


ORIGINAL ARTICLE

# Sodium-glucose cotransporter 2 inhibition does not reduce hepatic steatosis in overweight, insulin-resistant patients without type 2 diabetes

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## Key words

fatty liver, lipogenesis, magnetic resonance spectroscopy, sodium-glucose transporter 2.

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## Abstract

**Background and Aim:** Non-alcoholic fatty liver disease (NAFLD) is rapidly becoming the leading indication for liver transplant and is associated with increased cardiovascular and liver mortality, yet there are no licensed therapies. Sodium-glucose cotransporter 2 (SGLT2) inhibitors are widely used for their glucose-lowering effects in patients with type 2 diabetes (T2D). Preclinical models have suggested a beneficial impact on NAFLD, but clinical data are limited, and there are currently no data on patients without T2D. We aimed to investigate the impact of SGLT2 inhibition on NAFLD in overweight, nondiabetic patients and establish the effect these agents may have on the processes that regulate hepatic steatosis in vivo.

**Methods:** We conducted an open-label, experimental medicine pilot study on insulin-resistant overweight/obese individuals ( $n = 10$ ) using gold-standard noninvasive assessments of NAFLD phenotype, including magnetic resonance spectroscopy, two-step hyperinsulinemic euglycemic clamps, and stable isotope tracers to assess lipid and glucose metabolism. Investigations were performed before and after a 12-week treatment with the SGLT2 inhibitor, dapagliflozin.

**Results:** Despite a body weight reduction of 4.4 kg, hepatic steatosis was unchanged following treatment. Hepatic glucose production increased, and there was impairment of glucose disposal during the low-dose insulin infusion. Although circulating, non-esterified, fatty acid levels did not change, the ability of insulin to suppress lipolysis was reduced.

**Conclusions:** SGLT2 inhibition for 12 weeks does not improve hepatic steatosis in patients without T2D. Additional studies in patients with established T2D or impairments of fasting or postprandial glucose homeostasis are needed to determine whether SGLT2 inhibition represents a viable therapeutic strategy for NAFLD. (ClinicalTrials.gov Number NCT02696941).

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## Introduction

Non-alcoholic fatty liver disease (NAFLD) has reached epidemic proportions affecting up to 30% of the general population and 70–90% of individuals with type 2 diabetes (T2D) and/or obesity.<sup>1,2</sup> It drives increased morbidity and mortality, both through a specific impact on the liver and through adverse cardiovascular outcomes<sup>3</sup>; by 2020, NAFLD will become the leading indication for liver transplantation worldwide.<sup>4</sup> NAFLD is a spectrum of disease ranging from simple steatosis to inflammation (non-alcoholic steatohepatitis [NASH]), cirrhosis, and hepatocellular carcinoma (HCC). Despite the burden of disease, there are currently no licensed treatments for NAFLD.

While the development of diabetes is a major driver for NAFLD progression,<sup>5</sup> up to 75% of those with NAFLD will not have T2D.<sup>6</sup> A crucial component of diabetes and NAFLD risk is both hepatic and peripheral insulin resistance. Using hyperinsulinemic euglycemic clamp techniques incorporating the use of stable isotope infusions, studies have identified the liver (with increased glucose production), muscle (with decreased glucose disposal), and adipose tissue (failure of insulin to suppress lipolysis) as important sites of insulin resistance in patients with NAFLD.<sup>7</sup>

The processes that govern hepatic triacylglycerol (TAG) accumulation are complex and represent a balance between synthesis and utilization. Nonesterified fatty acids (NEFAs), largely from adipose tissue, are delivered to the liver via the portal circulation and are esterified to form TAG. While this is the largest contributor to the hepatic TAG pool, the synthesis of fatty acids from nonlipid precursors (most commonly glucose) in the process of *de novo* lipogenesis (DNL) becomes increasingly important in established NAFLD.<sup>8,9</sup> Based on these observations, there is significant clinical interest in targeting DNL as a potential therapeutic strategy in the treatment of NAFLD.<sup>10</sup>

