
Cell cycle regulation has shaped replication origins in budding yeast

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Supplementary Information

EcoRI site

XmaI site

Matches to the ORC consensus sequence on the top or bottom strand

LacO

Nucleosome binding site

Abf1

These sequences were cloned in pUC19 at EcoRI and XmaI sites

pCTL40 10bp

gaattcCTCGA TTTT TTTATGTTTAGTT TCGCGGACGACGGTTTCGAGGTGGCGAA
AAGCGTCTGAAGCGG AACTAAACATAAAAAA GCGGTGTTCCGCCGAGATCGGCTC
GCGCATGGCCGAGTTGAGCGGTTCCAGGCTGGCCACGCAGCATCAGATGGTA
GGCCTCCTGGCGCCGCACCGGCCTCAGCATCCGGTACCTCAGCTGGCCACCG
TCGGCGTCTCGCACGACCACCAAGTGCAAGGGTCTGAGCAGCGCCGTCTGTGCT
CCTCGGAGTGGAGGCAGCCGAGCGCGACGGTGTGCCCCGCTTCCTGGAGACC
TCCGCGCTCCGCAACCTCCACTTCTACGAGCGGCTCGGCTTCACCGTCACCGC
CGACGTCTGAGGTGCCCCGAAGGACCGCGCACCTGGTGCATGACCCGCAAGCCC
GGTGCCTGACGCTCGCCACACGACCCGCAGCGCCCGACCGAAAGGAGCGCAC
GA cccggg

pCTL9 12bp

gaattcCTCGA TTTT TTTATGTTTAGTT TCGCGGACGACGGTTTCGAGGTGGCGGT
AAAAGCGTCTGAAGCGG AACTAAACATAAAAAA GGTGTTCCGCCGAGATCGGCTC
GCGCATGGCCGAGTTGAGCGGTTCCAGGCTGGCCACGCAGCATCAGATGGTA
GGCCTCCTGGCGCCGCACCGGCCTCAGCATCCGGTACCTCAGCTGGCCACCG
TCGGCGTCTCGCACGACCACCAAGTGCAAGGGTCTGAGCAGCGCCGTCTGTGCT
CCTCGGAGTGGAGGCAGCCGAGCGCGACGGTGTGCCCCGCTTCCTGGAGACC
TCCGCGCTCCGCAACCTCCACTTCTACGAGCGGCTCGGCTTCACCGTCACCGC
CGACGTCTGAGGTGCCCCGAAGGACCGCGCACCTGGTGCATGACCCGCAAGCCC
GGTGCCTGACGCTCGCCACACGACCCGCAGCGCCCGACCGAAAGGAGCGCAC
GA cccggg

pCTL86 ARS317

gaattcACTAGTACTTAAAAAACTGTAGTTTCAGTGCAAAAAAGTTTAAACATTAC
GTATCTTGTACCCTTTTTATTGCATATAGAAAGGTCAAATAATCCTTCACATCATG
AAATATAAGCTAAATCGCATTTCTTTTCGTCCACATTTGCAAACAAAACCTTTTCAA
TAATAATTTTATAAATAGTATCAATATATATATATATATATATATATTTTATTGTTTACT
TTTTCTATCAGTGTTTTCAATTTTTTATTAACAATGTTTGATTTTTTAAATCGCAA
TTTAAT ACCTAAATATAAAAAA GTTATTATATTGCAAAAACCCATCAACCTTGAA
AAAAAGTAGAAACGTTTTTATTTAATTCTATCAATACATCATAAAATACGAACGATC
CCCGTCCAAGTTATGAGCTTAATCTTCCATAAAAAATTTGAAAGCAATAGATCA
TGTAATAAATAAATCAGGGAAATT cccggg

pCTL91 90bp-B2

gaattcCTCGA TTTT TTTATGTTTAGTT TCGCGGACGACGGTTTCGAGGTGGCGGT
CTGGACCACGCCGGAGAGCGTCTGAAGCGGAGGCGGTGTTCCGCCGAGATCGG
CTCGCGCATGGCCGAGTTCAAAAGGCCTGCAGGCAAGTGCACAAACAATACTT
AAATAAATACTACTCAGTAATAACCTATTTCTTAGCATTTTTTGACGAAATTCCGTC

