

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39

Galué, et al,

**Contrasting ecological patterns of Venezuelan equine encephalitis and
Madariaga viruses in small mammal and mosquito populations from two
enzootic regions of Panama**

Josefrancisco Galué^{1,2,3}, William M. de Souza⁴, Rolando Torres-Cosme⁵, Carlos
Lezcano-Coba^{1,2}, Robert B. Tesh⁶, Hilda Guzman⁶, Scott C. Weaver⁷, Zeuz Capitan-
Barrios^{1,8}, Anayansi Valderrama⁵, Rafael Samudio⁹, Amy Y. Vittor¹⁰, Nikos
Vasilakis^{6,11}, Lorenzo Cáceres Carrera⁵, Christl A. Donnelly¹², Nuno R. Faria¹³ and
Jean-Paul Carrera^{1,2,14*}

1. Carson Centre for Health and Ecosystem Research, La Peñita, Darién, Panama. Carson Center for Health and Ecosystem Research, La Peñita, Darién, Panama
2. Department of Research in Virology and Biotechnology, Gorgas Memorial Institute of Health Studies, Panamá City, Panamá
3. Emerge, Emerging Diseases and Climate Change Research Unit, School of Public Health and Administration, Universidad Peruana Cayetano Heredia (UPCH), Lima Peru
4. Department of Microbiology, Immunology, and Molecular Genetics, University of Kentucky, College of Medicine, Lexington, Kentucky, USA
5. Department of Medical Entomology, Gorgas Memorial Institute of Health Studies, Panamá City, Panamá
6. Department of Pathology, University of Texas Medical Branch, Galveston Texas, USA
7. Department of Microbiology and Immunology, University of Texas Medical Branch, Galveston, Texas, USA
8. Departamento de Microbiología y Parasitología, Facultad de Ciencias Naturales, Exactas y Tecnología, Universidad de Panama, Panama
9. Centro de Investigación y Capacitación en Conservación de la Biodiversidad, Sociedad Mastozoológica de Panamá, Panamá City, Panamá
10. Department of Medicine and Global Health, University of Florida, Gainesville, Florida
11. Center for Vector-Borne and Zoonotic Diseases, University of Texas Medical Branch (UTMB), Galveston, TX, USA
12. Department of Statistics, University of Oxford, Oxford United Kingdom
13. Department of Infectious Disease Epidemiology, MRC Centre for Global Infectious Disease Analysis, J-IDEA, Imperial College London, London, UK
14. Department of Biology, University of Oxford, Oxford, United Kingdom

*Correspondence: jpcarrera@centrocarson.org

6

7

8

40 Abstract

41

42 **Background:** Understanding enzootic cycles is key to arbovirus control. Rodents
43 are primary reservoirs of Venezuelan equine encephalitis virus (VEEV), while the
44 vertebrate host of Madariaga virus (MADV) remains unknown. *Culex (Melanoconion)*
45 mosquitoes are considered primary vectors. We studied small mammals and
46 mosquito ecology in the Darién province and Panamá Oeste province, two enzootic
47 regions of Panama. Additional alphavirus seroprevalence was assessed in small
48 mammals.

49 **Methods:** Small mammals were trapped using Sherman and Tomahawk traps, and
50 blood samples were tested for MADV and VEEV neutralizing antibodies using plaque
51 reduction neutralization test and attempt viral isolation and generic alphavirus RT-
52 PCR from small mammal tissues. Mosquitoes were collected concurrently using
53 CDC light, Trinidad, and resting traps. Diversity and similarity were analyzed using
54 standard ecological indices.

55 **Results:** We sampled 599 small mammals (16 species). MADV seroprevalence was
56 highest in Los Pavitos (9.0%) and restricted to Darién; VEEV was more widespread,
57 peaking in El Cacao (27.3%) and El Real (20.4%). *Oryzomys couesi*
58 and *Transandinomys bolivararis* showed elevated MADV seroprevalence; *T.*
59 *bolivararis* and *Proechimys semispinosus* had higher VEEV seroprevalence. Among
60 4,118 mosquitoes, *Coquillettidia venezuelensis* was most abundant. El Real had the
61 highest species richness and diversity. Geographic proximity influenced community
62 similarity.

63 **Conclusions:** Spatial differences in host and vector communities may reflect distinct
64 transmission patterns of MADV and VEE. These findings could contribute to better
65 understanding of alphavirus ecology and potentially inform public health strategies in
66 endemic regions.

67

68

69 Introduction

70 Madariaga virus (MADV) and Venezuelan equine encephalitis virus (VEEV), both
71 members of the *Alphavirus* genus within the *Togaviridae* family, are emerging
72 zoonotic arboviruses distributed across Latin America (Navarro et al., 2017, chaps.
73 169–192). VEEV causes widespread epizootics in the Americas, with notable impact
74 on humans and equids. Epizootic subtypes IAB and IC of VEEV are believed to arise
75 from enzootic subtype ID ancestors through mosquito- or equine-adaptive
76 mutations (Kinney et al., 1992). Enzootic subtypes ID and IE are known to circulate
77 primarily in the tropical forests of Latin America, within a sylvatic transmission cycle
78 between rodents and mosquitoes of the *Culex (Melanoconion)* subgenus, particularly
79 within the *Spissipes* section (Aguilar et al., 2011a; Weaver et al., 2004). A vertebrate
80 reservoir is defined as a host species that carries a pathogen, often without showing
81 disease symptoms. It plays a critical role in maintaining and transmitting the
82 pathogen in natural cycles (Deardorff et al., 2009a). Experimental and field studies
83 suggest that several rodent species, including those from the genera *Sigmodon*,
84 *Oryzomys*, *Zygodontomys*, *Heteromys*, *Peromyscus*, and *Proechimys*, may serve as
85 key reservoir hosts for VEEV (Aguilar et al., 2011a; Deardorff et al., 2009b;
86 Sotomayor-Bonilla et al., 2017).

87

11
12
13

88 Human infections with VEEV typically result from spillover events from the enzootic
89 cycle. While the majority of human VEEV infections result in nonspecific flu-like
90 outcomes (Aguilar et al., 2011a; Carrera et al., 2018), around 14% of symptomatic
91 VEEV cases develop neurological complications, with an estimated 10% fatality rate
92 (Aguilar et al., 2011a). In contrast, human infections with MADV are rare and are
93 typically reported in the context of outbreaks among equids. Unlike VEEV, MADV
94 lacks evidence of amplification in equids or epidemic potential (Aguilar et al., 2007).
95 Notably, MADV is widely distributed across the Americas, with its enzootic range
96 overlapping that of VEEV (Arrigo et al., 2010a). This broad distribution suggests that
97 MADV may circulate primarily through silent enzootic cycles, with infrequent spillover
98 into humans. The first documented outbreak of MADV occurred in the eastern Darién
99 region of Panamá in 2010, an area where VEEV also circulates (Carrera et al.,
100 2013a).

101

102 MADV infections result from spillover events originating in enzootic cycle within
103 forested regions. Similar to VEEV, mosquitoes from the *Culex (Melanoconion)*
104 subgenus are believed to be the primary vectors of MADV (Magalhaes et al., 2024).
105 MADV has been isolated from *Culex (Melanoconion)* species in Peru and Panama,
106 though its vector ecology remains poorly understood (Turell et al., 2005) (Dietz et al.,
107 1980; Srihongse and Galindo, 1967) (Turell et al., 2008). Notably, *Aedes*
108 *taeniorhynchus* has been found naturally infected during outbreaks (Causey et al.,
109 1961), suggesting other mosquitoes species could also play a role in MADV
110 transmission.

111

112 The primary vertebrate host for MADV remains unclear, though evolutionary,
113 epidemiological, and experimental studies suggest rodents as likely reservoirs,
114 similar to VEEV (Arrigo et al., 2010a, 2010c; Vittor et al., 2016b). Rodents and other
115 vertebrates, including birds and reptiles, have shown evidence of MADV exposure,
116 though the primary reservoir remains unidentified (Arrigo et al., 2010c). In Panamá, a
117 serosurvey in Darién showed predominant MADV seropositivity in *Zygodontomys*
118 *brevicauda* (short-tailed cane mouse) and VEEV in *Transandinomys bolivaris* (long-
119 whiskered rice rat) (Vittor et al., 2016b) suggesting exposure to these viruses.
120 However, this study was limited to Darién and focused exclusively on mammals,
121 leaving other enzootic area and potential vertebrates hosts unexamined. Despite
122 these findings, conclusive evidence regarding the primary vertebrate reservoir for
123 MADV remains lacking.

124

125 To assess small mammal exposure to MADV and VEEV, we conducted small
126 mammal trapping during 2011 and 2012 across five distinct site in the Darién and
127 Panamá Oeste provinces and measured alphavirus antibody seroprevalence. In
128 parallel, we surveyed mosquito community assemblages in these areas to document
129 the presence and overlap of small mammals and known alphavirus vector species.
130 We also examined alphavirus seroprevalence and explored its potential link with
131 small mammal and mosquito vector diversity.

