

15 **ABSTRACT**

16 **Gastropods are commonly preserved as steinkerns (internal casts), a mode of**
17 **fossilization that leads to loss of external morphological features. This loss of**
18 **information is problematic for taxonomic identification and ecological inference in**
19 **evaluating assemblages where original shell material is not preserved. We seek to**
20 **quantify how closely gastropod steinkerns represent the morphology of their original**
21 **shells. We investigated this relationship experimentally by fabricating steinkerns in**
22 **silicone from modern gastropod shells and comparing their geometry to that of the**
23 **shells we used to create them. In addition to recording traces of ornamentation such as**
24 **ribs and spines, we used a theoretical morphospace framework to evaluate the fidelity**
25 **of shell-coiling parameters in steinkerns. Our results show that some morphotypes**
26 **reflect their taxonomic identification more accurately than others, indicating that**
27 **steinkern fidelity is highly variable. Experimental steinkerns consistently cluster less**
28 **reliably by morphotype than their original shell counterparts. Additionally, we find**
29 **that shell thickness is an important factor in determining steinkern fidelity. The fidelity**
30 **of the high-spired *Duplicaria duplicata*, for example, is significantly lower than the**
31 **average value for the morphotypes investigated whereas the fidelity of planispiral**
32 ***Haplotrema concavum* and open-coiling *Epitonium* is significantly higher, a trend**
33 **related to shell thickness. Thus, taxonomic identification and subsequent analyses, such**
34 **as community composition, of steinkern assemblages must recognize this differential**
35 **fidelity to counter preservational biases.**

36 **INTRODUCTION**

37 Morphology is a first-order line of evidence for paleontologists. The shapes of fossils reveal
38 evolutionary histories of ancient organisms and allow investigators to analyze form and function,
39 phylogenetic relationships, and preservational conditions. Yet fossil morphology is subject to
40 taphonomic bias, as clearly evidenced by two types of fossilization for gastropods: preservation
41 of the shell (biomineralized part), and preservation of internal casts called steinkerns, which
42 occurs when sedimentary matrix or precipitated material infiltrates a shell forming a cast that
43 remains when the shell is destroyed. We focus on gastropod steinkerns but other mollusks,
44 including bivalves and cephalopods, are also subject to steinkern-style preservation (Henderson
45 and McNamara 1985). Steinkerns have been used as an alternative to biomineralized gastropod
46 fossils to study a wide range of topics including assemblage composition (Harzhauser and Fehse
47 2007), evolutionary dynamics (Roy 1994), trace fossils (Vinn et al. 2014), ecology and ontogeny
48 (Nützel 2014), body size (Brayard et al. 2010), and taphonomic biases (Dattilo et al. 2016). A
49 full understanding of the nature of steinkerns is necessary to establish a complete picture of
50 gastropod paleoecology and evolution because the likelihoods of original shell and steinkern
51 preservation vary between depositional environments and are generally inversely correlated. This
52 differential preservation is controlled by a variety of factors including salinity (sulfate content),
53 bioturbation, aragonite saturation, energy, and rate of burial (Wright et al. 2018) and has the
54 potential to substantially impact evidence of diversity. Bush and Bambach's (2004) model of
55 aragonite dissolution bias, for example, showed that in 79 level-bottom Paleozoic assemblages,
56 alpha diversity may be underestimated by up to 29% due to the preferential dissolution of
57 aragonitic gastropods and other fossils composed of the mineral. Thus, steinkerns serve as
58 important evidence for interpreting community composition under conditions where original or

59 mineral-replaced shell material is not preserved, and systematic exclusion or misidentification of
60 steinkerns leads to bias (Cherns and Wright 2009).

61 Although gastropod steinkerns are common in shallow marine sediments and have been used
62 historically for identification and taxonomy (Knight 1941), they remain under-utilized due to the
63 challenges inherent in the loss of external features typically used for identification. When
64 gastropods are preserved as internal casts, details of their external morphology such as spines
65 and ribs are lost. Without experimental data it is not known to what extent traces of these
66 features are evident on a steinkern. Additionally, taphonomic trends vary by shell morphotype,
67 resulting in bias, but the nature and extent of this bias remains unexplored for most gastropods.
68 Small Cambrian conical shells (not necessarily gastropods), for example, are preferentially
69 preserved as phosphatic steinkerns (Creveling et al. 2014). The influence of morphotype on ease
70 of sediment infill results in distortions of relative abundance among ammonoid and nautiloid
71 steinkerns, a comparable example of the problem investigated here, although those shells contain
72 septa throughout the body chamber that most gastropods lack (Henderson and McNamara 1985;
73 Anderson and Allmon 2018). Thus, even though gastropods are the most diverse group of
74 mollusks with over 60,000 modern species and an extensive fossil record (Bouchet et al. 2017),
75 our understanding of their past diversity is compromised because the fidelity of key
76 morphological characteristics in steinkerns is unknown. Experimental approaches are crucial to
77 quantifying bias in steinkern assemblages.