GGCGTCTCGCACGACCACCAGTGCAAGGGTCTGAGCAGCGCCGTCGTGCTCC
TCGGAGTGGAGGCAGCCGAGCGCGACGGTGTGCCCCGCCTTCCTGGAGACCTC
CGCGCTCCGCAACCTCCACTTCTACGAGCGGCTCGGCTTCACCGTCACCGCCG
ACGTCGAGGTGCCCCGAAGGACCGCGCACCTGGTGCATGACCCGCAAGCCCGG
TGCCTGACGCTCGCCACACGACCCGCGAGCGCCCGACCGAAAGGAGCGCACGA
cccg

pCTL93 300bp[*lacO*]

gaattcCTCGAATTTTTTATGTTTAGTTTCGCGGACGACGGTTTCGAGGTGGCGGT
CTGGACCACGCCGGAGAGCGTCTGAAGCGGAGGCGGTGTTCCGCGAGATCGG
GAGCTCTCACACCTACAAGGGATGTACATCAATTGTGAGCGGATAACAATTGTT
AGGGAGGAATTGTGAGCGGATAACAATTTGGAGTTGATAATTGTGAGCGGATAA
CAATTGGCTTCAACGTAATTGTGAGCGGATAACAATTTCCGTACGAATGTGCCG
AACTTATGGTACCGCCGAGCGCGACGGTGTGCCCCGCCTTCCTGGAGACCTCC
GCGCTCCGCAACCTCCACTTCTAAAAGCGGCTCGGCTTCAACTAAACATAAAAA
tACGAGGTGCCCCGAAGGACCGCGCACCTGGTGCATGACCCGCAAGCCCGGTG
CCTGACGCTCGCCACACGACCCGCGAGCGCCCGACCGAAAGGAGCGCACGAccc
ggg

pCTL94 One site[*lacO*]

gaattcCTCGAATTTTTTATGTTTAGTTTCGCGGACGACGGTTTCGAGGTGGCGGT
CTGGACCACGCCGGAGAGCGTCTGAAGCGGAGGCGGTGTTCCGCGAGATCGG
GAGCTCTCACACCTACAAGGGATGTACATCAATTGTGAGCGGATAACAATTGTT
AGGGAGGAATTGTGAGCGGATAACAATTTGGAGTTGATAATTGTGAGCGGATAA
CAATTGGCTTCAACGTAATTGTGAGCGGATAACAATTTCCGTACGAATGTGCCG
AACTTATGGTACCGCCGAGCGCGACGGTGTGCCCCGCCTTCCTGGAGACCTCC
GCGCTCCGCAACCTCCACTTCTACGAGCGGCTCGGCTTCACCGTCACCGCCGA
CGTCGAGGTGCCCCGAAGGACCGCGCACCTGGTGCATGACCCGCAAGCCCGGT
GCCTGACGCTCGCCACACGACCCGCGAGCGCCCGACCGAAAGGAGCGCACGAC
ccggg

pTM0005 N-ARS1-N

ccgcccttgagaatcGcggtgccgaggccgctcaattggctcgtagacagctctagcaccgctaaacgcacgtacg
cgctgtccccgcgtttaaccgccaaggggattactccctagctccaggcacgtgtcagatatatacatcctgtATT
TTACAGATTTTATGTTTAGATCTTTTATGCTTGCTTTTCAAAGGCCTGCAGGCA
AGTGCACAAACAATACTTAAATAAATACTACTCAGTAATAACCTATTTCTTAGCAT
TTTTGACGAAATTTGCTATTTTCCCAGTTCGCGCGCCACCTACCGTGTGAAGT
CGTCACTCGGGCTTCTAAGTACGCTTAGCGCACGGTAGAGCGCAATCCAAGGC
TAACCACCGTGCATCGATGTTGAAAGAGGCCCTCCGTCCTTATTACTTCAAGTC
CCTGGGGTgcagg

