

# The clinical profile of NMOSD in Australia and New Zealand

Wajih Bukhari, MD,<sup>1\*</sup> Laura Clarke, MD,<sup>1,2\*</sup> Cullen O’Gorman, MD, PhD,<sup>1,2</sup> Elham Khalilidehkordi, MD,<sup>1,3</sup> Simon Arnett, MD,<sup>1,3</sup> Kerri M Prain,<sup>4</sup> Mark Woodhall, PhD,<sup>5</sup> Roger Silvestrini,<sup>6</sup> Christine S Bundell,<sup>7</sup> Sudarshini Ramanathan, MD,<sup>8</sup> David Abernethy, MD,<sup>9</sup> Sandeep Bhuta, MD,<sup>1</sup> Stefan Blum, MD, PhD,<sup>10</sup> Mike Boggild, MD,<sup>11</sup> Karyn Boundy, MD,<sup>12</sup> Bruce J Brew, MD,<sup>13</sup> Wallace Brownlee, MD,<sup>14</sup> Helmut Butzkueven, MD, PhD,<sup>15</sup> William M Carroll, MD,<sup>16</sup> Celia Chen, MD, PhD,<sup>17</sup> Alan Coulthard, MD, PhD,<sup>18</sup> Russell C Dale, MD, PhD,<sup>19</sup> Chandi Das, MD,<sup>20</sup> Keith Dear, PhD,<sup>21</sup> Marzena J Fabis-Pedrini, PhD,<sup>22</sup> David Fulcher, MD, PhD,<sup>23</sup> David Gillis, MD,<sup>18</sup> Simon Hawke, MD, PhD,<sup>23</sup> Robert Heard, MD,<sup>19</sup> Andrew P D Henderson, MD,<sup>24</sup> Saman Heshmat, MD,<sup>1</sup> Suzanne Hodgkinson, MD, PhD,<sup>25</sup> Sofia Jimenez-Sanchez,<sup>1</sup> Trevor J Kilpatrick, MD, PhD,<sup>26</sup> John King, MD,<sup>27</sup> Chris Kneebone, MD,<sup>11</sup> Andrew J Kornberg, MD,<sup>28</sup> Jeannette Lechner-Scott, MD, PhD,<sup>29</sup> Ming-Wei Lin, MD,<sup>23</sup> Christopher Lynch, MD,<sup>30</sup> Richard A L Macdonnell, MD,<sup>31</sup> Deborah F Mason, MD,<sup>32</sup> Pamela A McCombe, MD, PhD,<sup>33</sup> Jennifer Pereira, MD,<sup>30</sup> John D Pollard, MD, PhD,<sup>23</sup> Stephen W Reddell, MD, PhD,<sup>34</sup> Cameron Shaw, MD,<sup>35</sup> Judith Spies, MD, PhD,<sup>23</sup> James Stankovich, PhD,<sup>36</sup> Ian Sutton, MD, PhD,<sup>37</sup> Steve Vucic, MD, PhD,<sup>19</sup> Michael Walsh, MD,<sup>9</sup> Richard C Wong, MD,<sup>18</sup> Eppie M Yiu, MD, PhD,<sup>28</sup> Michael H Barnett, MD, PhD,<sup>34</sup> Allan G Kermode, MD,<sup>16</sup> Mark P Marriott, MD, PhD,<sup>15</sup> John Parratt, MD,<sup>23</sup> Mark Slee, MD, PhD,<sup>17</sup> Bruce V Taylor, MD,<sup>35</sup> Ernest Willoughby, MD,<sup>14</sup> Robert J Wilson,<sup>2</sup> Fabienne Brilot, PhD,<sup>7</sup> Angela Vincent, MD, FRS,<sup>4</sup> Patrick Waters, PhD,<sup>4</sup> Simon A Broadley, MD, PhD.<sup>1,3</sup>

\* = co-first authors

*Affiliations*

<sup>1</sup> Menzies Health Institute Queensland, Gold Coast Campus, Griffith University QLD 4222, AUSTRALIA

<sup>2</sup> Department of Neurology, Princess Alexandra Hospital, Woolloongabba QLD 4102, AUSTRALIA

<sup>3</sup> Department of Neurology, Gold Coast University Hospital, Southport QLD 4215, AUSTRALIA

<sup>4</sup> Department of Immunology, Pathology Queensland, Royal Brisbane and Women's Hospital, Herston QLD 4006, AUSTRALIA

<sup>5</sup> Nuffield Department of Clinical Neurosciences, John Radcliffe Infirmary, University of Oxford, Oxford OX3 9DU, UK

<sup>6</sup> Department of Immunopathology, Westmead Hospital, Westmead NSW 2145, AUSTRALIA

<sup>7</sup> School of Pathology and Laboratory Medicine, University of Western Australia, Nedlands WA 6009, AUSTRALIA

<sup>8</sup> Brain Autoimmunity Group, Institute for Neuroscience and Muscle Research, The Kids Research Institute at the Children's Hospital, Westmead, NSW 2145, AUSTRALIA

<sup>9</sup> Department of Neurology, Wellington Hospital, Newtown 6021, NEW ZEALAND

<sup>10</sup> Department of Neurology, Princess Alexandra Hospital, Woolloongabba QLD 4102, AUSTRALIA

<sup>11</sup> Department of Neurology, Townsville Hospital, Douglas QLD 4814, AUSTRALIA

<sup>12</sup> Department of Neurology, Royal Adelaide Hospital, Adelaide SA 5000, AUSTRALIA

