

## Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

### Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- The exact sample size ( $n$ ) for each experimental group/condition, given as a discrete number and unit of measurement
- A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided  
*Only common tests should be described solely by name; describe more complex techniques in the Methods section.*
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- For null hypothesis testing, the test statistic (e.g.  $F$ ,  $t$ ,  $r$ ) with confidence intervals, effect sizes, degrees of freedom and  $P$  value noted  
*Give  $P$  values as exact values whenever suitable.*
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's  $d$ , Pearson's  $r$ ), indicating how they were calculated

*Our web collection on [statistics for biologists](#) contains articles on many of the points above.*

### Software and code

Policy information about [availability of computer code](#)

Data collection	N/A
Data analysis	Biacore 8K Control Software(version 3.0.12.15655) Biacore Insight Evaluation(version 3.0.12.15655) SoftMax Pro (version 7.0) GraphPad Prism software (version 10) SnapGene (6.1.1) ChimeraX (1.6.1) CryoSPARC (v3.3.1) Coot (0.9.4.1) CCP-EM (1.5.0)

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio [guidelines for submitting code & software](#) for further information.

## Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our [policy](#)

*Provide your data availability statement here.*

## Research involving human participants, their data, or biological material

Policy information about studies with [human participants or human data](#). See also policy information about [sex, gender \(identity/presentation\), and sexual orientation](#) and [race, ethnicity and racism](#).

Reporting on sex and gender	N/A
Reporting on race, ethnicity, or other socially relevant groupings	N/A
Population characteristics	N/A
Recruitment	N/A
Ethics oversight	N/A

Note that full information on the approval of the study protocol must also be provided in the manuscript.

## Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

- Life sciences     Behavioural & social sciences     Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://www.nature.com/documents/nr-reporting-summary-flat.pdf)

## Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	Sample size was determined based on common standards for animal, cell biology and biochemistry experiments, with three biological replicates. The number of animals used for each experiment was estimated based on previous and pilot studies in vivo assays. Normally, 6-8 mice for each group were analyzed to ensure the statistical significance. We performed more than three independent biological replicates for most of in vitro experiments.
Data exclusions	No data were excluded.
Replication	All experiments in this study had a minimum of three biological replicates.
Randomization	Mice were randomly grouped for different samples.
Blinding	N/A

## Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

## Materials &amp; experimental systems

n/a	Involvement
<input type="checkbox"/>	<input checked="" type="checkbox"/> Antibodies
<input type="checkbox"/>	<input checked="" type="checkbox"/> Eukaryotic cell lines
<input checked="" type="checkbox"/>	<input type="checkbox"/> Palaeontology and archaeology
<input type="checkbox"/>	<input checked="" type="checkbox"/> Animals and other organisms
<input checked="" type="checkbox"/>	<input type="checkbox"/> Clinical data
<input checked="" type="checkbox"/>	<input type="checkbox"/> Dual use research of concern
<input checked="" type="checkbox"/>	<input type="checkbox"/> Plants

## Methods

n/a	Involvement
<input checked="" type="checkbox"/>	<input type="checkbox"/> ChIP-seq
<input checked="" type="checkbox"/>	<input type="checkbox"/> Flow cytometry
<input checked="" type="checkbox"/>	<input type="checkbox"/> MRI-based neuroimaging