The sodium-glucose cotransporter type 2 (SGLT2) inhibitors are a recently licensed class of antidiabetic agents that promote glucosuria through decreased renal glucose reabsorption, which in turn improves insulin sensitivity and glycemic control.<sup>11,12</sup> In addition, the EMPA-REG study has demonstrated the positive impact that these agents may have on cardiovascular outcomes.<sup>13</sup> SGLT2 inhibitors have been shown to be effective in several rodent models of NAFLD. Using a choline-deficient diet to recapitulate some of the histological features of the NAFLD while maintaining normoglycemia, 5 weeks of SGLT2 inhibition led to significant reductions in hepatic TAG content and improved markers of liver fibrosis.<sup>14</sup> Similarly, 4 weeks of treatment in obese mice with T2D reduced hepatic steatosis and markers of liver oxidative stress.<sup>15</sup> Data from human studies are more limited, but recent randomized control studies have demonstrated that SGLT2 inhibitors reduce liver fat and improve liver function tests

in patients with T2D and NAFLD.<sup>16–19</sup> A small uncontrolled prospective study has suggested that SGLT2 inhibition may improve liver histology in NAFLD patients with T2D.<sup>20</sup>

Studies investigating the impact of glucose-lowering therapies to treat NAFLD are often challenging to interpret due to confounding factors that include the use of additional medications (e.g. insulin, thiazolidinediones [TZDs], glucagon-like peptide-1 [GLP-1] analogues, or dipeptidyl peptidase 4 [DPP-4] inhibitors) and changes in weight alongside improvements in glycemic control in patients with T2D. To address some of these issues, we have performed an experimental medicine study using gold-standard noninvasive techniques to assess the impact of SGLT2 inhibition in treatment-naïve, overweight, and obese individuals without T2D. Our aim was to determine if SGLT2 inhibition might represent a valid therapeutic strategy in these individuals, as well as identify the mechanisms by which these agents might be acting to regulate hepatic steatosis *in vivo*.

## Methods

**Study design and participants.** We performed a single-center, open-label trial of 12 weeks of therapy using the SGLT2 inhibitor dapagliflozin at 10 mg administered once daily in treatment-naïve, insulin-resistant, overweight/obese patients at risk of NAFLD. Participants were aged 18–70 years; had a body mass index (BMI) between 25 and 45 kg/m<sup>2</sup>; did not have T2D (HbA1C 32–48 mmol/mol); and were not taking medications known to alter hepatic, blood glucose, or weight. Participants consuming excess alcohol (females >14 units/week and males >21 units/week) or with comorbid liver disease were excluded. Patients were excluded if they self-reported significant attempts to lose weight within the last 3 months or if they had enrolled in any medically or commercially available weight reduction programs. The clinical study protocol received full ethical approval from the North of Scotland Research Ethics Committee (REC ref. 15/NS/0117) and local institutional review board. All adult subjects gave informed written consent prior to participation.

**Clinical protocol.** Baseline investigations included the assessment of hepatic TAG content using magnetic resonance spectroscopy (MRS) and a two-step hyperinsulinemic euglycemic clamp incorporating the use of stable isotope tracers. Imaging and metabolic assessments were performed within 1 week, after which time, treatment with 10 mg dapagliflozin once daily was started. All investigations were repeated after 12 weeks of treatment, after which all treatment was stopped. Metabolic equivalent of task (MET) minutes of exercise were calculated for the week before baseline and follow-up investigations using the

international physical activity questionnaire (IPAQ). Average daily energy requirements were also calculated for the week before baseline and follow-up investigations using data from a food diary self-completed by participants. All authors had access to the study data and reviewed and approved the final manuscript.

**Magnetic resonance protocol.** Hepatic steatosis was quantified using localized cardiac-triggered proton spectroscopy.<sup>21</sup> NAFLD was defined by steatosis >5.6%, which represents the 95th percentile of hepatic TAG content across the general population.<sup>22</sup> Hepatic water and iron content were quantified using T1 and T2\* mapping, respectively, and an “iron-corrected T1” (cT1) calculated as surrogate marker of inflammation and fibrosis.<sup>21</sup>

**De novo lipogenesis.** Participants were provided with deuterated water (<sup>2</sup>H<sub>2</sub>O) to drink the evening before and throughout the two-step hyperinsulinemic euglycemic clamp. Incorporation of <sup>2</sup>H from the <sup>2</sup>H<sub>2</sub>O into very low-density lipoprotein (VLDL) TAG-palmitate was used to determine hepatic DNL.