78 In addition to biodiversity, gastropod ecology can be inferred from morphology (Pietsch et
79 al. 2021, Vermeij 2022). Experiments have shown that surface varices help *Ceratostoma*
80 *foliatum* land upright from a fall via hydrodynamic stabilization (Palmer 1977) and observations
81 of extant turritelliforms, i.e., “high-spined, many-whorled shells lacking an aperture elongated by

105 belonged to old, under-utilized collections with inadequate locality data and all individuals
106 selected are represented by multiples. In contrast to CT scanning, our experimental method has
107 the advantage of producing high-fidelity, physical 3D objects in a way that mechanically
108 emulates natural steinkern preservation (Fig. 2). Further, data collection from 2D photos of 3D
109 objects allows this method to be readily applied to fossil specimens (Figs. S1, S2). To sample
110 specimens with a wide range of morphological properties that might affect preservation
111 potential, we chose a broad but non-exhaustive list of morphotypes—morphologies with
112 stereotyped, generalizable coiling patterns (Table 1). Although there is no consistent set of
113 gastropod morphotypes defined in the literature, the morphotypes included in this study are
114 loosely based on those outlined by Tryon (1882). Shells were imaged with the coiling axis both
115 parallel (apertural view) and perpendicular (umbilical view) to the camera (Fig. 1). To create
116 silicone internal casts, a hole was made in the apex of each shell to allow silicone to fully
117 infiltrate the hollow space. The shells were put in cups filled with Anchor Seal tin-cured GI-1000
118 silicone molding rubber and placed in a vacuum chamber. After the silicone had set, the shells
119 were cut out of the silicone and the original shell material was dissolved using hydrochloric acid
120 thereby creating artificial steinkerns (here termed “silikerns”). The silikerns were subsequently
121 imaged in the same views as the original shells (Fig. 3).

122 Morphometric landmarks were placed on images of shells and silikerns at homologous points
123 on each whorl (along sutures for closed coiling geometries) using the ruler tool in Adobe
124 Photoshop (Fig. 1). Raw data were collected in apertural view to calculate translation and whorl
125 expansion rate, and in umbilical view to calculate distance from the coiling axis. Ratios of
126 landmark coordinates were used to calculate translation (T), distance from the coiling axis (D),
127 and whorl expansion rate (W) following Raup (1966):

128
$$T = (y_{\theta+2\pi} - y_{\theta} / x_{\theta+2\pi} - x_{\theta}) \quad (1)$$

129
$$D = a/b \quad (2)$$

130
$$W = (x_{\theta}/x_o)^{2\pi/\theta} \quad (3)$$

131 where y is vertical distance between apex and landmark, x is horizontal distance between apex
132 and landmark, a is distance between coiling axis and inner edge of whorl, and b is distance
133 between coiling axis and outer edge of whorl. Differences in measurements within individuals
134 were averaged over successive whorls to produce a single value for T , D , and W for each
135 specimen. This dataset was then plotted in 3D morphospace (sensu Raup 1966) to facilitate
136 analysis (Figs. S3, S4). We were not able to place landmarks on the original shell images of
137 Morphotype 12 (*Monetaria caputserpentis*) because only the final whorl was visible, so this
138 species was excluded from quantitative analysis. However, the artificial steinkern it produced
139 could be measured and its unusual morphology makes it an interesting case to demonstrate the
140 range of information that can be obtained from steinkerns.

141 The differences in T , D , and W between all shells and silikerns were quantified using the
142 Gower coefficient, which produces a square distance matrix. Two sets of analyses were
143 conducted on this distance matrix. The first tested the null hypothesis (H_0) that there were no
144 significant differences in silikern fidelity between the different morphotypes. To achieve this, the
145 distance separating each silikern from its source shell was isolated and randomly subsampled
146 with replacement 10,000 times, generating 10 silikern-shell distances each time to match the
147 number of silikerns for each morphotype in the dataset. These 10-distance subsamples were then
148 averaged, producing a null distribution of 10,000 silikern-shell distances. This was used to derive
149 the probability of randomly sampling 10 silikerns with equivalent or worse fidelity than the 10-
150 silikern samples of each morphotype (i.e., the p-value), defined as the proportion of random

151 subsamples with equivalent or greater average silikern-shell distances than each morphotype. We
152 then repeated this analysis to test for significant differences in silikern fidelity between genera.
153 However, as most of the genera we sampled belong to a unique morphotype, these results mostly
154 duplicate those of the previous analysis. As such, we do not discuss them (but see Figs. S5, S6).