132

133 **Materials and methods**

134 **Ethics statement**

135 The capture, use, and euthanasia of wild rodents was evaluated and approved by
136 the Institutional Animal Care and Use Committee of the Gorgas Memorial Institute for

16

17

18

137 Health Studies (010/ CIUCAL/ICGES18) and the Panamanian Ministry of
138 Environment (SC/A-21-17, ANAM) using the criteria established in the "International
139 Guiding Principles for Biomedical Research Involving Animals" developed by the
140 Council for International Organizations of Medical Sciences (CIOMS). The study was
141 conducted in accordance with Law No. 23 of January 15, 1997 (Animal Welfare
142 Guarantee) of the Republic of Panama.

143 **Collection sites**

144

145 Rodent and mosquito collection were conducted in 2011 and 2012 in the Darién and
146 Panamá Oeste provinces (**Figure 1**). Darién province, located in eastern Panama, is
147 home to Central America's largest national park and is known for its rich ecological
148 and cultural diversity. The province receives between 1,800 and 4,500 mm of rainfall
149 annually, with average temperatures ranging from 28 °C to 31 °C. In contrast,
150 Panamá Oeste, located along the Pacific coast, is more urbanized, with annual
151 rainfall between 1,500 and 3,500 mm and temperatures ranging from 24 °C to 32 °C
152 (Autoridad Nacional del Ambiente, 2010). The main economic activities in both
153 regions are agriculture and cattle farming. Collection sites were selected based on
154 previous reports of confirmed human and equine encephalitic alphavirus infections in
155 2001, 2004 and 2010. A total of three locations were selected for rodent sampling in
156 Darién province: Los Pavitos, El Real and Santa Librada (**Figure 1**). Two sites were
157 selected in Panamá Oeste province: El Cacao and Cirí Grande (**Figure 1**).

158

159 **Small mammal trapping**

160 Small mammals were collected from June to November 2011 (rainy season) and
161 from March to April 2012 (dry season) using Sherman and Tomahawk traps baited
162 with a mixture of rice, corn, sorghum, and peanut butter. In the field, traps were
163 placed and maintained from 6:00 PM and then checked soon after 6:00 AM. For this
164 study, three distinct sampling points, each separated by 500 meters, were selected
165 within each collection site. At each point, a linear transect approximately 125 meters
166 in length was established. Each site was sampled for 8 to 14 nights, depending on
167 logistical feasibility and field conditions. On average, 100 Sherman and 10
168 Tomahawk traps were deployed per night, resulting in a total sampling effort of 5,940
169 trap-nights over 54 trapping days. Traps were placed within houses and in the peri-
170 domiciliary area of previously confirmed VEEV cases. The peri-domiciliary setting
171 includes grasslands, and crop fields as well as wooded areas near homes in each of
172 the selected locations. Trapped animals were euthanized using halothane and
173 identified using taxonomic keys or the Field Guide to the Mammals of Central
174 America & Southeast Mexico (Reid, 2009). Blood samples were collected from the
175 retro-orbital sinus. Heart, liver, spleen, lung, and kidney tissues were then harvested.
176 All samples were immediately placed into liquid nitrogen before transportation to the
177 Gorgas Memorial Institute (GMI) for testing. Animal carcasses were deposited in the
178 Vertebrate Museum of the University of Panamá and the Zoological Collection of the
179 GMI (Panamá City, Republic of Panama).

180 **Mosquito collection**

181

182 Mosquito collections were undertaken from June to November 2011 (rainy season)
183 and from March to April 2012 (dry season) at each collection site in Darien (Los
184 Pavitos, El Real and Santa Librada) and Panamá Oeste Provinces (El Cacao and

19 4

20

21
22
23

185 Cirí Grande) using three types of mosquito traps for three consecutive nights, from
186 18:00 to 06:00. Traps were positioned approximately 1.5 meters above the ground in
187 peridomestic areas and adjacent forested habitats—referred to as extradomiciliary
188 settings. In every site we selected a single trapping area, where we used 10 CDC light
189 traps baited with octanol, eight modified Trinidad 17 (TT-17) traps containing live
190 hamsters as bait, and eight encephalitis vector surveillance (EVS) traps baited with
191 carbon dioxide. Mosquitoes were euthanized by flash-freezing immediately after
192 collection and preliminarily identified to the genus level in the field. Subsequently,
193 specimens were sorted by trap type and sampling date, placed in labeled plastic
194 vials, and stored in liquid nitrogen. Samples were then transported to the Gorgas
195 Memorial Institute in Panama City, where species-level identification was carried out
196 using standard taxonomic keys. A preliminary report of mosquito data and methods
197 was published previously (Torres et al., 2017).

198

199 **Laboratory methods**

200 **Alphavirus serology in small mammals**

201 Rodent blood samples were screened in a 1:20 dilution using virus-specific plaque
202 reduction neutralization tests (PRNTs) for VEEV and MADV and then titrated. A
203 positive sample was considered as the reciprocal of the highest dilution that reduced
204 plaque counts by >80% (PRNT₈₀), as previously described (Vittor et al., 2016a). For
205 PRNT, we used the wild-type MADV strain GML-267113, isolated from a fatal human
206 case in Panamá in 2017 which belong to the MADV lineage enzootic subtype (Carrera
207 et al., 2020a), and the VEEV vaccine strain TC83 which is closely related to the
208 VEEV-ID Panamanian subtype (Quiroz et al., 2009).

209

210 **Viral isolation and molecular testing**

211 Small mammal tissues and mosquito pools were used to prepare a 10% suspension
212 with 2 mL of minimum essential medium supplemented with penicillin and
213 streptomycin, and 20% fetal bovine serum and homogenized using a Tissue Lyser
214 (Qiagen, Hidden, Germany). After centrifugation at 17,709 x g for 10 minutes, 200 μ L
215 of the supernatant was inoculated into each of two 12.5 cm² flasks of Vero cells
216 (African green monkey-ATCC CCL-81, USA) and passaged twice for cytopathic
217 effect confirmation.

218

219 Small mammal tissue, mosquito homogenates and cell culture supernatant were
220 used for viral RNA extraction using the QIAamp RNA viral extraction kit (Qiagen,
221 Valencia, CA) and tested for alphaviruses using reverse transcription-polymerase
222 chain reaction (RT-PCR) assays, as previously described (Sánchez-Seco et al.,
223 2001).

224

225 **Statistical and ecological methods**

226

227 **Diversity analysis of small mammals and mosquitos**

228 We assessed the diversity and species richness of small mammals and mosquitos
229 across collection sites in the Darién and Panamá Oeste provinces during 2011 and
230 2012. To estimate species richness, we calculated Chao-1 and iChao-1 indices for
231 five study sites—El Real, Los Pavitos, Santa Librada, El Cacao and Cirí Grande.
232 The Chao-1 index provides a lower-bound estimate of species richness based on the
233 frequency of rare species, while the iChao-1 index incorporates additional
234 information from low-frequency data, offering a more refined estimate, particularly in

24 5
25

26

27

28

235 datasets with sparse observations. To further evaluate diversity, we calculated
236 the Shannon-Wiener index (H) (Ortiz-Burgos, 2016) and Simpson's Diversity Index
237 (SDI, 1-D) (Gregorius and Gillet, 2008). The Shannon-Wiener index reflects both
238 species richness and evenness, with higher values indicating greater diversity,
239 whereas Simpson's Diversity Index ranges from 0 (no diversity) to 1 (maximum
240 diversity) and emphasizes the dominance of the most abundant species.
241 Additionally, we used Margalef's index as a complementary measure of species
242 richness, where higher values correspond to greater species richness (Death, 2021).
243 All diversity metrics, including richness estimates and evenness measures, were
244 analyzed using the statistical software PAST version 4.03 (Hammer et al., 2001).

245

246 **Estimation of small mammal and mosquito community similarity**

247 To evaluate small mammal and mosquito community similarity across collection
248 sites, we calculated the Jaccard and Sørensen-Dice indices, which measure the
249 proportion of shared species between sites. The Jaccard index considers the ratio of
250 shared species to the total observed species, while the Sørensen-Dice index, giving
251 greater weight to shared species, calculates twice the number of shared species
252 divided by the total species across sites. Five sites—El Real, Los Pavitos, Santa
253 Librada, El Cacao, and Cirí Grande—were analyzed. All analyses were conducted
254 using PAST version 4.03 (Hammer et al., 2001). Additionally, pairwise comparisons
255 using Tukey's Honestly Significant Difference (HSD) test was used to adjust p-values
256 and 95% confidence intervals (CIs) for multiple comparisons, based on the possible
257 pairs of means and studentized range distribution (Nanda et al., 2021).

258

259 **Factors associated with alphavirus seroprevalence in small mammals**

260 Small mammal species were grouped at the genus level to account for the small
261 sample size. Small mammal genus-specific, VEEV (n=296) and MADV (n=292)
262 seropositivity, and land use and land cover (LULC) (**Supplementary Figure 1**),
263 classification were used for univariate logistic regression analysis. To evaluate risk
264 factors at the community and genus level, we conducted separate univariate
265 analyses for MADV and VEEV; in each case, the outcome variable was the
266 presence/absence of antibodies against the virus, as determined by a PRNT₈₀ titer
267 >1:20. The associations between each outcome and independent variable (collection
268 site and genus) were estimated using logistic regression and were expressed as
269 odds ratios (ORs). Univariable ORs were calculated with 95% CIs. Statistical
270 analyses were undertaken using the package STATA version 14.1 (College Station,
271 TX).