pTM0023 N-90bp-N

ccgcccttgagaatcGcggtgccgaggccgctcaattggctcgtagacagctctagcaccgctaaacgcacgtacg
cgctgtccccgcgtttaaccgccaaggggattactccctagctccaggcacgtgtcagatatatacatcctgtATC
TCGAATTTTTTATGTTTAGTTTCGCGGACGACGGTTTCGAGGTGGCGGTCTGGACCACG
CCGGAGAGCGTGTTCAGGCTGGCCACGCGAGCGAAGCGGAGGCGGTGTTCCGCGAG
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GCGCGCCACCTACCGTGTGAAGTCGTCACTCGGGCTTCTAAGTACGCTTAGC
GCACGGTAGAGCGCAATCCAAGGCTAACCACCGTGCATCGATGTTGAAAGAGG
CCCTCCGTCCTTATTACTTCAAGTCCCTGGGGTgcagg

pCTL68 ARS1-Abf1

catatgcggtgtgaaataaccgcacagatgcgtaaggagaaaataaccgcatcaggcgccattcgccattcaggctgc
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gcgattaagttgggtaacTATTTCCTTAGCATTTTTGACGAAATTgtaaaacgacggccagtgaaattc
ATTTTACAGATTTTATGTTTAGATCTTTTATGCTTGCTTTTCAAAGGCCTGCAGG
CAAGTGCACAAACAATACTTAAATAAATACTACTCAGTAATAACCATTTCCTTAGC
ATTTTTGACGAAATTGCTATTTTGTAGAGTCTTTTACACCATTTGTCTCCACAC
CTCCGCTTACATCAACACCAATAACGCCATTTAATCTAAGCGCATCACCAACATT
TTCTGGCGTCAGTCCACCAGCTAACATAAAATGTAAGCTTTTCGGGGCTCTCTTG
CCTTCCAACCCAGTCAGAAATCGAGTTCCAATCCAAAAGTTCACCTGTCCCACC
TGCTTCTGAATCAAACAAGGGAATAAACGAATGAGGTTTCTGTGAAGCTGCACT
GAGTAGTATGTTGCAcccggggatcc

pCTL67 90bp-Abf1

catatgcggtgtgaaataaccgcacagatgcgtaaggagaaaataaccgcatcaggcgccattcgccattcaggctgc
gcaactgttggaagggcgatcgggtgcgggcctcttcgctattacgccagctggcgaaagggggatgtgctgcaag
gcgattaagttgggtaacgccagggtTATTTCCTTAGCATTTTTGACGAAATTacggccagtgaaattc
CTCGATTTTTTATGTTTAGTTTCGCGGACGACGGTTTCGAGGTGGCGGTCTGG
ACCACGCCGGAGAGCGTCAAGCGGAGGCGGTGTTTCGCCGAGATCGGCTCGC
GCATGGCCGAGTTGAGCGGTTCAAAGCTGGCCACGCAGAACTAAACATAAAAAA
ACCTGGCGCCGCTATTTCCTTAGCATTTTTGACGAAATTCTGGCCACCGTCGGCG
TCTCGCACGACCACAGTGCAAGGGTCTGAGCAGCGCCGTCGTGCTCCTCGG
AGTGGAGGCAGCCGAGCGCGACGGTGTGCCCGCCTTCCTGGAGACCTCCGCG
CTCCGCAACCTCCACTTCTACGAGCGGCTCGGCTTCACCGTCACCGCCGACGT
CGAGGTGCCCCGAAGGACCGCGCACCTGGTGCATGACCCGCAAGCCCGGTGCC
TGACGCTCGCCACACGACCCGCAGCGCCCGACCGAAAGGAGCGCACGAcccgg
ggatcc