155 The second set of analyses sought to quantify the impact of using casts instead of shells in
156 studies of fossil taxonomy, which usually rely on comparative anatomy to group
157 morphologically similar organisms into taxa. Agglomerative hierarchical clustering algorithms
158 apply a similar logic; at each step, they identify the pair of elements, which can be individual
159 points or clusters, separated by the shortest distance and group them together in a new cluster.
160 This process is repeated in a stepwise fashion until the entire dataset has been classified. When
161 applied to morphological distance matrices, these algorithms group taxa together based upon
162 their anatomical similarity, just as taxonomists do when classifying fossil taxa. As such, we
163 consider it reasonable to assume that anything that compromises the performance of hierarchical
164 clustering algorithms will also increase the likelihood of misclassification in taxonomic studies
165 restricted to morphological data.

166 To test the impact of this effect on casts versus shells, we separated the distance matrix into
167 two submatrices: one containing all silikern-silikern distances, the other containing all shell-shell
168 distances. 10,000 subsamples of the shell-shell submatrix, each of which contained 55 shells
169 (i.e., 50% of the total number), were randomly drawn without replacement to ensure the
170 dendrograms produced by the hierarchical clustering analysis contained equal numbers of tips.
171 Subsamples of the silikern-silikern submatrix were then drawn to match. Complete linkage
172 hierarchical clustering (also known as farthest neighbor clustering) was then used to analyze
173 these subsamples and the complete shell-shell submatrix. The result was 10,000 dendrograms

174 representing the silikern subsamples, 10,000 representing the shell subsamples, and a final
175 dendrogram representing all shells in the dataset that we used as our reference point for
176 evaluating how well subsamples of casts and shells preserve a clustering pattern. The difference
177 between the reference dendrogram and that of each subsample was then quantified using the
178 clustering information distance (CID), a reliable, efficient, and unbounded metric that quantifies
179 differences in branching order (Smith 2020a). The greater the CID, the greater the number of
180 topological discrepancies between the dendrograms, and so the greater the likelihood of
181 erroneous classification. The CID distributions of the two sets of subsamples were then
182 compared. These analyses were then repeated to test whether using casts differentially
183 compromised the ability of hierarchical clustering analyses to correctly group genera and
184 morphotypes. However, these results (Figs. S7, S8, S9) were not informative due to the
185 representation of most morphotypes by a single genus, and we do not discuss them. All analyses
186 were conducted in R (R Development Core Team 2022) using base R, Plotly (Plotly
187 Technologies, Inc.), vegan (Oksanen et al. 2022), ape (Paradis and Schliep 2019), and TreeDist
188 packages (Smith 2020b) functions (see Supplementary Information for further details).

189

190 Repositories and institutional abbreviations— Yale Peabody Museum Invertebrate Paleontology
191 (YPM IP), Yale Peabody Museum Invertebrate Zoology (YPM IZ).

192

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RESULTS

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Surface Features

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Our experiments provide evidence that surface traces on steinkerns can reflect external shell morphology. Two main types of ornamental features, spines and ridges, appear on silikerns as

197 three-dimensional structures (Fig. 2). Where a vertical row of three prominent spines is present
198 on one whorl of a *Siratus articulatus* specimen, the corresponding silikern shows only a small,
199 vertically oriented raised feature. Inflection points of the curvature of this feature represent
200 horizontal ridges on the original shell where each spine meets the body of the shell, so the
201 precise location of spines can be determined from the silikern (Fig. 2A). There is no evidence of
202 spines on whorls closer to the apex of the silikern, where spines are smaller on the original shell.
203 Observations of fossil steinkerns reveal analogous surface features, which indicate the presence,
204 general nature, and location of external ornamentation on the original shell, though, as in our
205 experimental silikern, this information is limited. The outermost whorls of *Sinistrofulgur*
206 *perversum* retain parallel grooves running perpendicular to the aperture (Fig. 2B). Although we
207 have not observed evidence of this feature on a fossil steinkern, it likely could be fossilized in a
208 similar manner to the spine rows. Previous observations of surface features on gastropod
209 steinkerns have yielded information on the morphology of the shell (Mackinnon 1985), but this
210 is the first experimental evidence linking steinkern features to external shell features.

211 Silikerns also reveal details beyond those of the original shell. Although Raup parameters
212 could not be determined from the original shell of *Monetaria caputserpentis*, silikerns reveal the
213 coiling pattern of this species (Fig. 3L). The columellar and labral teeth of the original shell are
214 reflected in grooves near the aperture of the silikern, and similar structures are evident on the
215 surface of some steinkerns (Fig. 2D).