- <sup>13</sup> Centre for Applied Medical Research, St Vincent's Hospital, University of New South Wales, Darlinghurst NSW 2010, AUSTRALIA
- <sup>14</sup> Department of Neurology, Auckland City Hospital, Grafton 1023, NEW ZEALAND
- <sup>15</sup> Melbourne Brain Centre, Royal Melbourne Hospital, University of Melbourne, Parkville VIC 3010, AUSTRALIA
- <sup>16</sup> Centre for Neuromuscular and Neurological Disorders, Queen Elizabeth II Medical Centre, University of Western Australia, Nedlands WA 6009, AUSTRALIA
- <sup>17</sup> Flinders Medical Centre, Flinders University, Bedford Park SA 5042, AUSTRALIA
- <sup>18</sup> School of Medicine, Royal Brisbane and Women's Hospital, University of Queensland, Herston QLD 4029, AUSTRALIA
- <sup>19</sup> The Children's Hospital at Westmead, Faculty of Medicine and Health, University of Sydney, Westmead NSW 2145 AUSTRALIA.
- <sup>20</sup> Department of Neurology, Canberra Hospital, Garran ACT 2605
- <sup>21</sup> Global Health Research Centre, Duke Kunshan University, Kunshan, Jiangsu, CHINA
- <sup>22</sup> Western Australian Neuroscience Research Institute, Queen Elizabeth II Medical Centre, University of Western Australia, Nedlands WA 6009, AUSTRALIA
- <sup>23</sup> Sydney Medical School, Royal Prince Alfred Hospital, University of Sydney, Camperdown NSW 2006, AUSTRALIA
- <sup>24</sup> Department of Neurology, Westmead Hospital, Westmead NSW 2145, AUSTRALIA
- <sup>25</sup> South Western Sydney Medical School, Liverpool Hospital, University of New South Wales, Liverpool NSW 2170, AUSTRALIA
- <sup>26</sup> Florey Institute of Neuroscience and Mental Health, University of Melbourne, Parkville VIC 3010, AUSTRALIA

<sup>27</sup> Department of Neurology, Royal Melbourne Hospital, Victoria 3010, AUSTRALIA

<sup>28</sup> School of Paediatrics, Royal Children's Hospital, University of Melbourne, Parkville VIC 3010, AUSTRALIA

<sup>29</sup> Hunter Medical Research Institute, University of Newcastle, New Lambton Heights NSW 2305, AUSTRALIA

<sup>30</sup> School of Medicine, University of Auckland, Grafton 1142, NEW ZEALAND

<sup>31</sup> Department of Neurology, Austin Health, Heidelberg VIC 3084, AUSTRALIA

<sup>32</sup> Department of Neurology, Christchurch Hospital, Christchurch 8140, NEW ZEALAND

<sup>33</sup> Centre for Clinical Research, Royal Brisbane and Women's Hospital, University of Queensland, Herston QLD 4029, AUSTRALIA

<sup>34</sup> Brain and Mind Research Institute, University of Sydney, Camperdown NSW 2006, AUSTRALIA

<sup>35</sup> School of Medicine, Deakin University, Waurn Ponds VIC 3217, AUSTRALIA

<sup>36</sup> Menzies Research Institute, University of Tasmania, Hobart TAS 7000, AUSTRALIA

<sup>37</sup> Department of Neurology, St Vincent's Hospital, Darlinghurst NSW 2010, AUSTRALIA

**Authorship**

DA, MHB, SBh, SBI, MBo, KB, BJB, SAB, MBr, WBr, HB, WMC, CC, AC, RCD, CD, KD, DG, SHa, RH, APDH, SHo, AGK, TJK, JK, CK, JL-S, CL, RALM, MPMa, DFM, PAMcC, CO'G, JPa, JPe, JDP, KMP, SWR, CS, MS, JSp, JSt, IS, BVT, AV, SV, MWa, PW, EW, RJW and RCW conceived and designed the study.

SA, SAB, WBU, CSB, LC, KD, MJF-P, DG, SHe, SJ-S, M-WL, KMP, RS, JSt, BVT, PW, RJW, MWO and EMY conducted the analyses.

WBU and LC prepared the initial draft and SA, MHB, BJB, FB, SAB, WMC, RCD, KD, MJF-P, DF, APDH, SHo, AJK, JL-S, M-WL, MPMa, PAMcC, MPMc, KMP, SR, RS, MS, BVT, AV, SV, MWa, PW, EW, RJW, RCW, MWO and EMY contributed to revisions.

All authors approved the final draft.

**Financial Disclosures Statement**

MHB has received honoraria for participation in advisory boards and travel sponsorship from Novartis, BioCSL, Genzyme and Biogen Idec.

MBo has received travel sponsorship and honoraria from Sanofi-Genzyme, Teva, Novartis, Biogen Idec and Roche.

BJB has received honoraria as a board member for GlaxoSmithKline, Biogen Idec, ViiV Healthcare and Merck Serono, has received speaker honoraria from ViiV Healthcare, Boehringer Ingelheim, Abbott, Abbvie, and Biogen Idec; has received travel sponsorship from Abbott and ViiV Healthcare, and has received research support funding from Eli Lilly, GlaxoSmithKline, ViiV Healthcare and Merck Serono.

SAB has received honoraria for attendance at advisory boards and travel sponsorship from Bayer-Scherring, Biogen-Idec, Merck-Serono, Novartis, and Sanofi-Genzyme, has received speakers honoraria from Biogen-Idec and Genzyme, is an investigator in clinical trials sponsored by Biogen Idec, Novartis and Genzyme, and was the recipient of an unencumbered research grant from Biogen-Idec.

HB has received honoraria for serving on scientific advisory boards for Biogen Idec, Novartis and Sanofi-Genzyme, has received conference travel sponsorship from Novartis and Biogen Idec, has received honoraria for speaking and acting as Chair at educational events organised by Novartis, Biogen Idec, Medscape and Merck Serono, serves on steering committees for trials conducted by Biogen Idec and Novartis, is chair (honorary) of the MSBase Foundation, which has received research support from Merck Serono, Novartis, Biogen Idec, Genzyme Sanofi and CSL Biopharma and has received research support form Merck Serono.

WMC has been the recipient of travel sponsorship from, and provided advice to, Bayer Schering Pharma, Biogen-Idec, Novartis, Genzyme, Sanofi-Aventis, BioCSL and Merck-Serono.

RCD has received research funding from the National Health and Medical Research Council, MS Research Australia, Star Scientific Foundation, Pfizer Neuroscience, Tourette Syndrome Association, University of Sydney, and the Petre Foundation and has received honoraria from Biogen-Idec and Bristol-Myers Squibb as an invited speaker

MjF-P has received travel sponsorship from Biogen Australia and New Zealand.

RH has received honoraria, educational support and clinic funding from Novartis, Biogen Idec, Genzyme and BioCSL.

AGK has received scientific consulting fees and/or lecture honoraria from Bayer, BioCSL, Biogen-Idec, Genzyme, Merck, Novartis, Sanofi-Aventis, and Teva.