### Two-step hyperinsulinemic euglycemic clamp.

Arterialized blood was sampled to determine the blood glucose concentration to be maintained (clamped) throughout the study. Infusions of U-2,2 D2-glucose, and [U-<sup>13</sup>C]palmitic acid (to label the plasma NEFA pool) were then commenced, and steady-state blood samples were taken after 2 h of this “basal phase.” Two phases of insulin were then infused: 2 h at “low dose” (20 mU/m<sup>2</sup>/min) and then 2 h at “high-dose” (100 mU/m<sup>2</sup>/min). The rate of a simultaneous infusion of 20% glucose enriched with 2,2 D2-glucose was titrated at 5-min intervals to maintain fasting glycemic levels. Steady-state blood was sampled at the end of each phase. Rates of hepatic endogenous glucose production (EGP) and glucose disposal (Gd) were calculated using modified versions of the Steele Equations.<sup>23</sup>

**Fatty acid oxidation.** Breath samples were collected at half hourly intervals throughout the clamp, and the generation of <sup>13</sup>CO<sub>2</sub> (from the infused <sup>13</sup>C-palmitate) was measured as a reflection of global fatty acid oxidation.

## Results

Ten patients (five male and five female) were enrolled in the study. The mean age was 50.8 ± 0.8 years, and all were either overweight or obese with a mean baseline BMI of 32.8 ± 1.2 kg/m<sup>2</sup>. At baseline, 8 of 10 participants had non-alcoholic fatty liver as defined by >5.6% steatosis on MRS. All participants completed the study protocol, and no significant adverse events were reported.

**Body composition and anthropometry.** Mean total body weight decreased by 4.5 ± 1.4 kg (−4.63%, *P* < 0.05) due to a reduction in both fat mass (0.9 ± 2.8 kg, NS) and lean mass (2.3 ± 2.1 kg, *P* < 0.01). Total body water also decreased by 1.6 ± 1.1 kg (*P* < 0.05). Systolic blood pressure reduced by 6 ± 7 mmHg (*P* = 0.57), and diastolic blood increased by 4 ± 2 mmHg (*P* = 0.31). Waist circumference remained

unchanged. There were no significant differences in self-reported caloric intake and exercise levels in the week preceding baseline and follow-up investigations as measured by a 7-day food diary and short-form IPAQ, respectively (Table 1).

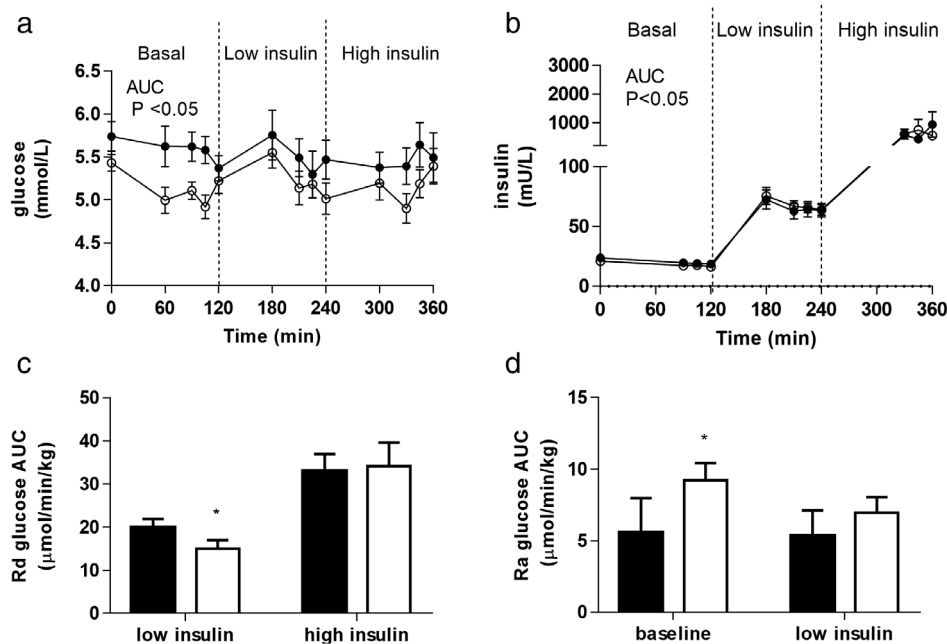
**Table 1** Study characteristics, magnetic resonance parameters, and fasting plasma biochemistry in 10 insulin-resistant, obese/overweight individuals before and after 12 weeks of SGLT2 inhibition with dapagliflozin 10 mg once daily