216 Steinkern Fidelity

217 All three coiling parameters show differences in fidelity as revealed in the silikerns of different
218 morphotypes (Figs. 3, 4, S3, S4, S10). Of the 11 morphotypes tested, only Morphotype 2, the
219 high-spined *Duplicaria duplicata* (Fig. 3B), presents significantly greater silikern-shell distances

220 relative to the null distribution ($p < 0.0001$), indicating that silikerns of this morphotype have
221 particularly low coiling parameter fidelity (Fig. 5A). In contrast two morphotypes, numbers 6
222 (*Haplotrema concavum*, a nearly planispiral taxon, Fig. 3F) and 11 (*Epitonium*, an open-coiling
223 taxon, Fig. 3K), present significantly smaller silikern-shell distances ($p = 0.0110$ and 0.0077
224 respectively), demonstrating relatively high coiling parameter fidelity (Fig. 5A) (applying a
225 threshold of $p < 0.05$ to the number of random subsamples with equivalent or smaller silikern-
226 shell distances instead of greater). Although it does not breach the $p < 0.05$ threshold,
227 Morphotype 8 (*Siratus articulatus*, Fig. 3H) also demonstrates good fidelity, as only 529 of the
228 10,000 random subsamples present smaller average silikern-shell distances ($p = 0.0529$).

229 Impact on Hierarchical Clustering

230 The clustering information distance (CID) separating the silikern clustering from the reference
231 pattern is greater than that separating the shell clustering across all subsamples, indicating that
232 silikerns consistently introduce more error into hierarchical clustering analyses than shells (Fig.
233 5B). On average, the silikern CIDs are about three times greater than the shell CIDs, with means
234 of 32.7 and 10.3 and medians of 32.7 and 10.4 respectively.

235

236 DISCUSSION

237 The power of steinkerns to record evidence of external ornamentation and reveal ecological
238 and evolutionary information has been historically underestimated. Our experiments demonstrate
239 that surface marks on steinkerns can be used to infer the presence and nature of external features,
240 which are often used in taxonomic identification. Additionally, our experiments revealed the
241 coiling pattern of *Monetaria caputserpentis*, which is obscured by the external shell morphology
242 (Fig. 2D). Fossil gastropod phylogenetics are heavily based on morphology (Wagner 2001), so

243 knowledge of the internal structures of taxa with similarly obscured features would add
244 characters of phylogenetic significance. For example, a surface marking similar to that left by the
245 aperture of *Monetaria caputserpentis* on many specimens of *Cassidaria cheops* fossil steinkerns
246 may indicate that this species had an outer whorl similar to that of the cowrie (Fig. 2D). Also,
247 marks that we identify as traces from encrusters that colonized a shell after death were observed
248 on both silikerns and steinkerns (Fig. 2C). If they were indeed created through similar processes,
249 these marks on steinkerns may reveal the biostratigraphic history of the specimen. This kind of
250 internal information preserved on steinkerns may provide additional data in some cases.

251 Calculations of assemblage diversity derived from steinkerns almost certainly underestimate
252 true diversity. Silikerns performed worse than original shells for morphotype clustering using
253 coiling parameters (Fig. 5B). These results likely underestimate the error in using real steinkerns
254 as silikerns are very high-resolution casts, maximizing the detail preserved. This discrepancy
255 illustrates the need for a full understanding of the different fidelity of shells versus steinkerns of
256 different morphotypes. Morphotype 2 (represented by *Duplicaria duplicata*) displayed the lowest
257 fidelity of the morphotypes we investigated. Due to its thick shell, the whorls of this high-spired
258 shell morphotype are in contact in the original shell but not in our silikerns. Open-coiling
259 patterns also occur in high-spired fossil steinkerns (Fig. S11). This major morphological
260 difference between original shell and steinkern is reflected in their measured parameters, causing
261 the discrepancy. This experiment shows that an open-coiled steinkern does not necessarily reflect
262 an open-coiled shell (although some gastropods are open-coiled in life), and open-coiling in the
263 steinkern may represent a significant transition away from the morphology of the original shell
264 and therefore low fidelity.

265 Both morphotypes with the highest fidelity are thin-shelled: Morphotype 6 (*Haplotrema*
266 *concauum*) is close-coiled whereas Morphotype 11 (*Epitonium*) is open-coiled but in both cases
267 this morphology is revealed by the silikern. This pattern suggests that thicker shells relative to
268 whorl size (e.g., Morphotype 2) likely lead to less representative steinkerns as opposed to thinner
269 shells (e.g., Morphotypes 6 and 11). Shell morphology in coiling-axis view, used to calculate
270 distance from the coiling axis, is consistently well preserved in Morphotypes 6 and 11, which
271 may also contribute to their high fidelity. In summary, steinkern identification at the assemblage
272 level has great potential but must be conducted in the context of morphological fidelity to avoid
273 the bias inherent in steinkern-style preservation. Identification and morphological reconstruction
274 could also be enhanced by surface features on steinkerns, such as those on our silikerns, that
275 represent external features on the original shell. However, not all types of sculpture are likely to
276 be replicated on steinkerns, as this is influenced by how sculpture develops in different
277 gastropods and whether internal or external remodeling are involved (Anderson and Allmon
278 2018).