272

273 **Ecological data and seroprevalence**

274 We conducted a site-level ecological analysis using data from five sites across the
275 Darién and Panamá Oeste provinces of Panamá. For each site, we computed
276 diversity and richness indices for small mammal and mosquito communities:
277 Richness: iChao-1 estimator. Diversity: Shannon and Simpson indices.
278 Seroprevalence data were derived from PRNT results targeting antibodies to VEEV
279 and MADV in small mammals.

280

281 **Correlation analysis**

282 We performed a Spearman rank correlation analysis to examine the association
283 between ecological diversity indices and alphavirus (VEEV and MADV)

29 6

30

31

32

33

284 seroprevalence across five study sites. The ecological indices included abundance,
285 Simpson, Shannon, Margalef, and Chao-1 indices for small mammals, mosquitoes,
286 and *Culex (Melanoconion)* mosquitoes. VEEV and MADV seroprevalence was
287 calculated as the percentage of seropositive individuals at each site. Correlation
288 coefficients (Spearman's ρ) and corresponding p-values were calculated using R
289 (v4.3.0) (Dalgaard, 2010).

290

291 **Results**

292 **Small mammal abundance across study sites**

293 We collected a total of 599 small mammals between 2011 and 2012, with specimens
294 belonging to 13 genera and 16 species (**Figure 2A and 2B, Supplementary Table**
295 **1**). Most animals were captured during 2011 (71.8% of all collections, $n = 430/599$).
296 In general, the majority of rodents were captured within Darién province (87.6% of all
297 collections, $n = 525/599$), specifically in El Real (33.7%, $n = 202/599$), followed by
298 Los Pavitos (27.6%, $n = 165/599$) and Santa Librada (26.4%, $n = 158/599$)
299 (**Supplementary Table 1**).

300

301 The short-tailed cane mouse (*Zygodontomys brevicauda*) was the most abundant
302 species identified across study sites (70.5% of trapped animals, $n = 402/599$),
303 followed by the Central American spiny rat (*Proechimys semispinosus*, 12.2%, $n =$
304 $73/599$) and the dusky rice rat (*Melanomys caliginosus*, 3.5%, $n = 21/599$). Species
305 with abundance $\leq 2\%$ are shown in **Supplementary Table 1**.

306

307 **Mosquito abundance across study sites**

308 A total of 4,118 mosquitoes were collected, and a few species accounted for the
309 majority of specimens. *Coquillettidia venezuelensis* was the most abundant,
310 comprising (19.6%, $n = 809/4,118$) of the total sample, followed by *Culex (Culex)*
311 sp. (14.9%, $n = 615/4,118$) and *Culex (Melanoconion)* sp. (14.2%, $n = 585/4,118$)
312 (**Figure 3A**). Less frequent species ($\leq 10\%$) are reported in (**Figure 3A**) and
313 (**supplementary table 2**).

314

315 When comparing abundance across sites, clear patterns of species dominance
316 emerged. In El Real, *Coquillettidia venezuelensis* was the most abundant species
317 (26.8%, $n = 777/2,900$; supplementary table 2). In Los Pavitos and Santa Librada,
318 *Culex (Culex)* sp. dominated (42.1%, $n = 90/214$) and (44.9%, $n = 336/749$) of
319 captures, respectively (**supplementary table 2**). Finally in Cirí Grande and El
320 Cacao, the mosquito fauna was dominated with *Culex* sp. (58.8%, $n = 87/148$) and
321 (43.9%, $n = 47/107$) of captures, respectively (**Figure 3A and Supplementary Table**
322 **2**).

323

324 **Highest small mammal diversity and richness across study sites**

325 Species richness and diversity varied across the five study sites in the Darién and
326 Panamá Oeste provinces. El Real exhibited the highest species richness (Chao-1 =
327 11.5; iChao-1 = 12.99), in contrast, Santa Librada demonstrated the lowest richness,
328 (Chao-1 = 5.99; iChao-1 = 6.48) (**Table 1**). Intermediate values were observed
329 for Cirí Grande, El Cacao and Los Pavitos. Diversity indices showed a similar
330 pattern, with El Real ranking highest (1-D = 0.60; H = 1.42; M = 1.88) and Santa
331 Librada lowest (1-D = 0.11; H = 0.29; M = 0.79) (**Table 1**). Estimates from all indices
332 —Chao-1, iChao-1, Simpson's, Shannon-Wiener, and Margalef's—showed

34 7

35

36

37

38

333 consistent patterns, reinforcing the observed gradients in species richness and
334 diversity across the study sites (**Table 1**).

335

336 **Highest mosquito diversity and richness across study sites**

337 Mosquito species richness and diversity varied across the five study sites in the
338 Darién and Panamá Oeste provinces. El Real exhibited the highest species richness,
339 (Chao-1 = 42.33; iChao-1 = 43.00). In contrast, Cirí Grande showed the lowest
340 richness, (Chao-1 = 16.99; iChao-1 = 19.74) (**Supplementary Table 3**). The
341 Shannon-Wiener index (H) and Simpson's diversity indices (1-D) also identified El
342 Real as the most diverse site (H = 2.58; 1-D = 0.87), while Cirí Grande showed the
343 lowest diversity (H = 1.66; 1-D = 0.64) (**Supplementary Table 3**). These patterns
344 were consistent with Margalef's index, which also ranked El Real highest in species
345 richness (M = 5.14) and Cirí Grande the lowest (M = 2.80) (**Supplementary Table**
346 **3**).

347

348 A subset analysis focusing exclusively on *Culex (Melanoconion)* species revealed a
349 distinct diversity pattern across the same sites. *Culex (Melanoconion)* species
350 richness and diversity varied across the five study sites in the Darién and Panamá
351 Oeste provinces (**Figure 3B**). El Real de exhibited the highest species richness,
352 (Chao-1 = 8.00; iChao-1 = 8.00) (**Supplementary Table 4**). In contrast, El Cacao
353 and Cirí Grande showed the lowest richness, with both (Chao-1 = iChao-1 = 1.00).
354 Shannon-Wiener (H) and Simpson's (1-D) indices showed the highest diversity in
355 Santa Librada (H = 1.48; 1-D = 0.75). El Cacao and Cirí Grande exhibited no
356 diversity. These patterns were consistent with Margalef's index, which ranked El
357 Real highest in species richness (M = 1.01) (**Supplementary Table 4**).

358

359 **Species similarity at the community level**

360 The Jaccard and Sørensen-Dice indices revealed distinct patterns of community
361 similarity among the five collection sites. The Jaccard index indicated the highest
362 similarity between El Real and Los Pavitos (78%). followed by Cirí Grande and El
363 Cacao (60%) (**Supplementary Table 5**). The Sørensen-Dice index, which is more
364 sensitive to shared species. The highest similarity was again between El Real and
365 Los Pavitos (88%), followed by Cirí Grande and El Cacao (75%) (**Supplementary**
366 **Table 6**).

367

368 Pairwise comparisons using Tukey's HSD test confirmed significant differences in
369 species composition across sites. Similarity was greatest within the Darién province
370 (e.g., Santa Librada and Los Pavitos, $p = 0.639$) and the Oeste province (e.g., El
371 Cacao and Cirí Grande). The largest differences were observed between the Darién
372 and Oeste provinces, though El Cacao and El Real, despite being in different
373 provinces, exhibited the smallest differences in species composition ($Contrast = -1.8$;
374 $p = 0.001$) (**Table 2**).

375

376 **Species similarity at the mosquito community level**

377 The Jaccard index, the highest similarity was observed between Cirí Grande and El
378 Cacao (50%), followed by Santa Librada and Los Pavitos (52%) (**Supplementary**
379 **Table 7**). The Sørensen-Dice index, which gives greater weight to shared species,
380 showed consistently higher similarity values. The highest similarity was found
381 between Cirí Grande and El Cacao (67%), followed by Santa Librada with El Real
382 (72%) (**Supplementary Table 8**).