	Pre-dapagliflozin	Post-dapagliflozin
Subject characteristics		
Gender (F/M)	5/5	5/5
Age (years)	50.8 ± 0.8	50.8 ± 0.8
Weight (kg)	95.1 ± 4.2	90.7 ± 3.9*
BMI (kg/m <sup>2</sup> )	32.8 ± 1.2	31.3 ± 1.3**
Body fat (%)	35.1 ± 2.9	34.2 ± 3.2
Absolute fat mass (kg)	33.8 ± 10.3	31.3 ± 10.9*
Total body water (kg)	44.9 ± 8.7	43.3 ± 7.6*
Lean mass (kg)	61.6 ± 3.7	59.3 ± 3.3**
HbA1c (mmol/mol)	39.3 ± 2.5	36.9 ± 1.7
Blood pressure (mmHg)	135 ± 7 (systolic)	129 ± 6 (systolic)
	77 ± 3 (diastolic)	81 ± 2 (diastolic)
Waist circumference (cm)	107.0 ± 3.5	107.7 ± 4
MET minutes of exercise a week	35 208 ± 2895.7	5289.7 ± 2570
Average daily energy consumption (kcal)	2558.3 ± 318	2698 ± 308
Magnetic resonance imaging		
Hepatic steatosis (%)	12.9 ± 2.9	13.2 ± 2.7
Intrahepatic iron (mg/g)	1.0 ± 0.04	1.0 ± 0.04
Iron-corrected T1 (cT1) (ms)	811.2 ± 21.9	813.1 ± 19.5
Fasting plasma biochemistry		
ALT (U/L)	35.0 ± 4.8	27.3 ± 1.9
Glucose (mmol/L)	5.7 ± 0.2	5.4 ± 0.1
NEFA (μmol/L)	624 ± 79	613 ± 76
Glycerol (mU/L)	58.7 ± 12.0	53.6 ± 9.4
Triglyceride (mmol/L)	1.1 ± 0.1	1.1 ± 0.2
3OHB (μmol/L)	141.1 ± 44.7	151.8 ± 25.3
Urea (mmol/L)	5.2 ± 0.4	5.6 ± 0.3
ApoB (g/L)	0.93 ± 0.05	0.93 ± 0.06
Total cholesterol (mmol/L)	4.8 ± 0.3	4.7 ± 0.3
HDL cholesterol (mmol/L)	1.2 ± 0.1	1.2 ± 0.1
Insulin (mU/L)	23.7 ± 2.8	21.0 ± 2.3*
HOMA-IR	5.1 ± 0.7	4.1 ± 0.5*

\**P* < 0.05, \*\**P* < 0.01 pre- versus post-dapagliflozin.

Data are presented as mean ± standard error.

3OHB, 3-hydroxy-butyrate; ALT, alanine aminotransferase; ApoB, apolipoprotein B; BMI, body mass index; HbA1c, hemoglobin A1c; HDL, high-density lipoprotein; HOMA-IR, homeostatic model assessment of insulin resistance; MET, metabolic equivalent of task; NEFA, nonesterified fatty acid.



**Figure 1** Plasma glucose (a) and plasma insulin (b) levels across a two-step hyperinsulinemic euglycemic clamp before (filled circles) and after (open circles) 12 weeks of treatment with the sodium-glucose cotransporter 2 inhibitor, dapagliflozin. Glucose disposal (Rd glucose) during the low-dose insulin infusion decreased following dapagliflozin treatment but was unchanged during high-dose insulin (c). Basal glucose production rates (Ra glucose) increased after 12 weeks of dapagliflozin treatment but were not different during low-dose insulin infusion (d) (data presented are mean  $\pm$  standard error of mean; \* $P < 0.05$ , pre-dapagliflozin = black bars, post-dapagliflozin = open bars). AUC, area under curve.

**Peripheral insulin sensitivity.** Although fasting glucose levels were unchanged following 12 weeks of treatment, fasting insulin levels decreased, as did the homeostatic model assessment of insulin resistance (HOMA-IR) (Table 1). Glucose and insulin levels across the clamp were the same before and after treatment (Fig. 1a,b). Glucose disposal (Rd glucose) following low-dose insulin infusion decreased following treatment indicative of impaired insulin action. However, after high-dose insulin, there was no difference in Rd glucose before or after treatment (Fig. 1c).