TJK has received travel sponsorship from Novartis, BioCSL, Novartis, Merck Serono and Biogen Idec, has received speaker honoraria from Biogen Idec, BioCSL, Merck Serono, Teva, Genzyme and Novartis, has received research support from Biogen Idec, Genzyme, GlaxoSmithKline, Bayer-Schering and Merck Serono, and has received scientific consulting fees from GlaxoSmithKline China, Biogen-Idec and Novartis.

JK has received remuneration for advisory board activities and presentations from Bayer Healthcare, Biogen Idec, BioCSL, Genzyme and Novartis.

CK has received travel support, honoraria and advisory board payments from Biogen Idec, Bayer, Genzyme, Novartis and Serono.

JL-S has received unencumbered funding as well as honoraria for presentations and membership on advisory boards from Sanofi Aventis, Biogen Idec, Bayer Health Care, CSL, Genzyme, Merck Serono, Novartis Australia and Teva.

RALM has received honoraria for attendance at advisory boards and travel sponsorship from Bayer, Biogen, CSL, Merck, Novartis, Roche, Celgene, Teva and Genzyme.

MPMa has received travel sponsorship, honoraria, trial payments, research and clinical support from Bayer Schering, Biogen Idec, BioCSL, Genzyme, Novartis and Sanofi Aventis Genzyme.

DFM has received honoraria for attendance at advisory boards from Biogen-Idec and Novartis, and travel sponsorship from Bayer-Scherring, Biogen-Idec, and Sanofi-Genzyme.

PAMcC has received honoraria or travel sponsorship from Novartis, Sanofi-Aventis and Biogen Idec.

JAP has received travel sponsorship, honoraria for presentations and membership on advisory boards from Biogen Idec and Novartis and Sanofi Aventis.

JDP has received honoraria for seminars or advisory boards from Teva, Biogen, Sanofi-Genzyme, Novartis, Merck, Bayer and research grants or fellowships from Merck, Novartis, Bayer, Biogen, Sanofi-Genzyme and Teva

SWR has received travel sponsorship, honoraria, trial payments, research and clinical support from Aspreva, Baxter, Bayer Schering, Biogen Idec, BioCSL, Genzyme, Novartis, Sanofi Aventis Genzyme and Servier, and is a director of Medical Safety Systems Pty Ltd.

CPS has received travel sponsorship from Biogen Idec, Novartis and Bayer-Schering.

IS has received remuneration for Advisory Board activities from Biogen, CSL, and Bayer Schering and educational activities with Biogen, CSL and travel sponsorship from Biogen, Novartis and Bayer Schering.

MS has received research support from Novartis, Biogen Idec and BioCSL.

JSp has received honoraria for lectures and participation in advisory boards, and travel sponsorship from Novartis, BioCSL, Genzyme and Biogen Idec.

BVT has received travel sponsorship from Novartis and Bayer Schering.

AV and the University of Oxford hold patents and receive royalties for antibody testing.

PW and the University of Oxford hold patents for antibody assays and have received royalties, has received speaker honoraria from Biogen Idec and Euroimmun AG, and travel grants from the Guthy-Jackson Charitable Foundation.

EW has received honoraria for participation in advisory boards from Biogen-Idec and Novartis, travel sponsorship from Biogen-Idec, Bayer-Schering and Teva and is an investigator in clinical trials funded by Biogen-Idec and Teva.

DA, SBh, SBI, KB, MBr, WBr, WBu, CSB, CCM, LC, AC, CD, KD, DF, DG, SHa, APDH, SHe, SHo, SJ-S, AJK, M-WL, CL, CO'G, MPM, CS, RS, JSt, AV, SV, MWa, RJW, RCW, MWo and EMY report no disclosures

**Running Title:** ANZ NMOSD Clinical Features

**Corresponding author:** Professor Simon Broadley  
 School of Medicine  
 Gold Coast Campus  
 Griffith University QLD 4222  
 AUSTRALIA  
 Tel.: +61 7 5678 0702  
 Fax. +61 7 5678 0708  
 Email [simon.broadley@griffith.edu.au](mailto:simon.broadley@griffith.edu.au)

**Principal Investigator:** As above

Characters in title = 58 Characters in running title = 27

Main text word count = 2,487 Abstract word count = 250

Figures = 7 Tables = 6

References = 32

**Statistical Analysis:** Was performed by SAB under the supervision of Prof Keith Dear, Director of Graduate Studies in Global Health, Global Health Research Centre, Duke Kunshan University, Jiangsu, CHINA.

**Keywords:** Neuromyelitis optica; Clinical features; Multiple sclerosis; Aquaporin; Autoimmune disease

**Study Funding:** Multiple Sclerosis Research Australia (11-038)  
 Brain Foundation  
 Griffith University/Gold Coast Hospital Foundation  
 Contributing to Australian Scholarship and Science  
 NHS National Specialised Commissioning Group for NMO

NIHR Oxford Biomedical Research Centre

**ABSTRACT**

Neuromyelitis optica spectrum disorders (NMOSD) are an inflammation of the central nervous system associated with autoantibodies to aquaporin-4. We have undertaken a clinic-based survey of NMOSD in the Australia and New Zealand populations with the aim of characterising the clinical features and establishing the value of recently revised diagnostic criteria. Cases of possible NMOSD and age and sex-matched controls with multiple sclerosis (MS) were referred from centres across Australia and New Zealand. Cases were classified as NMOSD if they met the 2015 IPND criteria and remained as suspected NMOSD if they did not. Clinical and paraclinical data were compared across the three groups. NMOSD was confirmed in 75 cases and 89 had suspected NMOSD. There were 101 controls with MS. Age at onset, relapse rates and EDSS scores were significantly higher in NMOSD than in MS. Lesions and symptoms referable to the optic nerve were more common in NMOSD whereas brainstem, cerebellar and cerebral lesions were more common in MS. Longitudinally extensive spinal cord lesions were seen in 48/71 (68%) of cases with NMOSD. Elevations of CSF white cell count and protein were more common in NMOSD. We have confirmed a clinical pattern of NMOSD that has been seen in several geographical regions. We have demonstrated the clinical utility of the current diagnostic criteria. Distinct patterns of disease are evident in NMOSD and MS, but there remains a large number of patients with NMOSD-like features who do not meet the current diagnostic criteria for NMOSD and remain a diagnostic challenge.