39 8

40

41
42
43

383

384 **Widespread alphavirus seroprevalence in small mammals across Panama**

385 The overall MADV and VEEV seroprevalence in small mammals were 3.8% (95% CI:
386 2.0-7.0; $n = 11/292$) and 12.5% (95% CI: 8.9-16.8; $n = 37/296$), respectively
387 (**Supplementary Table 9 and 10**). VEEV seroprevalence was higher in 2011
388 (16.2%, 95% CI: 11.4-22.1; $n = 32/197$) compared to 2012 (5.1%, 95% CI: 1.6-11.3;
389 $n = 5/99$) (**Supplementary Table 10**). MADV seroprevalence dropped from 4.6%
390 (95% CI: 2.1-8.6; $n = 9/194$) in 2011 to 2.0% (95% CI: 0.2-7.0; $n = 2/98$) in 2012
391 (**Supplementary Table 9**). VEEV seroprevalence was widespread across the Oeste
392 and Darién provinces with the highest seroprevalence found in El Cacao (27.3%,
393 95% CI: 16.1-40.9; $n=15/55$) in the Oeste province, followed by El Real (20.4%, 95%
394 CI: 12.6-29.7; $n=19/94$) in the Darién province (**Table 1 and 3**). MADV
395 seroprevalence was higher in rodents collected in Los Pavitos (9.0%, 95% CI: 3.6-
396 17.6; $n=7/78$), followed by El Real (3.2%, 95% CI: 1.0-9.0; $n = 3/94$) (**Table 1 and 3**).
397 No evidence of MADV infection isolation or antibodies was found in small mammals
398 collected in the Oeste province. *Transandinomys bolivaris* (80.0%, 95% CI: 28.3-
399 99.4; $n=4/5$) and *Proechimys semispinosus* (27.3%, 95% CI: 17.0-39.6; $n=18/66$)
400 had the highest VEEV seroprevalence (**Supplementary Table 9**), while *Oryzomys*
401 *couesi* (23.1%, 95% CI: 5.0-53.8; $n=3/13$) and *Transandinomys bolivaris* (20.0%,
402 95% CI: 0.5-71.6; $n=1/5$) had the highest MADV seroprevalence (**Supplementary**
403 **Table 10**). No evidence of antibodies against MADV or VEEV was observed in
404 *Marmosa* sp. or *Didelphis marsupialis*.

405

406 **Alphaviral active circulation**

407 No evidence of active alphavirus circulation was detected in any small mammal
408 samples or mosquito pools, as determined by RT-PCR or viral isolation.

409

410 **Factors associated with alphavirus seroprevalence in small mammals**

411 MADV seroprevalence was independent of collection site, but Los Pavitos (OR=0.1;
412 95% CI: 0.0-0.4; $p=0.002$) and Santa Librada (OR=0.1; 95% CI: 0.0-0.6; $p=0.017$)
413 were protective factors for VEEV seropositivity when compared with El Real.
414 Univariate analysis by rodent taxa revealed that the odds of MADV seropositivity
415 were 9.0 times in *Oryzomys* (OR=9.0; 95%CI: 1.9-43.2; $p=0.006$) compared to the
416 reference *Zygodontomys*. The odds of VEEV seropositivity in *Proechimys* were
417 significantly higher than in the reference *Zygodontomys* (OR=4.6; 95%CI: 2.1-10.2;
418 $p<0.001$).

419

420 **Ecological correlates of VEEV exposure across study sites**

421 Within the small mammal community, VEEV seroprevalence showed strong positive
422 correlations with diversity indices. The Simpson and Shannon indices both showed a
423 correlation of $\rho = 0.80$, while the Margalef index also exhibited a notable association
424 ($\rho = 0.60$). In contrast, a moderate negative correlation was observed with small
425 mammal abundance ($\rho = -0.40$) (**Figure 4**). Regarding mosquitoes, the correlations
426 were generally weak or negative. Mosquito abundance was negatively correlated
427 with VEEV seroprevalence ($\rho = -0.30$), while richness estimators such as Chao-1
428 and Margalef showed low correlations ($\rho = 0.30$). In the *Culex*
429 (*Melanoconion*) group, the pattern was even more distinct. *Culex* abundance was
430 moderately negatively correlated with VEEV ($\rho = -0.30$), and diversity indices
431 (Simpson, Shannon, Chao-1, and Margalef) showed marked negative correlations,
432 particularly Simpson and Shannon ($\rho = -0.72$) (**Figure 4**).

44 9

45

46
47
48
433

434 For MADV, the relationship with the small mammal community was notably different.
435 There was a very strong positive correlation between mammal abundance and
436 MADV seroprevalence ($\rho = 0.87$), along with a high correlation with the Chao-1
437 index ($\rho = 0.72$). Unlike VEEV, diversity indices (Simpson, Shannon, and Margalef)
438 showed slightly negative correlations ($\rho = -0.15$). In the mosquito community, MADV
439 showed moderate to strong positive correlations. Mosquito abundance was positively
440 associated with seroprevalence ($\rho = 0.67$), as was the Shannon index ($\rho = 0.67$).
441 Richness estimators such as Chao-1 and Margalef also showed consistent positive
442 associations ($\rho = 0.31$). Within *Culex (Melanoconion)* subgenus, correlations were
443 also consistently positive and high. *Culex* abundance showed a correlation of ($\rho =$
444 0.67) with MADV seroprevalence. All diversity indices (Simpson, Shannon,
445 Margalef, Chao-1, and corrected Chao-1) exhibited similar correlations ($\rho = 0.68$)
446 (Figure 4).

447
448
449

Discussion

450 Patterns of species richness, diversity, and community similarity revealed spatial
451 structuring in both small mammal and mosquito taxa across study sites in Darién
452 and Panamá Oeste. El Real (Darién) consistently ranked highest in small mammal
453 richness and diversity across all indices. El Cacao (Panamá Oeste) followed as the
454 second most diverse site for small mammals, whereas Santa Librada exhibited the
455 lowest richness and evenness. Within Darién, Los Pavitos showed moderate
456 richness and high compositional similarity with El Real, indicating a shared species
457 pool and likely habitat continuity. These findings of high rodent small mammals and
458 mosquito high diversity align with previous research demonstrating that primary
459 (intact) forests support greater species richness and a higher proportion of specialist
460 taxa compared to disturbed environments (Barlow et al., 2007).

461
462
463
464
465
466
467
468

Mosquito diversity mirrored this pattern, with El Real again showing the highest
richness and diversity. Dominant taxa such as *Coquillettidia venezuelensis* and *Culex*
(*Melanoconion*) spp. have been implicated in arbovirus transmission, underscoring
the potential public health relevance of these ecological patterns. Western sites such
as Cirí Grande and El Cacao were less diverse, although they showed high similarity
with each other, suggesting regional cohesion in species composition.

469 For the *Culex (Melanoconion)* subgroup, species were more restricted
470 geographically. While El Real had the highest richness, Santa Librada exhibited the
471 highest diversity, indicating suitable conditions for species coexistence. Western
472 sites, including El Cacao and Cirí Grande, recorded only a single *Melanoconion*
473 species and no diversity, supporting their limited presence in more disturbed or
474 fragmented habitats.

475
476

477 Our findings on MADV and VEEV seroprevalence in small mammals, particularly
478 rodents, provide descriptive support for the hypothesis that these vertebrates
479 contribute to the maintenance of both alphaviruses (Arrigo et al., 2010a, 2010c;
480 Vittor et al., 2016a). However, although seropositivity indicates prior exposure, it
481 alone does not confirm that these species function as reservoirs. MADV was
482 confined to Darién, where outbreaks and human cases have also been reported,

51

52

53

483 while VEEV was detected in both Darién and Panamá Oeste provinces, reflecting its
484 broader distribution (Carrera et al., 2020a, 2013b). This pattern aligns with recent
485 evidence of MADV being restricted to Darién but contrasts with historical reports
486 from 1948–1970, which indicated widespread circulation across Panamá (Dietz et
487 al., 1980; Medina et al., 1965; Obaldía et al., 1991). The current geographic
488 limitation of MADV may reflect a stable enzootic habitats, with earlier outbreaks
489 pontentially representing epizootic expansions (Brault et al., 1999). (Vittor et al., 2016a)

490

491 We found lower MADV than VEEV seroprevalence in rodents (3.8% vs. 12.5%),
492 consistent with previous findings and human data showing more frequent VEEV
493 infections (Vittor et al., 2016a). This pattern mirrors human seroprevalence studies
494 and outbreaks, where VEEV infections are more frequent than MADV (Carrera et al.,
495 2020b; Rivera et al., 2024). MADV exposure was highest in *O. couesi*, *T. bolivaris*,
496 and *H. alfaroi*, while VEEV was most frequent in *T. bolivaris* and *P. semispinosus*,
497 both recognized as potential reservoirs (Aguilar et al., 2011a). The adaptability of *O.*
498 *couesi* and *T. bolivaris*, which inhabit forests, grasslands, and agricultural areas, may
499 enable enzootic cycles to expand into rural settings (del Campo et al., 2022;
500 Eubanks et al., 2011). Larvae of *Culex (Melanoconion)* spp. often develop with
501 floating plants, potentially overlapping with rodent reservoirs and facilitating
502 alphavirus transmission. While no statistical associations with land use were
503 detected, habitat differences among sites may influence host–vector distributions
504 and warrant further study (Galindo and Adames, 1973).