**Hepatic insulin sensitivity and TAG content.** Alanine aminotransferase (ALT) decreased, but this did not reach statistical significance ( $P = 0.06$ ) (Table 1). Basal glucose production rate (Ra glucose) increased ( $5.6 \pm 2.4$  vs  $9.2 \pm 1.2$   $\mu\text{mol/min/kg}$ ,  $P < 0.05$ ), but there was no difference in the ability of low-dose insulin to suppress Ra glucose (Fig. 1d).

There were no differences in fasting total circulating TAG or VLDL-TAG before or after treatment. Both decreased across the clamp, and this suppression was unaffected by dapagliflozin (Fig. 2a,b). Incorporation of  $^{13}\text{C}$ -palmitate into VLDL-TAG reflecting re-esterification was not different before or after treatment (Fig. 3c). Incorporation of  $^2\text{H}$  in VLDL-TAG-palmitate (reflecting the contribution of DNL to the hepatic lipid pool) was also unaltered by dapagliflozin (Fig. 2d).

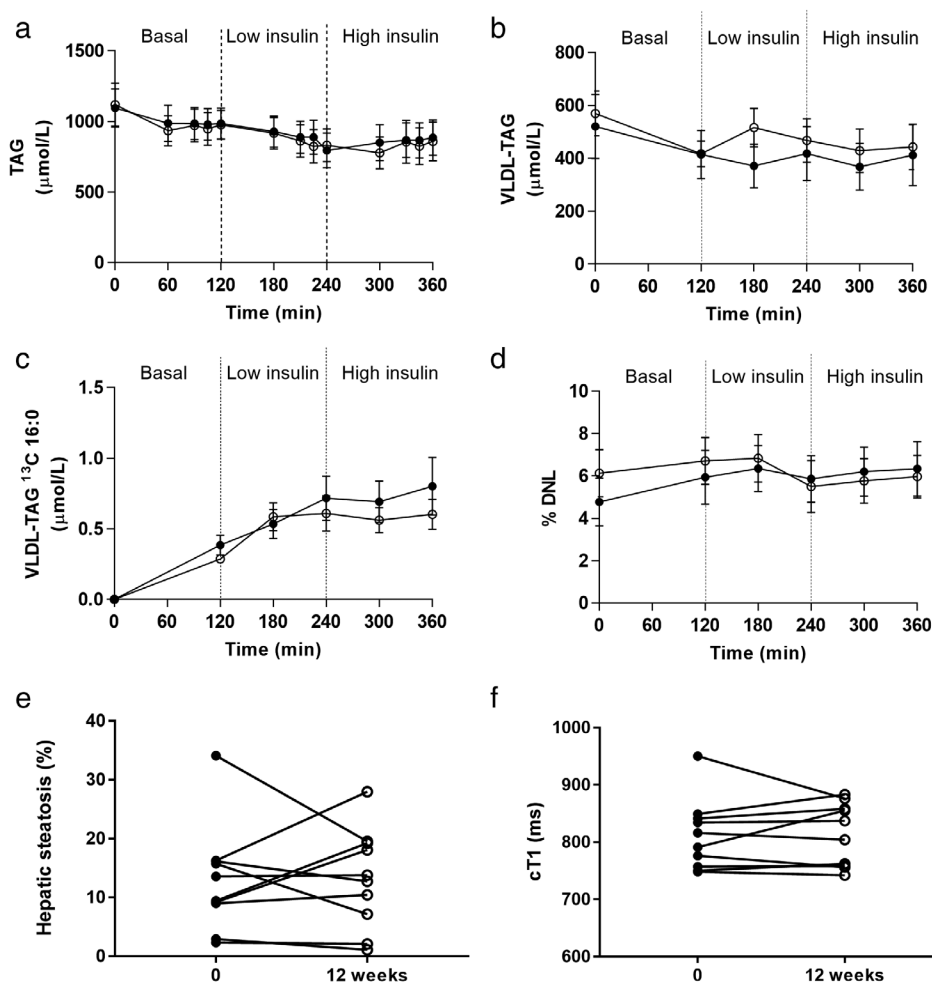
**Magnetic resonance imaging and spectroscopy.** Mean fat fraction at baseline was  $12.9 \pm 2.9\%$  compared with  $13.2 \pm 2.7\%$  after 12 weeks of treatment ( $P = 0.7$ ). Mean hepatic

iron content ( $1 \pm 0.04$  vs  $1 \pm 0.04$  mg/g) and whole liver cT1 values ( $811.2 \pm 21.9$  vs  $813.1 \pm 19.5$  ms,  $P = 0.9$ ) remained unchanged pre- and posttreatment (Fig. 2e,f).