279 Modern gastropod shells are composed of aragonite or aragonite and calcite (Hall and
280 Kennedy 1967). Aragonite is preferentially and frequently lost due to dissolution, especially in
281 high-salinity environments, so much of the gastropod fossil record relies on steinkern-style
282 preservation (Cherns and Wright 2009). A relationship between morphotype and shell
283 composition has previously been reported in the Pennsylvanian of north-central Texas, where
284 calcified, low-spired euomphalids preserved as shells while aragonitic, medium-spired
285 pleurotomarian gastropods are preserved as steinkerns (Yochelson et al. 1967; Batten 1984;
286 Wagner and Erwin 2006). Furthermore, shifts in dominant aragonite microstructures (from
287 primarily nacreous to crossed-lamellar) that evolved during the Mesozoic Marine Revolution

288 may have contributed to the dominance of high-spined shells (Salinas and Kisailus 2013), but the
289 susceptibility of different aragonite microstructures to dissolution is unknown. Biases in
290 steinkern preservation owing to mineral composition and morphology create additional
291 challenges for investigators who seek to understand assemblage composition and test
292 macroevolutionary hypotheses. However, experiments can be used to understand bias and
293 identify potential identification errors resulting from a lack of external features, thereby
294 increasing the rigor of such interpretations.

295 Our results identify several areas for further exploration in the use of gastropod steinkerns for
296 ecological and evolutionary studies. We tested Raup's (1966) classic morphometric model which
297 provides a first-order description of shell shape but is not all-encompassing. It is likely that some
298 parameters are more useful for steinkern identification than others. Different approaches that
299 have since been used to quantify shell shape could also be applied to steinkern identification
300 (Urduy et al. 2010; Noshita 2014; Collins et al. 2021). Future work could incorporate additional
301 parameters such as aperture shape and ontogenetic change in non-isometric taxa. Applying
302 similar methods to diverse and disparate modern taxa would allow the impact of shell thickness
303 on the fidelity of coiling parameters to be better quantified, as experiments would reveal which
304 taxa are most likely to leave evidence of shell sculpture on their steinkerns. We investigated
305 patterns of fidelity at the level of shell morphotype, but future studies at the genus or species
306 level might help produce a framework for calculating bias at different taxonomic levels, since
307 closer relatives may be more similar in morphology and thus analyses at lower taxonomic levels
308 may be more susceptible to bias. This is likely a complex trend, as our preliminary analysis
309 shows that the nature of generic differences varies depending on morphotype (Fig. S12).
310 Additional experimentally determined constraints on steinkern fidelity would allow investigators

311 to quantify bias in calculations of community composition across different taxa and
312 morphologies in steinkern assemblages more accurately. Steinkerns represent a valuable and
313 relatively untapped resource for researching the geological history and evolution of gastropods.

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DECLARATION OF COMPETING INTERESTS

326 The authors declare no competing interests.

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REFERENCES

330

331 ANDERSON, B.M. and ALLMON, W.D., 2018, When domes are spandrels: on septation in
332 turritellids (Cerithioidea) and other gastropods: *Paleobiology*, v. 44(3), p. 444-459, doi:
333 10.1017/pab.2018.12

334

335 BATTEN, R.L., 1984, The calcitic wall in the Paleozoic Families Euomphalidae and
336 Platyceratidae (Archeogastropoda): *Journal of Paleontology*, v. 58(5), p. 1186-1192.

337

338 BOUCHET, P., ROCROI, J.P., HAUSDORF, B., KAIM, A., KANO, Y., NÜTZEL, A., PARKHAEV, P.,
339 SCHRÖDL, M. and STRONG, E.E., 2017, Revised classification, nomenclator and typification of
340 gastropod and monoplacophoran families: *Malacologia* v. 61(1-2), p. 1-526, doi:
341 10.4002/040.061.0201

342 BRAYARD, A., NÜTZEL, A., STEPHEN, D.A., BYLUND, K.G., JENKS, J. and BUCHER, H., 2010,
343 Gastropod evidence against the Early Triassic Lilliput effect: *Geology*, v. 38(2), p. 147–150,
344 doi:10.1130/G30553.1.

345 BUSH, A.M., and BAMBACH, R.K., 2004, Did alpha diversity increase during the Phanerozoic?
346 Lifting the veils of taphonomic, latitudinal, and environmental biases: *The Journal of Geology*, v.
347 112(6), 625–642, doi:10.1086/424576.