505

506 Our findings of higher seroprevalence in certain rodent species may, in part, reflect
507 their greater abundance in regions where viral circulation is known to occur. For
508 example, species such as *Z. brevicauda* and *O. couesi* were more frequently trapped
509 in Darién, a region with confirmed MADV activity. This spatial overlap could increase
510 the likelihood of exposure and subsequent seroconversion. This also suggests that
511 exposure risk could also be influenced by spatial overlap with virus-endemic zones
512 rather than intrinsic host susceptibility (Plowright et al., 2017). Further studies
513 incorporating quantitative measures of host density and spatial modeling are needed
514 to better distinguish between exposure-related and host-specific factors driving
515 seroprevalence patterns (Viana et al., 2014)

516

517 MADV seroprevalence in rodents was highest in Los Pavitos, the site of the first
518 reported human and equine cases in 2010 (Carrera et al., 2013a). In contrast, VEEV
519 exposure was greater in El Cacao and El Real, where higher rodent diversity and
520 forest-related activities support sylvatic transmission, MADV appears more
521 associated with agriculture and cattle ranching (Vittor et al., 2016b). Differences in
522 rodent abundance and seroprevalence between 2011 (rainy) and 2012 (dry) may
523 reflect seasonal effects on hosts and vectors (Aguilar et al., 2011b).

524

525 These transitions may influence vector activity and host exposure, potentially
526 explaining spatial differences in MADV and VEEV circulation (Vasilakis and Weaver,
527 2008). Mosquito abundance and diversity varied across sites: Cirí Grande showed
528 low richness and dominance by *Culex* spp., while similarity analyses indicated
529 localized assemblages, with overlap between Cirí Grande and El Cacao, and
530 between El Real and Santa Librada. The presence of known and suspected vector
531 species, particularly in more diverse sites, supports the plausibility of site-specific

54 11

55

56

57

58

532 enzootic cycles, similar to those described for *Culex*-borne EEEV in forested
533 wetlands of the southeastern United States (Burkett-Cadena et al., 2022a)

534

535 A focused analysis of *Culex (Melanoconion)*, a subgenus that includes potential
536 vectors of MADV and proven vectors of VEEV, showed greater species richness and
537 diversity in Darién sites, particularly El Real and Santa Librada, while sites in
538 Panamá Oeste were typically dominated by a single species. These patterns may
539 reflect site-level differences in viral exposure risk, although their relevance requires
540 further investigation through viral detection and vector competence studies (Burkett-
541 Cadena et al., 2022b).

542

543 Despite the use of molecular and viral isolation methods, no active infection was
544 detected in small mammal tissues or mosquitos pool. This may reflect the short
545 viremic period in rodents or sampling outside the window of active transmission.
546 While seropositivity indicates past exposure, the absence of viral RNA limits our
547 ability to confirm the role of these species in the enzootic cycle, particularly for
548 MADV, whose reservoirs remain poorly defined (Arrigo et al., 2010b; Calisher et al.,
549 2006). Although our serological analyses did not allow VEEV subtyping or strain-
550 specific MADV identification, the PRNT strains used were supported by previous
551 studies. TC83 shows strong antigenic similarity to subtype ID, the lineage circulating
552 in Darién and Panamá Oeste (Quiroz et al., 2009). Similarly, MADV isolates from
553 2010 and 2017 showed minimal divergence, validating the continued use of strain
554 GML-267113 for serological screening (Carrera et al., 2020a).

555 The contrasting ecological patterns observed for VEEV and MADV seroprevalence
556 highlight potentially divergent exposure dynamics between these two alphaviruses,
557 despite their co-circulation in similar environments. VEEV seroprevalence was
558 positively associated with diversity metrics in the small mammal community—
559 particularly Shannon and Simpson indices—while negatively associated with both
560 host and vector abundance. These results suggest that more balanced and species-
561 rich host communities may facilitate VEEV exposure, possibly through increased
562 contact with competent reservoirs, whereas high host density may not be a driving
563 factor. Interestingly, greater diversity within the *Culex (Melanoconion)* community
564 correlated negatively with VEEV, which may reflect a dilution effect, where increased
565 vector diversity reduces the probability of efficient transmission.

566 In contrast, MADV seroprevalence showed a strong positive correlation with host
567 abundance and moderate to high correlations with both mosquito and *Culex* richness
568 and abundance. This pattern may indicate that MADV transmission benefits from
569 environments where both vector and host densities are high, potentially reflecting
570 more focal or opportunistic cycles of transmission. The opposing trends between
571 VEEV and MADV suggest that even closely related arboviruses may exploit distinct
572 ecological conditions to persist in shared landscapes.

573 Our study has several limitations. While the overall sample size was relatively large,
574 stratifying by species or genus created small subgroups, which reduced statistical
575 power and limited our ability to perform thorough analyses of associations between
576 seroprevalence and potential variables. Additionally, the lack of precise
577 environmental data where the rodents were collected prevented us from examining
578 the micro-ecological conditions associated with rodent distribution and alphavirus

59 12

60

61

62

63

579 seroprevalence. Finer-scale analyses exploring the impact of land use and land
580 cover on diversity and alphavirus seroprevalence are currently underway, utilizing
581 additional rodent data from Darién. Furthermore, the small volume of available
582 samples limited alphavirus testing in small mammals especially for less abundant
583 species or individuals with limited sample volume.

584

585 Another key limitation is the temporal gap between data collection (2010–2012) and
586 the present. Over the past decade, environmental changes, anthropogenic
587 pressures, and shifts in host or vector populations may have altered the eco-
588 epidemiological landscape of MADV and VEEV in the region. While these factors
589 may influence current transmission dynamics, our findings remain valuable as they
590 provide a historical baseline for understanding spatial patterns of exposure and
591 identifying potential vertebrate hosts. These data are essential for informing future
592 surveillance efforts and comparative studies in the context of changing ecological
593 conditions.

594

595 Seroprevalence studies in animal populations constitute a critical initial approach in
596 identifying potential reservoir hosts by indicating prior exposure to a pathogen.
597 However, serological evidence alone is insufficient to confirm reservoir competence.
598 A comprehensive assessment requires the integration of additional methodologies,
599 including pathogen isolation, molecular diagnostics, longitudinal surveillance to
600 evaluate persistence and shedding, ecological and behavioral investigations to
601 elucidate host-pathogen interactions, and experimental infection studies to determine
602 susceptibility, transmission potential, and pathogenesis under controlled conditions.
603 Moreover, it is essential to investigate a broader range of vertebrate species to
604 capture the full spectrum of potential reservoirs and better understand the ecological
605 complexity of pathogen maintenance and transmission.

606

607 In summary, our findings support the hypothesis that wild rodents may play a role in
608 the enzootic maintenance of both MADV and VEEV, as suggested by distinct
609 seroprevalence patterns across species and regions. These patterns appear to be
610 influenced by a combination of spatial factors, species-specific traits, and ecological
611 conditions, indicating the complex dynamics of arbovirus circulation in Panama.

612

613 **Acknowledgments**

614 We wish to express appreciation to Yaneth Pittí, Isela Guerrero, Davis Beltran and
615 Julio Cisneros for technical with laboratory testing. Alberto Cumbreira for helping with
616 the map. We also thank Fatima Rodriguez for funding administration. Special thanks
617 go to the health technicians from Herrera—Ricardo Rodríguez, Juan Bosco Navarro,
618 Mario Ávila, and Rigoberto Fernández (ZOMASPA)—whose invaluable contributions
619 were instrumental in the collection and processing of biological material in the field.
620 We extend our heartfelt gratitude to the entire staff of the Vector Control Department
621 in the Darién and Eastern Panamá Health Regions for their unwavering support and
622 dedication. We also remember and honor Santos Vegas (†), zone coordinator in
623 Darién, for his contributions.

624

625 **Funding**

626 JPC was supported by the Clarendon Scholarship from University of Oxford and
627 Lincoln-Kingsgate Scholarship from Lincoln College, University of Oxford [grant

64 13

65

66
67
68

628 number SFF1920_CB2_MPLS_1293647]. JFG is a masters student studying
629 Epidemiological Research at Universidad Peruana Cayetano Heredia supported by
630 training grant D43 TW007393 awarded by the Fogarty International Center of the US
631 National Institutes of Health. This work was supported by SENACYT [grant number
632 FID- 16-201 and FID23-147] grant to JPC. Proyecto: Estudio de las Enfermedades
633 Emergentes y Síndromes Febriles en la Población Migrante, Ministerio de Economía
634 y Finanzas de Panamá (Grant: 019911.013). Proyecto de Caracterización del subtipo
635 circulante de las Encefalitis equinas, Ministerio de Economía y Finanzas de Panamá
636 (Grant: 9044-027) to LCC. This research was in part funded by the Centers for
637 Research in Emerging Infectious Diseases “The Coordinating Research on
638 Emerging Arboviral Threats Encompassing the Neotropics (CREATE-NEO)” grant
639 1U01AI151807 by the National Institutes of Health (NIH/USA) to NV. CAD
640 acknowledges funding the National Institute of Health Research for support of the
641 Health Protection Research Unit in Emerging and Zoonotic Infections. WMS was
642 supported by the Global Virus Network fellowship and the NIH (AI12094) Global
643 Virus Network fellowship, Burroughs Wellcome fund (#1022448) and Wellcome
644 Trust-Digital Technology Development award (Climate Sensitive Infectious Disease
645 Modelling; (226075/Z/22Z). NRF acknowledges support from Wellcome Trust and
646 Royal Society Sir Henry Dale Fellowship (204311/Z/16/Z), Bill and Melinda Gates
647 Foundation (INV034540) and Medical Research Council-Sao Paulo Research
648 Foundation (FAPESP) CADDE partnership award (MR/S0195/1 and FAPESP
649 18/14389-0). LC is member of the Sistema Nacional de Investigación (SNI),
650 SENACYT, Panama. SCW and WMS were supported by NIH grant R24 AI120942.
651