**Fatty acid oxidation.** Total body oxidation of fatty acid was measured by the appearance of  $^{13}\text{CO}_2$  in the breath, and this increased over time. There was no difference in  $^{13}\text{CO}_2$  appearance rate before or after dapagliflozin treatment (Fig. 3a). 3-hydroxy-butyrate (3OHB) is an end-product of fatty acid oxidation. As expected, insulin suppressed 3OHB generation across the clamp, but this was unaffected by dapagliflozin (Fig. 3b).

**Circulating NEFA and glycerol.** Fasting circulating NEFA levels were not different before and after dapagliflozin treatment (Table 1). In addition, dapagliflozin did not alter the ability of insulin to suppress circulating NEFA levels during the hyperinsulinemic clamp (Fig. 3c). However, although fasting glycerol levels were not different before and after treatment, across the clamp, there was enhanced insulin-mediated suppression of glycerol after dapagliflozin treatment, potentially reflecting enhanced adipose tissue insulin sensitivity [glycerol area under curve (AUC): low insulin  $30.3 \pm 5.5$  vs  $22.2 \pm 2.9$  mU/L.h,  $P < 0.06$ ; high insulin  $14.7 \pm 2.2$  vs  $9.5 \pm 1.9$  mU/L.h,  $P < 0.05$ , before vs after Dapagliflozin] (Fig. 3d,e).

A subgroup analysis was performed excluding the two participants without clinically significant hepatic steatosis ( $<5.6\%$  on MRS). The significant findings were unaltered by their exclusion.



**Figure 2** Plasma total TAG (a) and VLDL TAG (b) decreased across the hyperinsulinemic clamp (both  $P < 0.001$  effect of time). There was no impact of treatment with dapagliflozin. The incorporation of  $^{13}\text{C}$ -palmitate into VLDL-TAG (c) and  $^2\text{H}$  from  $^2\text{H}_2\text{O}$  into palmitate (d) did not change. Hepatic TAG content as measured by magnetic resonance spectroscopy (e) and cT1 values (f) did not change with dapagliflozin treatment. Data presented are mean  $\pm$  standard error of mean; pre-dapagliflozin = filled circles/bars, post-dapagliflozin = open circles/bars. DNL, de novo lipogenesis; TAG, triacylglycerol; VLDL, very low-density lipoprotein.