348
349 CHERNS, L. and WRIGHT, V.P., 2009, Quantifying the impacts of early diagenetic aragonite
350 dissolution on the fossil record: *PALAIOS*, v. 24(11), 756–771, doi:10.2110/palo.2008.p08-134r.
351

352 CLEMENTS, R., LIEW, T.S., VERMEULEN, J.J. and SCHILTHUIZEN, M., 2008, Further twists in
353 gastropod shell evolution: *Biology Letters*, v. 4(2), p. 179–182, doi: 10.1098/rsbl.2007.0602.

354 COLLINS, K.S., KLAPAUKH, R., CRAMPTON, J.S., GAZLEY, M.F., SCHIPPER, C.I., MAKSIMENKO, A.
355 and HINES, B.R., 2021. Going round the twist—an empirical analysis of shell coiling in
356 helicospiral gastropods: *Paleobiology*, v. 47(4), p.648-665, doi: [10.1017/pab.2021.8](https://doi.org/10.1017/pab.2021.8)
357

358 CREVELING, J.R., KNOLL, A.H. and JOHNSTON, D.T., 2014, Taphonomy of Cambrian phosphatic
359 small shelly fossils: *PALAIOS*, v. 29(6), p. 295–308, doi:10.2110/palo.2014.002.

360 DATTILO, B.F., FREEMAN, R.L., PETERS, W.S., HEIMBROCK, W.P., DELINE, B., MARTIN, A.J.,
361 KALLMEYER, J.W., REEDER, J. and ARGAST, A., 2016, Giants among micromorphs: were
362 Cincinnati (Ordovician, Katian) small shelly phosphatic faunas dwarfed?: *PALAIOS*, v. 31(3),
363 p. 55–70, doi:10.2110/palo.2015.040.

364 GERBER, S., 2017, The geometry of morphospaces: lessons from the classic Raup shell coiling
365 model: *Biological Reviews*, v. 92(2), p.1142-1155, doi: 10.1111/brv.12276.
366

367 GOWER, J.C. (1971) General coefficient of similarity and some of its properties, *Biometrics*, v.
368 27, p. 857-871, doi: 10.2307/2528823.
369

370 HALL, A., KENNEDY, W.J., and TAYLOR, J.H., 1967, Aragonite in fossils: *Proceedings of the*
371 *Royal Society of London. Series B. Biological Sciences*, v. 168(1013), p. 377-412.
372

373 HARZHAUSER, M. and FEHSE, D., 2007, Oligocene and Aquitanian gastropod faunas from the
374 Sultanate of Oman and their biogeographic implications for the early western Indo-Pacific:
375 *Palaeontographica Abteilung A*, v. 280(4–6), p. 75–121, doi:10.1127/pala/280/2007/75.

376 HENDERSON, R.A. and MCNAMARA, K.J., 1985, Taphonomy and ichnology of cephalopod shells
377 in a Maastrichtian chalk from Western Australia: *Lethaia*, v. 18(4), p. 305–322,
378 doi:10.1111/j.1502-3931.1985.tb00710.x.

379 HICKMAN, C.S., 1985, Gastropod morphology and function: Notes for a Short Course: Studies in
380 Geology, v. 13, p. 138–156, doi:10.1017/S0271164800001147
381
382 JOHNSON, E.H., 2020, Experimental tests of bivalve shell shape reveal potential tradeoffs
383 between mechanical and behavioral defenses: Scientific Reports, v. 10(1), p. 19425,
384 doi:10.1038/s41598-020-76358-x.

385 KNIGHT, J.B., 1941, Paleozoic gastropod genotypes: Geological Society of America Special
386 Papers, v. 32, 510 p.

387 LINSLEY, R.M., 1977, Some “laws” of gastropod shell form: Paleobiology, v. 3(2), p. 196–206,
388 doi:10.1017/S0094837300005261
389

390 MACKINNON, D.I., 1985, New Zealand late middle Cambrian molluscs and the origin of
391 Rostroconchia and Bivalvia: Alcheringa: An Australasian Journal of Palaeontology, v. 9(1), p.
392 65–81, doi:10.1080/03115518508618959
393

394 NOSHITA, K., 2014. Quantification and geometric analysis of coiling patterns in gastropod shells
395 based on 3D and 2D image data: Journal of theoretical biology, v. 363, p. 93-104.
396

397 NÜTZEL, A., 2014, Larval ecology and morphology in fossil gastropods: Palaeontology. v. 57(3),
398 p. 479–503, doi:10.1111/pala.12104.