652 **References**

653 Aguilar P V., Estrada-Franco JG, Navarro-Lopez R, et al. Endemic Venezuelan Equine
654 Encephalitis in the Americas: Hidden under the Dengue Umbrella. *Future Virol* 2011a; doi:
655 10.2217/fvl.11.50.
656 Aguilar P V, Estrada-Franco JG, Navarro-Lopez R, et al. Endemic Venezuelan Equine
657 Encephalitis in the Americas: Hidden under the Dengue Umbrella. *Future Virol*
658 2011b;6(6):721–740.
659 Aguilar P V., Robich RM, Turell MJ, et al. Endemic Eastern Equine Encephalitis in the
660 Amazon Region of Peru. *American Journal of Tropical Medicine and Hygiene* 2007; doi:
661 10.4269/ajtmh.2007.76.293.
662 Reid, FA. FIELD GUIDE TO THE MAMMALS OF CENTRAL AMERICA AND
663 SOUTHEAST MEXICO. 2009.
664 Arrigo NC, Adams AP and Weaver SC. Evolutionary Patterns of Eastern Equine Encephalitis
665 Virus in North versus South America Suggest Ecological Differences and Taxonomic
666 Revision. *J Virol* 2010a; doi: 10.1128/jvi.01586-09.
667 Arrigo NC, Adams AP and Weaver SC. Evolutionary Patterns of Eastern Equine Encephalitis
668 Virus in North versus South America Suggest Ecological Differences and Taxonomic
669 Revision. *J Virol* 2010b;84(2):1014–1025; doi:
670 10.1128/JVI.01586-09/SUPPL_FILE/SUPPLEMENTAL_VEEV_TABLE.DOC.
671 Arrigo NC, Paige Adams A, Watts DM, et al. Cotton Rats and House Sparrows as Hosts for
672 North and South American Strains of Eastern Equine Encephalitis Virus. *Emerg Infect Dis*
673 2010c; doi: 10.3201/eid1609.100459.
674 Autoridad Nacional del Ambiente. Atlas Ambiental. 2010.

71
72
73

- 675 Barlow J, Gardner TA, Araujo IS, et al. Quantifying the Biodiversity Value of Tropical
676 Primary, Secondary, and Plantation Forests. *Proc Natl Acad Sci U S A* 2007;104(47):18555–
677 18560; doi: 10.1073/PNAS.0703333104/SUPPL_FILE/03333TABLE4.PDF.
- 678 Brault AC, Powers AM, Villarreal Chavez CL, et al. Genetic and Antigenic Diversity among
679 Eastern Equine Encephalitis Viruses from North, Central, and South America. *American*
680 *Journal of Tropical Medicine and Hygiene* 1999;61(4); doi: 10.4269/ajtmh.1999.61.579.
- 681 Burkett-Cadena ND, Day JF and Unnasch TR. Ecology of Eastern Equine Encephalitis Virus
682 in the Southeastern United States: Incriminating Vector and Host Species Responsible for
683 Virus Amplification, Persistence, and Dispersal. *J Med Entomol* 2022a;59(1):41–48; doi:
684 10.1093/JME/TJAB076.
- 685 Burkett-Cadena ND, Day JF and Unnasch TR. Ecology of Eastern Equine Encephalitis Virus
686 in the Southeastern United States: Incriminating Vector and Host Species Responsible for
687 Virus Amplification, Persistence, and Dispersal. *J Med Entomol* 2022b;59(1):41–48; doi:
688 10.1093/JME/TJAB076.
- 689 Calisher CH, Childs JE, Field HE, et al. Bats: Important Reservoir Hosts of Emerging
690 Viruses. *Clin Microbiol Rev* 2006;19(3):531; doi: 10.1128/CMR.00017-06.
- 691 del Campo JTF, Olvera-Vargas M, Silla-Cortés F, et al. Composition and Structure of
692 Vegetation and Tide Regulate the Occurrence of *Oryzomys Couesi* and *Hodomys Alleni* in
693 Mangrove Forests of Laguna de Cuyutlán, West-Central Mexico. *Wetl Ecol Manag*
694 2022;30(1); doi: 10.1007/s11273-021-09831-7.
- 695 Carrera JP, Bagamian KH, Travassos Da Rosa AP, et al. Human and Equine Infection with
696 Alphaviruses and Flaviviruses in Panamá during 2010: A Cross-Sectional Study of
697 Household Contacts during an Encephalitis Outbreak. *American Journal of Tropical*
698 *Medicine and Hygiene* 2018; doi: 10.4269/ajtmh.17-0679.
- 699 Carrera JP, Cucunuba ZM, Neira K, et al. Endemic and Epidemic Human Alphavirus
700 Infections in Eastern Panama, An Analysis of Population-Based Cross-Sectional Surveys.
701 *bioRxiv* 2020a;10(01):901462; doi: 10.1101/2020.01.10.901462.
- 702 Carrera J-P, Forrester N, Wang E, et al. Eastern Equine Encephalitis in Latin America. *New*
703 *England Journal of Medicine* 2013a; doi: 10.1056/nejmoa1212628.
- 704 Carrera J-P, Forrester N, Wang E, et al. Eastern Equine Encephalitis in Latin America. *New*
705 *England Journal of Medicine* 2013b;369(8); doi: 10.1056/nejmoa1212628.
- 706 Carrera J-P, Cucunuba ZM, Neira K, et al. Endemic and Epidemic Human Alphavirus
707 Infections in Eastern Panama: An Analysis of Population-Based Cross-Sectional Surveys.
708 *American Journal of Tropical Medicine and Hygiene* 2020b;10(01):901462; doi:
709 10.4269/ajtmh.20-0408.
- 710 Causey OR, Causey CE, Maroja OM, et al. The Isolation of Arthropod-Borne Viruses,
711 Including Members of Two Hitherto Undescribed Serological Groups, in the Amazon Region
712 of Brazil. *Am J Trop Med Hyg* 1961;10; doi: 10.4269/ajtmh.1961.10.227.
- 713 Dalgaard P. R Development Core Team (2010): R: A Language and Environment for
714 Statistical Computing. 2010.
- 715 Deardorff ER, Forrester NL, Travassos Da Rosa AP, et al. Experimental Infection of
716 Potential Reservoir Hosts with Venezuelan Equine Encephalitis Virus, Mexico. *Emerg Infect*
717 *Dis* 2009a;15(4):519; doi: 10.3201/EID1504.081008.
- 718 Deardorff ER, Forrester NL, Travassos Da Rosa AP, et al. Experimental Infection of
719 Potential Reservoir Hosts with Venezuelan Equine Encephalitis Virus, Mexico. *Emerg Infect*
720 *Dis* 2009b;15(4); doi: 10.3201/eid1504.081008.
- 721 Death R. Margalefs Index - Population Dynamics - Ecology Center. Ecology Center - Massey
722 University 2021.