## Introduction

Neuromyelitis optica spectrum disorders (NMOSD) encompasses clinical presentations associated with antibodies to aquaporin-4 (AQP4), a water channel expressed on astrocytes [1]. Without treatment NMOSD is frequently a disabling, sometimes fatal condition, with high annualised relapse rates and poor recovery after each attack. Studies suggest that the treatment response in NMOSD is quite distinct to that seen in multiple sclerosis (MS) with NMOSD being highly steroid responsive and to some extent steroid dependent [2]. Acute treatment with plasma exchange appears to be particularly helpful in NMOSD. Several immunomodulatory treatments for MS ( $\beta$ -interferon, fingolimod and natalizumab) may worsen NMOSD [3-5] and alemtuzumab has shown no discernible benefit in a small number of cases of NMOSD [6]. NMOSD appears to be responsive to immunosuppressive therapy and anti-B-cell therapies (e.g. rituximab, inebilizumab) [7,8]. More recently treatment efficacy with anti-IL6 receptor (tocilizumab) [9] and anti-complement component 5 (eculizumab) [10] monoclonal antibodies has been demonstrated. Due to this prognostic and therapeutic divergence between NMOSD and MS it is important to recognise NMOSD early.

Since the discovery of pathogenic antibodies that are present in the majority of patients with NMSOD, the disease has been more clearly defined and the clinical phenotype has expanded to include cases of area postrema dysfunction, brainstem disease, hypothalamic dysfunction, encephalopathic presentations and cerebral lesions in addition to the originally described optic neuritis and longitudinally extensive spinal cord disease [11]. The most recent diagnostic criteria for NMOSD

emphasise the value of positive AQP4 antibodies in patients with a core clinical presentation, but also accommodate cases where AQP4 antibodies are negative or unknown [11]. There are few studies of cohorts of patients with NMOSD as defined by the 2015 International Panel for NMO Diagnosis (IPND) diagnostic criteria [11]. We have therefore undertaken a clinical survey of NMOSD across 23 central nervous system (CNS) demyelination clinics in Australia and New Zealand with the aim of outlining the clinical profile of the disease in this region and evaluating the new diagnostic criteria. We have also compared the NMOSD patients with a cohort of contemporaneously recruited patients with classical MS and the cases of suspected NMOSD who did not meet the 2015 IPND criteria.

## **Methods**

### ***Data Collection***

Cases of “possible NMOSD” were identified by clinicians experienced in the diagnosis of inflammatory CNS disease at 23 adult and paediatric clinics covering the major urban centres of Australia and New Zealand. Possible NMOSD was determined using the clinical and para-clinical features of NMOSD as defined in the 2006 Wingerchuk criteria as being features associated with a high risk for NMOSD (Table 1) [12]. Age and sex-matched cases with MS were also recruited from the same locations. Cases were collected between 1 January 2011 and 31 December 2013. Demographic and clinical features were collected for all cases and controls using a questionnaire designed to provide information regarding the patient’s age, gender, ethnic ancestry, age at onset, relapse profile, associated medical conditions,

past history of glandular fever, smoking history, family history of demyelinating disease, medication history, disease course and current symptoms. Symptom severity was rated using a self-reported four-point scale of 0 = symptom not present; 1 = intermittent or mild symptoms; 2 = persistent symptoms that are moderate in severity; and 3 = severe symptoms that significantly impact activities of daily living. Expanded disability status scale (EDSS) scoring was undertaken by a neurologist experienced in the assessment of demyelinating disease was used to assess disability. Cases were defined retrospectively as “NMOSD” if they met the 2015 IPND criteria (incorporating AQP4 antibody results) [11], “suspected NMOSD” if they were referred as possible NMOSD but did not meet these criteria and “MS” if they were referred as MS, met the 2010 McDonald criteria [13] and were seronegative for AQP4 antibodies. Institutional ethics approval for this study was obtained for all participating centres and all patients gave written, informed consent.

### ***Paraclinical Information***

Where available the results of cerebrospinal fluid (CSF) analysis, visual evoked potentials (VEP) and prior laboratory and radiological screening for potential NMOSD mimics and associated autoimmune disease were also recorded. DICOM formatted files of magnetic resonance (MR) imaging of the brain and spine were obtained wherever possible and downloaded to a central reading centre using eFilm® software (IBM Watson Health®, US). Images were reviewed using RadiForce® MX270W high resolution screens (EIZO®, California USA). MR images were reviewed by a single blinded assessor with access available for review by a second blinded assessor in cases of uncertainty. Final decisions were made by consensus.

### ***Serological Testing***

Testing for AQP4 antibodies was performed using standard immunofluorescence techniques in all cases and in a subset using a combination of ELISA and cell-based assays for AQP-4 as previously described [14]. All testing was performed blind to the clinical status of the patient.

### ***Statistical Analysis***

Frequencies are expressed as n/N (%) and continuous data are presented as median (range) if not normally distributed or mean (SD) if normally distributed. Simple comparisons have been made using appropriate parametric or non-parametric tests for categorical and continuous data. As comparisons were purely exploratory, no correction for multiple testing has been undertaken.

## **RESULTS**

### ***Diagnostic characterization***

In total, 296 patients were recruited for inclusion in the study. Of these 179 had possible NMOSD and 117 had MS. Of the NMOSD cases 15/179 (8%) were excluded because of insufficient clinical data (9), study inclusion criteria not met (3), no serum sample available (1), withdrawal from study (1) or were given an alternative diagnosis (1 - Leber's hereditary optic neuropathy). Of the remaining 164 cases,

75/164 (46%) met the 2015 IPND criteria for a diagnosis of NMOSD (68/75 [91%] were AQP4 seropositive) [11], leaving 89 cases with suspected NMOSD. Of the MS controls 16/117 (14%) were excluded, leaving 101 controls. Reasons for exclusion in this group were insufficient clinical data (6) and no serum sample (10). All of the suspected NMOSD cases and MS controls were negative for AQP4 antibodies. The final distribution of cases and MS controls is summarised in Figure 1. MR imaging of the brain was available for 255/265 (96%) and of the spine in 239/265 (90%) cases and controls. There were only 4 instances where lack of appropriate imaging may have resulted in failure for a possible NMOSD case to be categorised as seronegative NMOSD.