399 OKSANEN, J., SIMPSON, G., BLANCHET, F., KINDT, R., LEGENDRE, P., MINCHIN, P., O'HARA, R.,
400 SOLYMOS, P., STEVENS, M., SZOECs, E., WAGNER, H., BARBOUR, M., BEDWARD, M., BOLKER, B.,
401 BORCARD, D., CARVALHO, G., CHIRICO, M., DE CACERES, M., DURAND, S., EVANGELISTA, H.,
402 FITZJOHN, R., FRIENDLY, M., FURNEAUX, B., HANNIGAN, G., HILL, M., LAHTI, L., MCGLINN, D.,
403 OUELLETTE, M., RIBEIRO CUNHA, E., SMITH, T., STIER, A., TER BRAAK, C. and WEEDON, J., 2022,
404 vegan: Community Ecology Package, R package version 2.6-2. Available at:
405 <https://CRAN.Rproject.org/package=vegan>.
406

407 PALMER, A.R., 1977, Function of shell sculpture in marine gastropods: Hydrodynamic
408 destabilization in *Ceratostoma foliatum*: Science, v. 197(4310), p. 1293–1295.
409 doi:10.1126/science.197.4310.1293
410

411 PARADIS, E. and SCHLIEP, K., 2019, ape 5.0: an environment for modern phylogenetics and
412 evolutionary analyses in R: Bioinformatics, v. 35(3), p. 526-528.
413

414 PIETSCH, C., ANDERSON, B.M., MAISTROS, L.M., PADALINO, E.C. and ALLMON, W.D., 2021.
415 Convergence, parallelism, and function of extreme parietal callus in diverse groups of Cenozoic
416 Gastropoda: Paleobiology, v. 47(2), p.337-362, doi: 10.1017/pab.2020.33
417

418 R Development Core Team, 2022, R: a language and environment for statistical computing: R
419 Foundation for Statistical Computing, Vienna, Austria. Available at: <https://www.R-project.org/>.
420

421 RAUP, D., 1966, Geometric analysis of shell coiling: General problems: Journal of Paleontology,
422 v. 40(5), p. 1178–1190.

- 423 ROY, K., 1994, Effects of the Mesozoic Marine Revolution on the taxonomic, morphologic, and
424 biogeographic evolution of a group: Aporrhaid gastropods during the Mesozoic: *Paleobiology*, v.
425 20(3), p. 274–296.
- 426 SALINAS, C. and KISAILUS, D., 2013, Fracture mitigation strategies in gastropod shells: *JOM*, v.
427 65(4), p. 473–480, doi: [org/10.1007/s11837-013-0570-y](https://doi.org/10.1007/s11837-013-0570-y)
428
- 429 SIGNOR, P.W., 1982, Resolution of life habits using multiple morphologic criteria: Shell form and
430 life-mode in turritelliform gastropods: *Paleobiology*, v. 8(4), 378–388,
431 doi:10.1017/S0094837300007120
432
- 433 SMITH, M.R., 2020a, Information theoretic Generalized Robinson-Foulds metrics for comparing
434 phylogenetic trees. *Bioinformatics*, v. 36(20), p. 5007–5013,
435 doi:org/10.1093/bioinformatics/btab200.
436
- 437 SMITH, M.R., 2020b, TreeDist: distance between phylogenetic trees. R package version 2.6.2'.
438 Available at: <https://zenodo.org/record/8098675>.
439
- 440 TRYON, G.W., 1882, *Structural and systematic conchology: an introduction to the study of the*
441 *mollusca*. The Author.
- 442 URDY, S., GOUEMAND, N., BUCHER, H. and CHIRAT, R., 2010. Allometries and the
443 morphogenesis of the molluscan shell: a quantitative and theoretical model: *Journal of*
444 *Experimental Zoology Part B: Molecular and Developmental Evolution*, v. 314(4), p.280-302,
445 doi: 10.1002/jez.b.21337
446
- 447 VERMEIJ, G.J., 1971, Gastropod evolution and morphological diversity in relation to shell
448 geometry: *Journal of Zoology*, v. 163(1), p. 15–23.
- 449 VERMEIJ, G.J., 2022. The balanced life: evolution of ventral shell weighting in
450 gastropods: *Zoological Journal of the Linnean Society*, v. 194 (1), p. 256-275, doi:
451 10.1093/zoolinlean/zlab019
452
- 453 VINN, O., WILSON, M.A., ZATOŃ, M. and TOOM, U., 2014, The trace fossil *Arachnostega* in the
454 Ordovician of Estonia (Baltica): *Palaeontologia Electronica*, 17.3.40A, p. 1-9,
455 doi:org/10.26879/477.
- 456 WAGNER, P.J., 2001, Gastropod phylogenetics: progress, problems, and implications: *Journal of*
457 *Paleontology*, v. 75(6), p. 1128–1140. doi:10.1666/0022-
458 3360(2001)075<1128:GPPPAI>2.0.CO;2.
- 459 WAGNER, P.J. and ERWIN, D.H., 2006, Patterns of convergence in general shell form among
460 Paleozoic gastropods: *Paleobiology*, v. 32(2), p. 316–337.
461
- 462 WRIGHT, V.P., CHERNS, L., AZERÊDO, A.C., and CABRAL, M.C., 2018, Testing whether early
463 diagenesis of skeletal carbonate is different in non-marine settings: Contrasting styles of