74 15
75

76
77
78

- 723 Dietz WH, Galindo P and Johnson KM. Eastern Equine Encephalomyelitis in Panama: The
724 Epidemiology of the 1973 Epizootic. *American Journal of Tropical Medicine and Hygiene*
725 1980; doi: 10.4269/ajtmh.1980.29.133.
- 726 Eubanks BW, Hellgren EC, Nawrot JR, et al. Habitat Associations of the Marsh Rice Rat
727 (*Oryzomys Palustris*) in Freshwater Wetlands of Southern Illinois. *J Mammal* 2011;92(3);
728 doi: 10.1644/10-MAMM-A-289.1.
- 729 Galindo P and Adames AJ. Ecological Profile of *Culex* (*Melanoconion*) *Aikenii* (Diptera:
730 Culicidae), Vector of Endemic Venezuelan Encephalitis in Panama1. *Environ Entomol*
731 1973;2(1); doi: 10.1093/ee/2.1.81.
- 732 Gregorius HR and Gillet EM. Generalized Simpson-Diversity. *Ecol Modell* 2008;211(1–2);
733 doi: 10.1016/j.ecolmodel.2007.08.026.
- 734 Hammer DAT, Ryan PD, Hammer Ø, et al. Past: Paleontological Statistics Software Package
735 for Education and Data Analysis. *Palaeontologia Electronica* 2001;4(1):178.
- 736 Kinney RM, Tsuchiya KR, Sneider JM, et al. Genetic Evidence That Epizootic Venezuelan
737 Equine Encephalitis (VEE) Viruses May Have Evolved from Enzoitic VEE Subtype I-D
738 Virus. *Virology* 1992;191(2); doi: 10.1016/0042-6822(92)90232-E.
- 739 Magalhaes T, Hamer GL, de Carvalho-Leandro D, et al. Uncertainties Surrounding
740 Madariaga Virus, a Member of the Eastern Equine Encephalitis Virus Complex. *Vector-*
741 *Borne and Zoonotic Diseases* 2024; doi: 10.1089/vbz.2023.0162.
- 742 Medina G, Gleiser CA and Mackenzi RB. Brote de Encefalomiélitis Equina En La
743 Republica de Panama. *Boletín de la Oficina Sanitaria Panamericana* 1965;58(5):390–394.
- 744 Nanda A, Mohapatra DrBB, Mahapatra APK, et al. Multiple Comparison Test by Tukey’s
745 Honestly Significant Difference (HSD): Do the Confident Level Control Type I Error.
746 *International Journal of Statistics and Applied Mathematics* 2021;6(1); doi:
747 10.22271/math.2021.v6.i1a.636.
- 748 Navarro JC, Carrera JP, Liria J, et al. Alphaviruses in Latin America and the Introduction of
749 Chikungunya Virus. In: *Human Virology in Latin America: From Biology to Control* 2017;
750 doi: 10.1007/978-3-319-54567-7_9.
- 751 Obaldía N, Dutary B, Clavel F, et al. Encefalomiélitis Equina Del Este, Epizootia de 1986 En
752 Panamá. *Notas veterinarias* 1991;1(3):4–7.
- 753 Ortiz-Burgos S. Shannon-Weaver Diversity Index. In: *Encyclopedia of Earth Sciences Series*
754 2016; doi: 10.1007/978-94-017-8801-4_233.
- 755 Plowright RK, Parrish CR, McCallum H, et al. Pathways to Zoonotic Spillover. *Nature*
756 *Reviews Microbiology* 2017 15:8 2017;15(8):502–510; doi: 10.1038/nrmicro.2017.45.
- 757 Quiroz E, Aguilar P V., Cisneros J, et al. Venezuelan Equine Encephalitis in Panama: Fatal
758 Endemic Disease and Genetic Diversity of Etiologic Viral Strains. *PLoS Negl Trop Dis*
759 2009;3(6).
- 760 Rivera LF, Lezcano-Coba C, Galué J, et al. Characteristics of Madariaga and Venezuelan
761 Equine Encephalitis Virus Infections, Panama. *Emerg Infect Dis* 2024;30(14):94–104; doi:
762 10.3201/EID3014.240182.
- 763 Sánchez-Seco MP, Rosario D, Quiroz E, et al. A Generic Nested-RT-PCR Followed by
764 Sequencing for Detection and Identification of Members of the Alphavirus Genus. *J Virol*
765 *Methods* 2001; doi: 10.1016/S0166-0934(01)00306-8.
- 766 Sotomayor-Bonilla J, Abella-Medrano CA, Chaves A, et al. Potential Sympatric Vectors and
767 Mammalian Hosts of Venezuelan Equine Encephalitis Virus in Southern Mexico. *J Wildl Dis*
768 2017;53(3); doi: 10.7589/2016-11-249.
- 769 Srihongse S and Galindo P. The Isolation of Eastern Equine Encephalitis Virus from *Culex*
770 (*Melanoconion*) *Taeniopus* Dyar and Knab in Panama. *Mosquito News* 1967;27(1):74–76.
- 771 Torres R, Young J, Maárquez R, et al. Enzoitic Mosquito Vector Species at Equine
772 Encephalitis Transmission Foci in the República de Panama. *PLoS One* 2017.

81

82

83

773 Turell MJ, O'Guinn ML, Dohm D, et al. Susceptibility of Peruvian Mosquitoes to Eastern
774 Equine Encephalitis Virus. *J Med Entomol* 2008; doi: 10.1603/0022-
775 2585(2008)45[720:SOPMTE]2.0.CO;2.

776 Turell MJ, O'Guinn ML, Jones JW, et al. Isolation of Viruses from Mosquitoes (Diptera:
777 Culicidae) Collected in the Amazon Basin Region of Peru. *J Med Entomol* 2005;42(5):891–
778 898; doi: 10.1093/jmedent/42.5.891.

779 Vasilakis N and Weaver SC. The History and Evolution of Human Dengue Emergence. *Adv*
780 *Virus Res* 2008;72:1–76; doi: 10.1016/S0065-3527(08)00401-6.

781 Viana M, Mancy R, Biek R, et al. Assembling Evidence for Identifying Reservoirs of
782 Infection. *Trends Ecol Evol* 2014;29(5):270–279; doi: 10.1016/J.TREE.2014.03.002/ASSET/
783 4F3900E0-87F9-44A0-AD17-388669C97681/MAIN.ASSETS/GR2B3.JPG.

784 Vittor AY, Armien B, Gonzalez P, et al. Epidemiology of Emergent Madariaga Encephalitis
785 in a Region with Endemic Venezuelan Equine Encephalitis: Initial Host Studies and Human
786 Cross-Sectional Study in Darien, Panama. *PLoS Negl Trop Dis* 2016a;10(4); doi:
787 10.1371/journal.pntd.0004554.

788 Vittor AYAY, Armien B, Gonzalez P, et al. Epidemiology of Emergent Madariaga
789 Encephalitis in a Region with Endemic Venezuelan Equine Encephalitis: Initial Host Studies
790 and Human Cross-Sectional Study in Darien, Panama. *PLoS Negl Trop Dis* 2016b;10(4); doi:
791 10.1371/journal.pntd.0004554.

792 Weaver SC, Ferro C, Barrera R, et al. Venezuelan Equine Encephalitis. *Annu Rev Entomol*
793 2004; doi: 10.1146/annurev.ento.49.061802.123422.

794

795

796

86
87
88
797

Table 1. Characteristics of collection sites, small mammal diversity and alphavirus seroprevalence. The total of small

798 mammals included in the diversity analysis was 599 from a total of 16 species.

Sites	Panamá Oeste Province			Darién Province	
	El Cacao	Cirí Grande	El Real de Santa María	Los Pavitos	Santa Librada
Latitude and Longitude	8,76613418; -80,0168149	8,8712848; -80,053276	8,13021585; -77,727379	8,47052; -77,9549	8,157787; -77,691809
*No. rodents captured (n)	56	18	202	165	158
**No. rodent species (n)	6	5	11	7	5
†VEEV seroprevalence	15/55 (27.3, 95% CI: 16.1-40.9)	0/17 (0.0%, 95% CI: 0.0-19.5)	19/94 (20.2%, 95% CI:12.6-29.7)	2/81 (2.4%, 95% CI:0.3-8.6)	1/49 (2.0%, 95% CI:0.0-10.8)
††MADV seroprevalence	0/55 (0.0%, 95% CI:0.0 - 6.0)	0/17 (0.0%, 95% CI: 0.0- 19.5)	3/94 (3.2%, 95% CI:0.6-9.0)	7/78 (9.0%, 95% CI:3.6-17.6)	1/48 (2.1%, 95% CI:0.0-11.8)
Simpson's	0.53	0.46	0.60	0.23	0.11

89 18
90

91
92
93

diversity (Ds)						
Shannon-Wiener						
(H)	1.13	0.96	1.42	0.57	0.29	
Margalef index						
(M)	1.24	1.38	1.88	1.18	0.79	
Chao-1	6.00	6.41	11.5	7.49	5.99	
iChao-1	6.47	9.25	12.9	9.35	6.48	

799 †VEEV: n=296

800 †† MADV: n=292

801

802

803

96
97
98
804

Table 2. Pairwise comparison of rodent species by collection site.

Sites	Contrast	95% CI	P-value*
Cirí Grande vs El Real	-2.3	-4.4 - - 0.2	0.023
El Cacao vs El Real	-1.8	-3.1- - 0.5	0.001
Los Pavitos vs El Real	2.1	1.2 - 3.0	<0.001
Santa Librada vs El Real	2.6	1.6 - 3.5	<0.001
El Cacao vs Cirí Grande	0.5	-1.8 - 2.8	0.974
Los Pavitos vs Cirí Grande	4.4	2.3 - 6.5	<0.001
Santa Librada vs Cirí Grande	4.9	2.8 - 7.0	<0.001
Los Pavitos vs El Cacao	3.9	2.6 - 5.2	<0.001
Santa Librada vs El Cacao	4.4	3.0 - 5.7	<0.001
Santa Librada vs Los Pavitos	0.5	-0.5 - 1.4	0.639

805 *P-value: Statistically significant values ($p < 0.05$) are shown in bold

806
807

101
102
103
808

809 **Table 3. Seroprevalences by virus, collection sites and year of trapping.**

Sites	MADV				VEEV			
	2011†		2012*		2011††		2012**	
	n/N (%)	95% CI	n/N (%)	95% CI	n/N (%)	95% CI	n/N (%)	95% CI
El Real	3/72 (4.2)	0.01 - 0.12	0/22 (0.0)	0.00 - 0.15	17/72 (23.6)	0.14 - 0.35	2/22 (9.1)	0.01 - 0.29
Los Pavitos	6/60 (10.0)	0.04 - 0.21	1/18 (5.6)	0.00 - 0.27	2/63 (3.2)	0.00 - 0.11	0/18 (0.0)	0.00 - 0.19
Santa Librada	0/9 (0.0)	0.00 - 0.34	1/39 (2.6)	0.00 - 0.13	0/9 (0.0)	0.00 - 0.34	1/40 (2.5)	0.00 - 0.13
El Cacao	0/41 (0.0)	0.00 - 0.86	0/14 (0.0)	0.00 - 0.23	13/41 (31.7)	0.18 - 0.48	2/14 (14.3)	0.18 - 0.43
Cirí Grande	0/12 (0.0)	0.00 - 0.26	0/5 (0.0)	0.00 - 0.52	0/12 (0.0)	0.00 - 0.26	0/5 (0.0)	0.00 - 0.52

