, seroprevalence suggests that B cell receptor (BCR) molecular mimicry
36 between HSV proteins and NMDARs is also alone insufficient to initiate NMDAR-antibody

1 encephalitis. Explaining this specificity and sequence from infection-through-
2 autoimmunity creates opportunities for research questions to elucidate key pathogenic
3 mechanisms of direct relevance to disease prevention and treatment.

4 Tumours

5 Autoantigen exposures may also arise in the context of systemic cancer. Indeed, the
6 expression of a shared antigen in both a tumour and in the CNS is a defining feature of a
7 paraneoplastic neurological syndrome (PNS).⁵⁷ Most such paraneoplastic antigens are
8 cytoplasmic or nuclear in localisation, suggesting the initiating event is likely to be a T cell
9 response to surface HLA-expressed antigens or a secondary B cell response to antigens
10 exposed after necrosis in the local tumour microenvironment.^{58,59} The activation of
11 lymphocytes directed against tumour-antigens may be beneficial, supported by evidence
12 that patients with PNS exhibit lower metastatic burden and improved survival compared to
13 cancer patients without PNS.^{58,60,61} Why only a minority of cancer patients develop PNS
14 remains unclear, and provides potential insights into NGSAb-disease immune initiation.
15 While tolerance mechanisms typically aim to constrain such autoreactivity, the
16 inflammatory tumour microenvironment and tumour-derived *neoantigens*, to which the
17 immune system has theoretically been ignorant, create ideal scenarios to evade self-
18 tolerance.⁶² For example, the presence of frequent intratumoural genomic alterations in the
19 Yo antigens, CDR2/CDR2L, are described in PNS-associated ovarian carcinomas.^{63,64} Also,
20 in tumours from patients with other PNS (e.g. GABA_B receptor- and NMDAR-antibody
21 encephalitis), the tumours show dysplastic neurons,⁶⁵ higher somatic mutation rates, and
22 chromosomal gains of antigen-coding genes,^{63,66} all features of autoantigens which may
23 preferentially break immune tolerance. Additionally, in some tumours, the presence of
24 dense immune infiltrates forming tertiary lymphoid structures containing the autoantigen,
25 suggest the organisation of cellular architecture may determine the development of PNS.⁶⁷⁻
26 ⁷⁰ The detection of NMDAR-reactive B cells in these tumours, and soluble NMDAR-
27 antibodies in their cystic components, provides a paradigm which suggests the tumour
28 contains all the necessary machinery to generate the end effectors of NGSAb-disease.

29 Medications

30 In addition, a common iatrogenic intervention in oncology – the administration of immune
31 checkpoint inhibitors (ICI) - can also trigger NGSAb-diseases, typically within 6-10 weeks of
32 ICI administration.⁷¹ As the main checkpoint molecular targets, CTLA4 and PD-1, are
33 principally expressed on regulatory T cells, the mechanism is thought to be via disinhibition
34 of these cells, with consequent B cell activation.⁷² This process can result in both typical
35 autoantigen-clinical associations and in well-recognised autoimmune neurological
36 syndromes without recognised autoantibodies. Hence, unleashing of endogenous

1 autoreactive lymphocytes with ICIs may be an opportune avenue to discover novel
2 autoantigens and, additionally, to explore novel emergent phenotypes which may not yet
3 associate with autoantibodies.^{73,74} The occurrence of NGSAb-mediated diseases in the
4 post-transplant immunosuppressed setting has also been described, suggesting that other
5 drugs which modify diverse immune functions can also induce these illnesses.⁷⁵

6 Microbiome

7 Finally, as robustly established in the pathogenesis of multiple sclerosis,^{9,76-80} the gut
8 microbiome is a plausible contributor to the multistep cascade initiating NGSAb-diseases.
9 However, limited evidence of this exists to date. Experimental data show that a HLA-DR
10 restricted immunodominant peptide from the AQP4 protein can activate patient T cells,
11 and shows high homology with a *Clostridium perfringens* derived peptide.⁸¹ Further, this
12 very organism was over-represented in AQP4⁺NMOSD patient stool.⁸² This concept
13 suggests potential for direct molecular mimicry, at the T cell level. Yet, faecal *Clostridium*
14 *perfringens* has a high prevalence, and hence this could only provide one step in the
15 cascade to pathogenicity for AQP4⁺NMOSD. Additionally, early data suggest reductions in
16 short chain fatty acid producers in both LGI1- and NMDAR-antibody patients,^{83,84} consistent
17 with a pro-inflammatory shift in the intestinal environment which may provide a less
18 tolerogenic milieu for disease initiation.

19 Other considerations

20 Overall, it is very challenging to analyse or estimate the number of hits or total duration of
21 this presymptomatic phase. Other than studying the conversion from HSVE to AE and
22 longitudinal cases administered ICIs, there are few realistic methods to enrich cohorts for
23 presymptomatic or 'at risk' individuals. Therefore, prospective studies in these well-defined
24 two cohorts may be fruitful in understanding features which tip the balance to NGSAb-
25 disorders. Does it take days, months or years to manifest a NGSAb-disease? In ICI-induced
26 disease, time from medication exposure to clinical features is rapid, suggesting a
27 preformed set of autoantigen-reactive T-B cells are primed by the release of inhibition. A
28 contrasting clue comes from patients with acetylcholine-receptor antibody positive
29 myasthenia gravis who develop AQP4⁺NMOSD after a median of 16 years.⁸⁵ Yet, in a case
30 report, AQP4⁺NMOSD occurred only ~2 months after a bone marrow transplant, with a
31 clear *de novo* detection of both AQP4-IgG and AQP4-IgM,⁸⁶ the latter consistent with a new
32 immune response as observed after exposure to novel pathogens. Overall, marked
33 potential variability exists in the duration from presymptomatic to symptomatic, and may
34 depend upon the nature and vigour of the immune priming event.

35

1 Peripheral autoantigen-reactive lymphocyte priming

2 After triggering events, autoreactive B cells must escape tolerance mechanisms to
3 generate antibody secreting cells (ASCs) that produce pathogenic NGSAbs.⁸⁷ The log fold
4 higher levels of autoantibodies seen in serum versus CSF strongly implicate the periphery,
5 rather than CNS, as the initiating site of this response.² Important questions which
6 influence our understanding of pathophysiology and potential therapeutic approaches
7 include: where in peripheral B cell development these checkpoint breaches occur, how
8 and if T cells are involved, which plasma cells secrete the autoantibodies and the relative
9 roles of anatomical neuroimmune compartments in driving these processes.²

10 Checkpoints and naïve B cells

11 In the bone marrow, developing B cells progress from preB- to immature B cell stages by
12 traversing a central tolerance checkpoint (Fig.2). Here, the ‘Goldilocks’ threshold level of
13 reactivity (which is ‘just right’) between the membrane-inserted BCR and autoantigens
14 determines whether a cell will: 1) modestly recognize its autoantigen and survive its
15 journey through B cell development, 2) react excessively and die by apoptosis, or 3) react
16 too much but subsequently successfully edit its light chain so its BCR acquires reduced
17 autoantigen reactivity, facilitating its survival.^{2,88,89} The relative role of long bones versus,
18 the recently rejuvenated importance of, skull bone marrow in this process have not been
19 investigated in humans, but recent animal work has suggested that physical
20 communication channels between skull bone marrow and CSF may permit early skull bone
21 marrow resident B cells to be rapidly mobilised to the CNS parenchyma and its borders.⁴⁶

22 After bone marrow- egress into the circulation as new emigrant B cells, peripheral
23 tolerance checkpoints further curate the repertoire (Fig.2). Here, BCR-autoantigen
24 interactions determine survival as a mature naïve B cell via apoptosis or anergy, a process
25 by which an autoantigen-reactive B cell becomes functionally quiescent.^{2,8,90} Hereafter,
26 other peripheral checkpoints, including selection processes in germinal centres, may
27 further modify the frequencies of autoreactive B cells.^{30,91,92}

28 The relative integrity of central and peripheral checkpoints have been predominantly
29 examined in primary immunodeficiencies and multiple sclerosis.^{8,90} These studies use
30 generic measurements of autoreactivity, reactivity to Hep-2 cells, dsDNA, insulin and
31 lipopolysaccharide, as their principal readouts. More recently, the presence of reactivity to
32 the specific NGS-autoantigens has been determined across these checkpoints. Naïve B
33 cells from CASPR2-antibody encephalitis patients and healthy controls both react with
34 CASPR2 at surprisingly high frequencies (~0.5%), suggesting promiscuous central
35 tolerance in both disease and health. However, CASPR2-reactive memory B cells are

1 identified exclusively in patients, implicating more rigorous later peripheral checkpoints
2 successfully curtail CASPR2-reactivities in health.⁵⁰ By contrast, while AQP4⁺NMOSD
3 patients harbour naïve B cells with AQP4-reactivities, consistent with a breach of the
4 central checkpoint, these are not observed in healthy controls. Hence, tolerance to NGS-
5 antigens may be differentially regulated in health.⁹³ It is tempting to speculate that B cells
6 in bone marrow rarely encounter the almost exclusively CNS-expressed proteins (such as
7 MOG, CASPR2 and LGI1) but are more likely tolerized against CNS autoantigens which are
8 also peripherally expressed (e.g. AQP4). Future experiments should address this question.
9 However, as no NGSAb-focused studies to date differentiate new emigrant from mature
10 naïve B cells, a precise analysis of the conventional checkpoints is currently limited.
11 Nevertheless, unmutated autoantigen-reactive B cells have been isolated in both MOGAD
12 and from NMDAR-antibody encephalitis,^{94,95} and when mutations are removed from
13 patient-derived BCRs against LGI1, CASPR2 and NMDARs, many retain binding against
14 these autoantigens.^{15,94} These collective findings identify naïve autoantigen-reactive B cells
15 as a feature common across NGSAb-diseases, suggesting a consistent feature of these
16 diseases is the very early loss of tolerance against the key disease-defining autoantigen.
17 This finding also directly implies that deletion of early B cell populations may represent an
18 under-appreciated, but important, mechanism of action of therapeutics, such as anti-
19 CD19 and anti-CD20 monoclonal antibodies (Fig.4). Moreover, this explain why HSCT and B
20 cell targeting chimeric antigen receptor T cell (CAR-T) strategies may offer longer-lasting
21 disease resolution (Fig.4).

22 T cell involvement and germinal centres

23 Yet, in AQP4⁺NMOSD, and other non-neurological autoantibody mediated diseases,
24 germline reversion of BCR mutations is reported to abrogate autoantigen-reactivity,⁹⁶⁻⁹⁸
25 strongly implying a necessity of SHM for autoantigen-recognition, arguing for insufficiency
26 of naïve B cells. In line with this, by comparison to mutated LGI1-, CASPR2- and NMDAR-
27 reactive BCRs, loss of mutations consistently decreases affinity for target autoantigens.
28 Hence, despite the importance of naïve B cells, SHM likely plays a major role in delivering
29 highly targeted pathogenic autoantibodies. B cells undergo SHM in germinal centres after
30 receiving T cell help via molecular interactions including the peptide-HLA-TCR complex,
31 and costimulatory pathways including CD80/86-CD28/CTLA4 and CD40-CD40L (Fig.1). T-
32 dependent pathways represent viable and, in the case of the former, highly-specific
33 potential therapeutic targets to terminate germinal centre reactions.⁹⁹ Deletion of the
34 professional antigen presenting cells may be an alternative method to therapeutically
35 terminate peptide-HLA presentation to T cells in germinal centres. Yet, and by comparison
36 to the more pronounced SHM observed in AQP4-, LGI1- and CASPR2-reactive BCRs, the
37 low mutation rates observed in many MOG- and NMDAR-BCRs suggest their receipt of

1 limited T cell help, and hence restricted participation in germinal centres (Table 1).
2 However, direct T cell studies to date do not linearly superimpose upon these predictions.
3 While AQP4-, MOG- and NMDAR-reactive CD4⁺ T cells have been directly identified,^{81,100-102}
4 LGI1-reactive T cells were undetectable, creating a vacuum regarding the identity of cells
5 which induce observed mutations in LGI1-reactive BCRs.¹⁰²

6 Nevertheless, classical germinal centres are typically formed in secondary lymphoid
7 organs, mostly lymph nodes. In mice, those which predominantly drain the CNS are the
8 CLNs,¹⁰³ and receive contents of meningeal lymphatics that carry mixed CNS extracellular
9 fluid and cerebrospinal fluid, and are hence enriched for CNS-derived autoantigens.¹⁰⁴
10 Direct sampling of human CLNs has identified high frequencies of NMDAR- and AQP4-
11 reactive B cells, alongside intranodal synthesis of AQP4-IgGs,^{105,106} implicating CLNs as
12 local sites of focused autoantigen-reactive immunisations. Further, human CLNs contain
13 enrichments of multiple CNS expressed proteins, including neurodegenerative biomarkers
14 such as tau.^{107,108} Taken together, CLNs may provide a hub of neuroimmune autoantigen-
15 lymphocyte communications which initiate and propagate the autoimmunisation that
16 culminates in generation of NGSABs.

17 Classical germinal centre activity can generate both memory B cells and ASCs.
18 Additionally, extrafollicular responses, which occur inside secondary lymphoid organs but
19 outside of germinal centres, more rapidly generate short-lived ASCs but these induce
20 relatively transient antibody responses which are typically unmutated, lower affinity and
21 often of the IgM isotype. However, both extrafollicular and germinal centre responses can
22 initiate Ig CSR,¹⁰⁹ in which IgD and IgM BCRs in naïve B cells express one of the downstream
23 immunoglobulin heavy chain genes encoding IgG3, IgG1, IgA1, IgG2, IgG4, IgE or IgA2.¹¹⁰
24 Patterns of immunoglobulin subclasses represent consistent signatures of each
25 autoantigen-specificity (Table 1): IgG4-dominant LGI1, CASPR2 and IgLON5 antibodies
26 versus IgG1 dominant NMDAR, MOG and AQP4-antibodies. Subclasses can endow BCRs
27 and soluble autoantibodies with different functional capabilities, such as efficient
28 complement activation with IgM, IgG1 and IgG3, raising the important question of how to
29 mechanistically link autoantigen-specificity with the predominant immunoglobulin
30 subclass, further discussed below. CSR can be strategically induced by certain cytokines
31 binding to DNA regions critical to the switching process,¹¹¹ and it may be that NGSAB-
32 diseases polarise to certain cytokine patterns.¹¹² This may also depend on the
33 immunisation microenvironment, for example NMDAR-IgAs predict an underlying ovarian
34 teratoma which typically contains substantial mucosal tissue.^{105,113} While mechanisms of
35 CSR are overall poorly understood they carry future therapeutic importance, as conversion
36 of one subclass to another may be sufficient to nullify pathogenicity of autoantigen-
37 reactive IgGs, for example in the case of complement fixing AQP4-IgG1s.

1 Antibody Secreting Cells

2 The ASCs generated by germinal centre activity are often classified by their longevity, as
3 either relatively short-lived or long-lived plasma cells (SLPCs and LLPCs, respectively).
4 SLPCs would typically require frequently renewed events of antigen exposure to create
5 sufficient IgG levels to maintain serum autoantibody titres for several years – a
6 phenomenon observed in most NSAb-diseases.¹¹⁴⁻¹¹⁶ In this model, either memory B cells
7 would re-enter germinal centres or de novo germinal centre activity would be initiated by
8 invasion of naïve B cells. In repeat animal immunizations, the latter replacement model
9 appears dominant.¹¹⁷ In humans with NGSAb-diseases, the frequent presence of NMDAR-
10 and AQP4-IgMs in patient sera may suggest new IgM⁺ naïve founder clones are continually
11 invading germinal centres.^{105,106} As AQP4-IgMs are closely correlated with relapse timings,
12 these naïve B cells may also have direct clinical pertinence.¹⁰⁶ Similarly, the balance
13 between a monophasic versus relapsing NGSAb-disease course may depend up on the
14 equilibrium established between generation of SLPCs versus LLPCs. LLPCs usually return
15 to a poorly-drug-accessible bone marrow niche where they are non-proliferative and
16 dedicate much energy to secretion of durable, high affinity- immunoglobulins against a
17 single target autoantigen.^{87,118,119} These LLPC-derived antibodies are thought to effectively
18 maintain natural and vaccine-induced immunity against varied organisms, potentially for
19 decades.^{2,120} However, their role in NGSAb-diseases, and other autoimmune conditions,
20 has received very little attention, likely as there is limited availability of patient bone
21 marrow for studies. By inference, serum autoantibody levels which do not change
22 substantially after administration of rituximab is consistent with the dominant contribution
23 of CD20⁻CD19⁺ LLPCs to serum IgGs. In AQP4⁺NMOSD, the antibody is considered as
24 lifelong in most patients, implicating its secretion by LLPCs. Clinical quiescence and
25 conversion to seronegative status is observed after haemopoietic stem cell transplantation
26 (HSCT), with virtually no other treatments rendering these patients seronegative.¹²¹ As
27 chemotherapy prior to HSCT is intended to deplete LLPCs, this observation may strengthen
28 a role for these cells in maintenance of AQP4-IgGs. However, HSCT has wide ranging
29 cellular effects and does not appear to be a permanent solution: exemplified by cases who
30 seroreverted after several years, likely indicating a fundamental predisposition to *de novo*
31 generation of this antibody in AQP4⁺NMOSD patients.¹²² CAR-T cell therapies which can
32 target both SLPC and LLPC populations, for example those with CD19 or BCMA
33 specificities, may more robustly reduce autoantibody levels, as suggested by early
34 studies.¹²³

35

1 Gaining CNS access

2 Fundamentally, NGSAbs must reach their target autoantigens in the CNS to be pathogenic.
3 An important question is whether NGSAbs are passively or actively transported into the
4 CNS or whether the peripherally primed B cells are the major traffickers. Also, knowledge
5 around the duration of lymphocyte and antibody retention in the CNS will provide
6 important potential therapeutic implications.