pCTL69 ARS317-Abf1

catatgcggtgtgaaataaccgcacagatgcgtaaggagaaaataaccgcatcaggcgccattcgccattcaggctgc
gcaactgttggaagggcgatcgggtgcgggcctcttcgctattacgccagctggcgaaagggggatgtgctgcaag
gcgattaagttgggtaacTATTTCCTTAGCATTTTTGACGAAATTgtaaaacgacggccagtgaaattc
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TTGTACCCTTTTTATTGCATATAGAAAGGTCAAATAATCCTTCACATCATGAAATA
TAAGCTAAATCGCATTTCTTTTCGTCCACATTTGCAAACAAAACCTTTTCAATAATA
ATTTTATAAATAGTATCAATATATATATATATATATATATATTTATTTGTTTACTTTTTTC
TATCAGTGTTTTCAATTTTTTATTAAACAATGTTTGATTTTTTAAATCGCAATTTAA
TACCTAAATATAAAAAAATGTTATTATATTGCAAAAACCCATCAACCTTGAAAAAAA
GTAGAAACGTTTTATTTAATTCTATCAATACATCATAAATACGAACGATCCCCG
TCCAAGTTATGAGCTTAATCTTCCATAAAAAATATTTGAAAGCAATAGATCATGTA
CTAAACTAAATCAGGGAAATTcccgggggatcc

Orc6-4A (S106A, S116A, S123A, T146A)

ATGCAACAAGTTCAACACTGTGTTGCTGAAGTTTTGAGATTGGACCCACAAGAA
AAGCCAGACTGGTCTTCTGGTACTTGAAGAAGTTGACTAACGCTACTTCTATTT
TGTACAACACTTCTTTGAACAAGGTTATGTTGAAGCAAGACGAAGAAGTTGCTA
GATGTCACATTTGTGCTTACATTGCTTCTCAAAGATGAACGAAAAGCACATGC
CAGACTTGTGTTACTACATTGACTCTATTCCATTGGAACCAAAGAAGGCTAAGC
ACTTGATGAACCTGTTTCAGACAATCTTTGTCTAACTCTGCTCCAATGAAGCAATT

CGCTTGGACTCCAGCTCCAAAGAAGAACAAGAGAGCTCCAGTTAAGAACGGTG
GTAGATTCACCTTCTTCTGACCCAAAGGAATTGAGAAACCAATTGTTCCGGTGCTC
CAACTAAGGTTAGAAAGTCTCAAAACAACGACTCTTTTCGTTATTCCAGAATTGCC
ACCAATGCAAACCTAACGAATCTCCATCTATTACTAGAAGAAAGTTGGCTTTTCGAA
GAAGACGAAGACGAAGACGAAGAAGAACCAGGTAACGACGGTTTGTCTTTGAA
GTCTCACTCTAACAAGTCTATTACTGGTACTAGAAACGTTGACTCTGACGAATAC
GAAAACCACGAATCTGACCCAACTTCTGAAGAAGAACCATTGGGTGTTCAAGAA
TCTAGATCTGGTAGAACTAAGCAAAACAAGGCTGTTGGTAAGCCACAATCTGAA
TTGAAGACTGCTAAGGCTTTGAGAAAGAGAGGTAGAATTCCAAACTCTTTGTTG
GTTAAGAAGTACTGTAAGATGACTACTGAAGAAATTATTAGATTGTGTAACGACT
TCGAATTGCCAAGAGAAGTTGCTTACAAGATTGTTGACGAATACAACATTAACG
CTTCTAGATTGGTTTGTCCATGGCAATTGGTTTGTGGTTTGGTTTGAACGTGAC
TTTCATTGTTTTCAACGAAAGAAGAAGAAAGGACCCAAGAATTGACCACTTCATT
GTTTCTAAGATGTGTTCTTTGATGTTGACTTCTAAGGTTGACGACGTTATTGAAT
GTGTTAAGTTGGTTAAGGAATTGATTATTGGTGAAAAGTGGTTCAGAGACTTGC
AAATTAGATACGACGACTTCGACGGTATTAGATACGACGAAATTATTTTCAGAAA
GTTGGGTTCTATGTTGCAAACCTACTAACATTTTGGTTACTGACGACCAATACAAC
ATTTGGAAGAAGAGAATTGAAATGGACTTGGCTTTGACTGAACCATTGTAA

Orc2-N-terminal truncation (delete a.a. 2-235)