### ***Demographic and Clinical Features***

The principal demographic and clinical features of NMOSD cases, suspected NMOSD and MS controls are shown in Table 2. There was no statistically significant difference in the current age and gender ratio for NMOSD and MS, indicating that the matching of MS controls to cases was successful. Median age of onset was higher in NMOSD than in MS ( $p=0.001$ ) and correspondingly disease duration was shorter ( $p<0.001$ ). The distribution of age of onset in the three groups is shown in Figure 2 which shows a flattened distribution and extended tail in the higher age ranges for NMOSD. Onset prior to the age of 10 years was not seen in this NMOSD cohort.

Mean annualised relapse rates and median EDSS at last assessment were both higher in NMOSD than in MS, despite the shorter disease duration. The distribution of EDSS scores is illustrated in Figure 3. A greater relative frequency of EDSS

scores of 4.0 and 6.0-6.5 was apparent for the NMOSD group. Clinical course was not significantly different between the groups, but as has been previously noted a secondary progressive course was rare in NMOSD (2%) and primary progressive disease was not seen [15]. There was no difference in the frequency of relapse types, reflecting the considerable overlap between NMOSD and MS. However, as has been previously noted there was a trend to more frequent episodes of optic neuritis and transverse myelitis in NMOSD and more brainstem and other relapse types in MS (Table 2). Initial MR imaging of the brain was more likely to be normal (not meeting Paty criteria) [16] in NMOSD compared to MS, but this was only the case in 17% of cases (Table 2). Longitudinally extensive spinal cord lesions were seen in 48/71 (68%) cases with NMOSD and 1/89 (1%) in MS ( $p < 0.001$ ). The longitudinally extensive spinal cord lesion seen in the MS case had the appearance of coalesced short segment lesions, but was included as it met our definition of continuous central cord lesion extending over 3 or more vertebral segments and was read as such by our blinded MR imaging readers. There were no significant differences in clinical features observed between seropositive and seronegative NMOSD cases (Table 3), but as has been previously reported there was a trend towards older age of onset and a higher frequency of longitudinally extensive spinal cord lesions in the antibody positive group [17].

Symptom severity (none, mild, moderate or severe) at the time of assessment is shown in Figure 4 and indicates that a relatively greater burden of disease in NMOSD was seen in the visual, sensory and motor (particularly lower limb) domains. Brainstem symptoms were more common in MS but were not frequently seen in any group. Complaints of fatigue and cognitive symptoms were similar in the three

groups. The profile of visual acuities is shown in Figure 5. Visual acuities were 6/12 or worse in 31% of eyes in NMOSD and 20% in MS, and counting fingers only (CFO) or worse in 11% of eyes in NMOSD and 0.6% in MS, despite a shorter disease duration in NMOSD. Suspected NMOSD fell between these two extremes with 25% of eyes 6/12 or worse and 2% CFO or worse.

### ***Relapse Data***

A total of 960 relapses were documented across all three groups over a total of 2015 patient-years. The pattern of relapses in the three groups is shown in Table 4. As has been previously noted, episodes of optic neuritis were more common in NMOSD ( $p<0.001$ ) while brainstem/cerebellar ( $p<0.001$ ) and cerebral relapses ( $p=0.009$ ) were more common in MS [18]. Suspected NMOSD had a relapse profile that resembles NMOSD more than MS, reflecting the recruitment criteria. Relapses in NMOSD were more likely to be treated with intravenous methylprednisolone than relapses in MS ( $p<0.001$ ), possibly reflecting greater relapse symptom severity in NMOSD.

The types of relapse encountered at first presentation for cases with NMOSD are given in Table 5. Initial presentations with transverse myelitis were the most common (44%), followed by optic neuritis (38%). Simultaneous optic neuritis and transverse myelitis was only seen in 2/75 (2%) of initial presentations of NMOSD. Area postrema syndrome was the initial presentation in 7/75 (9%).

### ***CSF and VEP Results***

Results of CSF analysis are summarised in Figure 6. CSF white cell count and protein levels were higher in NMOSD compared to MS ( $p < 0.001$ ), with suspected NMOSD falling in between. Evidence of local synthesis of oligoclonal bands in CSF was more common in MS than in NMOSD or suspected NMOSD and systemic synthesis of oligoclonal bands was seen in a small proportion of NMOSD and suspected NMOSD cases. P100 latency on VEP was significantly longer in MS compared to NMOSD and evidence of bilateral delay was more common in MS than in NMOSD (Figure 7).

### ***Co-occurrence of autoimmune disease***

The frequency of co-occurring autoimmune diseases is shown in Table 6. Other autoimmune diseases were more commonly seen in NMOSD than in MS or suspected NMOSD, but these differences were not statistically significant. Although the numbers are small they do suggest a specific association for NMOSD with systemic lupus erythematosus and Sjögren's syndrome and for MS with psoriasis and similar specific associations have been noted previously [19-21].

### **Discussion**

This comprehensive clinical survey of NMOSD across Australia and New Zealand using the most recent diagnostic criteria included 39% of the estimated 193 cases of NMOSD identified in our recent prevalence study [22]. This is therefore likely to be a representative sample. This cohort was identified on the basis of clinical features

previously described as being predictive of NMOSD thereby removing the potential bias of identifying cases through seropositive status alone. This methodology increased the likelihood of identifying seronegative cases. It is noteworthy that there were a large number of suspected cases of NMOSD referred (n=89) with at least one typical NMO feature that did not meet the new diagnostic criteria for NMOSD and all were negative for AQP4 antibodies.

The clinical features observed in this NMOSD cohort are similar to those previously described in other populations from a variety of geographical locations [23-26]. As with other studies performed using the 2015 IPND diagnostic criteria from NMOSD [18,25] the proportion of seropositive cases was high at 91% compared to studies using earlier criteria (27 – 88%) [27,28]. Noting that any criteria that include the AQP4 antibody test as a key component of disease classification will result in high sensitivity for the antibody, the high sensitivity noted in studies using the 2015 IPND criteria suggests that the new diagnostic criteria are appropriately permitting only seronegative cases with two typical presentations and supportive paraclinical features to be included as NMOSD.