7 Cells in CSF

8 Access mechanisms may vary based on the integrity of the blood brain barrier. Gadolinium
9 enhancement suggests this is highly permeable in, for example, AQP4⁺NMOSD where most
10 acute lesions enhance: this creates potential for substances with large molecular weights,
11 such as IgG and IgM to enter the brain parenchyma. Whereas in LGI1-, CASPR2-, IgLON5-
12 and NMDAR-antibody diseases, and 50% of MOGAD cases, no or very little gadolinium
13 enhancement is observed,¹²⁴ suggesting a more stringent and selective mechanism of
14 entry may be necessary. Yet, in all these conditions studied to date, CSF contains a
15 remarkably high frequency of autoantigen-reactive B cells and ASCs, reported at between
16 ~50-80% in AQP4-, LGI1- and CASPR2-antibody diseases and ~10% in NMDAR-antibody
17 disease.^{13,15,125} This suggests a deliberate process is selectively homing these autoantigen-
18 reactive cells to the CSF, where they can then secrete pathogenic antibodies in the close
19 vicinity of their targets. It may be a high local IgG concentration is required for disease. This
20 chemoattraction process may involve soluble factors including cytokines and chemokines,
21 such as CXCL13. CXCL13 is elevated in CSF of patients with NMDAR-antibody encephalitis
22 and its levels correlate with the degree of intrathecal NMDAR-antibody synthesis, a
23 surrogate for the number of CSF NMDAR-reactive plasma cells.¹²⁶ Blocking CXCL13, or
24 other chemoattractants, may represent an attractive therapeutic avenue. Yet, cytokines
25 would not be predicted to specifically retain the autoantigen-specific cells in CSF, but
26 rather retain cells agnostic to their BCR specificity. Hence, the observed enrichments of
27 autoantigen-specific cells in CSF strongly indicates the hitherto unproven presence of
28 soluble autoantigens in CSF. While LGI1, CASPR2, MOG and IgLON5 can be secreted,¹²⁷⁻¹³⁰
29 it is less clear how this process may operate for non-secreted proteins such as AQP4 or the
30 NMDAR.

31 Retention of cells in CNS

32 While attractant signals remain unproven, once in the CSF, B cells may establish tertiary
33 lymphoid structures and a plasma cell niche, as observed in the meninges of patients with
34 multiple sclerosis.¹³¹ If so, CNS penetrant therapies may be required for treatment of
35 NGSAb-diseases. Yet, at least in LGI1- and CASPR2-antibody diseases, most mutations

1 and affinity for the autoantigen appear peripherally acquired, prior to CSF access,
2 suggesting limited autoantigen-reactive B cell maturation is observed in CSF in NGSAb-
3 diseases.¹⁵ However, given the vast diversity of BCRs, cross-sectional assessments may
4 not detect maturation of individual NGSAb-reactive B cells, implicating longitudinal CSF
5 assessments are required in future studies. Histology may be a gold standard for
6 determining the presence of tertiary lymphoid structures in these diseases, although as
7 mortality is thankfully low, few specimens are available. Yet, to date, histology in MOGAD,
8 AQP4⁺NMOSD and NMDAR-antibody encephalitis have all reported the absence of
9 meningeal follicles.¹³²⁻¹³⁴

10 The role of CSF T cells is less well established and initial studies suggest overall limited
11 clonality in the CSF of LGI1- and CASPR2-antibody patients, suggesting few autoantigen-
12 reactive T cells and aligning with the notion that there is limited help for BCR affinity
13 maturation within the CNS.^{15,135}

14 Antibody access to CSF

15 It is also possible that soluble serum IgGs directly access the CSF or CNS parenchyma. In
16 AQP4⁺NMOSD, lesions can occur around the ventricles, where the blood-brain barrier is
17 incomplete, suggesting that the constitutively expressed antigen may be exposed to
18 pathogenic systemic IgGs.¹³⁶ Yet, for AQP4⁺NMOSD, a substantial proportion of the CSF
19 immunoglobulin proteome overlaps with CSF BCR sequences, with limited stand-alone
20 contributions from blood, indicating that intrathecal B cells provide much of the soluble
21 CSF IgG.¹⁴ Nevertheless, the far higher levels of autoantibodies in serum versus CSF
22 provide a suitable gradient for diffusion into CSF, and perhaps this contribution to CSF is
23 greater in scenarios with more disruption of the blood brain barriers. Yet, even if IgG does
24 access the CNS in high enough concentrations to elicit disease, it is likely that efflux
25 mechanisms will – often rapidly – remove it, an important consideration in development of
26 CNS-retained monoclonal antibodies.¹³⁷

27

28 Autoantibody-induced end-organ dysfunction

29 General principles

30 Once autoantibodies access the exposed extracellular domains of antigenic targets, they
31 can exert pathogenic effects through several distinct and overlapping mechanisms, each
32 with potential therapeutic implications. Strikingly, the main NGSAb-mediated diseases
33 conveniently divide into those with dominant IgG4 versus IgG1 subclasses of the

1 autoantigen-specific autoantibodies (Table 1), each with fundamentally different biological
2 properties. IgG1 antibodies are bivalent molecules which can both activate the classical
3 complement cascade, culminating in chemoattraction via C3a and C5a and in C5b-9-
4 mediated membrane attack complex (MAC) pore formation with resultant target cell lysis,
5 and antibody-dependent cellular cytotoxicity (ADCC) when the immunoglobulin Fc
6 domains interact with Fc receptors on innate immune cells.^{138,139} In contrast, IgG4
7 antibodies mediate very limited complement activation or ADCC but possess the peculiar
8 ability to exchange their two halves: a phenomenon termed Fab-arm exchange, which
9 renders many IgG4 molecules functionally monovalent and therefore markedly diversifies
10 their range of potential antigenic targets.¹⁴⁰ IgG4-dominant NGSAb-diseases rarely only
11 show autoantigen-reactive IgG4s, but these are the least prevalent subclass in human
12 serum (~3% of total IgG), emphasising the striking skew observed in these conditions.
13 Perhaps given these discrete reported effects, there are no studies which attempt to
14 quantify these relative effects across multiple NGSAb-mediated diseases. Table 1 aims to
15 estimate relative effects of complement activation, antigen internalization, steric
16 hinderance and ADCC.

17 For the big six NGSAb-mediated diseases, intraventricular or intrathecal 'passive' transfer
18 of the autoantibodies to experimental animals can reproduce a phenotype and/or
19 histological changes consistent with those observed in the corresponding patients, a
20 traditional confirmation of their pathogenicity.^{1,3,141-145} For AQP4⁺NMOSD, this requires an
21 additional source of complement, and for MOG the marked differences between rodent
22 and human MOG inevitably limit modelling to the cross-reactive IgGs.^{144,145} While
23 convenient proof-of-concept studies, these simplistic models fail to capture the upstream
24 cellular events, triggers and contributions of factors other than soluble IgG. To partly
25 reproduce such key elements of the human disease, further active immunisation models
26 are required to initiate disease at the level of the provoking autoantigen.^{146,147}

27 AQP4

28 Despite generalisable characteristics, each of the IgG1 dominant diseases appears to
29 mediate neuroglial dysfunction via different dominant mechanisms. AQP4-antibodies
30 show the capacity to internalise surface AQP4, a property dependent on an intact bivalent
31 IgG molecule facilitating AQP4 cross-linking.¹⁴⁸ The densely-packed arrays of AQP4 on
32 astrocyte end feet facilitate binding of C1q to multimerised Fc domains.¹⁴⁹ C1q activation
33 subsequently initiates classical complement pathways resulting in MAC deposition on
34 both astrocyte and neighbouring neuron cell membranes (Fig.3).^{148,150} Marked complement
35 deposition, alongside abundant IgG and IgM, is observed in AQP4⁺NMOSD pathology,¹⁵¹
36 tissue which also contains neutrophils, eosinophils and NK cells, providing ideal

1 substrates for ADCC (Fig.3B). Activated NK cells can release perforin and granzymes,
2 inducing apoptosis, whereas macrophages and neutrophils can phagocytose cells and
3 release cytotoxic mediators such as reactive oxygen species and proteases. Despite all
4 these possible mechanisms, the almost absolute efficacy of complement inhibitors in
5 AQP4⁺NMOSD suggests this is the major mechanism operating in patients.¹⁵²

6 MOG

7 MOG is expressed on the outer surface of the myelin sheath in the CNS. MOGAD
8 histopathological studies typically demonstrate more limited deposition of complement
9 components than in AQP4⁺NMOSD, likely because MOG is sparsely expressed on the outer
10 myelin sheath, representing approximately 0.05% of the surface proteins, limiting ability of
11 MOG-IgGs to form the hexameric assemblies required for C1q binding.¹⁵³ Yet, patient
12 derived MOG-IgGs can strongly induce CDC *in vitro* using cell lines expressing MOG,¹³⁹
13 perhaps as *in vitro* overexpression produces unnaturally high antigen densities which
14 enable hexamer formation. MOGAD tissue also shows marked infiltration and activation of
15 macrophages and microglia and MOG-IgGs can promote NK cell-mediated cytotoxicity
16 against MOG-expressing cells, where the extent of ADCC correlated strongly with serum
17 MOG IgG levels (Fig.3C).^{154,155} Further, MOGAD patient tissue contains active peripherally-
18 derived macrophages with early cytoplasmic myelin degradation products, including
19 MOG,¹⁵⁶⁻¹⁵⁹ further implicating the contribution of innate immune cell-mediated
20 mechanisms to demyelination in MOGAD.

21 NMDAR

22 The other common IgG1-dominant NGSAb-disease is NMDAR-antibody encephalitis. In
23 this condition, there is limited observed neuronal loss and pathology rarely shows
24 complement products.¹⁶⁰ Yet, the patient IgGs can deposit complement *in vitro* suggesting
25 that, like MOG-IgGs, the human antigen density and distribution may fail to promote C1q
26 binding. Rather, the dominant mechanism of NMDAR-antibodies appears to be NMDAR-
27 hypofunction induced by internalisation by the IgGs, an effect maintained with bivalent Fc-
28 deficient F(ab')₂ fragments but not with monovalent Fab fragments, indicating necessity of
29 both IgG arms to cross-link NMDARs (Fig.3D).¹⁶¹ This effect can occur very rapidly in
30 cultured neurons with markedly altered postsynaptic currents within just 30 minutes of
31 incubation with patient derived NMDAR-reactive monoclonal antibodies.¹⁶² It is plausible
32 that the combination of rapid direct channel modulation together with subsequent
33 internalisation of the NMDAR is sufficient for the complex psychiatric and multifocal
34 neurological dysfunction observed in patients with NMDAR-antibody encephalitis.^{19,161,163}

1 Studies of NMDAR-IgGs raise an intriguing broader point regarding IgG access to the
2 autoantigen (Fig.3A). Reaching targets in the synaptic cleft may be challenging because the
3 cleft is only 10–40 nm wide, and additionally filled with cell-adhesion molecules and
4 extracellular matrix proteins that physically bridge pre- and post-synaptic membranes. Yet
5 IgG is ~15 nm in size (Fig.3A). Indeed, a recent study demonstrated that, after 30 minutes,
6 NMDAR-autoantibodies target and internalize *extrasynaptic* NMDARs, inducing a major
7 reorganization of extrasynaptic membrane proteins which was sufficient to, within hours,
8 destabilise the synapse itself.¹⁶⁴ This concept of a ‘secondary synaptopathy’ may also apply
9 to the other NGSAb-diseases.^{164,165}

10 At a more macroscopic level, antigen distribution likely dictates the observed clinical
11 features. For example, AQP4 density is reported as highest in the optic nerve, spinal cord
12 and area postrema,¹⁶⁶ the three major sites of symptomatic pathology in AQP4⁺NMOSD.
13 Less intuitively, this disease manifests with few other CNS sites of pathology and no
14 apparent peripheral involvement, despite widespread AQP4 expression in the CNS,
15 stomach and kidney. It is plausible that these unaffected sites show lower antigen density
16 which prevents sufficient complement fixation, or that endogenous complement regulatory
17 proteins are expressed in these sites.¹⁶⁷ For other autoantigens, such as LGI1 or NMDAR,¹⁶⁸
18 their density is highest in the hippocampus, and many, but not all clinical features of these
19 diseases can be ascribed to hippocampal dysfunction, suggesting wider network
20 involvement is responsible for many of the observed clinical features.¹⁶⁹

21 LGI1

22 For the IgG4-dominant diseases, complement and ADCC are less likely effector
23 mechanisms. More often, these autoantibodies typically modulate protein-protein
24 interactions. Internalisation may be an additional effect of these autoantibodies but
25 perhaps limited if they commonly become functionally monovalent after Fab-arm
26 exchange.¹⁴⁰ LGI1 is a secreted protein, composed of an N-terminal leucine-rich repeats
27 (LRR) domain and a C-terminal epitempin (EPTP) domain, which forms a dimer or trimer
28 within a trans-synaptic complex, linking postsynaptic ADAM22 to presynaptic ADAM23
29 (Fig.3E).¹⁷⁰ Patient-derived monoclonal antibodies have been used to dissect the molecular
30 pathogenesis of this condition: antibodies which bind the LRR domain lead to
31 internalization of LGI1 and its neighbouring proteins, like presynaptic K_v1.1 and
32 postsynaptic AMPARs, whereas, EPTP binders block the docking of LGI1 with ADAM22/23,
33 disrupting downstream signalling without internalization (Fig.3E).¹⁷¹ Similar molecular
34 effects have been observed after transfer of patient serum IgGs to the ventricular system of
35 mice, alongside a phenotype consistent with memory loss.¹⁴² Further, when LGI1-reactive
36 antibodies are intraventricularly infused, rats appear to develop clinical and electrographic

1 seizures alongside altered Kv1.1 expression in the hippocampus.¹⁷² Non-synaptic
2 mechanisms may also play a role in LGI1-antibody pathogenesis, with proven effects at the
3 axon initial segment conferring the ability to disrupt action potential initiation and
4 integration of synaptic signals.^{173,174} This complex biology of LGI1, and its multiple targeted
5 domains, make it challenging to propose a simple method to therapeutically counter these
6 pathogenic antibodies. Further, it may be that the minority proportion of LGI1-IgG1
7 antibodies are more pathogenic. This is suggested by their correlation with cognitive
8 performance and hippocampal sclerosis, and the presence of complement deposition in a
9 few studied post-mortem brain tissues.¹⁷⁵⁻¹⁷⁷

10 CASPR2

11 CASPR2 is a membrane protein with a large extracellular domain and strongly expressed at
12 the juxtaparanodes of myelinated neurons, in both the CNS and PNS. It is the only one of
13 the big six with a prominent PNS phenotype. Functionally, CASPR2 assists Kv1 channels to
14 correctly localise and mediate repolarisation. However, despite dense expression of
15 CASPR2 at juxtaparanodes, shielding of this region by myelin appears to prevent access of
16 antibodies (Fig.3F). Indeed, a passive transfer model has identified CASPR2 antibodies do
17 not pass the paranodal barrier.¹⁷⁸ Peripheral unmyelinated sensory nerve terminals may be
18 a more likely target of CASPR2-antibodies, particularly as pain is a common feature in
19 these patients, a strong predictor of long-term disability and these patients often show loss
20 of epidermal small nerve fibres (Fig.3F).^{114,179-182} CASPR2 antibodies can also cause a pain
21 syndrome when transferred to mice, likely through dysfunction of the dorsal root ganglia via
22 Kv1 channels.¹⁸³ These subcellular localisations cannot account for the seizures, cognitive
23 disturbances and movement disorders observed in most patients with CASPR2-antibody
24 encephalitis. Here, it is thought CASPR2 expression at synapses is the pathological target
25 (Fig.3G). The main proposed mechanisms include internalisation of CASPR2, disruption of
26 the proposed interaction between contactin-2 and CASPR2, and the effect of CASPR2-
27 antibodies on AMPARs and post-synaptic hyperexcitability, observed within just a few
28 hours *in vitro* with dependence on the subclass of the CASPR2 monoclonal antibody.^{50,141}
29 Finally, as with LGI1, a role for complement has been raised from human histological
30 findings and may be consistent with the dominance of CASPR2-IgG1s in patients with
31 forms of CASPR2-antibody disease and encephalitis.^{184,185}

32 IgLON5

33 Like CASPR2, IgLON5 is a putative neuronal cell adhesion protein with a large extracellular
34 portion and 45–93% of IgLON5-antibodies are of the IgG4 subclass.¹⁸⁶ The IgG1 fraction of
35 IgLON5-antibodies reduces surface IgLON5 clusters and, with prolonged exposure,
36 neurofilament disorganization can be induced, an effect not observed with the IgG4

1 antibodies. It may be that the IgLON5- IgG4 antibodies modify or block neighbouring
2 protein-protein interactions, a more conventional mechanism for IgG4 antibodies
3 (Fig.3H).¹²⁹ Indeed, the marked deposition of IgG4s in brain tissue from patients early in the
4 disease, without overt tau deposition, is consistent with its key role in initiating neuronal
5 pathogenesis.¹⁸⁷ Later in the disease course, IgLON5-antibody disease patient postmortem
6 brains typically show tau deposition,¹⁷ and tau accumulation is reproduced in cultured
7 human and rodent neurons exposed to patient IgG and in passive transfer mouse models,
8 suggesting these antibodies are sufficient to induce features typical of neurodegeneration.
9 This paradigm hence provides a rare and compelling example of molecularly precise
10 immune-to-degeneration directionality in neurology.^{143,188}

11

12 How biology shapes resolution, relapse and chronicity

13 The natural disease trajectory is an additional fundamental feature which is likely best
14 modelled by understanding propagation of the immune processes. Although all these
15 conditions can relapse, tendencies to relapse are likely different, despite being potentially
16 confounded by immunotherapies (Table 1). At one extreme, AQP4⁺NMOSD is considered a
17 lifelong syndrome, given high recurrence risks after rituximab withdrawal and even after
18 ^{123, 122, 189} The return of memory B cells and the presence of AQP4-IgMs and AQP4-IgG
19 subclass shifts, represent promising indicators of imminent relapses in
20 AQP4⁺NMOSD,^{106, 190} consistent with the concept that clinical attacks arise secondary to
21 recurrent, dynamic germinal centre reactions with continued autoantigen availability and
22 incomplete restoration of tolerance.² Perhaps these reactions reflect T cell activation
23 licencing escape of AQP4-reactive B cells, and hence both AQP4-reactive T and B cells
24 need to interact for relapses to occur.¹⁰⁰ By contrast, it is rare for post-HSVE to relapse,
25 even by comparison to NMDAR-antibody encephalitis. Perhaps in this scenario the
26 immune milieu created by acute infection and the infectious trigger itself have been
27 eliminated. Similarly, relapse rates in NMDAR-antibody encephalitis are markedly reduced
28 by teratoma removal,^{19, 161} suggesting elimination of a pivotal germinal centre and the
29 associated aberrantly expressed autoantigen are key to remaining monophasic.
30 Alternatively, in this and other long-lived diseases such as NMDAR-antibody encephalitis
31 and some forms of MOGAD, autonomous actions of long-lived plasma cells could be key to
32 persistent autoantibodies and disease activity. However, it is a consistent observation
33 across NGSAb-diseases that the NGSAb levels per se show minimal correlations with
34 clinical outcomes, suggesting this population of cells are not driving disease
35 activity.^{4, 114, 115, 191} It is also unclear whether recurrent CSF entry is required for relapses or

1 ongoing disease activity, or if immune cells take up more permanent residency in the CNS,
2 a concept which feeds into the tertiary lymphoid structure discussions above.