ATGGCTTCTTCTTTCTTGGACACTTTCGAAGGTTACTTCGACCAAAGAAAGATTG
TTAGAATAACGCTAAGTCTAGACACACTATGTCTATGGCTCCAGACGTTACTA
GAGAAGAATTCTCTTTGGTTTCTAACTTCTTCAACGAAAACCTCCAAAAGAGACC
AAGACAAAAGTTGTTTCGAAATTCAAAGAAGATGTTCCCACTACTGGTTCGAA
TTGACTCAAGGTTTCTCTTTGTTGTTCTACGGTGTGGTTCTAAGAGAACTTCT
TGGAAGAATTCGCTATTGACTACTTGTCTCCAAAGATTGCTTACTCTCAATTGGC
TTACGAAAACGAATTGCAACAAAACAAGCCAGTTAACTCTATTCCATGTTTGATT
TTGAACGGTTACAACCCATCTTGTAACTACAGAGACGTTTTCAAGGAAATTACTG
ACTTGTGGTTCCAGCTGAATTGACTAGATCTGAAACTAAGTACTGGGGTAACC
ACGTTATTTTGCAAATTCAAAGATGATTGACTTCTACAAGAACCAACCATTGGA
CATTAAAGTTGATTTTGGTTGTTTCACACTTGGACGGTCCATCTATTAGAAAGAAC
ACTTTCCAACTATGTTGTCTTTCTTGTCTGTTATTAGACAAATTGCTATTGTTGC
TTCTACTGACCACATTTACGCTCCATTGTTGTGGGACAACATGAAGGCTCAAAA
CTACAACCTTCGTTTTCCACGACATTTCTAACTTCGAACCATCTACTGTTGAATCT
ACTTTCCAAGACGTTATGAAGATGGGTAACTCTGACACTTCTTCTGGTGCTGAA
GGTGCTAAGTACGTTTTGCAATCTTTGACTGTTAACTCTAAGAAGATGTACAAGT
TGTTGATTGAACTCAAATGCAAAACATGGGTAACTTGTCTGCTAACACTGGTC
CAAAGAGAGGTACTCAAAGAAGTGGTGTGATTGAAGTTGTTCAACCACTTGT
GTGCTGCTGACTTCATTGCTTCTAACGAAATTGCTTTGAGATCTATGTTGAGAGA
ATTCATTGAACACAAGATGGCTAACATTACTAAGAACAACCTCTGGTATGGAAATT
ATTTGGGTTCCATACACTTACGCTGAATTGGAAAAGTTGTTGAAGACTGTTTTGA
ACACTTTGTAA

Supplementary Table 1: Primers

Primers	Sequence	Note	Source	Company
GC227	Biotin-gctgcaatgataccgcgagaccac	Amplify 3 kb template	Coster and Diffley, 2017	Sigma
GC228	Biotin-gggccagatggaagccctc	Amplify 3 kb template	Coster and Diffley, 2017	Sigma
GC117	Biotin-atgcttcggctgctatgttg	Amplify 0.6 kb template	Coster and Diffley, 2017	Sigma