## Antibodies

## Antibodies used

## Antibodies used in HDX, negative staining and cryo-electron microscopy analysis:

Anti CD73 antibody: HB0038, in-house

Anti CD73 antibody: HB0039, in-house

## Antibodies used in animal experiments:

Anti CD73 antibody: HB0038, in-house

Anti CD73 antibody: HB0039, in-house

Anti CD73 antibody: HB0045, in-house

Anti CD73 antibody (AstraZeneca): Oleclumab, in-house

Anti 4-1BB antibody: HB0027, in-house

Anti-PD-L1/VEGF bispecific antibody: HB0025, in house

IgG control: in-house

## Antibodies used in SPR binding assay:

Anti CD73 antibody: HB0038, in-house

Anti CD73 antibody: HB0039, in-house

## Antibodies used in CD4+ or CD8+ T cell proliferation:

Anti CD73 antibody: HB0038, in-house

Anti CD73 antibody: HB0039, in-house

Anti CD73 antibody: HB0045, in-house

Anti CD73 antibody (AstraZeneca): Oleclumab, in-house

Ultra-LEAF™ Purified anti-human CD3 Antibody: Cat NO: 317326, 1 µg/mL, Bilegend

CD28 Monoclonal antibody, Cat NO: 14-0289-82, 1 µg/mL, Thermo Fisher Scientific

FITC anti-human CD4 Antibody, Cat NO: 317408, 1:100, Bilegend

APC anti-human CD4 Antibody, Cat NO: 317416, 1:200, Bilegend

APC/Cyanine7 anti-human CD4 Antibody, Cat NO: 317418, 1:100, Bilegend

Brilliant Violet 510™ anti-human CD8 Antibody, Cat NO: 344732, 1:100, Bilegend

APC anti-human CD8 Antibody, Cat NO: 344722, 1:200, Bilegend

Propidium iodide, Cat NO: P4170, 2 µg/mL, Sigma

## Antibodies used in intro CD73 binding, enzymatic inhibition activity:

Anti CD73 antibody: HB0038, in-house

Anti CD73 antibody: HB0039, in-house

Anti CD73 antibody: HB0045, in-house

Anti CD73 antibody (AstraZeneca): Oleclumab, in-house

Anti CD73 antibody (Corvus): Mupadolimab, in-house

Anti CD73 antibody (Akeso): AK119, in-house

Anti CD73 antibody: HB0038 Fab, in-house

Anti CD73 antibody: HB0039 Fab, in-house

IgG control: in-house

PE Goat anti-Human IgG, Fcy: # 109-115-098, 1:200, Jackson

## Antibodies used in antibody-mediated internalization:

Anti CD73 antibody: HB0038, in-house

Anti CD73 antibody: HB0039, in-house

Anti CD73 antibody: HB0045, in-house

IgG control: in-house

## Antibodies used in flow cytometric analysis of CD73 expression in CD4+ cell and CD8+ cell:

CD73 Monoclonal Antibody (AD2), PE, Cat NO: 12-0739-42, eBioscience™, Invitrogen

Mouse IgG1 kappa Isotype Control (P3.6.2.8.1), PE, eBioscience™, Invitrogen

Validation

HB0038, HB0039, HB0045, HB0027, and HB0025 were validated in this study or our previous work ([doi.org/10.3389/fimmu.2021.778978](https://doi.org/10.3389/fimmu.2021.778978)) by a number of approaches such as SPR binding assay and structural analyses. Oleclumab, Mupadolimab, and AK119 targeting CD73 were developed by AstraZeneca, Corvus, and Akeso, respectively. The rest of antibodies were validated by manufacturers.

## Eukaryotic cell lines

Policy information about [cell lines and Sex and Gender in Research](#)

Cell line source(s)

B-hCD73 MC38: CD73 humanized MC38 cell line B-hCD73 MC38 was prepared by Biocytogen Pharmaceutical (Beijing) Co. MDA-MB-231: MDA-MB-231 cells were purchased from Cell Bank, Chinese Academy of Science, catalog number: TCHu227 BxPC-3: BxPC-3 cells were purchased from Nanjing Cbioer Biosciences Co. LTD. catalog number: CBP60542 A375: A375 cells were purchased from Cell Bank, Chinese Academy of Science, catalog number: TCHu155 HepG2: HepG2 cells were purchased from the American Type Culture Collection (ATCC). CHO-K1-CD73-3F4: CHO-K1 cell line stably expressing full-length CD73 (CHO-K1-CD73-3F4) was made in-house. CHO-K1 was purchased from ATCC. Human PBMC: Human PBMC were purchased from Milestone (Shanghai) Biological Science & Technology Co. Ltd. (Reference number: PB010C)

Authentication

All these cell lines were authenticated by corresponding manufacturers.

Mycoplasma contamination

We routinely conducted mycoplasma contamination checks on cell lines utilized in this study, and the results consistently confirmed their negativity.

Commonly misidentified lines  
(See [ICLAC](#) register)

N/A

## Animals and other research organisms

Policy information about [studies involving animals; ARRIVE guidelines](#) recommended for reporting animal research, and [Sex and Gender in Research](#)

Laboratory animals

1. The B-hCD73/hCD39(v3) humanized C57BL/6 mice (CD73 and CD39 humanized C57BL/6) used in Supplemental Figure 1B-D; female, 7-8 weeks.
2. The immunodeficient Prkdcscid Il2rnull (NPG) mice used in Figure 5B-D, and Supplemental Figure 8A-E; female, 7-8 weeks.
3. The immunodeficient NOD-Prkdcem26Cd52Il2rgem26Cd22/NjuCrl (NCG) mice used in Figure 6A-D female, 9-10 weeks.
4. The immunodeficient NOD-Prkdcscid Il2rgem1/Cyagen (C-NKG) mice used in Figure 6E, F, female, 5-8 weeks.
5. The immunodeficient Prkdcscid Il2rnull (NPG) mice used in Supplemental Figure 8I-K; male, 5-8 weeks.

Wild animals

N/A

Reporting on sex

Female mice were used in all in vivo animal efficacy experiments. Compared with female mice, male mice are more aggressive and prone to fight. This has a negative impact on the animal's living conditions, and might also lead to tumor rupture in the later stages. This difference in survival status will indirectly affect the body weight and tumor volume of the mice. Therefore, we chose to use female mice for our in vivo animal efficacy experiments. Male mice were used in pharmacokinetics studies, this avoids the effects of changes in hormone levels on pharmacokinetics.

Field-collected samples

Our study did not involve field-collected samples.

Ethics oversight

Experiments complied with the relevant laws and institutional guidelines, as overseen by the Animal Studies Committee of the Association for Assessment and Accreditation of Laboratory Animal Care (AAALAC).

Note that full information on the approval of the study protocol must also be provided in the manuscript.

## Plants

Seed stocks

N/A

Novel plant genotypes

N/A

Authentication

N/A