3 While the autoantigenic cascade forms the core of the pathology in most NGSAb-diseases,
4 additional complexity may exist beyond these observations. The pathogenic pathways may
5 begin as autoantigen specific but can evolve to affect large numbers of neurons or glia,
6 leading to a cascade of more generic events associated with cell damage and network
7 dysfunction. Examples include the diffuse involvement of white matter tracts and regions
8 outside of the hippocampus in LGI1-antibody encephalitis,^{169,192} the move from an
9 astrocyte focused injury to neuronal damage in AQP4⁺NMOSD,^{148,150} and, in NMDAR-
10 antibody encephalitis, the development of clinical features which may be more loosely
11 related NMDAR dysfunction, including the movement disorders. These downstream
12 molecular cascades have received limited study to date but may represent important
13 methods to assess mechanisms of longer-term injury which markedly impair quality of life
14 across these diseases.^{114,193,194}

15

16 Therapeutic targets along the cascade

17 Current practice

18 As mentioned above, these fundamental immunological and neurobiological observations
19 can provide targeted avenues for precision therapeutics (Fig.4). This is of key importance as
20 clinical trials have only yielded FDA approved therapies for one of the NGSAb-mediated
21 disorders, AQP4⁺NMOSD, and several clinical trials ongoing in the other diseases have
22 already failed to recruit adequately.¹⁹⁵

23 Historically, the treatment of autoimmune neurological diseases has relied upon drugs
24 with broad mechanisms of action, across lymphoid and myeloid lineages and, often, both
25 neurons and glia. These medications often target the generic machinery of cell replication
26 (e.g. azathioprine, mycophenolate mofetil) or downstream signalling pathways (e.g.
27 corticosteroids) common to multiple cell lineages, resulting in both broad
28 immunosuppression, CNS activity and significant multi-organ adverse effects.¹⁹⁶ In the
29 acute phase of treatment, the myriad actions of corticosteroids, intravenous
30 immunoglobulins, and plasma exchange may be advantageous in inhibiting multiple
31 pathways, particularly if the precise diagnosis is not yet known. Second-line acute or
32 maintenance therapies, often in cases of more severe and refractory presentations,
33 include other broadly acting agents including deletion of the whole B cell lineage with
34 CD19 or CD20 targeting medications (e.g. inebilizumab or rituximab, respectively),

1 blockade of the highly pleotropic IL-6 receptor (tocilizumab and satralizumab) or the anti-
2 proliferative cyclophosphamide.^{196,197} The best example of a more targeted therapeutic
3 used in routine clinical practice are the C5 complement protein inhibitors, which almost
4 entirely eliminate relapses in patients with AQP4⁺NMOSD.^{152,198}

5 However, many of these drugs remain broadly immunosuppressive or, if more targeted, are
6 associated with specific and/or serious side effects.^{195,199} In an attempt to reduce these
7 adverse effects, the underlying biology has illuminated current and future options with
8 likely greater precision which work by targeting: (1) Pathways involved in autoantibody
9 production and maintenance, (2) Autoantibody effector functions (Fig. 4).

10 Targeting pathways of autoantibody production and maintenance

11 Many utilised and proposed medications for NGSAb-diseases target the pathways of B cell
12 differentiation and function as this cell line is most directly implicated in autoantibody
13 production and therefore, disease pathogenicity. Anti-CD20 monoclonal antibodies (e.g.
14 rituximab, ocrelizumab, ublitixumab, ofatumumab) target B cells in their life cycle from
15 pre-B cells to memory cells and some ASCs which retain CD20, therefore not deleting
16 plasma cells or some of the earliest bone marrow resident B cells (Fig. 2).²⁰⁰ The extent to
17 which these agents can mitigate disease activity may depend on several immunobiological
18 factors, particularly which aberrant B cell population is implicated in driving the ongoing
19 generation of NGSAb, the immunoglobulin subclass, and the disease process. For
20 example, the success of rituximab in AQP4⁺NMOSD may relate to its ability to abrogate
21 germinal centre B cell activity,⁹⁸ while in LGI1-antibody encephalitis, its more limited
22 reported efficacy may be due to its inability to delete most ASCs or penetrate CSF to target
23 B cells.^{12,177} Intriguingly, there are also some reports of CD20 expression on select T cells,
24 an additional unforeseen mechanism by which these drugs may show clinical efficacy.²⁰¹
25 Anti-CD19 therapeutics target a broader range of the B cell lineage than anti-CD20 agents,
26 creating value added in diseases where plasmablasts, SLPCs and LLPCs are implicated in
27 pathogenic autoantibody production.²⁰² Similarly, anti-CD38 medications (e.g.
28 daratumumab) and proteasome inhibitors (e.g. bortezomib) can somewhat selectively
29 target these plasma cell populations, making them useful in pathologies where the activity
30 of LLPCs and CSF-resident ASCs may drive ongoing disease.^{195,203,204}

31 Natalizumab targets the alpha4beta1 integrin to block activated lymphocytes from entering
32 the CNS, a process which appears to be important across NGSAb-disease pathogenesis
33 given the enrichments of autoantigen-specific B cells observed across multiple NGSAb-
34 diseases.^{13,15,125} Additionally, as the alpha4beta1 integrin is expressed by B and T cells, it
35 may be broadly effective against multiple immunological pathways which contribute to
36 pathology.²⁰⁵

1 More recent attention has turned towards CAR-T therapies targeting CD19, CD20 and B cell
2 maturation antigen (BCMA). This intervention has reported success in individual
3 neurological cases with myasthenia gravis, MOGAD, AQP4⁺NMOSD and stiff person
4 syndrome, plus a promising phase 1 trial of BCMA CAR-T in AQP4⁺NMOSD and an ongoing
5 clinical trial of CD19 CAR-T for refractory stiff person syndrome (NCT06588491).²⁰⁶ By
6 comparison to therapeutic antibodies, CAR-T cells targeting the same proteins may offer
7 greater penetration into immunologically privileged spaces, such as CSF and secondary
8 lymphoid organs, resulting in a deeper, more complete depletion of the B cell lineage.
9 Indeed, frequent occurrence of CAR-T induced immune effector cell-associated
10 neurotoxicity syndrome (ICANS) is considered a significant adverse effect, although may be
11 less frequent outside of haematological cancer settings as substantially fewer targets cells
12 are depleted. Nevertheless, as ICANS associates with CAR-T penetration into the CSF it
13 suggests a mechanism by which CAR-T cells may, somewhat paradoxically, provide
14 enhanced efficacy in CNS diseases.²⁰⁷ Despite this enthusiasm, side effects and off-target
15 effects of CAR-T can be myriad, including neurological sequelae of encephalopathy,
16 seizures, aphasia and Parkinsonism.²⁰⁸

17 Therapeutic monoclonal antibodies against cytokines have been used throughout multiple
18 medical disciplines. Interleukin-6 is a highly pleiotropic cytokine with effects including
19 modification of blood-brain permeability, neuronal and glial functions and T cells and
20 plasmablasts.²⁰⁹ Blockade of the IL-6 pathway reduced relapses in a randomised trial of
21 AQP4⁺NMOSD, and has shown promise in refractory MOGAD and AE.^{209,210} Hence, IL-6-IL-
22 6R pathway blockade provides an example of a broad mechanism of action which confers
23 efficacy across several diseases. Identifying cytokines more specific to NGSAb-diseases
24 may provide more targeted future opportunities.¹¹²

25 Rather than target autoantibody production mechanisms, Fc receptor neonatal (FcRn)
26 blockers, such as efgartigimod and rozanolixizumab, aim to treat disease by blocking
27 antibody uptake into monocytes and/or endothelial cells and hence, increase antibody
28 degradation in circulation.²¹¹ Clinically, they have proven efficacy in refractory myasthenia
29 gravis and in an exploratory cohort of NMDAR-antibody encephalitis, and are the subject of
30 a Phase 3 clinical trial in MOGAD (NCT05063162).²¹² These drugs reduce autoantibodies
31 proportional to total IgG levels, typically inducing ~70% reductions in serum IgG levels.
32 Hence, while they should be effective across all autoantibody-mediated conditions, they
33 also lack a truly precision approach to exclusively targeting the *autoantibodies*.
34 Additionally, reports of infections on FcRn inhibitors, such as enterovirus
35 meningoencephalitis, mandate future vigilance.²¹¹

1 One idea to achieve greater precision is targeting of the CD4⁺-HLA-peptide interaction,
2 particularly when the HLA is homogenous within a disease population (Table 1). A
3 consistent HLA-peptide complex may provide exquisite selectivity without impairing the
4 other physiological functions of that HLA molecule, and may effectively terminate the T cell
5 help required for germinal centre activity.²¹³ This approach should be considered within
6 future T cells studies in NGSAb-diseases. Similarly, through limiting T-B cell interactions,
7 blockade of the CD40-CD40L pathway may turn off germinal centre reactions, and has
8 already shown efficacy in rheumatological diseases.²¹⁴

9 Another precision concept has been to express the autoantigen on T cells, building on the
10 CAR-T approach, this cellular therapy is termed chimeric autoantibody receptor (CAAR)-T
11 cells. The autoantigen expressed on the surface of a CAAR-T should selectively interact
12 with and delete autoantigen-reactive B cells. Preclinical models have been promising in
13 desmoglein-3 associated pemphigus, muscle specific kinase myasthenia and in NMDAR-
14 antibody encephalitis.²¹⁵⁻²¹⁷ However, it is plausible that autoantigen-expressing cells will
15 be overwhelmed by soluble serum autoantibodies, rendering the CAAR-Ts inert. Also, the
16 administration of exogenous human autoantigens may inadvertently re-invigorate the
17 immune response, leading to re-immunisation of the patient against the autoantigen.
18 Clinical studies are awaited to evaluate these theoretical benefits and risks of this
19 theoretically elegant approach.

20 Targeting pathogenic effector functions of autoantibodies

21 To date, some of the most refined precision therapy opportunities stem from targeting the
22 autoantibody-autoantigen interactions and the autoantibody effector functions as these
23 represent a highly discrete process in NGSAb-diseases (Fig.4). An exemplary case of this
24 concept comes from the development of complement inhibitors in NMOSD, which
25 leveraged the importance of the complement pathway observed in preclinical studies^{145,218}
26 to pave the way for the success of the C5 inhibitors eculizumab and ravulizumab.^{152,198} As
27 with most therapeutic monoclonals, only a tiny percentage can be detected in CSF and this
28 concentration appears sufficient to mediate complement inhibition.²¹⁹

29 Analogous to a more precision form of FcRn inhibition, selective degradation of the
30 autoantigen-reactive antibodies may be achieved by creating autoantigen-Fc fusion
31 proteins which spare all IgGs, rather focusing on depleting the autoantigen-specific IgGs.²²⁰
32 This concept, as recently demonstrated as feasible for NMDAR-antibodies,²²¹ would
33 theoretically represent therapeutic opportunities with very few adverse effects which could
34 be applied across all NGSAb-diseases.

1 The autoantibody-autoantigen complex itself may also be a therapeutic target.
2 Aquaporin is a non-complement fixing monoclonal antibody with sufficiently high
3 affinity for the extracellular domain of AQP4 that it prevents binding of endogenous patient
4 AQP4-IgGs and, by not binding C1q, protects from complement dependent cytotoxicity.²²²
5 A similar, yet alternative approach has been taken for the NMDAR using a monovalent
6 engineered antibody which binds with high affinity to the NR1 subunit of the NMDAR, the
7 known immunodominant region of patient IgGs, but without internalising the receptor or
8 inducing electrophysiological effects.²²³ Importantly, its peripheral administration to
9 marmosets appears to reverse patient-IgG induced behavioural deficits. Again, while
10 clinical efficacy is awaited, this concept is of great interest. Yet, it remains to be seen
11 whether it shows sufficient CNS penetration and can out-compete preexisting antibodies
12 formed secondary to an ongoing vigorous downstream cellular immune response.
13 Nevertheless, it may prove valuable in preventing the action of preformed autoantibodies
14 while simultaneously tackling the cellular response with corticosteroids, rituximab and
15 other more traditional approaches. In a similar vein, the use of an allosteric activator of the
16 NMDAR has been shown to have efficacy in a preclinical model of established NMDAR-
17 antibody encephalitis, and can antagonise the effects of patient IgGs on NMDAR
18 membrane organisation.¹⁴⁶ Hence, emerging options to exploit the inherently fastidious
19 interaction between autoantibody and autoantigen are exciting novel approaches to
20 identify precision therapeutics in NGSAb-diseases.

21 Finally, there is increasing interest in exploring how to re-deploy innate surveillance
22 mechanisms to restore tolerance. In this vein, peptide-loaded tolerogenic dendritic cells
23 are an additional autoantigen specific approach which has been proposed in MS and
24 AQP4⁺NMOSD,²²⁴ but could be of interest in other NGSAb-diseases with probable ongoing
25 peripheral immune drivers.

26

27 Missing pieces and future directions

28 The detection of NGSAbs confers a molecularly precise diagnosis, multiple coherent
29 fundamental clinical observations and potential for clinical reversibility, elevating the
30 importance of these conditions. Their discovery has not only revolutionized neurological
31 diagnosis in “not to miss” conditions, but also enabled the development of FDA approved
32 therapies in AQP4⁺NMOSD. In addition, it is likely they will have wider implications for other
33 associated illnesses, for example cancers, as the anti-tumour paraneoplastic response
34 may be protective.^{57,225} However, the recruitment of patients with rare diseases and the
35 interpretation of clinical trials with differing prior immunotherapies and stages of disease

1 will likely prove a major challenge to interpret or apply in real world practice. Hence, a
2 major opportunity presented by these conditions is the development of precision
3 therapies, based on the autoantigenic target. Such autoantigen specific approaches are
4 numerous and would ideally aim to restore immune tolerance, fundamentally creating an
5 exclusive reset of the autoantigen-dedicated immune system and avoiding less specific
6 therapies. The optimal method to reset each of these diseases may differ or show parallels,
7 a future aim to resolve as more research is undertaken into studying their relative
8 immunobiologies. We optimistically anticipate a future akin to molecular diagnostics in
9 oncology, where specific immune findings such as HLA associations, rogue B cell
10 populations and immunodominant epitopes can be defined and exploited in patients for
11 diagnostic, prognostic and therapeutic benefits. These experiences can be applied to the
12 multiple emergent new autoantibodies in neurological and non-neurological diseases,
13 towards the ultimate, and increasingly realistic, aim of cure.

14

15 Acknowledgement

16 The thumbnail image for the online table of contents was created in BioRender. Montini, F.
17 (2026) <https://BioRender.com/ds10p4k>.

18

19 Funding

20 SRI is supported by the Wellcome (104079/Z/14/Z), the Medical Research Council (MRC)
21 (MR/V007173/1), by the National Institute for Health Research (NIHR) Oxford Biomedical
22 Research Centre (BRC) and the Mayo Clinic Robert and Arlene Kogod Center on Aging
23 [Aging Nervous System, 2024]. The views expressed are those of the authors and not
24 necessarily those of the NHS, the NIHR, or the Department of Health. For the purpose of
25 Open Access, the author has applied a CC BY public copyright licence to any Author
26 Accepted Manuscript (AAM) version arising from this submission. The views expressed are
27 those of the author(s) and not necessarily those of the NHS, the NIHR or the Department of
28 Health.

29

1 Competing interests

2 SRI has received honoraria/research support from Amgen, Argenx, Alexion, Atheneum,
3 AstraZeneca, UCB, Roche, Janssen, IQVIA, Clarivate, Slingshot Insights, Slingshot,
4 Cerebral therapeutics, BioHaven therapeutics, CSL Behring, and ONO Pharma. Mayo Clinic
5 Center for Multiple Sclerosis and Autoimmune Neurology, and receives licensed royalties
6 on patent application WO/2010/046716 entitled 'Neurological Autoimmune Disorders', and
7 has filed two other patents entitled "Diagnostic method and therapy" (WO2019211633 and
8 US app 17/051,930; PCT application WO202189788A1) and "Biomarkers"
9 (WO202189788A1, US App 18/279,624; PCT/GB2022/050614). S.J.P. is a named inventor on
10 patent #9,891,219B2, application #12-573,942, methods for treating neuromyelitis optica
11 (NMO) by administration of Eculizumab to an individual that is AQP4-IgG positive, which
12 has been issued and for which he had received royalties. S.J.P. has received grants or
13 research support from Alexion/Astra Zeneca Rare, Horizon/Amgen, F.Hoffmann-La Roche
14 AG, UCB, and Adimune. Since receiving research funding, S.J.P. has received no personal
15 compensation from these companies. In the event of consultation, all compensation for
16 consulting activities is paid directly to Mayo Clinic as per Mayo Clinic policy. S.J.P. has
17 received personal compensation for consultation for Arianys. E.F has served on advisory
18 boards for Alexion, Genentech and Horizon Therapeutics. He has received speaker
19 honoraria from Pharmacy Times. He received royalties from UpToDate. Dr Flanagan was a
20 site primary investigator in a randomized clinical trial on Inebilizumab in neuromyelitis
21 optica spectrum disorder run by Medimmune/Viela-Bio/Horizon Therapeutics. Dr Flanagan
22 has received funding from the NIH (R01NS113828). Dr Flanagan is a member of the
23 medical advisory board of the MOG project. Dr Flanagan is an editorial board member of
24 the Journal of the Neurological Sciences and Neuroimmunology Reports. A patent has
25 been submitted on DACH1-IgG as a biomarker of paraneoplastic autoimmunity.