GC298	gatgtgctgcaaggcgattaag	Amplify 0.6 kb template	Coster and Diffley, 2017	Sigma
EACS.B1 F	attcCTCGATTTTTTTATGTTTAGTTTCGCGGACGACGGTTTCGAGGTGG	Competitor DNA	Coster and Diffley, 2017	Sigma
EACS.B1 R	CCACCTCGAAACCGTCGTCCGCGAAACTAAACATAAAAAATCGAGgaat	Competitor DNA	Coster and Diffley, 2017	Sigma
NCP F	Desthiobiotin – CGAtagaaCTCGGCGcgccctggagaatCGcggtgcg	Amplify nucleosome template	Miller et al., 2020	IDT
NCP R	CctgcACCCAGGGACTTGAAGTAATAAGGAC	Amplify nucleosome template	Miller et al., 2020	IDT
CTL38	ATCTTTTGAATTTGCAAAATAACGTGGTTACCCAGTACTTAG	Insertion for ORC2ΔN	This study	Sigma
CTL39	GCAAAATCAAAAGATGATTGACTTCTACAAGAAC	Vector for ORC2ΔN	This study	Sigma
CTL40	TATAGTTTTTCTCCTTGACGTTAAAGTATAGAGG	Vector for ORC2ΔN	This study	Sigma
CTL61	ATCAGTGTTTTCAATTTTTTATTAACAATGTTTGATTTTTTAAATCGCAATT TAATACCTggtcaccggccgagcg	Replace endogenous ARS317::kanMX6	This study	Sigma
CTL62	ATGATCTATTGCTTTCAAATATTTTTATGGAAGATTAAGCTCATAACTTGA CGGGGATCGTTTAACTGGATGGCGGCGTTAG	Replace endogenous ARS317::kanMX6	This study	Sigma
CTL98	TGCTACTCATCCTAGTCTGCTTCCAGCTATTTAATATCATGCACGA AAAGCAAAACcgacatggaggccag	Replace URA3::hphNT1	This study	IDT
CTL99	GGCCGCATCTTCTCAAATATGCTTCCAGCTGCTTTTCTGTAACGTTTAC CCTCTACCTccttcgagcgctcccaaac	Replace URA3::hphNT1	This study	IDT
CTL86	GCCTTTTATATACCGATTAAATATTTATTTCTGACTTTTTCTTTACCGACA CCAAcaggcctgtttaacGCTTTTCAATTCATCTTTTTTTTTTTG	Insertion of URA3 in ARS419	This study	IDT
CTL87	TTCTATTCTGAAGAACTGTTTATAGAATACTCTTTTTTTATGTTTTGAGCC TACACCTtctagaagctcatatgTTAGTTTGTCTGGCCGCATC	Insertion of URA3 in ARS419	This study	IDT
CTL125	GGAGAAAAAATATACACCGGTGTATAAAACAATGGCTTCTTCTTTCTTG ACACTTTGGAAGGTTACTTC	Insertion for ORC2ΔN	This study	IDT
CTL141	gggtgaaataccgcacagatg	qPCR detection site	This study	IDT
CTL142	gctggcgaatagcgaagag	qPCR detection site	This study	IDT
CTL143	GTGCATGGCCTTCTTTCTAATTG	Internal control for qPCR	This study	IDT
CTL144	GCAGGATTAGGTAGCGTTTAATTTATCC	Internal control for qPCR	This study	IDT
CTL147	TATTTCTTAGCATTTTTGACGAAATTacggccagtgattcCTCG	Insert for pCTL67	This study	IDT
CTL148	AATTCGTCAAAAATGCTAAGAAATAGCGGCGCCAGGTaTTTTATG	Insert for pCTL67	This study	IDT
CTL149	ATTTTTGACGAAATCTGGCCACCGTCGGC	Vector for pCTL67	This study	IDT
CTL150	AAATGCTAAGAAATAcacctggcggttacccaac	Vector for pCTL67	This study	IDT
CTL156	TATTTCTTAGCATTTTTGACGAAATTacggccagtgattcACTAGTAC	Insert for pCTL68 and pCTL69	This study	IDT
CTL157	AATTCGTCAAAAATGCTAAGAAATActagagatgccccggg	Insert for pCTL68 and pCTL69	This study	IDT
CTL158	AAATGCTAAGAAATAgttaccacctaatacgccctgc	Vector for pCTL68 and pCTL69	This study	IDT
CTL159	ATTTTTGACGAAATtgtaatcatggtcatagctgttcctg	Vector for pCTL68 and pCTL69	This study	IDT
CTL160	GCAGCTGGTGTTCATCGAACATGATGAGCTTTAAACAAGGAGGAGTAT AAACTTTTTAAcatatcggtgtgaaataccgc	Amplify sequence for ARS419 insertion	This study	IDT
CTL161	ATAACCGATGACTTGGCAAGAAAAGGATTTAAGAGTTGTTCTATTCTGTA GAACGTGTTTcaggaaacagctatgaccatgattac	Amplify sequence for ARS419 insertion	This study	IDT
CTL172	TAGGTGTGAGAGCTCCCGATCTCGGCGAACAC	Amplify sequence for LacO plasmid	This study	IDT
CTL173	CGAACTTATGGTACCGCCGAGCGCGACG	Amplify sequence for LacO plasmid	This study	IDT
CTL174	GAGCTCTCACACCTACAAGGGATGTACATCAATTGTGAGCGGATAACAAT TGTTAGGGAGGAATTGTGAGCGGATAACAATTTGGAGTTGATAATTGTGA GCGGATAACAATTGGCTTCAACGTAATTGTGAGCGGATAACAATTTCCGTA CGAATGTGCCGAACCTATGGTACC	Duplex DNA of LacO	This study	IDT
CTL208	TCAAAAGGCCTGCAGGCAAGTGCACAAACAATACTTAAATAAATACTACTC AGTAATAACCTATTTCTTAGCATTTTTGACGAAATT	oligo for ARS1_B2	This study	IDT
CTL209	AATTCGTCAAAAATGCTAAGAAATAGGTTATTACTGAGTAGTATTTATTTA AGTATTGTTTGTCACTTGCCTGCAGGCCTTTTGA	oligo for ARS1_B2	This study	IDT
CTL214	CTTGCTGTCAGGCCTTTTGAACCTCGGCATGCGC	Amplify sequence for 90bp-B2	This study	IDT
CTL215	TTAGCATTTTTGACGAAATTCGTCGGCTCTCG	Amplify sequence for 90bp-B2	This study	IDT
50bp up 209	TTAAGTTGGTAACGCCAGGG	Amplify sequence for EMSA	This study	Sigma
50bp down 209	CCAGCTGAGGTACCGGATGCT	Amplify sequence for EMSA	This study	Sigma