As with previous studies [29,18,30] this cohort demonstrates an older age of onset, higher relapse rates, higher EDSS scores, higher CSF white cell counts and protein levels, lower frequency of oligoclonal bands in CSF, less delayed P100 latency on VEP and a higher frequency of co-occurring autoimmune diseases in NMOSD compared to MS. For some clinical features (age at onset, annualized relapse rate, EDSS, CSF white cell count, CSF protein, VEP results) suspected NMOSD cases were intermediate between NMOSD and MS but others were closer to NMSOD

(frequency of oligoclonal bands). The profile of symptoms and relapse locations, with optic neuritis being more common in NMOSD and brainstem, cerebellar and cerebral lesions being more common in MS, is not surprising and is consistent with prior studies [29,18,30]. The frequency of longitudinally extensive spinal cord lesions was most common in NMOSD and was a little lower in suspected NMOSD. Initial MR imaging of the brain was most likely to be normal in cases of suspected NMOSD. These conflicting patterns for cases of suspected NMOSD suggests that they constitute an admixture of pathologies.

The numbers of seronegative NMOSD in the present study were too small to provide statistically significant differences in comparison to seropositive cases. Other studies from the pre-2015 IPND diagnostic criteria, have suggested a number of differences between seronegative and seropositive cases of NMOSD [31].

Strengths of the present study are that we have used the 2015 IPND diagnostic criteria to define cases of NMOSD and that cases have been identified from across Australia and New Zealand using clinical features only to identify potential cases. One drawback in this approach is that only clinical features identified in the 2006 Wingerchuk criteria, which were the criteria at the time of participant recruitment, were used to identify cases and therefore the more recently identified typical presentations of NMOSD (area postrema, acute brainstem, diencephalic and cerebral syndromes) may be under-represented. However, the frequency of area postrema syndrome as a first relapse in our cohort (9%) was similar to that identified in a recently reported figure from a large international database (7.1 – 10.3%) [32].

In summary, this clinically ascertained cohort of NMOSD patients confirms a clinical profile of disease that is consistent with other studies and has been replicated across many geographical locations. New diagnostic criteria are both sensitive and specific for NMOSD and therefore represent a practical way to ascertain cases for clinical studies and will assist diagnosis in routine clinical practice. Distinctly different profiles of clinical features between NMOSD and MS are of considerable assistance in identifying potential cases, but a significant number of cases with NMOSD-like features are antibody negative and do not meet current clinical criteria for a diagnosis of NMOSD, indicating that there is a persistent grey-zone between NMOSD and MS.

## Figure Legends

### Figure 1

Distribution of cases and controls as defined by diagnostic criteria used in this study.

NMOSD = neuromyelitis optica spectrum disorders

MS = multiple sclerosis

AQP4 = aquaporin-4 antibodies

+ve = positive

-ve = negative

### Figure 2

Age at onset distribution of NMOSD cases, suspected NMOSD and MS controls expressed as relative frequency (percentage) within each diagnostic group.

NMOSD = neuromyelitis optica spectrum disorders

Susp NMOSD = suspected NMOSD

MS = multiple sclerosis

### Figure 3

Distribution of EDSS scores at last assessment for NMOSD cases, suspected NMOSD and MS controls expressed as relative frequency (percentage) within each diagnostic group.

NMOSD = neuromyelitis optica spectrum disorders

Susp NMOSD = suspected NMOSD

MS = multiple sclerosis

**Figure 4**

Symptom severity for NMOSD cases, suspected NMOSD and MS controls expressed as relative frequency (percentage) within each symptom cluster for each clinical group.

NMOSD = neuromyelitis optica spectrum disorders

Susp NMOSD = suspected NMOSD

MS = multiple sclerosis

**Figure 5**

Visual acuity measurements in NMOSD cases, suspected NMOSD and MS controls expressed as relative frequency (percentage) within each clinical group. Visual acuities recorded according to metric Snellen chart for individual eyes. Number of eyes examined: NMOSD = 136, Susp NMOSD = 164; MS = 154.

NMOSD = neuromyelitis optica spectrum disorders

Susp NMOSD = suspected NMOSD

MS = multiple sclerosis

NPL = no perception of light

LPO = light perception only

HMO = hand movements only

CFO = counting fingers only

**Figure 6**

Summary of CSF examination in NMOSD cases, suspected NMOSD and MS controls showing (A) box and whisker plot of CSF white cell counts (cells per mm<sup>3</sup>), (B) box and whisker plot of CSF protein (g/L) and (C) bar chart of frequency

(percentage) of local, systemic, negative or unknown oligoclonal bands results.

Numbers (N) of examinations are indicated above each chart. Box and whisker plots shows median (solid bar), interquartile range (boxes) and range (whiskers).

NMOSD = neuromyelitis spectrum disorders

Susp NMOSD = suspected NMOSD

MS = multiple sclerosis

CSF = cerebrospinal fluid

### **Figure 7**

Summary of VEP results for NMOSD cases, suspected NMOSD and MS controls showing (A) box and whisker plot of VEP P100 latency (ms) and (B) bar chart of relative frequency (percentage) of bilateral delay, unilateral delay or normal VEP within each clinical group. Numbers (N) of examinations are indicated above each chart. Box and whisker plot shows median (solid bar), interquartile range (boxes) and range (whiskers).