27 References

- 28 1. Segal Y, Soltys J, Clarkson BDS, Howe CL, Irani SR, Pittock SJ. Toward curing
29 neurological autoimmune disorders: Biomarkers, immunological mechanisms, and therapeutic
30 targets. *Neuron*. Jan 8 2025;doi:10.1016/j.neuron.2024.12.006
- 31 2. Sun B, Ramberger M, O'Connor KC, Bashford-Rogers RJM, Irani SR. The B cell
32 immunobiology that underlies CNS autoantibody-mediated diseases. *Nat Rev Neurol*. Sep
33 2020;16(9):481-492. doi:10.1038/s41582-020-0381-z

- 1 3. de Bruijn M, Leypoldt F, Dalmau J, *et al.* Autoimmune encephalitis. *Nat Rev Dis Primers*.
2 Sep 11 2025;11(1):65. doi:10.1038/s41572-025-00650-1
- 3 4. Varley JA, Strippel C, Handel A, Irani SR. Autoimmune encephalitis: recent clinical and
4 biological advances. *J Neurol*. Aug 2023;270(8):4118-4131. doi:10.1007/s00415-023-11685-3
- 5 5. Irani SR. Autoimmune Encephalitis. *Continuum (Minneap Minn)*. Aug 1 2024;30(4):995-
6 1020. doi:10.1212/CON.0000000000001448
- 7 6. Pittock SJ, Lucchinetti CF. Neuromyelitis optica and the evolving spectrum of autoimmune
8 aquaporin-4 channelopathies: a decade later. *Ann N Y Acad Sci*. Feb 2016;1366(1):20-39.
9 doi:10.1111/nyas.12794
- 10 7. Banwell B, Bennett JL, Marignier R, *et al.* Diagnosis of myelin oligodendrocyte
11 glycoprotein antibody-associated disease: International MOGAD Panel proposed criteria. *Lancet*
12 *Neurol*. Mar 2023;22(3):268-282. doi:10.1016/S1474-4422(22)00431-8
- 13 8. Meffre E. The establishment of early B cell tolerance in humans: lessons from primary
14 immunodeficiency diseases. *Ann N Y Acad Sci*. Dec 2011;1246:1-10. doi:10.1111/j.1749-
15 6632.2011.06347.x
- 16 9. Montini F, Weiner HL. The Innate Immune System in CNS Diseases; Overview. 2024;
- 17 10. Binks SNM, Klein CJ, Waters P, Pittock SJ, Irani SR. LGI1, CASPR2 and related
18 antibodies: a molecular evolution of the phenotypes. *J Neurol Neurosurg Psychiatry*. May
19 2018;89(5):526-534. doi:10.1136/jnnp-2017-315720
- 20 11. Orozco E, Valencia-Sanchez C, Britton J, *et al.* Autoimmune Encephalitis Criteria in
21 Clinical Practice. *Neurol Clin Pract*. Jun 2023;13(3):e200151.
22 doi:10.1212/cpj.000000000000200151
- 23 12. Kunchok A, McKeon A, Zekeridou A, *et al.* Autoimmune/Paraneoplastic Encephalitis
24 Antibody Biomarkers: Frequency, Age, and Sex Associations. *Mayo Clin Proc*. Mar
25 2022;97(3):547-559. doi:10.1016/j.mayocp.2021.07.023
- 26 13. Bennett JL, Lam C, Kalluri SR, *et al.* Intrathecal pathogenic anti-aquaporin-4 antibodies
27 in early neuromyelitis optica. *Ann Neurol*. Nov 2009;66(5):617-29. doi:10.1002/ana.21802

- 1 14. Kowarik MC, Dzieciatkowska M, Wemlinger S, *et al.* The cerebrospinal fluid
2 immunoglobulin transcriptome and proteome in neuromyelitis optica reveals central nervous
3 system-specific B cell populations. *J Neuroinflammation*. Jan 28 2015;12:19. doi:10.1186/s12974-
4 015-0240-9
- 5 15. Theorell J, Harrison R, Williams R, *et al.* Ultrahigh frequencies of peripherally matured
6 LGI1- and CASPR2-reactive B cells characterize the cerebrospinal fluid in autoimmune
7 encephalitis. *Proc Natl Acad Sci U S A*. Feb 13 2024;121(7):e2311049121.
8 doi:10.1073/pnas.2311049121
- 9 16. Irani SR, Alexander S, Waters P, *et al.* Antibodies to Kv1 potassium channel-complex
10 proteins leucine-rich, glioma inactivated 1 protein and contactin-associated protein-2 in limbic
11 encephalitis, Morvan's syndrome and acquired neuromyotonia. *Brain*. 2010;133(9):2734-2748.
12 doi:10.1093/brain/awq213
- 13 17. Sabater L, Gaig C, Gelpi E, *et al.* A novel non-rapid-eye movement and rapid-eye-
14 movement parasomnia with sleep breathing disorder associated with antibodies to IgLON5: a case
15 series, characterisation of the antigen, and post-mortem study. *Lancet Neurol*. Jun 2014;13(6):575-
16 86. doi:10.1016/S1474-4422(14)70051-1
- 17 18. Dalmau J, Tuzun E, Wu HY, *et al.* Paraneoplastic anti-N-methyl-D-aspartate receptor
18 encephalitis associated with ovarian teratoma. *Ann Neurol*. Jan 2007;61(1):25-36.
19 doi:10.1002/ana.21050
- 20 19. Irani SR, Bera K, Waters P, *et al.* N-methyl-D-aspartate antibody encephalitis: temporal
21 progression of clinical and paraclinical observations in a predominantly non-paraneoplastic
22 disorder of both sexes. *Brain*. Jun 2010;133(Pt 6):1655-67. doi:10.1093/brain/awq113
- 23 20. Bastiaansen AEM, de Bruijn M, Schuller SL, *et al.* Anti-NMDAR Encephalitis in the
24 Netherlands, Focusing on Late-Onset Patients and Antibody Test Accuracy. *Neurol*
25 *Neuroimmunol Neuroinflamm*. Mar 2022;9(2)doi:10.1212/NXI.0000000000001127
- 26 21. Hor JY, Fujihara K. Epidemiology of myelin oligodendrocyte glycoprotein antibody-
27 associated disease: a review of prevalence and incidence worldwide. *Frontiers in Neurology*.
28 2023;14doi:10.3389/fneur.2023.1260358

- 1 22. Ramanathan S, Brilot F, Irani SR, Dale RC. Origins and immunopathogenesis of
2 autoimmune central nervous system disorders. *Nat Rev Neurol*. Mar 2023;19(3):172-190.
3 doi:10.1038/s41582-023-00776-4
- 4 23. Vorasoot N, Kunchok A, Edmond MC, *et al*. Age and Sex Distributions of MOG-IgG and
5 AQP4-IgG Positive Sera from a Large Neuroimmunology Laboratory Database. *Neurology*. Apr
6 25 2023;100(17)doi:10.1212/Wnl.0000000000204098
- 7 24. Kim TJ, Lee ST, Moon J, *et al*. Anti-LGI1 encephalitis is associated with unique HLA
8 subtypes. *Ann Neurol*. Feb 2017;81(2):183-192. doi:10.1002/ana.24860
- 9 25. van Sonderen A, Roelen DL, Stoop JA, *et al*. Anti-LGI1 encephalitis is strongly associated
10 with HLA-DR7 and HLA-DRB4. *Ann Neurol*. Feb 2017;81(2):193-198. doi:10.1002/ana.24858
- 11 26. Binks S, Varley J, Lee W, *et al*. Distinct HLA associations of LGI1 and CASPR2-antibody
12 diseases. *Brain*. Aug 1 2018;141(8):2263-2271. doi:10.1093/brain/awy109
- 13 27. Muniz-Castrillo S, Joubert B, Elsensohn MH, *et al*. Anti-CASPR2 clinical phenotypes
14 correlate with HLA and immunological features. *J Neurol Neurosurg Psychiatry*. Oct
15 2020;91(10):1076-1084. doi:10.1136/jnnp-2020-323226
- 16 28. Yogeshwar SM, Muniz-Castrillo S, Sabater L, *et al*. HLA-DQB1*05 subtypes and not
17 DRB1*10:01 mediates risk in anti-IgLON5 disease. *Brain*. Mar 1
18 2024;doi:10.1093/brain/awae048
- 19 29. Hammer J, Sturniolo T, Sinigaglia F. HLA class II peptide binding specificity and
20 autoimmunity. *Adv Immunol*. 1997;66:67-100. doi:10.1016/s0065-2776(08)60596-9
- 21 30. Victora GD, Nussenzweig MC. Germinal Centers. *Annu Rev Immunol*. Apr 26
22 2022;40:413-442. doi:10.1146/annurev-immunol-120419-022408
- 23 31. Binks SNM, Elliott KS, Muniz-Castrillo S, *et al*. Novel risk loci in LGI1-antibody
24 encephalitis: genome-wide association study discovery and validation cohorts. *Brain*. Oct 26
25 2024;doi:10.1093/brain/awae349
- 26 32. Moseley N, King J, Van Dort B, *et al*. Anti-voltage-Gated Potassium Channel (VGKC)
27 Antibodies and Acquired Neuromyotonia in Patients with Immune Dysregulation,

- 1 Polyendocrinopathy, Enteropathy X-Linked (IPEX) Syndrome. *J Clin Immunol.* Nov
2 2021;41(8):1972-1974. doi:10.1007/s10875-021-01128-x
- 3 33. Peris Sempere V, Luo G, Muñoz-Castrillo S, *et al.* HLA and KIR genetic association and
4 NK cells in anti-NMDAR encephalitis. *Front Immunol.* 2024;15:1423149.
5 doi:10.3389/fimmu.2024.1423149
- 6 34. Liu X, Zheng X, Shu Y, *et al.* Genome-Wide Association Study Identifies IFIH1 and HLA-
7 DQB1*05:02 Loci Associated With Anti-NMDAR Encephalitis. *Neurology Neuroimmunology*
8 *& Neuroinflammation.* 2024;11(3):e200221. doi:doi:10.1212/NXI.0000000000200221
- 9 35. Hyun JW, Kim S, Moon J, *et al.* HLA Association With AQP4-IgG-Positive Neuromyelitis
10 Optica Spectrum Disorder in the Korean Population. *Neurol Neuroimmunol Neuroinflamm.* May
11 2025;12(3):e200366. doi:10.1212/nxi.0000000000200366
- 12 36. Estrada K, Whelan CW, Zhao F, *et al.* A whole-genome sequence study identifies genetic
13 risk factors for neuromyelitis optica. *Nat Commun.* May 16 2018;9(1):1929. doi:10.1038/s41467-
14 018-04332-3
- 15 37. Ghafouri-Fard S, Azimi T, Taheri M. A Comprehensive Review on the Role of Genetic
16 Factors in Neuromyelitis Optica Spectrum Disorder. *Front Immunol.* 2021;12:737673.
17 doi:10.3389/fimmu.2021.737673
- 18 38. Pittock SJ, Berthele A, Fujihara K, *et al.* Eculizumab in Aquaporin-4-Positive
19 Neuromyelitis Optica Spectrum Disorder. *New England Journal of Medicine.* 2019;381(7):614-
20 625. doi:doi:10.1056/NEJMoa1900866
- 21 39. Blackburn KM, Wang C. Post-infectious neurological disorders. *Therapeutic Advances in*
22 *Neurological Disorders.* 2020-8-30 2020;13:1756286420952901.
23 doi:10.1177/1756286420952901
- 24 40. Armangué T, Olivé-Cirera G, Martínez-Hernandez E, *et al.* Neurologic complications in
25 herpes simplex encephalitis: clinical, immunological and genetic studies. *Brain.* Oct 3
26 2023;146(10):4306-4319. doi:10.1093/brain/awad238
- 27 41. Armangué T, Spatola M, Vlăgea A, *et al.* Frequency, symptoms, risk factors, and outcomes
28 of autoimmune encephalitis after herpes simplex encephalitis: a prospective observational study

- 1 and retrospective analysis. *The Lancet Neurology*. 2018-09 2018;17(9):760-772.
2 doi:10.1016/S1474-4422(18)30244-8
- 3 42. Pruss H, Finke C, Holtje M, *et al*. N-methyl-D-aspartate receptor antibodies in herpes
4 simplex encephalitis. *Ann Neurol*. Dec 2012;72(6):902-11. doi:10.1002/ana.23689
- 5 43. Hacoen Y, Deiva K, Pettingill P, *et al*. N-methyl-D-aspartate receptor antibodies in post-
6 herpes simplex virus encephalitis neurological relapse. *Mov Disord*. Jan 2014;29(1):90-6.
7 doi:10.1002/mds.25626
- 8 44. Nibber A, Wills B, Pettingill P, *et al*. Evolution and significance of neuronal surface
9 autoantibodies after Japanese encephalitis. *J Neuroimmunol*. Sep 15 2025;406:578671.
10 doi:10.1016/j.jneuroim.2025.578671
- 11 45. Luo H, Ding X, Li Y, *et al*. Clinical Characteristics of Children With Anti-N-Methyl-d-
12 Aspartate Receptor Encephalitis After Japanese Encephalitis. *Pediatr Neurol*. May 2022;130:46-
13 52. doi:10.1016/j.pediatrneurol.2022.02.004
- 14 46. Smyth LCD, Kipnis J. Redefining CNS immune privilege. *Nature Reviews Immunology*.
15 2025;doi:10.1038/s41577-025-01175-0
- 16 47. Cleaver J, Jeffery K, Klenerman P, *et al*. The immunobiology of herpes simplex virus
17 encephalitis and post-viral autoimmunity. *Brain*. Apr 4 2024;147(4):1130-1148.
18 doi:10.1093/brain/awad419
- 19 48. Armangue T, Spatola M, Vlagea A, *et al*. Frequency, symptoms, risk factors, and outcomes
20 of autoimmune encephalitis after herpes simplex encephalitis: a prospective observational study
21 and retrospective analysis. *Lancet Neurol*. Sep 2018;17(9):760-772. doi:10.1016/S1474-
22 4422(18)30244-8
- 23 49. Armangue T, Olive-Cirera G, Martinez-Hernandez E, *et al*. Neurologic complications in
24 herpes simplex encephalitis: clinical, immunological and genetic studies. *Brain*. Oct 3
25 2023;146(10):4306-4319. doi:10.1093/brain/awad238
- 26 50. Sun B, Fernandes D, Soltys J, *et al*. Permissive central tolerance plus defective peripheral
27 checkpoints license pathogenic memory B cells in CASPR2-antibody encephalitis. *Sci Adv*. Apr
28 18 2025;11(16):eadr9986. doi:10.1126/sciadv.adr9986

- 1 51. Kleeman SO, Michalski K, Zhao X, *et al.* Ectopic NMDAR expression in cancer unmasks
2 germline-encoded autoimmunity. *Nature*. Mar 25 2026;doi:10.1038/s41586-026-10278-0
- 3 52. Linnoila JJ, Binnicker MJ, Majed M, Klein CJ, McKeon A. CSF herpes virus and
4 autoantibody profiles in the evaluation of encephalitis. *Neurology(R) Neuroimmunology &*
5 *Neuroinflammation*. 2016-08 2016;3(4):e245. doi:10.1212/NXI.0000000000000245
- 6 53. Prüss H. Postviral autoimmune encephalitis: manifestations in children and adults. *Current*
7 *Opinion in Neurology*. 2017-06 2017;30(3):327-333. doi:10.1097/WCO.0000000000000445
- 8 54. Armangue T, Leypoldt F, Málaga I, *et al.* Herpes Simplex Virus Encephalitis is a Trigger
9 of Brain Autoimmunity. *Annals of neurology*. 2014-2 2014;75(2):317-323. doi:10.1002/ana.24083
- 10 55. Izzy S, Yahya T, Albastaki O, *et al.* Nasal anti-CD3 monoclonal antibody ameliorates
11 traumatic brain injury, enhances microglial phagocytosis and reduces neuroinflammation via IL-
12 10-dependent T(reg)-microglia crosstalk. *Nat Neurosci*. Mar 2025;28(3):499-516.
13 doi:10.1038/s41593-025-01877-7
- 14 56. Salovin A, Glanzman J, Roslin K, Armangue T, Lynch DR, Panzer JA. Anti-NMDA
15 receptor encephalitis and nonencephalitic HSV-1 infection. *Neurology® Neuroimmunology &*
16 *Neuroinflammation*. 2018-4-05 2018;5(4):e458. doi:10.1212/NXI.0000000000000458
- 17 57. Graus F, Vogrig A, Muniz-Castrillo S, *et al.* Updated Diagnostic Criteria for Paraneoplastic
18 Neurologic Syndromes. *Neurol Neuroimmunol Neuroinflamm*. Jul
19 2021;8(4)doi:10.1212/NXI.0000000000001014
- 20 58. Binks S, Uy C, Honnorat J, Irani SR. Paraneoplastic neurological syndromes: a practical
21 approach to diagnosis and management. *Pract Neurol*. Feb 2022;22(1):19-31.
22 doi:10.1136/practneurol-2021-003073
- 23 59. Dalmau J, Rosenfeld MR. Paraneoplastic syndromes of the CNS. *Lancet Neurol*. Apr
24 2008;7(4):327-40. doi:10.1016/S1474-4422(08)70060-7
- 25 60. Albert ML, Darnell RB. Paraneoplastic neurological degenerations: keys to tumour
26 immunity. *Nat Rev Cancer*. Jan 2004;4(1):36-44. doi:10.1038/nrc1255
- 27 61. Hetzel DJ, Stanhope CR, O'Neill BP, Lennon VA. Gynecologic cancer in patients with
28 subacute cerebellar degeneration predicted by anti-Purkinje cell antibodies and limited in