464 molluscan preservation in the Upper Jurassic of Portugal: Palaeogeography, Palaeoclimatology,
465 Palaeoecology, v. 492, p. 1–9, doi:10.1016/j.palaeo.2017.11.014.

466

467 YOCHELSON, E.L., WHITE JR., J.S. and GORDON JR., M. 1967. Aragonite and calcite in mollusks
468 from the Pennsylvanian Kendrick Shale: U.S. Geological Survey Professional Paper, v. 575, p.
469 76-78.

470

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FIGURE CAPTIONS

474 FIG. 1.—Measurement scheme. A) Aperture view used to calculate translation (T) and whorl

475 expansion rate (W). B) Coiling axis view used to calculate distance from the coiling axis (D).

476 Red dots represent the coiling axis, blue dots represent landmarks.

477

478 FIG. 2.—Original shells (left), corresponding silikerns (middle), and steinkerns showing similar

479 characteristics (right). A) Morphotype 8 *Siratus articulatus* (YPM IZ 110139) showing evidence

480 of shell spines on the silikern compared to *Cassidaria cheops* (YPM IP 255138) which preserves

481 similar surface features. B) Morphotype 7 *Sinistrofulgur perversum* (YPM IZ 110127) showing

482 evidence of shell ridges on the silikern. C) Morphotype 4 *Naticarius canrena* (YPM IZ 110149)

483 and *Lanistes sodaensis* (YPM IP 255119) showing traces of encrustation. D) Morphotype 12

484 *Monetaria caputserpentis* (YPM IZ 110163) showing columellar and labral teeth corresponding

485 to grooves on the silikern compared to *Cassidaria cheops* (YPM IP 255138), which preserves

486 similar grooves. Both fossils are from the Eocene of the Fayum Depression, Egypt.

487

488 FIG. 3.—Side-by-side comparisons of the shell and silikern of each morphotype. A) Morphotype

489 1, *Conus brunneus*, YPM IZ 110115. B) Morphotype 2, *Duplicaria duplicata*, YPM IZ 110121.

490 C) Morphotype 3, *Oliva* sp., YPM IZ 1110133. D) Morphotype 4, *Naticarius canrena*, YPM IZ

491 110147. E) Morphotype 5, *Rumina decollata*, YPM IZ 110152. F) Morphotype 6, *Haplotrema*
492 *concovum*, YPM IZ 110157. G) Morphotype 7, *Sinistrofulgur perversum*, YPM IZ 110127. H)
493 Morphotype 8, *Siratus articulatus*, YPM IZ 110139. I) Morphotype 9, *Astraea americana*, YPM
494 IZ 110356. J) Morphotype 10, *Fasciolaria tulipa*, YPM IZ 110348. K) Morphotype 11,
495 *Epitonium* sp., YPM IZ 110192. L) Morphotype 12, *Monetaria caputserpentis*, YPM IZ 110163.
496

497 FIG. 4.—Raup parameters for all shells and silikerns of each morphotype. A) Translation. B)
498 Distance from the coiling axis. C) Whorl expansion rate. Differences between the values of
499 shells (blue circles) and silikerns (orange squares) represent differential fidelity by morphotype.
500

501 FIG. 5.—Quantitative results. A) Distribution of average silikern-shell distances of 10,000
502 random samples each consisting of 10 silikerns. Average silikern-shell distances are bounded
503 between 0 and 1. Dashed lines indicate average distances for each morphotype (each represented
504 by 10 specimens). P = p-values. M = morphotype. P-values defined as number of random
505 samples with average distances equal to or greater than that of the morphotype. B) Clustering
506 information distances (CIDs) separating the clustering of subsamples of shells and silikerns from
507 the reference clustering pattern derived using all shells. Each boxplot summarizes 10,000 CIDs:
508 the box delimits the interquartile range, the central line denotes the median, and the whiskers
509 encompass the CIDs that are no more than 1.5 times the interquartile range beyond the upper and
510 lower quartiles.

511
512 TABLE. 1.—Taxa used in this study. Twelve morphotypes are represented either by ten
513 specimens each of one genus or five specimens each of two genera, based on specimen

514 availability, with the exception of *Monetaria caputserpentis*, which posed unique difficulties and
515 was excluded from quantitative analysis (see text).