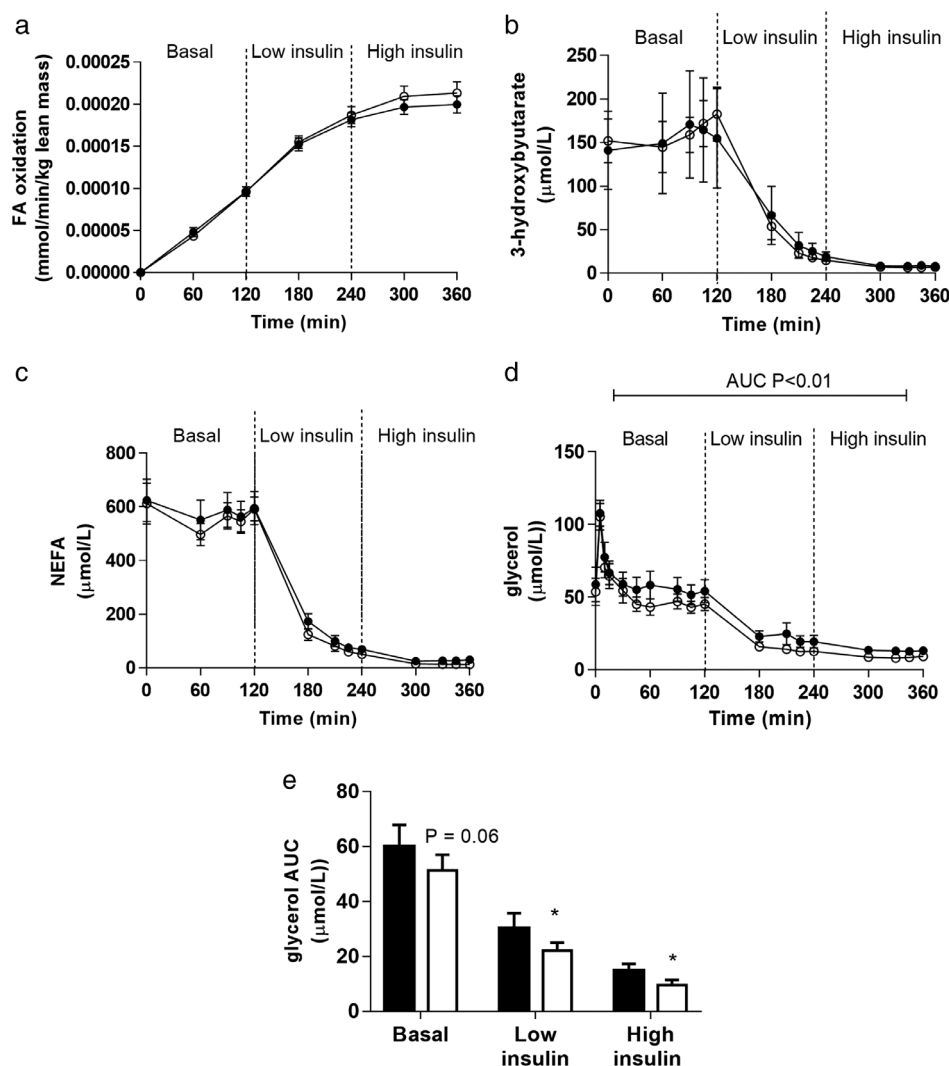
## Discussion

Using an experimental medicine approach incorporating currently accepted gold-standard noninvasive metabolic assessments in patients with NAFLD, we have assessed the impact of SGLT2 inhibition using dapagliflozin on the tissue-specific insulin-regulated processes that drive the accumulation of TAG within the liver. This is the first study to evaluate the effect of SGLT2 inhibition on NAFLD in patients without T2D and to assess hepatic steatosis using MRS.

We have shown that 12 weeks of treatment with dapagliflozin did not alter hepatic steatosis on MRS. In agreement with these data, we did not observe any significant change in the contribution of DNL to VLDL-TAG, suggesting that there would be no change in the contribution of DNL to hepatic TAG accumulation. Furthermore, we found no change in global measures of lipid oxidation or any beneficial impact on hepatic

insulin sensitivity. While we did observe enhanced suppression of glycerol following insulin infusion, there were no significant changes in NEFA, and therefore, it is likely that NEFA delivery to the liver with subsequent re-esterification to TAG would be unaltered. The apparent lack of effect on lipolysis in our study contrasts with data in patients with T2D where SGLT2 inhibition increased circulating NEFA and beta-hydroxybutyrate.<sup>18</sup> It has been postulated that increased lipolysis with SGLT2 inhibitors may be mediated through increased glucagon secretion,<sup>24</sup> although circulating glucagon levels were not measured in the current study.

The evidence base for the potential benefits of SGLT2 inhibition in the context of NAFLD treatment is growing. Recent randomized control data from the E-LIFT trial demonstrated reduced hepatic steatosis on MRS with empagliflozin.<sup>16</sup> Several groups have also recently demonstrated a benefit on liver fat fraction with dapagliflozin and canagliflozin.<sup>17–19</sup> However, another



**Figure 3** Whole-body fatty acid oxidation as measured by  $^{13}\text{CO}_2$  in breath samples from infused  $^{13}\text{C}$ -palmitate (a), hepatic fatty acid oxidation determined by 3-hydroxybutyrate (b), and plasma NEFA levels (c) were unaltered by treatment with dapagliflozin. While fasting plasma glycerol levels were the same pre- and post-dapagliflozin treatment, there was a more pronounced suppression of glycerol during the basal and low- and high-dose insulin infusions after dapagliflozin treatment (d and e). Data presented are mean  $\pm$  standard error of mean; pre-dapagliflozin = filled circles/bars, post-dapagliflozin = open circles/bars. AUC, area under curve; FA, fatty acid; NEFA, nonesterified fatty acid.

human study, which included an exploratory subgroup analysis using magnetic resonance imaging to look at changes in hepatic steatosis in patients with poorly controlled T2D, did not identify a significant effect.<sup>25</sup> All of these data, however, are derived from patients with comorbid T2D and NAFLD, with no dedicated studies, to our knowledge, conducted on NAFLD patients without T2D.