- 1 metastatic volume. *Mayo Clin Proc.* Dec 1990;65(12):1558-63. doi:10.1016/s0025-
2 6196(12)62189-2
- 3 62. Ely ZA, Kulstad ZJ, Gunaydin G, *et al.* Pancreatic cancer–restricted cryptic antigens are
4 targets for T cell recognition. *Science.* 2025;388(6747)doi:10.1126/science.adk3487
- 5 63. Small M, Treilleux I, Couillault C, *et al.* Genetic alterations and tumor immune attack in
6 Yo paraneoplastic cerebellar degeneration. *Acta Neuropathologica.* 2018/04/01 2018;135(4):569-
7 579. doi:10.1007/s00401-017-1802-y
- 8 64. Herdlevaer I, Kråkenes T, Schubert M, Vedeler CA. Localization of CDR2L and CDR2 in
9 paraneoplastic cerebellar degeneration. *Ann Clin Transl Neurol.* Nov 2020;7(11):2231-2242.
10 doi:10.1002/acn3.51212
- 11 65. Jiang X-Y, Lei S, Zhang L, *et al.* Co-expression of NMDA–receptor subunits NR1, NR2A,
12 and NR2B in dysplastic neurons of teratomas in patients with paraneoplastic NMDA-receptor-
13 encephalitis: a retrospective clinico-pathology study of 159 patients. *Acta Neuropathologica*
14 *Communications.* 2020-08-08 2020;8(1):130. doi:10.1186/s40478-020-00999-2
- 15 66. Vogrig A, Pegat A, Villagrán-García M, *et al.* Different Genetic Signatures of Small-Cell
16 Lung Cancer Characterize Anti-GABAB R and Anti-Hu Paraneoplastic Neurological Syndromes.
17 *Annals of Neurology.* 2023-12 2023;94(6):1102-1115. doi:10.1002/ana.26784
- 18 67. Darnell JC, Albert ML, Darnell RB. Cdr2, a target antigen of naturally occurring human
19 tumor immunity, is widely expressed in gynecological tumors. *Cancer Research.* 2000-04-15
20 2000;60(8):2136-2139.
- 21 68. Raspotnig M, Haugen M, Thorsteinsdottir M, *et al.* Cerebellar degeneration-related
22 proteins 2 and 2-like are present in ovarian cancer in patients with and without Yo antibodies.
23 *Cancer immunology, immunotherapy: CII.* 2017-11 2017;66(11):1463-1471. doi:10.1007/s00262-
24 017-2041-8
- 25 69. Rosenberg MI, Greenstein E, Buchkovich M, *et al.* Polyclonal lymphoid expansion drives
26 paraneoplastic autoimmunity in neuroblastoma. *Cell Rep.* Aug 29 2023;42(8):112879.
27 doi:10.1016/j.celrep.2023.112879

- 1 70. Makuch M, Wilson R, Al-Diwani A, *et al.* N-methyl-D-aspartate receptor antibody
2 production from germinal center reactions: Therapeutic implications. *Annals of Neurology*. 2018-
3 03 2018;83(3):553-561. doi:10.1002/ana.25173
- 4 71. Farina A, Villagran-Garcia M, Vogrig A, *et al.* Neurological adverse events of immune
5 checkpoint inhibitors and the development of paraneoplastic neurological syndromes. *Lancet*
6 *Neurol*. Jan 2024;23(1):81-94. doi:10.1016/S1474-4422(23)00369-1
- 7 72. Das R, Bar N, Ferreira M, *et al.* Early B cell changes predict autoimmunity following
8 combination immune checkpoint blockade. *J Clin Invest*. Feb 1 2018;128(2):715-720.
9 doi:10.1172/JCI96798
- 10 73. Willis MD, Schroeder B, Marandino L, Turajlic S, Carr AS. Neurological immune-related
11 adverse events with checkpoint inhibitor therapy: challenges for the neurologist. *Journal of*
12 *Neurology, Neurosurgery & Psychiatry*. 2025;96(11):1024. doi:10.1136/jnnp-2025-
13 335998
- 14 74. Wilson R, Menassa DA, Davies AJ, *et al.* Seronegative antibody-mediated neurology after
15 immune checkpoint inhibitors. *Ann Clin Transl Neurol*. May 2018;5(5):640-645.
16 doi:10.1002/acn3.547
- 17 75. Cohen DA, Lopez-Chiriboga AS, Pittock SJ, *et al.* Posttransplant autoimmune
18 encephalitis. *Neurol Neuroimmunol Neuroinflamm*. Nov 2018;5(6):e497.
19 doi:10.1212/NXI.0000000000000497
- 20 76. Schwerdtfeger LA, Montini F, Lanser TB, *et al.* Gut microbiota and metabolites are linked
21 to disease progression in multiple sclerosis. *Cell Rep Med*. Apr 15 2025;6(4):102055.
22 doi:10.1016/j.xcrm.2025.102055
- 23 77. Antonini Cencicchio M, Montini F, Palmieri V, *et al.* Microbiota-produced immune
24 regulatory bile acid metabolites control central nervous system autoimmunity. *Cell Rep Med*. Apr
25 15 2025;6(4):102028. doi:10.1016/j.xcrm.2025.102028
- 26 78. Martinelli V, Albanese M, Altieri M, *et al.* Gut-oriented interventions in patients with
27 multiple sclerosis: fact or fiction? *Eur Rev Med Pharmacol Sci*. Feb 2022;26(3):935-946.
28 doi:10.26355/eurev_202202_28003

- 1 79. Schwerdtfeger LA, Montini F, Chitnis T, Cox LM, Weiner HL. Faecal mucoprotein MUC2
2 is decreased in multiple sclerosis and is associated with mucin degrading bacteria. *EBioMedicine*.
3 Jun 2025;116:105721. doi:10.1016/j.ebiom.2025.105721
- 4 80. Li S, Montini F, Song A, *et al*. Alterations of the nasal and oral microbiota in multiple
5 sclerosis. *eBioMedicine*. 2025;121doi:10.1016/j.ebiom.2025.105959
- 6 81. Varrin-Doyer M, Spencer CM, Schulze-Topphoff U, *et al*. Aquaporin 4-specific T cells in
7 neuromyelitis optica exhibit a Th17 bias and recognize Clostridium ABC transporter. *Ann Neurol*.
8 Jul 2012;72(1):53-64. doi:10.1002/ana.23651
- 9 82. Cree BA, Spencer CM, Varrin-Doyer M, Baranzini SE, Zamvil SS. Gut microbiome
10 analysis in neuromyelitis optica reveals overabundance of Clostridium perfringens. *Ann Neurol*.
11 Sep 2016;80(3):443-7. doi:10.1002/ana.24718
- 12 83. Wei J, Zhang X, Yang F, *et al*. Gut microbiome changes in anti-N-methyl-D-aspartate
13 receptor encephalitis patients. *BMC Neurol*. Jul 25 2022;22(1):276. doi:10.1186/s12883-022-
14 02804-0
- 15 84. Gilbert E, Binks S, Damato V, *et al*. The gut microbiome associated with LGI1-antibody
16 encephalitis. *Epilepsia*. Aug 6 2025;doi:10.1111/epi.18556
- 17 85. Leite MI, Coutinho E, Lana-Peixoto M, *et al*. Myasthenia gravis and neuromyelitis optica
18 spectrum disorder: a multicenter study of 16 patients. *Neurology*. May 15 2012;78(20):1601-7.
19 doi:10.1212/WNL.0b013e31825644ff
- 20 86. McNaughton P, Payne R, Michael S, *et al*. Naive B cells followed by aquaporin-4
21 antibodies characterise the onset of neuromyelitis optica: evidence from stem cell transplantation.
22 *J Neurol Neurosurg Psychiatry*. May 23 2022;93(11):1234-6. doi:10.1136/jnnp-2022-328982
- 23 87. Sun B, Ramberger M, O'Connor KC, Bashford-Rogers RJM, Irani SR. The B cell
24 immunobiology that underlies CNS autoantibody-mediated diseases. *Nature Reviews Neurology*.
25 2020/09/01 2020;16(9):481-492. doi:10.1038/s41582-020-0381-z
- 26 88. Nemazee D. Mechanisms of central tolerance for B cells. *Nat Rev Immunol*. May
27 2017;17(5):281-294. doi:10.1038/nri.2017.19

- 1 89. Pelanda R, Torres RM. Central B-cell tolerance: where selection begins. *Cold Spring Harb*
2 *Perspect Biol.* Apr 1 2012;4(4):a007146. doi:10.1101/cshperspect.a007146
- 3 90. Meffre E, O'Connor KC. Impaired B-cell tolerance checkpoints promote the development
4 of autoimmune diseases and pathogenic autoantibodies. *Immunol Rev.* Nov 2019;292(1):90-101.
5 doi:10.1111/imr.12821
- 6 91. Tiller T, Tsuiji M, Yurasov S, Velinzon K, Nussenzweig MC, Wardemann H.
7 Autoreactivity in human IgG+ memory B cells. *Immunity.* Feb 2007;26(2):205-13.
8 doi:10.1016/j.immuni.2007.01.009
- 9 92. Tsuiji M, Yurasov S, Velinzon K, Thomas S, Nussenzweig MC, Wardemann H. A
10 checkpoint for autoreactivity in human IgM+ memory B cell development. *J Exp Med.* Feb 20
11 2006;203(2):393-400. doi:10.1084/jem.20052033
- 12 93. Wilson R, Makuch M, Kienzler AK, *et al.* Condition-dependent generation of aquaporin-
13 4 antibodies from circulating B cells in neuromyelitis optica. *Brain.* Apr 1 2018;141(4):1063-1074.
14 doi:10.1093/brain/awy010
- 15 94. Wenke NK, Kreye J, Andrzejak E, *et al.* N-methyl-D-aspartate receptor dysfunction by
16 unmutated human antibodies against the NR1 subunit. *Ann Neurol.* May 2019;85(5):771-776.
17 doi:10.1002/ana.25460
- 18 95. Wetzel NS, Kulsvehagen L, Lecourt AC, *et al.* Patient-Derived Monoclonal Myelin
19 Oligodendrocyte Glycoprotein Autoantibodies Mediate Cytotoxicity. *Neurol Neuroimmunol*
20 *Neuroinflamm.* Jan 2026;13(1):e200520. doi:10.1212/NXI.0000000000200520
- 21 96. Cotzomi E, Stathopoulos P, Lee CS, *et al.* Early B cell tolerance defects in neuromyelitis
22 optica favour anti-AQP4 autoantibody production. *Brain.* Jun 1 2019;142(6):1598-1615.
23 doi:10.1093/brain/awz106
- 24 97. Di Zenzo G, Di Lullo G, Corti D, *et al.* Pemphigus autoantibodies generated through
25 somatic mutations target the desmoglein-3 cis-interface. *J Clin Invest.* Oct 2012;122(10):3781-90.
26 doi:10.1172/JCI64413
- 27 98. Piccoli L, Campo I, Fregni CS, *et al.* Neutralization and clearance of GM-CSF by
28 autoantibodies in pulmonary alveolar proteinosis. *Nat Commun.* Jun 16 2015;6:7375.
29 doi:10.1038/ncomms8375

- 1 99. Dhanik A, R. Kirshner J, Macdonald D, *et al.* In-silico discovery of cancer-specific
2 peptide-HLA complexes for targeted therapy. *BMC Bioinformatics*.
3 2016;17(1)doi:10.1186/s12859-016-1150-2
- 4 100. Saggau C, Bacher P, Esser D, *et al.* Autoantigen-specific CD4(+) T cells acquire an
5 exhausted phenotype and persist in human antigen-specific autoimmune diseases. *Immunity*. Aug
6 26 2024;doi:10.1016/j.immuni.2024.08.005
- 7 101. Ono H, Misu T, Namatame C, *et al.* CD4-Positive T-Cell Responses to MOG Peptides in
8 MOG Antibody-Associated Disease. *International Journal of Molecular Sciences*.
9 2025;26(8):3606. doi:10.3390/ijms26083606
- 10 102. Dao LM, Machule ML, Bacher P, *et al.* Decreased inflammatory cytokine production of
11 antigen-specific CD4(+) T cells in NMDA receptor encephalitis. *J Neurol*. Jun 2021;268(6):2123-
12 2131. doi:10.1007/s00415-020-10371-y
- 13 103. Da Mesquita S, Fu Z, Kipnis J. The Meningeal Lymphatic System: A New Player in
14 Neurophysiology. *Neuron*. Oct 24 2018;100(2):375-388. doi:10.1016/j.neuron.2018.09.022
- 15 104. Kipnis J. The anatomy of brainwashing. *Science*. Jul 26 2024;385(6707):368-370.
16 doi:10.1126/science.adp1705
- 17 105. Al-Diwani A, Theorell J, Damato V, *et al.* Cervical lymph nodes and ovarian teratomas as
18 germinal centres in NMDA receptor-antibody encephalitis. *Brain*. Aug 27 2022;145(8):2742-
19 2754. doi:10.1093/brain/awac088
- 20 106. Damato V, Theorell J, Al-Diwani A, *et al.* Rituximab abrogates aquaporin-4-specific
21 germinal center activity in patients with neuromyelitis optica spectrum disorders. *Proc Natl Acad*
22 *Sci U S A*. Jun 14 2022;119(24):e2121804119. doi:10.1073/pnas.2121804119
- 23 107. Al-Diwani A, Provine NM, Murchison A, *et al.* Neurodegenerative fluid biomarkers are
24 enriched in human cervical lymph nodes. *Brain*. Feb 3 2025;148(2):394-400.
25 doi:10.1093/brain/awae329
- 26 108. Provine NM, Al-Diwani A, Agarwal D, *et al.* Fine needle aspiration of human lymph nodes
27 reveals cell populations and soluble interactors pivotal to immunological priming. *Eur J Immunol*.
28 May 2024;54(5):e2350872. doi:10.1002/eji.202350872

- 1 109. Roco JA, Mesin L, Binder SC, *et al.* Class-Switch Recombination Occurs Infrequently in
2 Germinal Centers. *Immunity*. Aug 20 2019;51(2):337-350 e7. doi:10.1016/j.immuni.2019.07.001
- 3 110. Horns F, Vollmers C, Croote D, *et al.* Lineage tracing of human B cells reveals the in vivo
4 landscape of human antibody class switching. *Elife*. Aug 2 2016;5doi:10.7554/eLife.16578
- 5 111. Hwang JK, Alt FW, Yeap LS. Related Mechanisms of Antibody Somatic Hypermutation
6 and Class Switch Recombination. *Microbiol Spectr*. Feb 2015;3(1):MDNA3-0037-2014.
7 doi:10.1128/microbiolspec.MDNA3-0037-2014
- 8 112. Aboseif A, Mangioris G, Yang B, *et al.* Cytokine, Chemokine, and Neurofilament Light
9 Chain Signatures in LGI1 Autoimmune Encephalitis. *Ann Clin Transl Neurol*. Nov
10 2025;12(11):2258-2270. doi:10.1002/acn3.70158
- 11 113. Desestret V, Chefdeville A, Viaccoz A, *et al.* CSF IgA NMDAR antibodies are potential
12 biomarkers for teratomas in anti-NMDAR encephalitis. *Neurol Neuroimmunol Neuroinflamm*.
13 Dec 2015;2(6):e166. doi:10.1212/NXI.0000000000000166
- 14 114. Ceronie B, Strippel C, Uy C, *et al.* Immunotherapy-Resistant Neuropathic Pain and Fatigue
15 Predict Quality-of-Life in Contactin-Associated Protein-Like 2 Antibody Disease. *Ann Neurol*. Jan
16 18 2025;doi:10.1002/ana.27177
- 17 115. Jitrapaikulsan J, Fryer JP, Majed M, *et al.* Clinical utility of AQP4-IgG titers and
18 measures of complement-mediated cell killing in NMOSD. *Neurol Neuroimmunol Neuroinflamm*.
19 Jul 2020;7(4)doi:10.1212/NXI.0000000000000727
- 20 116. Ciano-Petersen NL, Robert M, Muniz-Castrillo S, *et al.* Prognostic Value of Persistent CSF
21 Antibodies at 12 Months in Anti-NMDAR Encephalitis. *Neurol Neuroimmunol Neuroinflamm*. Jul
22 2023;10(4)doi:10.1212/NXI.0000000000200108
- 23 117. de Carvalho RVH, Ersching J, Barbulescu A, *et al.* Clonal replacement sustains long-lived
24 germinal centers primed by respiratory viruses. *Cell*. Jan 5 2023;186(1):131-146 e13.
25 doi:10.1016/j.cell.2022.11.031
- 26 118. Tellier J, Nutt SL. Plasma cells: The programming of an antibody-secreting machine. *Eur*
27 *J Immunol*. Jan 2019;49(1):30-37. doi:10.1002/eji.201847517