NMOSD = neuromyelitis optica spectrum disorders

Susp NMOSD = suspected NMOSD

MS = multiple sclerosis

VEP = visual evoked potential

## References

1. Pittock SJ, Lucchinetti CF (2016) Neuromyelitis optica and the evolving spectrum of autoimmune aquaporin-4 channelopathies: a decade later. *Ann N Y Acad Sci* 1366 (1):20-39. doi:10.1111/nyas.12794
2. Trebst C, Jarius S, Berthele A, Paul F, Schippling S, Wildemann B, Borisow N, Kleiter I, Aktas O, Kumpfel T, Neuromyelitis Optica Study G (2014) Update on the diagnosis and treatment of neuromyelitis optica: recommendations of the Neuromyelitis Optica Study Group (NEMOS). *J Neurol* 261 (1):1-16. doi:10.1007/s00415-013-7169-7
3. Palace J, Leite MI, Nairne A, Vincent A (2010) Interferon Beta treatment in neuromyelitis optica: increase in relapses and aquaporin 4 antibody titers. *Arch Neurol* 67 (8):1016-1017. doi:67/8/1016 [pii] 10.1001/archneurol.2010.188
4. Jacob A, Hutchinson M, Elson L, Kelly S, Ali R, Saukans I, Tubridy N, Boggild M (2012) Does natalizumab therapy worsen neuromyelitis optica? *Neurology* 79 (10):1065-1066. doi:10.1212/WNL.0b013e31826845fe
5. Kira J, Itoyama Y, Kikuchi S, Hao Q, Kurosawa T, Nagato K, Tsumiyama I, von Rosenstiel P, Zhang-Auberson L, Saida T (2014) Fingolimod (FTY720) therapy in Japanese patients with relapsing multiple sclerosis over 12 months: results of a phase 2 observational extension. *BMC Neurol* 14:21. doi:10.1186/1471-2377-14-21
6. Kowarik MC, Hoshi M, Hemmer B, Berthele A (2016) Failure of alemtuzumab as a rescue in a NMOSD patient treated with rituximab. *Neurol Neuroimmunol Neuroinflamm* 3 (2):e208. doi:10.1212/NXI.0000000000000208

7. Huang W, Wang L, Zhang B, Zhou L, Zhang T, Quan C (2019) Effectiveness and tolerability of immunosuppressants and monoclonal antibodies in preventive treatment of neuromyelitis optica spectrum disorders: A systematic review and network meta-analysis. *Multiple sclerosis and related disorders* 35:246-252. doi:10.1016/j.msard.2019.08.009
8. Cree BAC, Bennett JL, Kim HJ, Weinshenker BG, Pittock SJ, Wingerchuk DM, Fujihara K, Paul F, Cutter GR, Marignier R, Green AJ, Aktas O, Hartung HP, Lublin FD, Drappa J, Barron G, Madani S, Ratchford JN, She D, Cimbora D, Katz E, investigators NMs (2019) Inebilizumab for the treatment of neuromyelitis optica spectrum disorder (N-MOmentum): a double-blind, randomised placebo-controlled phase 2/3 trial. *Lancet*. doi:10.1016/S0140-6736(19)31817-3
9. Araki M, Matsuoka T, Miyamoto K, Kusunoki S, Okamoto T, Murata M, Miyake S, Aranami T, Yamamura T (2014) Efficacy of the anti-IL-6 receptor antibody tocilizumab in neuromyelitis optica: a pilot study. *Neurology* 82 (15):1302-1306. doi:10.1212/WNL.0000000000000317
10. Pittock SJ, Berthele A, Fujihara K, Kim HJ, Levy M, Palace J, Nakashima I, Terzi M, Totolyan N, Viswanathan S, Wang KC, Pace A, Fujita KP, Armstrong R, Wingerchuk DM (2019) Eculizumab in Aquaporin-4-Positive Neuromyelitis Optica Spectrum Disorder. *N Engl J Med* 381 (7):614-625. doi:10.1056/NEJMoa1900866
11. Wingerchuk DM, Banwell B, Bennett JL, Cabre P, Carroll W, Chitnis T, de Seze J, Fujihara K, Greenberg B, Jacob A, Jarius S, Lana-Peixoto M, Levy M, Simon JH, Tenenbaum S, Traboulsee AL, Waters P, Wellik KE, Weinshenker BG, International Panel for NMOSD (2015) International consensus diagnostic criteria for neuromyelitis optica spectrum disorders. *Neurology* 85 (2):177-189. doi:10.1212/WNL.0000000000001729

12. Wingerchuk DM, Lennon VA, Pittock SJ, Lucchinetti CF, Weinshenker BG (2006) Revised diagnostic criteria for neuromyelitis optica. *Neurology* 66 (10):1485-1489. doi:66/10/1485 [pii]  
10.1212/01.wnl.0000216139.44259.74
13. Polman CH, Reingold SC, Banwell B, Clanet M, Cohen JA, Filippi M, Fujihara K, Havrdova E, Hutchinson M, Kappos L, Lublin FD, Montalban X, O'Connor P, Sandberg-Wollheim M, Thompson AJ, Waubant E, Weinshenker B, Wolinsky JS (2011) Diagnostic criteria for multiple sclerosis: 2010 revisions to the McDonald criteria. *Ann Neurol* 69 (2):292-302. doi:10.1002/ana.22366
14. Prain K, Woodhall M, Vincent A, Ramanathan S, Barnett MH, Bundell CS, Parratt JDE, Silvestrini RA, Bukhari W, The Australian and New Zealand NMO Collaboration, Brilot F, Waters P, Broadley SA (2019) AQP4 antibody assay sensitivity comparison in the era of the 2015 diagnostic criteria for NMOSD. *Front Neurol* 10:1028
15. Wingerchuk DM, Pittock SJ, Lucchinetti CF, Lennon VA, Weinshenker BG (2007) A secondary progressive clinical course is uncommon in neuromyelitis optica. *Neurology* 68 (8):603-605. doi:10.1212/01.wnl.0000254502.87233.9a
16. Paty DW, Oger JJ, Kastrukoff LF, Hashimoto SA, Hooge JP, Eisen AA, Eisen KA, Purves SJ, Low MD, Brandeys V (1988) MRI in the diagnosis of MS: a prospective study with comparison of clinical evaluation, evoked potentials, oligoclonal banding, and CT. *Neurology* 38 (2):180-185
17. Yang CS, Zhang DQ, Wang JH, Jin WN, Li MS, Liu J, Zhang CJ, Li T, Shi FD, Yang L (2014) Clinical features and sera anti-aquaporin 4 antibody positivity in patients with demyelinating disorders of the central nervous system from Tianjin, China. *CNS Neurosci Ther* 20 (1):32-39. doi:10.1111/cns.12156