Supplementary Table 2: Plasmids

Plasmid number	Plasmid	Vector	Insert source	Insert sites	Source
pGC199	No site	pUC19			From Coster <i>et al.</i> , Science 2017.
pGC200	One site	pUC19			From Coster <i>et al.</i> , Science 2017.
pGC201	25bp	pUC19			From Coster <i>et al.</i> , Science 2017.

pGC202	50bp	pUC19			From Coster <i>et al.</i> , Science 2017.
pGC211	90bp	pUC19			From Coster <i>et al.</i> , Science 2017.
pCTL8	pRS304-Orc5/Orc6-4A	pJF18	synthetic GeneArt strings (Thermo Fisher Scientific)	BplI/EcoRI	pJF18 from Frigola <i>et al.</i> , Nature 2013.
pCTL9	12bp	pUC19	synthetic GeneArt strings (Thermo Fisher Scientific)	BsoBI/EcoRI	This study
pCTL40	10bp	pUC19	synthetic GeneArt strings (Thermo Fisher Scientific)	BsoBI/EcoRI	This study
pCTL64	pRS306-Orc1/Orc2ΔN	PCR from pJF19 with CTL39 and CTL40	PCR from pJF19 with CTL125 and CTL38	In-fusion	pJF19 from Frigola <i>et al.</i> , Nature 2013.
pCTL67	pUC19-90bp-Abf1	PCR from pGC211 with CTL149 and CTL150	PCR from pGC211 with CTL147 and CTL148	In-fusion	pGC211 from Coster <i>et al.</i> , Science 2017.
pCTL68	pUC19-ARS1-Abf1	PCR from pGC403 with CTL158 and CTL159	PCR from pGC403 with CTL156 and CTL157	In-fusion	pGC403 from Coster <i>et al.</i> , Science 2017.
pCTL69	pUC19-ARS317-Abf1	PCR from pCTL45 with CTL158 and CTL159	PCR from pCTL45 with CTL156 and CTL157	In-fusion	This study
pCTL86	pUC19-ARS317	pUC19	synthetic GeneArt strings 495bp (Thermo Fisher Scientific)	EcoRI/BsoBI	This study
pCTL91	pUC19-90bp-B2	pGC211	PCR pGC211 with primer CTL214 and CTL215; ARS1_B2 sequence generated from IDT as oligo CTL208 and CTL209	In-fusion	pGC211 from Coster <i>et al.</i> , Science 2017.
pCTL93	pUC19-300bp+LacO	pGC205	PCR pGC200 with primer CTL172 and CTL173; LacO sequence generated from IDT as duplex DNA CTL174	In-fusion	pGC 205 from Coster <i>et al.</i> , Science 2017.
pCTL94	pUC-One site+LacO	pGC200	PCR pGC200 with primer CTL172 and CTL173; LacO sequence generated from IDT as duplex DNA CTL174	In-fusion	pGC200 from Coster <i>et al.</i> , Science 2017.