- 1 119. Liu L, Zhang X, Chai Y, Zhang J, Deng Q, Chen X. Skull bone marrow and skull meninges
2 channels: redefining the landscape of central nervous system immune surveillance. *Cell Death &*
3 *Disease*. 2025/01/29 2025;16(1):53. doi:10.1038/s41419-025-07336-2
- 4 120. Turner JS, Kim W, Kalaidina E, *et al*. SARS-CoV-2 infection induces long-lived bone
5 marrow plasma cells in humans. *Nature*. Jul 2021;595(7867):421-425. doi:10.1038/s41586-021-
6 03647-4
- 7 121. Burt RK, Balabanov R, Han X, *et al*. Autologous nonmyeloablative hematopoietic stem
8 cell transplantation for neuromyelitis optica. *Neurology*. Oct 29 2019;93(18):e1732-e1741.
9 doi:10.1212/WNL.0000000000008394
- 10 122. Vorasoot N, Blackburn KM, Nguyen L, *et al*. Late Relapse After Autologous
11 Hematopoietic Stem Cell Transplantation in AQP4-IgG-Positive NMOSD. *JAMA Netw Open*. Apr
12 1 2025;8(4):e255989. doi:10.1001/jamanetworkopen.2025.5989
- 13 123. Qin C, Tian DS, Zhou LQ, *et al*. Anti-BCMA CAR T-cell therapy CT103A in relapsed or
14 refractory AQP4-IgG seropositive neuromyelitis optica spectrum disorders: phase 1 trial interim
15 results. *Signal Transduct Target Ther*. Jan 4 2023;8(1):5. doi:10.1038/s41392-022-01278-3
- 16 124. Kelly MJ, Grant E, Murchison AG, *et al*. Magnetic Resonance Imaging Characteristics of
17 LGI1-Antibody and CASPR2-Antibody Encephalitis. *JAMA Neurol*. Mar 18
18 2024;doi:10.1001/jamaneurol.2024.0126
- 19 125. Kreye J, Wenke NK, Chayka M, *et al*. Human cerebrospinal fluid monoclonal N-methyl-
20 D-aspartate receptor autoantibodies are sufficient for encephalitis pathogenesis. *Brain*. Oct
21 2016;139(Pt 10):2641-2652. doi:10.1093/brain/aww208
- 22 126. Leypoldt F, Hoftberger R, Titulaer MJ, *et al*. Investigations on CXCL13 in anti-N-methyl-
23 D-aspartate receptor encephalitis: a potential biomarker of treatment response. *JAMA Neurol*. Feb
24 2015;72(2):180-6. doi:10.1001/jamaneurol.2014.2956
- 25 127. Martin-de-Saavedra MD, Dos Santos M, Culotta L, *et al*. Shed CNTNAP2 ectodomain is
26 detectable in CSF and regulates Ca(2+) homeostasis and network synchrony via PMCA2/ATP2B2.
27 *Neuron*. Feb 16 2022;110(4):627-643 e9. doi:10.1016/j.neuron.2021.11.025

- 1 128. Ramberger M, Berretta A, Tan JMM, *et al.* Distinctive binding properties of human
2 monoclonal LGI1 autoantibodies determine pathogenic mechanisms. *Brain*. Jun 1
3 2020;143(6):1731-1745. doi:10.1093/brain/awaa104
- 4 129. Landa J, Serafim AB, Gaig C, *et al.* Patients' IgLON5 autoantibodies interfere with
5 IgLON5-protein interactions. *Frontiers in Immunology*.
6 2023;14doi:10.3389/fimmu.2023.1151574
- 7 130. Peschl P, Bradl M, Höftberger R, Berger T, Reindl M. Myelin Oligodendrocyte
8 Glycoprotein: Deciphering a Target in Inflammatory Demyelinating Diseases. *Frontiers in*
9 *Immunology*. 2017;8doi:10.3389/fimmu.2017.00529
- 10 131. Mitsdoerffer M, Peters A. Tertiary Lymphoid Organs in Central Nervous System
11 Autoimmunity. *Frontiers in Immunology*. 2016;7doi:10.3389/fimmu.2016.00451
- 12 132. Valencia-Sanchez C, Guo Y, Krecke KN, *et al.* Cerebral Cortical Encephalitis in Myelin
13 Oligodendrocyte Glycoprotein Antibody-Associated Disease. *Ann Neurol*. Feb 2023;93(2):297-
14 302. doi:10.1002/ana.26549
- 15 133. Zrzavy T, Endmayr V, Bauer J, *et al.* Neuropathological Variability within a Spectrum of
16 NMDAR-Encephalitis. *Ann Neurol*. Nov 2021;90(5):725-737. doi:10.1002/ana.26223
- 17 134. Nakajima A, Yanagimura F, Saji E, *et al.* Stage-dependent immunity orchestrates AQP4
18 antibody-guided NMOSD pathology: a role for netting neutrophils with resident memory T cells
19 in situ. *Acta Neuropathol*. Apr 24 2024;147(1):76. doi:10.1007/s00401-024-02725-x
- 20 135. Esser D, Muller-Miny L, Heming M, *et al.* Activated alphabeta T- and reduced mucosa-
21 associated invariant T cells in LGI1- and CASPR2-encephalitis. *Brain*. Mar 17
22 2025;doi:10.1093/brain/awaf096
- 23 136. Pittock SJ, Lennon VA, Krecke K, Wingerchuk DM, Lucchinetti CF, Weinshenker BG.
24 Brain abnormalities in neuromyelitis optica. *Arch Neurol*. Mar 2006;63(3):390-6.
25 doi:10.1001/archneur.63.3.390
- 26 137. Schellhammer L, Beffinger M, Salazar U, Laman JD, Buch T, Vom Berg J. Exit pathways
27 of therapeutic antibodies from the brain and retention strategies. *iScience*. Nov 17
28 2023;26(11):108132. doi:10.1016/j.isci.2023.108132

- 1 138. Hinson SR, Pittock SJ, Lucchinetti CF, *et al.* Pathogenic potential of IgG binding to water
2 channel extracellular domain in neuromyelitis optica. *Neurology*. Dec 11 2007;69(24):2221-31.
3 doi:10.1212/01.WNL.0000289761.64862.ce
- 4 139. Yandamuri SS, Filipek B, Obaid AH, *et al.* MOGAD patient autoantibodies induce
5 complement, phagocytosis, and cellular cytotoxicity. *JCI Insight*. Jun 8
6 2023;8(11)doi:10.1172/jci.insight.165373
- 7 140. Wollenweber L, Bondt A, Vidarsson G, Huijbers MG, Heck AJR. Widespread Fab-arm
8 exchange affects all endogenous serum IgG4. *Nat Commun*. Dec 8 2025;doi:10.1038/s41467-025-
9 67105-9
- 10 141. van Hoof S, Kreye J, Cordero-Gomez C, *et al.* Human cerebrospinal fluid monoclonal
11 CASPR2 autoantibodies induce changes in electrophysiology, functional MRI, and behavior in
12 rodent models. *Brain Behav Immun*. Nov 2024;122:266-278. doi:10.1016/j.bbi.2024.08.027
- 13 142. Petit-Pedrol M, Sell J, Planaguma J, *et al.* LGI1 antibodies alter Kv1.1 and AMPA
14 receptors changing synaptic excitability, plasticity and memory. *Brain*. Nov 1 2018;141(11):3144-
15 3159. doi:10.1093/brain/awy253
- 16 143. Gao Y, Li H, Luo H, *et al.* Purified Serum IgG from a Patient with Anti-IgLON5 Antibody
17 Cause Long-Term Movement Disorders with Impaired Dopaminergic Pathways in Mice.
18 *Biomedicines*. 2023;11(9):2483. doi:10.3390/biomedicines11092483
- 19 144. Spadaro M, Winklmeier S, Beltrán E, *et al.* Pathogenicity of human antibodies against
20 myelin oligodendrocyte glycoprotein. *Annals of Neurology*. Aug 2018;84(2):315-328.
21 doi:10.1002/ana.25291
- 22 145. Saadoun S, Waters P, Bell BA, Vincent A, Verkman AS, Papadopoulos MC. Intra-cerebral
23 injection of neuromyelitis optica immunoglobulin G and human complement produces
24 neuromyelitis optica lesions in mice. *Brain*. Feb 2010;133(Pt 2):349-61.
25 doi:10.1093/brain/awp309
- 26 146. Maudes E, Planaguma J, Marmolejo L, *et al.* Neuro-immunobiology and treatment
27 assessment in a mouse model of anti-NMDAR encephalitis. *Brain*. Dec 24
28 2024;doi:10.1093/brain/awae410

- 1 147. Serizawa K, Miyake S, Katsura Y, *et al.* Intradermal AQP4 peptide immunization induces
2 clinical features of neuromyelitis optica spectrum disorder in mice. *J Neuroimmunol.* Jul 15
3 2023;380:578109. doi:10.1016/j.jneuroim.2023.578109
- 4 148. Hinson SR, Clift IC, Luo N, Kryzer TJ, Lennon VA. Autoantibody-induced internalization
5 of CNS AQP4 water channel and EAAT2 glutamate transporter requires astrocytic Fc receptor.
6 *Proc Natl Acad Sci U S A.* May 23 2017;114(21):5491-5496. doi:10.1073/pnas.1701960114
- 7 149. Soltys J, Liu Y, Ritchie A, *et al.* Membrane assembly of aquaporin-4 autoantibodies
8 regulates classical complement activation in neuromyelitis optica. *J Clin Invest.* Apr 8
9 2019;129(5):2000-2013. doi:10.1172/JCI122942
- 10 150. Duan T, Smith AJ, Verkman AS. Complement-dependent bystander injury to neurons in
11 AQP4-IgG seropositive neuromyelitis optica. *Journal of Neuroinflammation.*
12 2018;15(1)doi:10.1186/s12974-018-1333-z
- 13 151. Lucchinetti CF, Mandler RN, McGavern D, *et al.* A role for humoral mechanisms in the
14 pathogenesis of Devic's neuromyelitis optica. *Brain.* Jul 2002;125(Pt 7):1450-61.
15 doi:10.1093/brain/awf151
- 16 152. Pittock SJ, Barnett M, Bennett JL, *et al.* Ravulizumab in Aquaporin-4-Positive
17 Neuromyelitis Optica Spectrum Disorder. *Ann Neurol.* Jun 2023;93(6):1053-1068.
18 doi:10.1002/ana.26626
- 19 153. Takai Y, Misu T, Fujihara K, Aoki M. Pathology of myelin oligodendrocyte glycoprotein
20 antibody-associated disease: a comparison with multiple sclerosis and aquaporin 4 antibody-
21 positive neuromyelitis optica spectrum disorders. *Front Neurol.* 2023;14:1209749.
22 doi:10.3389/fneur.2023.1209749
- 23 154. Brilot F, Dale RC, Selter RC, *et al.* Antibodies to native myelin oligodendrocyte
24 glycoprotein in children with inflammatory demyelinating central nervous system disease. *Annals*
25 *of Neurology.* 2009/12/01 2009;66(6):833-842. doi:https://doi.org/10.1002/ana.21916
- 26 155. Takai Y, Misu T, Suzuki H, *et al.* Staging of astrocytopathy and complement activation in
27 neuromyelitis optica spectrum disorders. *Brain.* 2021;144(8):2401-2415.
28 doi:10.1093/brain/awab102

- 1 156. Takai Y, Misu T, Kaneko K, *et al.* Myelin oligodendrocyte glycoprotein antibody-
2 associated disease: an immunopathological study. *Brain.* 2020;143(5):1431-1446.
3 doi:10.1093/brain/awaa102
- 4 157. Kwon YN, Kim B, Kim J-S, *et al.* Myelin Oligodendrocyte Glycoprotein-Immunoglobulin
5 G in the CSF. *Neurology Neuroimmunology & Neuroinflammation.* 9(1):e1095.
6 doi:10.1212/NXI.0000000000001095
- 7 158. Spadaro M, Gerdes LA, Mayer MC, *et al.* Histopathology and clinical course of MOG-
8 antibody-associated encephalomyelitis. *Annals of Clinical and Translational Neurology.*
9 2015/03/01 2015;2(3):295-301. doi:https://doi.org/10.1002/acn3.164
- 10 159. Ikeda T, Yamada K, Ogawa R, *et al.* The pathological features of MOG antibody-positive
11 cerebral cortical encephalitis as a new spectrum associated with MOG antibodies: A case report.
12 *Journal of the Neurological Sciences.* 2018/09/15/ 2018;392:113-115.
13 doi:https://doi.org/10.1016/j.jns.2018.06.028
- 14 160. Martinez-Hernandez E, Horvath J, Shiloh-Malawsky Y, Sangha N, Martinez-Lage M,
15 Dalmau J. Analysis of complement and plasma cells in the brain of patients with anti-NMDAR
16 encephalitis. *Neurology.* Aug 9 2011;77(6):589-93. doi:10.1212/WNL.0b013e318228c136
- 17 161. Dalmau J, Gleichman AJ, Hughes EG, *et al.* Anti-NMDA-receptor encephalitis: case series
18 and analysis of the effects of antibodies. *Lancet Neurol.* Dec 2008;7(12):1091-8.
19 doi:10.1016/S1474-4422(08)70224-2
- 20 162. Michalski K, Abdulla T, Kleeman S, *et al.* Structural and functional mechanisms of anti-
21 NMDAR autoimmune encephalitis. *Nature Structural & Molecular Biology.*
22 2024;31(12):1975-1986. doi:10.1038/s41594-024-01386-4
- 23 163. Al-Diwani A, Theorell J, Zghoul T, *et al.* The distinctive psychopathology of NMDAR-
24 antibody encephalitis compared with primary psychoses: an international, multicentre,
25 retrospective phenotypic analysis. *The Lancet Psychiatry.* 2026;13(1):47-61. doi:10.1016/s2215-
26 0366(25)00305-0
- 27 164. Jamet Z, Mergaux C, Meras M, *et al.* NMDA receptor autoantibodies primarily impair the
28 extrasynaptic compartment. *Brain.* 2024;147(8):2745-2760. doi:10.1093/brain/awae163

- 1 165. Zhao M, Lynch DR, Irani SR. Autoimmune 'secondary synaptopathies': do NMDAR
2 antibodies cause a primary extra-synaptopathy? *Brain*. Aug 1 2024;147(8):2601-2603.
3 doi:10.1093/brain/awae236
- 4 166. Chihara N, Yamamura T. Immuno-pathogenesis of neuromyelitis optica and emerging
5 therapies. *Semin Immunopathol*. Sep 2022;44(5):599-610. doi:10.1007/s00281-022-00941-9
- 6 167. Saadoun S, Papadopoulos MC. Role of membrane complement regulators in neuromyelitis
7 optica. *Multiple Sclerosis Journal*. 2015/11/01 2015;21(13):1644-1654.
8 doi:10.1177/1352458515571446
- 9 168. Ramirez-Franco J, Debreux K, Extremet J, *et al*. Patient-derived antibodies reveal the
10 subcellular distribution and heterogeneous interactome of LGI1. *Brain*. Nov 21
11 2022;145(11):3843-3858. doi:10.1093/brain/awac218
- 12 169. Heine J, Pruss H, Kopp UA, *et al*. Beyond the limbic system: disruption and functional
13 compensation of large-scale brain networks in patients with anti-LGI1 encephalitis. *J Neurol*
14 *Neurosurg Psychiatry*. Nov 2018;89(11):1191-1199. doi:10.1136/jnnp-2017-317780
- 15 170. Yamagata A, Miyazaki Y, Yokoi N, *et al*. Structural basis of epilepsy-related ligand-
16 receptor complex LGI1-ADAM22. *Nat Commun*. Apr 18 2018;9(1):1546. doi:10.1038/s41467-
17 018-03947-w
- 18 171. Ramberger M, Berretta A, Tan JMM, *et al*. Distinctive binding properties of human
19 monoclonal LGI1 autoantibodies determine pathogenic mechanisms. *Brain*. 2020;143(6):1731-
20 1745. doi:10.1093/brain/awaa104
- 21 172. Upadhy M, Kirmann T, Wilson MA, *et al*. Peripherally-derived LGI1-reactive
22 monoclonal antibodies cause epileptic seizures in vivo. *Brain*. Aug 1 2024;147(8):2636-2642.
23 doi:10.1093/brain/awae129
- 24 173. Extremet J, El Far O, Ankri N, Irani SR, Debanne D, Russier M. An Epitope-Specific
25 LGI1-Autoantibody Enhances Neuronal Excitability by Modulating Kv1.1 Channel. *Cells*. Aug
26 31 2022;11(17)doi:10.3390/cells11172713
- 27 174. Sell J, Rahmati V, Kempfer M, *et al*. Comparative Effects of Domain-Specific Human
28 Monoclonal Antibodies Against LGI1 on Neuronal Excitability. *Neurol Neuroimmunol*
29 *Neuroinflamm*. May 2023;10(3)doi:10.1212/NXI.0000000000200096

- 1 175. Thompson J, Bi M, Murchison AG, *et al.* The importance of early immunotherapy in
2 patients with faciobrachial dystonic seizures. *Brain*. Feb 1 2018;141(2):348-356.
3 doi:10.1093/brain/awx323
- 4 176. Bien CG, Rada A, Mertens M, *et al.* LGI1 encephalitis: potentially complement-activating
5 anti-LGI1-IgG subclasses 1/2/3 are associated with the development of hippocampal sclerosis. *J*
6 *Neurol*. Sep 2024;271(9):6325-6335. doi:10.1007/s00415-024-12594-9
- 7 177. Bien CG, Vincent A, Barnett MH, *et al.* Immunopathology of autoantibody-associated
8 encephalitides: clues for pathogenesis. *Brain*. May 2012;135(Pt 5):1622-38.
9 doi:10.1093/brain/aws082
- 10 178. Manso C, Querol L, Mekaouche M, Illa I, Devaux JJ. Contactin-1 IgG4 antibodies cause
11 paranode dismantling and conduction defects. *Brain*. Jun 2016;139(Pt 6):1700-12.
12 doi:10.1093/brain/aww062
- 13 179. Ramanathan S, Tseng M, Davies AJ, *et al.* Leucine-Rich Glioma-Inactivated 1 versus
14 Contactin-Associated Protein-like 2 Antibody Neuropathic Pain: Clinical and Biological
15 Comparisons. *Ann Neurol*. Oct 2021;90(4):683-690. doi:10.1002/ana.26189
- 16 180. Laurencin C, Andre-Obadia N, Camdessanche JP, *et al.* Peripheral small fiber dysfunction
17 and neuropathic pain in patients with Morvan syndrome. *Neurology*. Dec 8 2015;85(23):2076-8.
18 doi:10.1212/WNL.0000000000002037
- 19 181. Irani SR, Alexander S, Waters P, *et al.* Antibodies to Kv1 potassium channel-complex
20 proteins leucine-rich, glioma inactivated 1 protein and contactin-associated protein-2 in limbic
21 encephalitis, Morvan's syndrome and acquired neuromyotonia. *Brain*. Sep 2010;133(9):2734-48.
22 doi:10.1093/brain/awq213
- 23 182. Irani SR, Pettingill P, Kleopa KA, *et al.* Morvan syndrome: clinical and serological
24 observations in 29 cases. *Ann Neurol*. Aug 2012;72(2):241-55. doi:10.1002/ana.23577
- 25 183. Dawes JM, Weir GA, Middleton SJ, *et al.* Immune or Genetic-Mediated Disruption of
26 CASPR2 Causes Pain Hypersensitivity Due to Enhanced Primary Afferent Excitability. *Neuron*.
27 2018;97(4):806-822.e10. doi:10.1016/j.neuron.2018.01.033