810 †Seroprevalence total of MADV by 2011: n=9/194; 4.6%, 95% CI (0.02 to 0.09)

104 21
105

106

107

108

811 *Seroprevalence total of MADV by 2012: n=2/98; 2.0 %, 95% CI (0.00 to 0.07)

812 ††Seroprevalence total of VEEV by 2011: n=32/197; 16.2%, 95% CI (0.11 to 0.22)

813 **Seroprevalence total of VEEV by 2012: n=5/99; 5.1%, 95% CI (0.02 to 0.11)

109 22

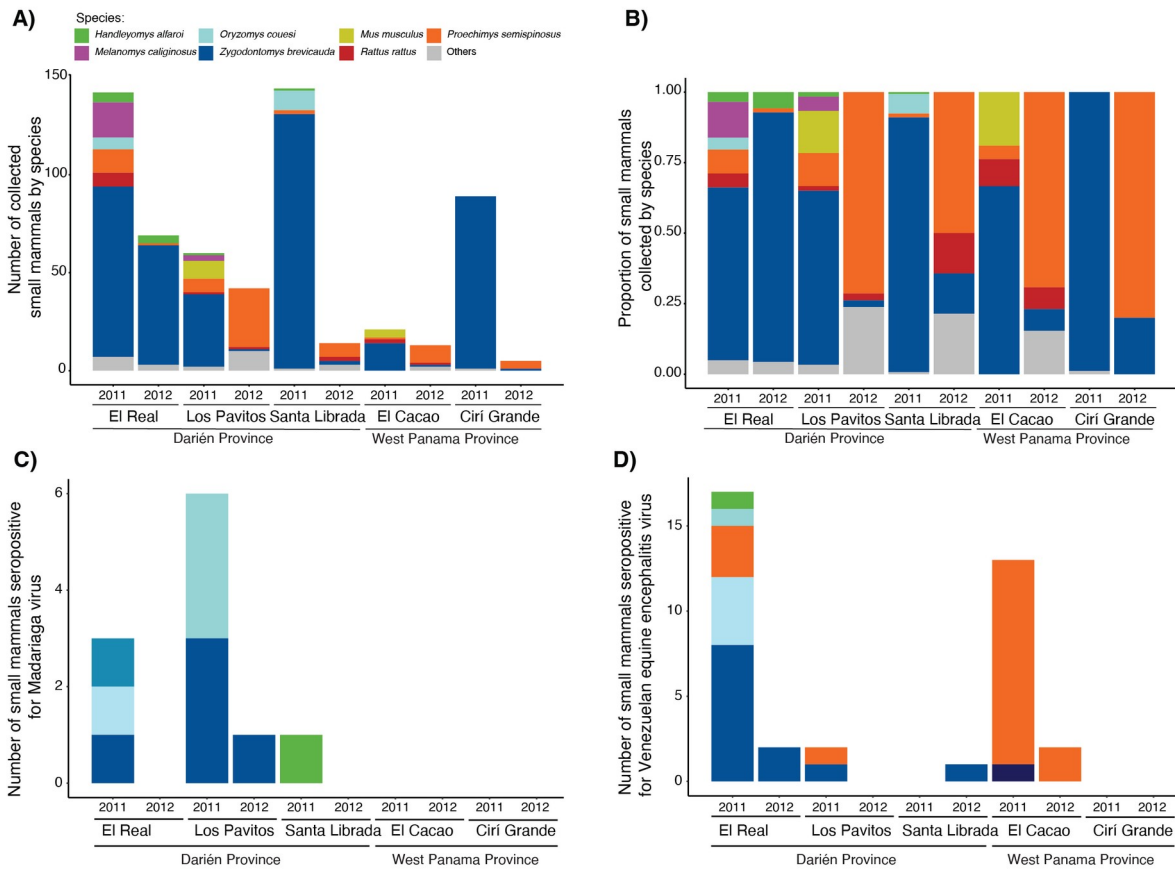
110

111
112
113
814



815
816
817

Figure 1. Study Areas and Collection Sites.



818

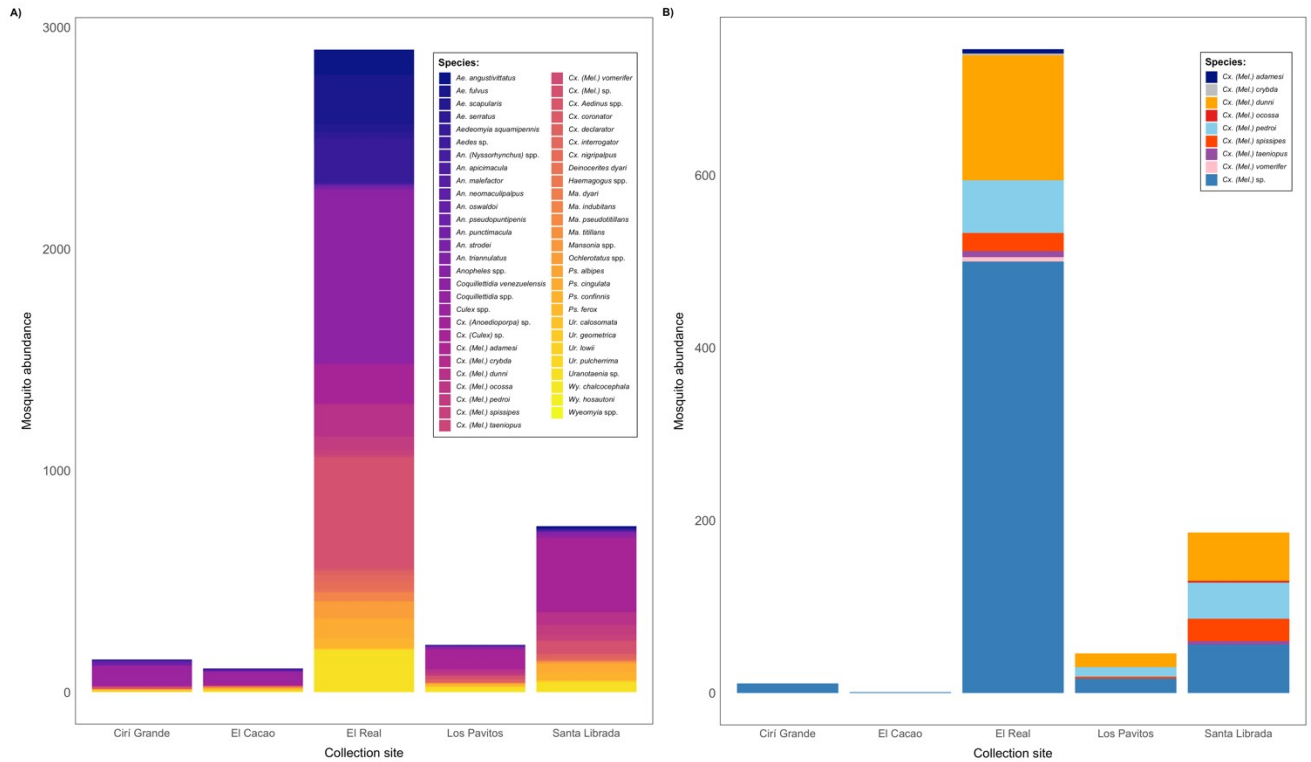
819 **Figure 2. Alphavirus seropositivity in small mammals collected across study**
820 **sites in Panama. A) Number of sampled small mammal species by site and year. B)**

821 Proportion of sampled small mammals by site and year. C) Number of small
822 mammals seropositive for Madariaga virus (MADV). D) Number of small mammals
823 seropositive for Venezuelan equine encephalitis virus (VEEV). Years/sites with no
824 bars indicate zero detections.

825

826

121
122
123



827

828 **Figure 3. Mosquitoes collected across study sites.** A) Mosquito species
829 composition and abundance by collection site in Darién and Panamá Oeste
830 provinces. B) Species composition and abundance of *Culex*
831 (*Melanoconion*) mosquitoes across five collection sites.

832

833

834

835

836

837

838 **Figure 4. Spearman correlation heatmap of ecological diversity indices and**
839 **alphavirus seroprevalence in Darién and Panamá Oeste provinces.**

840 Heatmap showing Spearman correlation coefficients (ρ) among ecological indicators:
841 abundance, diversity, and richness indices for small mammals, mosquitoes,
842 and *Culex (Melanoconion)* mosquitoes and site-level seroprevalence of VEEV and
843 Madariaga virus MADV. Color intensity represents the strength and direction of the
844 correlation (red = positive, blue = negative).