18. Srikajon J, Siritho S, Ngamsombat C, Prayoonwiwat N, Chirapapaisan N, Siriraj Neuroimmunology Research G (2018) Differences in clinical features between optic neuritis in neuromyelitis optica spectrum disorders and in multiple sclerosis. *Mult Scler J Exp Transl Clin* 4 (3):2055217318791196. doi:10.1177/2055217318791196
19. Pittock SJ, Lennon VA, de Seze J, Vermersch P, Homburger HA, Wingerchuk DM, Lucchinetti CF, Zephir H, Moder K, Weinshenker BG (2008) Neuromyelitis optica and non organ-specific autoimmunity. *Arch Neurol* 65 (1):78-83. doi:65/1/78 [pii]
- 10.1001/archneurol.2007.17
20. Jarius S, Jacobi C, de Seze J, Zephir H, Paul F, Franciotta D, Rommer P, Mader S, Kleiter I, Reindl M, Akman-Demir G, Seifert-Held T, Kristoferitsch W, Melms A, Wandinger KP, Wildemann B (2011) Frequency and syndrome specificity of antibodies to aquaporin-4 in neurological patients with rheumatic disorders. *Mult Scler* 17 (9):1067-1073. doi:10.1177/1352458511403958
21. Shahmohammadi S, Doosti R, Shahmohammadi A, Mohammadianinejad SE, Sahraian MA, Azimi AR, Harirchian MH, Asgari N, Naser Moghadasi A (2019) Autoimmune diseases associated with Neuromyelitis Optica Spectrum Disorders: A literature review. *Multiple sclerosis and related disorders* 27:350-363. doi:10.1016/j.msard.2018.11.008
22. Bukhari W, Prain KM, Waters P, Woodhall M, O'Gorman CM, Clarke L, Silvestrini RA, Bundell CS, Abernethy D, Bhuta S, Blum S, Boggild M, Boundy K, Brew BJ, Brown M, Brownlee WJ, Butzkueven H, Carroll WM, Chen C, Coulthard A, Dale RC, Das C, Dear K, Fabis-Pedrini MJ, Fulcher D, Gillis D, Hawke S, Heard R, Henderson APD, Heshmat S, Hodgkinson S, Jimenez-Sanchez S, Killpatrick T, King J, Kneebone C, Kornberg AJ, Lechner-Scott J, Lin MW, Lynch C, Macdonell R, Mason

- DF, McCombe PA, Pender MP, Pereira JA, Pollard JD, Reddel SW, Shaw C, Spies J, Stankovich J, Sutton I, Vucic S, Walsh M, Wong RC, Yiu EM, Barnett MH, Kermod AG, Marriott MP, Parratt JDE, Slee M, Taylor BV, Willoughby E, Wilson RJ, Vincent A, Broadley SA (2017) Incidence and prevalence of NMOSD in Australia and New Zealand. *J Neurol Neurosurg Psychiatry*. doi:10.1136/jnnp-2016-314839
23. Adoni T, Lino AM, da Gama PD, Apostolos-Pereira SL, Marchiori PE, Kok F, Callegaro D (2010) Recurrent neuromyelitis optica in Brazilian patients: clinical, immunological, and neuroimaging characteristics. *Mult Scler* 16 (1):81-86. doi:10.1177/1352458509353651
24. Eskandarieh S, Nedjat S, Azimi AR, Moghadasi AN, Sahraian MA (2017) Neuromyelitis optica spectrum disorders in Iran. *Multiple sclerosis and related disorders* 18:209-212. doi:10.1016/j.msard.2017.10.007
25. Houzen H, Kondo K, Niino M, Horiuchi K, Takahashi T, Nakashima I, Tanaka K (2017) Prevalence and clinical features of neuromyelitis optica spectrum disorders in northern Japan. *Neurology* 89 (19):1995-2001. doi:10.1212/WNL.0000000000004611
26. Hyun JW, Jeong IH, Joung A, Kim SH, Kim HJ (2016) Evaluation of the 2015 diagnostic criteria for neuromyelitis optica spectrum disorder. *Neurology* 86 (19):1772-1779. doi:10.1212/WNL.0000000000002655
27. Pandit L, Kundapur R (2014) Prevalence and patterns of demyelinating central nervous system disorders in urban Mangalore, South India. *Mult Scler* 20 (12):1651-1653. doi:10.1177/1352458514521503
28. Jacob A, Panicker J, Lythgoe D, Elson L, Mutch K, Wilson M, Das K, Boggild M (2013) The epidemiology of neuromyelitis optica amongst adults in the Merseyside

county of United Kingdom. *J Neurol* 260 (8):2134-2137. doi:10.1007/s00415-013-6926-y

29. Chen H, Liu SM, Zhang XX, Liu YO, Li SZ, Liu Z, Dong HQ (2016) Clinical Features of Patients with Multiple Sclerosis and Neuromyelitis Optica Spectrum Disorders. *Chin Med J (Engl)* 129 (17):2079-2084. doi:10.4103/0366-6999.189046

30. Jurynczyk M, Craner M, Palace J (2015) Overlapping CNS inflammatory diseases: differentiating features of NMO and MS. *J Neurol Neurosurg Psychiatry* 86 (1):20-25. doi:10.1136/jnnp-2014-308984

31. Jarius S, Ruprecht K, Wildemann B, Kuempfel T, Ringelstein M, Geis C, Kleiter I, Kleinschnitz C, Berthele A, Brettschneider J, Hellwig K, Hemmer B, Linker RA, Lauda F, Mayer CA, Tumani H, Melms A, Trebst C, Stangel M, Marziniak M, Hoffmann F, Schippling S, Faiss JH, Neuhaus O, Ettrich B, Zentner C, Guthke K, Hofstadt-van Oy U, Reuss R, Pellkofer H, Ziemann U, Kern P, Wandinger KP, Bergh FT, Boettcher T, Langel S, Liebetrau M, Rommer PS, Niehaus S, Munch C, Winkelmann A, Zettl UU, Metz I, Veauthier C, Sieb JP, Wilke C, Hartung HP, Aktas O, Paul F (2012) Contrasting disease patterns in seropositive and seronegative neuromyelitis optica: A multicentre study of 175 patients. *J Neuroinflammation* 9:14. doi:10.1186/1742-2094-9-14

32. Shosha E, Dubey D, Palace J, Nakashima I, Jacob A, Fujihara K, Takahashi T, Whittam D, Leite MI, Misu T, Yoshiki T, Messina S, Elson L, Majed M, Flanagan E, Gadoth A, Huebert C, Sagen J, Greenberg BM, Levy M, Banerjee A, Weinshenker B, Pittock SJ (2018) Area postrema syndrome: Frequency, criteria, and severity in AQP4-IgG-positive NMOSD. *Neurology* 91 (17):e1642-e1651. doi:10.1212/WNL.0000000000006392