- 1 184. Kortvelyessy P, Bauer J, Stoppel CM, *et al.* Complement-associated neuronal loss in a
2 patient with CASPR2 antibody-associated encephalitis. *Neurol Neuroimmunol Neuroinflamm.* Apr
3 2015;2(2):e75. doi:10.1212/NXI.0000000000000075
- 4 185. Terroba-Navajas P, Spatola M, Chuquisana O, *et al.* Humoral signatures of Caspr2-
5 antibody spectrum disorder track with clinical phenotypes and outcomes. *Med.* Feb 14
6 2025;6(2):100515. doi:10.1016/j.medj.2024.09.004
- 7 186. Sabater L, Planaguma J, Dalmau J, Graus F. Cellular investigations with human antibodies
8 associated with the anti-IgLON5 syndrome. *J Neuroinflammation.* Sep 1 2016;13(1):226.
9 doi:10.1186/s12974-016-0689-1
- 10 187. Reinecke R, Nitsch S, Faber F, *et al.* Brainstem pathology in anti-IgLON5 disease: new
11 insights into early events and tau progression. *Brain.* Jan 17 2026;doi:10.1093/brain/awag015
- 12 188. Ryding M, Gamre M, Nissen MS, *et al.* Neurodegeneration Induced by Anti-IgLON5
13 Antibodies Studied in Induced Pluripotent Stem Cell-Derived Human Neurons. *Cells.* Apr 8
14 2021;10(4)doi:10.3390/cells10040837
- 15 189. Paul F, Marignier R, Palace J, *et al.* International Delphi Consensus on the Management
16 of AQP4-IgG+ NMOSD: Recommendations for Eculizumab, Inebilizumab, and Satralizumab.
17 *Neurol Neuroimmunol Neuroinflamm.* Jul 2023;10(4)doi:10.1212/NXI.0000000000200124
- 18 190. Kim SH, Huh SY, Lee SJ, Joung A, Kim HJ. A 5-year follow-up of rituximab treatment in
19 patients with neuromyelitis optica spectrum disorder. *JAMA Neurol.* Sep 1 2013;70(9):1110-7.
20 doi:10.1001/jamaneurol.2013.3071
- 21 191. Sechi E, Buciu M, Pittock SJ, *et al.* Positive Predictive Value of Myelin Oligodendrocyte
22 Glycoprotein Autoantibody Testing. *JAMA Neurol.* Jun 1 2021;78(6):741-746.
23 doi:10.1001/jamaneurol.2021.0912
- 24 192. Krohn S, Muller-Jensen L, Kuchling J, *et al.* Cognitive Deficits in Anti-LGI1 Encephalitis
25 Are Linked to Immunotherapy-Resistant White Matter Network Changes. *Neurol Neuroimmunol*
26 *Neuroinflamm.* Mar 2025;12(2):e200360. doi:10.1212/NXI.0000000000200360
- 27 193. Binks SNM, Veldsman M, Handel AE, *et al.* Fatigue predicts quality of life after leucine-
28 rich glioma-inactivated 1-antibody encephalitis. *Ann Clin Transl Neurol.* Feb 1
29 2024;doi:10.1002/acn3.52006

- 1 194. Hummert MW, Schoppe LM, Bellmann-Strobl J, *et al.* Costs and Health-Related Quality
2 of Life in Patients With NMO Spectrum Disorders and MOG-Antibody-Associated Disease:
3 CHANCE(NMO) Study. *Neurology*. Mar 15 2022;98(11):e1184-e1196.
4 doi:10.1212/WNL.0000000000200052
- 5 195. Abboud H, Clardy SL, Dubey D, *et al.* The Clinical Trial Landscape in Autoimmune
6 Encephalitis: Challenges and Opportunities. *Neurology*. 2025;104(8):e213487.
- 7 196. Sechi E, Flanagan EP. Antibody-Mediated Autoimmune Diseases of the CNS: Challenges
8 and Approaches to Diagnosis and Management. *Front Neurol*. 2021;12:673339.
9 doi:10.3389/fneur.2021.673339
- 10 197. Ciano-Petersen NL, Muniz-Castrillo S, Birzu C, *et al.* Cytokine dynamics and targeted
11 immunotherapies in autoimmune encephalitis. *Brain Commun*. 2022;4(4):fcac196.
12 doi:10.1093/braincomms/fcac196
- 13 198. Pittock SJ, Berthele A, Fujihara K, *et al.* Eculizumab in Aquaporin-4-Positive
14 Neuromyelitis Optica Spectrum Disorder. *N Engl J Med*. Aug 15 2019;381(7):614-625.
15 doi:10.1056/NEJMoa1900866
- 16 199. Trewin BP, Freeman I, Ramanathan S, Irani SR. Immunotherapy in autoimmune
17 encephalitis. *Curr Opin Neurol*. Jun 1 2022;35(3):399-414.
18 doi:10.1097/WCO.0000000000001048
- 19 200. Forsthuber TG, Cimbora DM, Ratchford JN, Katz E, Stuve O. B cell-based therapies in
20 CNS autoimmunity: differentiating CD19 and CD20 as therapeutic targets. *Ther Adv Neurol*
21 *Disord*. 2018;11:1756286418761697. doi:10.1177/1756286418761697
- 22 201. Palanichamy A, Jahn S, Nickles D, *et al.* Rituximab efficiently depletes increased CD20-
23 expressing T cells in multiple sclerosis patients. *J Immunol*. Jul 15 2014;193(2):580-586.
24 doi:10.4049/jimmunol.1400118
- 25 202. Suan D, Moore J, Goodnow CC. Can autoimmune disease be cured by deep CD19+ cell
26 depletion? *J Immunol*. Jun 1 2025;214(6):1075-1092. doi:10.1093/jimmun/vkaf008
- 27 203. Dinoto A, Cheli M, Bratina A, Sartori A, Manganotti P. Bortezomib in anti-N-Methyl-d-
28 Aspartate-Receptor (NMDA-R) encephalitis: A systematic review. *J Neuroimmunol*. Jul 15
29 2021;356:577586. doi:10.1016/j.jneuroim.2021.577586

- 1 204. Holzer MT, Ruffer N, Huber TB, Kotter I, Ostendorf L, Krusche M. Daratumumab for
2 autoimmune diseases: a systematic review. *RMD Open*. Dec 14 2023;9(4)doi:10.1136/rmdopen-
3 2023-003604
- 4 205. Lehmann-Horn K, Sagan SA, Bernard CC, Sobel RA, Zamvil SS. B-cell very late antigen-
5 4 deficiency reduces leukocyte recruitment and susceptibility to central nervous system
6 autoimmunity. *Ann Neurol*. May 2015;77(5):902-8. doi:10.1002/ana.24387
- 7 206. Chinas NA, Alexopoulos H. CAR T-cells meet autoimmune neurological diseases: a new
8 dawn for therapy. *Frontiers in Immunology*. 2025;16doi:10.3389/fimmu.2025.1604174
- 9 207. Berger SC, Fehse B, Akyüz N, *et al*. Molecular monitoring of T-cell kinetics and migration
10 in severe neurotoxicity after real-world CD19-specific chimeric antigen receptor T cell therapy.
11 *Haematologica*. 2022;108(2):444-456. doi:10.3324/haematol.2022.281110
- 12 208. Sanderson JB, Lin YH, Su K, *et al*. Treatment-Emergent Parkinsonism in Four Patients
13 Treated with Chimeric Antigen Receptor T-Cell Therapy. *Mov Disord Clin Pract*. Nov 19
14 2025;doi:10.1002/mdc3.70452
- 15 209. Grebenciucova E, VanHaerents S. Interleukin 6: at the interface of human health and
16 disease. *Front Immunol*. 2023;14:1255533. doi:10.3389/fimmu.2023.1255533
- 17 210. Lee WJ, Lee ST, Shin YW, *et al*. Teratoma Removal, Steroid, IVIG, Rituximab and
18 Tocilizumab (T-SIRT) in Anti-NMDAR Encephalitis. *Neurotherapeutics*. Jan 2021;18(1):474-
19 487. doi:10.1007/s13311-020-00921-7
- 20 211. Alfaidi N, Karmastaji S, Matic A, Bril V. FcRn Inhibitor Therapies in Neurologic Diseases.
21 *CNS Drugs*. Jun 2024;38(6):425-441. doi:10.1007/s40263-024-01090-3
- 22 212. Liu J, Li M, Liu J, *et al*. Multicenter experience with Efgartigimod in the treatment of anti-
23 NMDAR encephalitis compared with IVIG and SPA-IA during acute attacks. *Life Sci*. Jun 15
24 2025;371:123597. doi:10.1016/j.lfs.2025.123597
- 25 213. Hassel JC, Piperno-Neumann S, Rutkowski P, *et al*. Three-Year Overall Survival with
26 Tebentafusp in Metastatic Uveal Melanoma. *New England Journal of Medicine*.
27 2023;389(24):2256-2266. doi:10.1056/nejmoa2304753

- 1 214. St. Clair EW, Baer AN, Ng W-F, *et al.* CD40 ligand antagonist dazodalibep in Sjögren's
2 disease: a randomized, double-blinded, placebo-controlled, phase 2 trial. *Nature Medicine*.
3 2024;30(6):1583-1592. doi:10.1038/s41591-024-03009-3
- 4 215. Ellebrecht CT, Bhoj VG, Nace A, *et al.* Reengineering chimeric antigen receptor T cells
5 for targeted therapy of autoimmune disease. *Science*. 2016;353(6295):179-184.
6 doi:10.1126/science.aaf6756
- 7 216. Oh S, Mao X, Manfredo-Vieira S, *et al.* Precision targeting of autoantigen-specific B cells
8 in muscle-specific tyrosine kinase myasthenia gravis with chimeric autoantibody receptor T cells.
9 *Nature Biotechnology*. 2023;41(9):1229-1238. doi:10.1038/s41587-022-01637-z
- 10 217. Reincke SM, von Wardenburg N, Homeyer MA, *et al.* Chimeric autoantibody receptor T
11 cells deplete NMDA receptor-specific B cells. *Cell*. Nov 9 2023;186(23):5084-5097 e18.
12 doi:10.1016/j.cell.2023.10.001
- 13 218. Hinson SR, Romero MF, Popescu BF, *et al.* Molecular outcomes of neuromyelitis optica
14 (NMO)-IgG binding to aquaporin-4 in astrocytes. *Proc Natl Acad Sci U S A*. Jan 24
15 2012;109(4):1245-50. doi:10.1073/pnas.1109980108
- 16 219. Singh P, Gao X, Kleijn HJ, Bellanti F, Pelto R. Eculizumab Pharmacokinetics and
17 Pharmacodynamics in Patients With Neuromyelitis Optica Spectrum Disorder. *Front Neurol*.
18 2021;12:696387. doi:10.3389/fneur.2021.696387
- 19 220. Sun W, Khare P, Wang X, *et al.* Selective Depletion of Antigen-Specific Antibodies for
20 the Treatment of Demyelinating Disease. *Mol Ther*. Mar 3 2021;29(3):1312-1323.
21 doi:10.1016/j.ymthe.2020.11.017
- 22 221. Steinke S, Kirmann T, Loi EA, *et al.* NMDA-receptor-Fc-fusion constructs neutralize anti-
23 NMDA receptor antibodies. *Brain*. May 2 2023;146(5):1812-1820. doi:10.1093/brain/awac497
- 24 222. Duan T, Tradtrantip L, Phuan PW, Bennett JL, Verkman AS. Affinity-matured
25 'aquaporin-4' anti-aquaporin-4 antibody for therapy of seropositive neuromyelitis optica
26 spectrum disorders. *Neuropharmacology*. Jan 1 2020;162:107827.
27 doi:10.1016/j.neuropharm.2019.107827

- 1 223. Kanno A, Kito T, Maeda M, *et al.* Monoclonal humanized monovalent antibody blocking
 2 therapy for anti-NMDA receptor encephalitis. *Nat Commun.* Jun 17 2025;16(1):5292.
 3 doi:10.1038/s41467-025-60628-1
- 4 224. Zubizarreta I, Flórez-Grau G, Vila G, *et al.* Immune tolerance in multiple sclerosis and
 5 neuromyelitis optica with peptide-loaded tolerogenic dendritic cells in a phase 1b trial.
 6 *Proceedings of the National Academy of Sciences.* 2019;116(17):8463-8470.
 7 doi:doi:10.1073/pnas.1820039116
- 8 225. Pérez-Bucio C, Behere A, Landegren N. Mechanisms of autoimmune-mediated
 9 paraneoplastic syndromes: immune tolerance and disease pathogenesis. *Front Immunol.*
 10 2025;16:1608934. doi:10.3389/fimmu.2025.1608934

11

12 Figure legends

13 **Figure 1 Cascade to pathogenicity in neuroglial-surface autoantibody**
 14 **(NGSAb)-mediated CNS diseases.** Schematic timeline from the pre-symptomatic
 15 phase through symptomatic disease to sequelae/recurrence, highlighting a series of events
 16 including: (1) Genetic predisposition; (2-3) Triggering events (e.g., infection, tumour, or
 17 immune checkpoint inhibitors [ICI]); (4) CNS entry of autoantigen-reactive
 18 lymphocytes/antibodies; (5-7) End-organ dysfunction leading to chronic sequelae and
 19 unknown factors precipitating relapse. NGSAb, neuroglial-surface autoantibody; ASC,
 20 antibody-secreting cell; HLA, human leukocyte antigen; ICI, immune checkpoint inhibitor;
 21 TCR, T-cell receptor; CTLA4, cytotoxic T-lymphocyte-associated protein 4. Created in
 22 BioRender. Montini, F. (2026) <https://BioRender.com/k8bgyg5>.

23

24 **Figure 2 Anatomical compartments and B cell checkpoints underlying** 25 **immune tolerance escape and access to the cerebrospinal fluid.**

26 Neuroimmune anatomical compartments, including bone marrow (skull and long bone),
 27 blood, lymph node, and cerebrospinal fluid, and the central and peripheral tolerance
 28 checkpoints which shape the B cell repertoire. In the bone marrow, B cell development
 29 proceeds through preB (surrogate light chain expressed, in red) and immature B cells
 30 (CD19⁺CD20⁺IgM⁺CD27⁻, that express a bone fide light chain and hence a mature B cell
 31 receptor, BCR), which flank a central checkpoint. The first B cells to reach blood are new

1 emigrant B cells (CD19⁺CD20⁺CD10⁺IgD⁺CD27⁻) and can interact in germinal centers
 2 (typically situated in lymph nodes) as mature naïve B cells (CD19⁺CD20⁺IgD⁺CD27⁻) after
 3 traversing a peripheral checkpoint. Here, their interaction with dendritic cells and
 4 autoantigen reactive CD4⁺ T cells signals via the TCR-HLA class II, CD80/86-CD28/CTLA4,
 5 and CD40-CD40L molecular pathways alongside cytokines (e.g., IFN γ , IL21). These
 6 pathways generate unswitched/switched memory B cells (e.g., CD27⁺ with or without IgD
 7 expression, respectively), short-lived plasmablasts/ASCs
 8 (CD19⁺CD20^{lo}CD27⁺⁺CD38⁺⁺CD138^{+/-}), and long-lived (CD19⁺CD20⁻
 9 CD27⁺⁺CD38⁺⁺CD138⁺) plasma cells, which secrete autoantibodies. The CSF compartment
 10 highlights the infiltration of B cells and ASCs, and contributions from T cells (both CD4⁺ and
 11 CD8⁺) and other rare types, including MAIT/NK cells. Soluble antigen (from
 12 secretion/ectodomain shedding or injury-related release) may engage BCRs and retain
 13 antigen-specific B cells/ASCs in CSF. BCR, B-cell receptor; ASC, antibody-secreting cell;
 14 LLPC, long-lived plasma cell; MAIT, mucosa-associated invariant T cell; NK, natural killer;
 15 HLA, human leukocyte antigen; IFN- γ , interferon-gamma; IL-21, interleukin-21; CSF,
 16 cerebrospinal fluid. Created in BioRender. Montini, F. (2026)
 17 <https://BioRender.com/187o6s1>.