Although not statistically significant, ALT fell by 20% in the current study. This is in line with data from randomized trials of SGLT2 inhibitors in T2DM, which demonstrate similar reductions in liver enzymes independent of changes in weight and HbA1c.<sup>26</sup> This may suggest a liver-specific effect of SGLT2 inhibitors, although ALT has been shown to be a poor biomarker for hepatic steatosis.<sup>27</sup>

There are several studies investigating SGLT2 inhibitors in NAFLD that are actively recruiting (<https://clinicaltrials.gov/ct2/home>), although all are exclusively on patients with T2D. While rodents provide compelling evidence, including in NAFLD models with normoglycaemia,<sup>14,15</sup> there are limitations with the use of rodent models,<sup>28</sup> and therefore, undertaking detailed mechanistic studies using treatments that have been shown to be safe and well tolerated is warranted.

Glucose is the major substrate for DNL in patients with T2D. Coexistent hepatic and global (skeletal muscle and adipose) insulin resistance results in unchecked NEFA delivery to the liver and, when combined with the impact on DNL, drives hepatic TAG accumulation. In patients with T2D, SGLT2 inhibitors, through their glycosuric effect of lowering blood glucose, would

therefore potentially decrease glucose delivery to the liver and limit DNL. In addition, any impact to enhance insulin sensitivity (and to promote weight loss) would decrease NEFA mobilization and delivery to the liver for re-esterification. However, it is important to note that up to 75% of patients with NAFLD do not have T2D<sup>6</sup> and therefore would not have access to medications licensed for the treatment of T2D, which have been shown to have histological benefit in NAFLD.<sup>29</sup>

There are perhaps several explanations regarding the lack of effect that we observed in our patients. Importantly, our patients did not have T2D, and we did not observe any change in blood glucose levels; this may be fundamental in understanding the lack of change in DNL. In patients with T2D, 2 weeks of treatment with SGLT2 inhibition enhanced peripheral glucose disposal but also increased EGP rate.<sup>30</sup> While it has been proposed that this may reflect the amelioration of glucotoxicity as a consequence of renal glucose loss and the consequent improvement in glycemic control, the precise mechanisms underpinning this observation remain to be determined. Recent evidence has also suggested that SGLT2 inhibition can augment the actions of GLP-1,<sup>31</sup> and this mechanism may be important when considering any potential benefit to patients with NAFLD. In our study, we did observe a significant increase in Ra glucose in the basal state following SGLT2 inhibition, although suppression by low-dose insulin was not changed. Contrary to findings in patients with T2D, glucose disposal was decreased after low-dose insulin. The contribution of the pathological processes that drive NAFLD will differ according to the presence or absence of T2D, and therefore, it is likely that the response to therapy, (especially one that relies on glucose lowering) will also differ.

The changes that we observed in body composition and blood pressure are in line with previous published studies.<sup>32</sup> On average, patients lost 4.4 kg in weight (4.6%). Despite this, there was no improvement in hepatic steatosis, which is potentially at odds with lifestyle intervention studies in the context of NAFLD.<sup>33</sup> Furthermore, weight loss did not correlate with change in steatosis as measured by MRS. This may be due to the relatively modest decrease in fat mass and the parallel changes in lean mass and total body water. This contrasts with previous literature on T2DM patients where SGLT2 inhibitors are associated with reductions in total body fat.<sup>34</sup> Despite a reduction in total body water measured by bioimpedance, there was no significant reduction in hepatic free water content as measured by cT1. Therapies that are able to achieve larger changes, specifically in fat mass, are likely to be more efficacious in the treatment of NAFLD.

In conclusion, we have demonstrated that, in a small cohort of patients without T2D, SGLT2 inhibition was not able to alter the hepatic TAG content or the processes that are crucial to its pathogenesis. We do acknowledge, however, the marked variability in the individual responses of hepatic steatosis to SGLT2 inhibition and that interpretations may be limited by small sample size. Further studies are needed to determine if there is clinical improvement in NAFLD in patients with T2D or impaired glucose handling (fasting or postprandial) and to define the mechanisms by which this might occur. Overall, it seems unlikely that SGLT2 inhibition will be of significant benefit to those patients with NAFLD who have normoglycemia.

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