Supplementary Table 3: Yeast strains

Yeast strain	Background	Genotype	Source	
ySDORC	W303-1a	MATa ade2-1 ura3-1 his3-11,15 trp1-1 leu2-3,112 can1-100 bar1::Hyg pep4::KanMx his3::pRS303-Gal1, 10-ORC3/4 (HIS3) ura3::pRS306-Gal1, 10-ORC1/2 CBP-Orc1 (URA3) trp1::pRS304-Gal1, 10-ORC5/6 (TRP1) MATa ade2-1 ura3-1 his3-11,15 trp1-1 leu2-3,112 can1-100 bar1::Hyg pep4::KanMx	Frigola <i>et al.</i> , Nature 2013.	
YGC229	W303-1a	his3::pRS303-Gal1, 10-Cdt1/Gal4 (HIS3) ura3::pRS306-Gal1, 10-Mcm2/3 (URA3) CBP-TEV-NanoLuc-Mcm3 trp1::pRS304-Gal1, 10-Mcm4/5 (TRP1) leu2::pRS305-Gal1, 10-Mcm6/7 (LEU2) MATa ade2-1 ura3-1 his3-11,15 trp1-1 leu2-3,112 can1-100 bar1::Hyg pep4::KanMx	Coster <i>et al.</i> , Science 2017.	
yCTL6 (Orc6-4A)	W303-1a	MATa ade2-1 ura3-1 his3-11,15 trp1-1 leu2-3,112 can1-100 bar1::Hyg pep4::KanMX Flag at WT ORC6 (Leu) c-terminal ura::URApRS306/SDORC1,2 his::HISpRS303/SDORC3,4 trp::TRPpRS304/SDORC5,ORC6-4S/T-A	This study	
yCTL17 (Orc2_deltaN)	W303-1a	MATa ade2-1 ura3-1 his3-11,15 trp1-1 leu2-3,112 can1-100 bar1::Hyg pep4::KanMX Flag at WT ORC2 (Nat) C-terminal ura::URApRS306/SDORC1,ORC2_del2-235 his::HISpRS303/SDORC3,4 trp::TRPpRS304/SDORC5,ORC6	This study	RE digestion with NcoI on pCTL64
yCTL28 (ARS1Δ)	A364a	MATa ade2 ade3 ura3-52 trp1-289 leu2-3,112 bar1::LEU2 ORC2 ORC6 ura3-52::GAL1p-Δntcdc6, URA3 ΔURA3::hph cdc20::MET3p-HA3-CDC20, TRP1 MCM7-2NLS ARS317::KanMX6 ΔARS419::ARS1Δ	This study	PCR on pGC404 with CTL160 and CTL161. pGC404 from Coster <i>et al.</i> , Science 2017.
yCTL29 (ARS1)	A364a	MATa ade2 ade3 ura3-52 trp1-289 leu2-3,112 bar1::LEU2 ORC2 ORC6 ura3-52::GAL1p-Δntcdc6, URA3 ΔURA3::hph	This study	PCR on pCTL68 with CTL160 and CTL161

		cdc20::MET3p-HA3-CDC20, TRP1 MCM7-2NLS ARS317::KanMX6 ΔARS419::ARS1		
yCTL30 (ARS317)	A364a	MATa ade2 ade3 ura3-52 trp1-289 leu2-3,112 bar1::LEU2 ORC2 ORC6 ura3-52::GAL1p-Δntcdc6, URA3 ΔURA3::hph cdc20::MET3p-HA3-CDC20, TRP1 MCM7-2NLS ARS317::KanMX6 ΔARS419::ARS317	This study	PCR on pCTL69 with CTL160 and CTL161
yCTL33 (90bp)	A364a	MATa ade2 ade3 ura3-52 trp1-289 leu2-3,112 bar1::LEU2 ORC2 ORC6 ura3-52::GAL1p-Δntcdc6, URA3 ΔURA3::hph cdc20::MET3p-HA3-CDC20, TRP1 MCM7-2NLS ARS317::KanMX6 ΔARS419::90bp	This study	PCR on pCTL67 with CTL160 and CTL161