18

19 **Figure 3 Mechanisms by which autoantibodies modulate neuroglial**

20 **surface antigens.** A) Schematic showing the potential discrepancy between size of the
 21 synaptic cleft compared to IgG antibodies. B) IgG1 binds AQP4 on the astrocyte end feet,
 22 triggering the classical complement pathway and ADCC via NK cells, and other cell types
 23 including neutrophils. C) MOG-IgG1s can function via complement fixation, and promote
 24 NK cell-mediated cytotoxicity plus infiltration and activation of macrophages and
 25 microglia. D) NMDAR-IgG1 induces receptor internalization leading to NMDAR-
 26 hypofunction. E) LRR domain-directed IgG4s can lead to internalization of LGI1 and its
 27 neighbouring proteins. Whereas EPTP domain-directed IgG4 blocks the docking of LGI1
 28 with ADAM22/23. Both effects may disrupt presynaptic Kv1 potassium channels or post-
 29 synaptic AMPARs. Additionally, the axon initial segment is a proven site of LGI1-antibody
 30 effects, with potential to modulate action potential frequency (inset). F) IgG access to
 31 CASPR2 in PNS juxtaparanodes may be limited by myelin. Pain and other PNS symptoms
 32 are more likely caused by effects on unmyelinated sensory nerve terminals. G) CASPR2-
 33 IgG4s may bind synaptic / extra-synaptic CASPR2 leading to internalization of CASPR2,
 34 plus disruption of the CASPR2-contactin-2 interaction. H) IgLON5-IgG1 induces
 35 internalization of IgLON5, whereas IgLON5-IgG4 likely acts as to block protein-protein
 36 interactions antibody; both mechanisms ultimately alter interactions with neighbouring

1 proteins. AQP4, aquaporin-4; ADCC, antibody-dependent cellular cytotoxicity; NK, natural
2 killer; MOG, myelin oligodendrocyte glycoprotein; NMDAR, N-methyl-D-aspartate
3 receptors; LRR, leucine-rich repeat; EPTP, epitempin; LGI1, leucine-rich glioma-inactivated
4 1; ADAM, A Disintegrin and metalloproteinase domain-containing protein; AMPAR, α -
5 Amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptor; CASPR2, contactin-
6 associated protein-like 2; PNS, peripheral nervous system; IgLON, immunoglobulin-like cell
7 adhesion molecule; CNS, central nervous system; MAC, membrane attack complex; RTK,
8 receptor tyrosine kinase. Created in BioRender. Montini, F. (2026)
9 <https://BioRender.com/7kbps9d>.

10

11 **Figure 4 Therapeutics with targeted mechanisms in autoantibody driven**

12 **disease.** Schematic of choice, tailored mechanisms employed in autoantibody
13 neurological disease. Many targeted therapeutics act chiefly in the periphery (left),
14 downregulating the immune cells and pathways involved in autoantibody production and
15 maintenance. Most of these (e.g. anti-CD19/20/38, CAR-T/CAAR-T) bind to membrane-
16 bound proteins on pathogenic B-cell or ASC populations to decrease autoantibody
17 production, while others like tocilizumab or FcRn inhibitors target soluble proteins or
18 pathways of antibody recycling to decrease autoantibody production and maintenance,
19 respectively. Natalizumab, acting at the blood brain barrier (center), is positioned between
20 the peripheral and CNS compartments to prevent B- and T-cell ingress. Medications that
21 act within the CNS at the target site (right) serve to prevent autoantibody-antigen
22 interaction and the effector functions therein. Included on the bottom of the figure are
23 emerging therapeutics, highlighting new uses of foundational therapies and emerging
24 medications. *Limited effect on SLPCs, LLPCs. APC, antigen-presenting cell; CAR-T,
25 chimeric antigen receptor T-cell; CAAR-T, chimeric autoantibody receptor T-cell; CNS,
26 central nervous system; CXCL13, chemokine ligand 13; FcRn, neonatal fragment
27 crystallizable receptor; HLA, human leukocyte antigen; IL-, interleukin; JAK, Janus kinase;
28 LLPC, long-lived plasma cell; SLPC, short-lived plasma cell; VLA-4, very late antigen-4.
29 Created in BioRender. Montini, F. (2026) <https://BioRender.com/4a73gbm>.

30

31

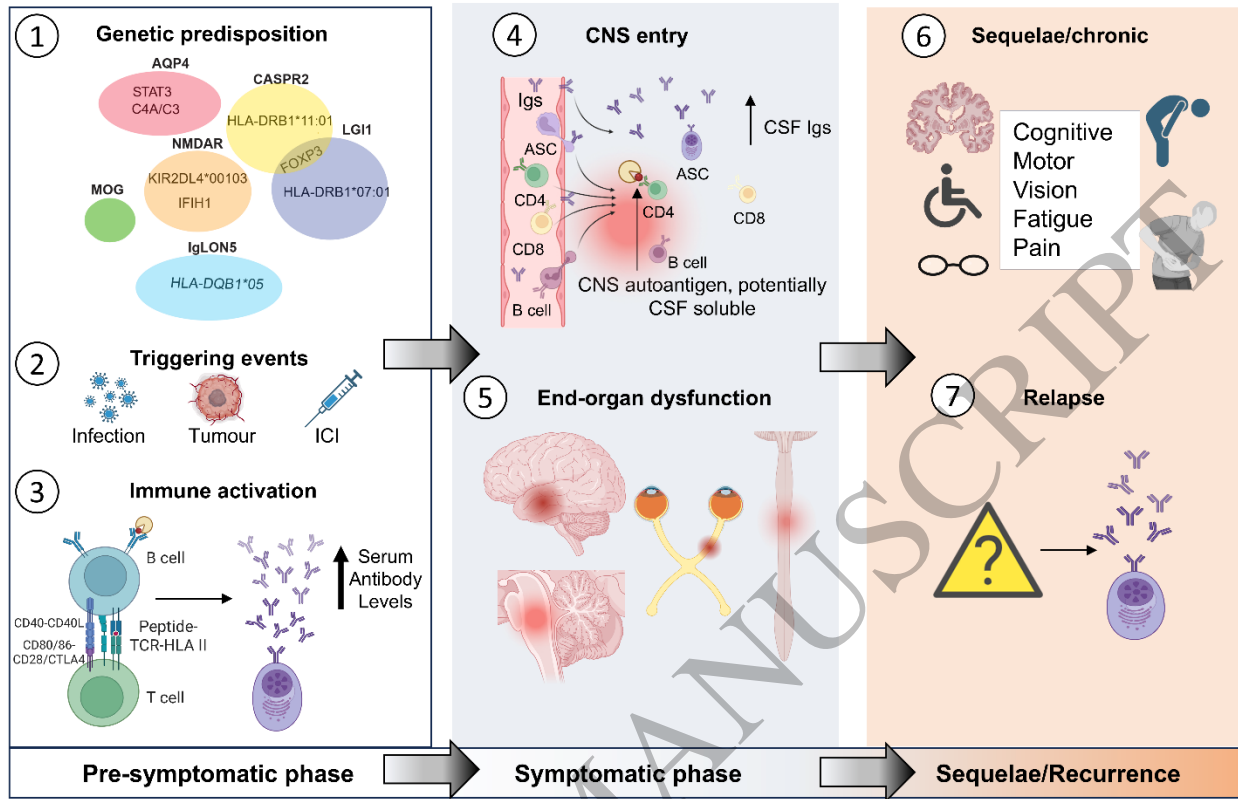


Figure 1
165x108 mm (x DPI)

1
2
3
4

ACCEPTED MANUSCRIPT

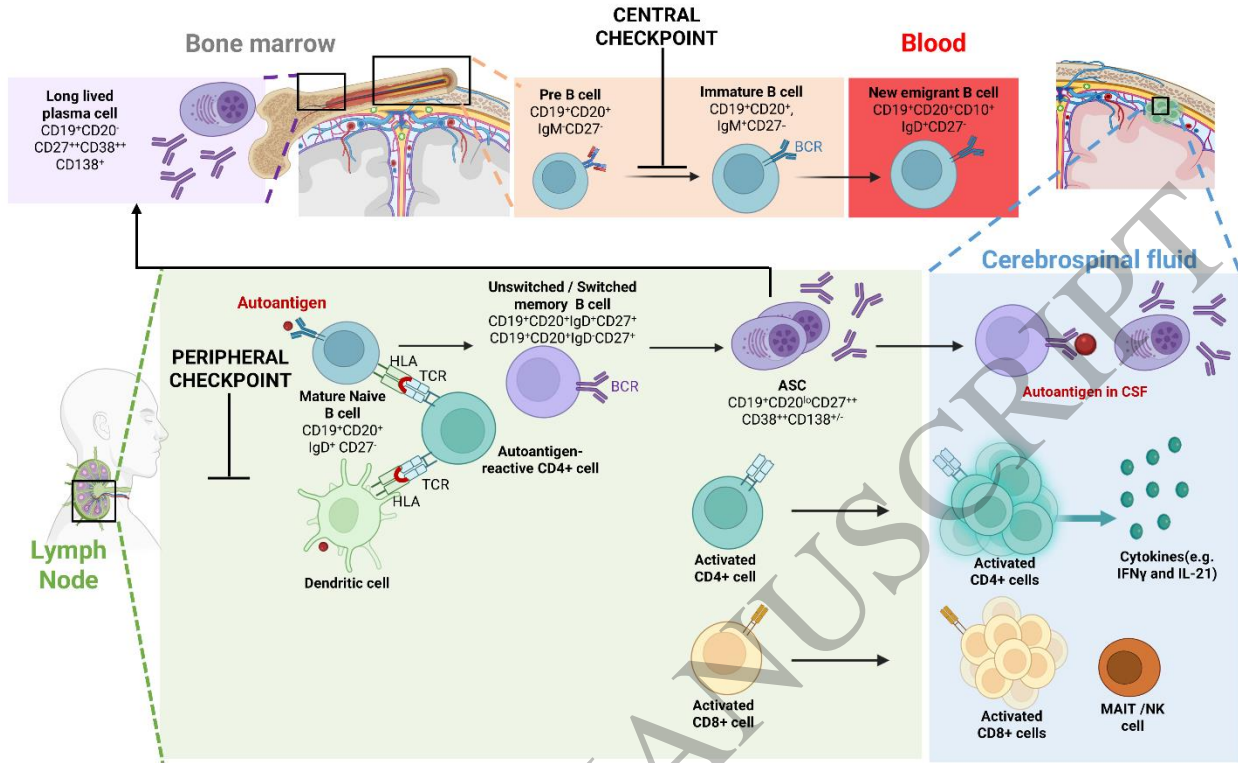


Figure 2
165x102 mm (x DPI)

1
2
3
4

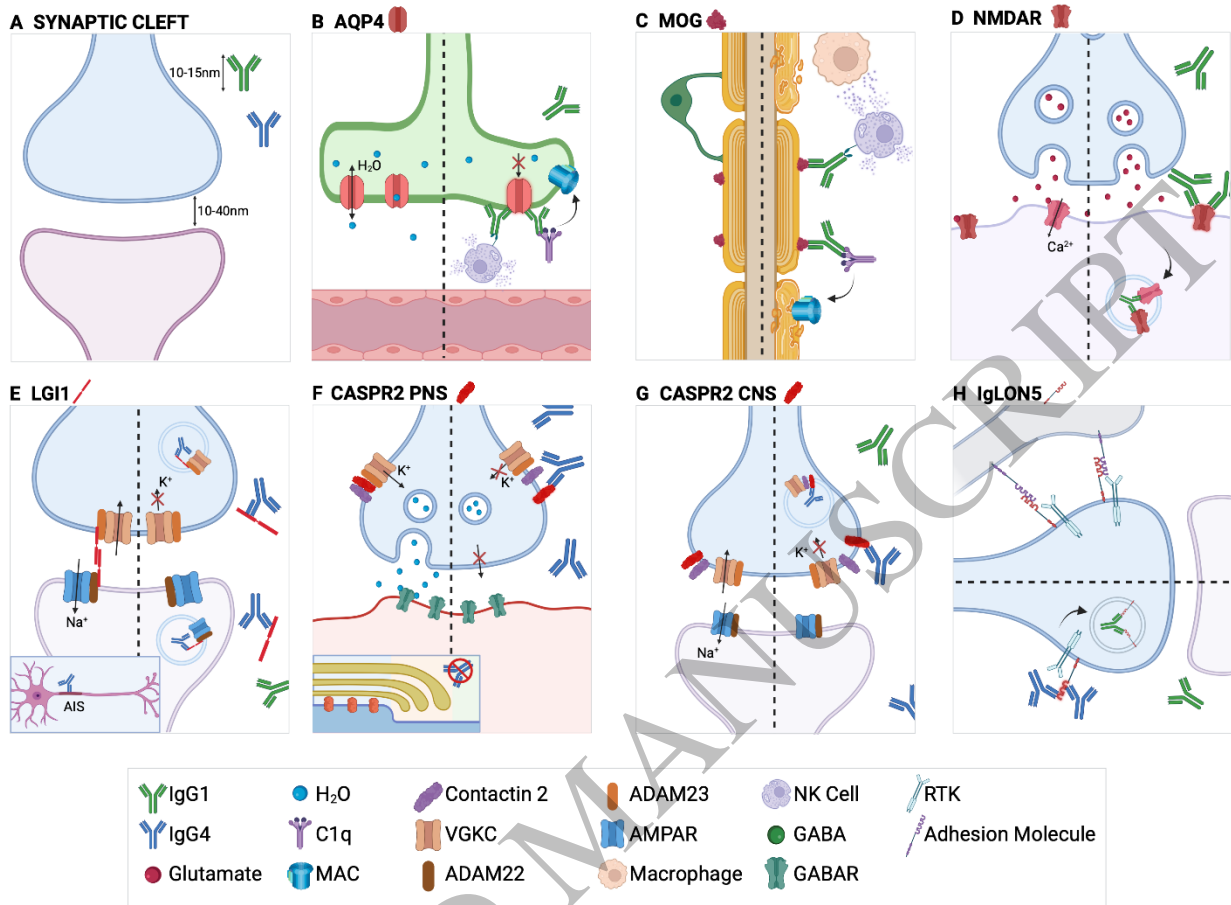
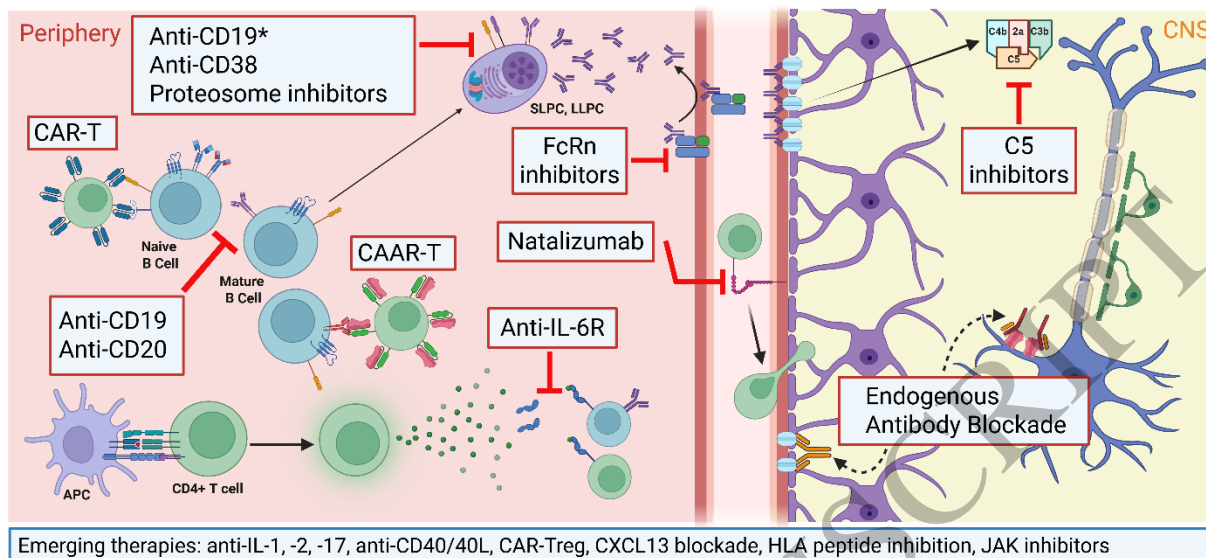


Figure 3
165x121 mm (x DPI)

1
2
3
4



- Emerging therapies: anti-IL-1, -2, -17, anti-CD40/40L, CAR-Treg, CXCL13 blockade, HLA peptide inhibition, JAK inhibitors
- Anti-CD19: inebelizumab
 - Anti-CD20: rituximab, ocrelizumab, ofatumumab, ublituximab
 - Anti-CD38: daratumumab
 - Anti-IL6R: tocilizumab, satralizumab
 - ART5803
 - Aquaporin-4
 - Aquaporumab
 - B-cell maturation antigen (BCMA)
 - CAR-T targets: CD19, BCMA; CAAR-T targets: NMDAR, MUSK
 - C5 inhibitors: eculizumab, ravulizumab, zilucoplan
 - FcRn inhibitors: efgartigimod, razanolixizumab
 - NMDAR
 - Proteasome inhibitors: bortezomib, carfilzomib and delanzomib
 - VLA-4 integrin

Figure 4
165x116 mm (x DPI)

Table I Clinical and biological features observed in each of the big six neuroglial-surface antibody mediated diseases

Autoantigen	Clinical features						Biological features						
	Age, median (range) ^a	Sex (M:F)	Tumors	Main Ethnicities	HSV Association	Relapses	Subclass	BCR Mutations	Complement Activation	HLA Restriction	Antigen Internalization	Steric Hindrance	ADCC
LGII	~60 (30-95)	2:1	<5%	White	-	++	IgG4 > I	+++	+	++++	++	+++	-
CASPR2	~60 (30-95)	7:1	10%	White	-	++	IgG4 > I	+++	+	+++	+	+++	-
IgLON5		1:1	<5%	White	-	+/-	IgG4 > I	N/A	-	++++	++	+++	-
NMDAR	~20 (1-80)	1:3	~30%	Afro-Caribbean and Asian	++	+++	IgG1	+/-	-	-	++++	-	-
AQP4	~40 (10-70)	1:7	<5%	Afro-Caribbean	-	++++	IgG1	+++	++++	+	+	-	++
MOG	~35 (5-70)	1:1.5	<5%	White	-	+++	IgG1	+/-	++	-	-	-	++

Strength of association estimated from nil (-) through low (+/++) to high (+++/++++). ADCC = antibody dependent cell cytotoxicity; BCR = B cell receptor; F = female; HLA = human leucocyte antigen; HSV = herpes simplex virus; M = male; N/A = no available data.

^aSimplified from primary data to represent summary statistics.