

Classification and nomenclature of metacaspases and paracaspases: no more confusion with caspases

Elena A. Minina^{1,2,*}, Jens Staal³, Vanina E. Alvarez⁴, John A. Berges⁵, Ilana Berman-Frank⁶, Rudi Beyaert³, Kay D. Bidle⁷, Frédéric Bornancin⁸, Magali Casanova⁹, Juan J. Cazzulo⁴, Chang Jae Choi¹⁰, Nuria S. Coll¹¹, Vishva M. Dixit¹², Marko Dolinar¹³, Nicolas Fasel¹⁴, Christiane Funk¹⁵, Patrick Gallois¹⁶, Kris Gevaert¹⁷, Emilio Gutierrez-Beltran¹⁸, Stephan Hailfinger¹⁹, Marina Klemenčič¹³, Eugene V. Koonin²⁰, Daniel Krappmann²¹, Anna Linusson¹⁵, Maurício F. M. Machado²², Frank Madeo²³, Lynn A. Megeney²⁴, Panagiotis N. Moschou^{25,26,27}, Jeremy C. Mottram²⁸, Thomas Nyström²⁹, Heinz D. Osiewacz³⁰, Christopher M. Overall³¹, Kailash C. Pandey³², Jürgen Ruland^{33,34,35}, Guy S. Salvesen³⁶, Yigong Shi³⁷, Andrei Smertenko³⁸, Simon Stael^{17,39}, Jerry Ståhlberg¹, María Fernanda Suárez⁴⁰, Margot Thome¹⁴, Hannele Tuominen⁴¹, Frank Van Breusegem³⁹, Renier A. L. van der Hoorn⁴², Assaf Vardi⁴³, Boris Zhivotovsky^{44,45}, Eric Lam⁴⁶, Peter V. Bozhkov^{1,*}

¹Department of Molecular Sciences, Uppsala BioCenter, Swedish University of Agricultural Sciences and Linnean Center for Plant Biology, Uppsala, Sweden

²COS, Heidelberg University, Heidelberg, Germany

³VIB Center for Inflammation Research; Department of Biomedical Molecular Biology, Ghent University; Ghent, Belgium

⁴Instituto de Investigaciones Biotecnológicas, Universidad Nacional de San Martín, San Martín, Buenos Aires, Argentina

⁵Department of Biological Sciences and School of Freshwater Sciences, University of Wisconsin-Milwaukee, Milwaukee, WI, USA

⁶Department of Marine Biology, Leon H. Charney School of Marine Sciences, University of Haifa, Israel

⁷Department of Marine and Coastal Sciences, Rutgers University, New Brunswick, NJ, USA

⁸Novartis Institutes for BioMedical Research, Basel, Switzerland

⁹Aix-Marseille Univ, CNRS, LISM, Institut de Microbiologie de la Méditerranée, Marseille, France

¹⁰The University of Texas at Austin, Marine Science Institute, Port Aransas, TX, USA

32 ¹¹Centre for Research in Agricultural Genomics (CRAG), CSIC-IRTA-UAB-UB, Campus UAB,
33 Bellaterra, Barcelona, Spain

34 ¹²Department of Physiological Chemistry, Genentech, South San Francisco, CA, USA

35 ¹³University of Ljubljana, Faculty of Chemistry and Chemical Technology, Ljubljana, Slovenia

36 ¹⁴Department of Biochemistry, University of Lausanne, Epalinges, Switzerland

37 ¹⁵Department of Chemistry, Umeå University, Umeå, Sweden

38 ¹⁶Faculty of Biology, Medicine and Health, University of Manchester, Manchester, UK

39 ¹⁷VIB Center for Medical Biotechnology; Department of Biomolecular Medicine, Ghent
40 University; Ghent, Belgium

41 ¹⁸Instituto de Bioquímica Vegetal y Fotosíntesis, Universidad de Sevilla and Consejo Superior de
42 Investigaciones Científicas, Sevilla, Spain

43 ¹⁹Interfaculty Institute for Biochemistry, Eberhard Karls University, Tübingen, Germany

44 ²⁰National Center for Biotechnology Information, National Library of Medicine, Bethesda, MD,
45 USA

46 ²¹Research Unit Cellular Signal Integration, Helmholtz Zentrum München - German Research
47 Center for Environmental Health, Neuherberg, Germany

48 ²²Interdisciplinary Center for Biochemical Research, University of Mogi das Cruzes, Mogi das
49 Cruzes, Brazil

50 ²³Institute of Molecular Biosciences, NAWI Graz, University of Graz; BioTechMed Graz, Graz,
51 Austria

52 ²⁴Sprott Centre for Stem Cell Research, Ottawa Hospital Research Institute and Departments of
53 Medicine and Cellular and Molecular Medicine, University of Ottawa, Ottawa, ON, Canada

54 ²⁵Institute of Molecular Biology and Biotechnology, Foundation for Research and Technology -
55 Hellas, Heraklion, Greece

56 ²⁶Department of Biology, University of Crete, Greece

57 ²⁷Department of Plant Biology, Uppsala BioCenter, Swedish University of Agricultural Sciences
58 and Linnean Center for Plant Biology, Uppsala, Sweden

59 ²⁸York Biomedical Research Institute, Department of Biology, University of York, York, UK

60 ²⁹Institute for Biomedicine, Sahlgrenska Academy, Centre for Ageing and Health – AgeCap,
61 University of Gothenburg, Gothenburg, Sweden

62 ³⁰Institute for Molecular Biosciences, Faculty of Biosciences, Goethe University,
63 Frankfurt/Main, Germany

64 ³¹Departments of Oral Biological and Medical Sciences / and Biochemistry and Molecular
65 Biology, Life Sciences Institute, University of British Columbia, Vancouver, BC, Canada

66 ³²ICMR-National Institute of Malaria Research, New Delhi, India

67 ³³Institute of Clinical Chemistry and Pathobiochemistry, School of Medicine, Technical
68 University of Munich, Munich, Germany

69 ³⁴German Cancer Consortium (DKTK), partner site Munich, Germany

70 ³⁵German Center for Infection Research (DZIF), partner site Munich, Germany

71 ³⁶Sanford Burnham Prebys Medical Discovery Institute, La Jolla, CA, USA

72 ³⁷School of Life Sciences, Westlake University, Xihu District, Hangzhou Zhejiang Province,
73 China

74 ³⁸Institute of Biological Chemistry, Washington State University, Pullman, WA, USA

75 ³⁹Department of Plant Biotechnology and Bioinformatics, Ghent University; VIB-UGent Center
76 for Plant Systems Biology; Ghent, Belgium

77 ⁴⁰Departamento de Biología Molecular y Bioquímica, Facultad de Ciencias, Universidad de
78 Málaga, Campus de Teatinos, Málaga, Spain

79 ⁴¹Umeå Plant Science Centre, Department of Plant Physiology, Umeå University, Umeå, Sweden

80 ⁴²Department of Plant Sciences, University of Oxford, Oxford, UK

81 ⁴³Department of Plant and Environmental Sciences, Weizmann Institute of Science, Rehovot,
82 Israel

83 ⁴⁴Division of Toxicology, Institute of Environmental Medicine, Karolinska Institutet, Stockholm,
84 Sweden

85 ⁴⁵Faculty of Fundamental Medicine, MV Lomonosov Moscow State University, Moscow, Russia

86 ⁴⁶Department of Plant Biology, Rutgers the State University of New Jersey, New Brunswick, NJ
87 USA

88 *Correspondence: Peter.Bozhkov@slu.se; Alena.Minina@slu.se

89

Metacaspases and paracaspases are proteases that were first identified as containing a caspase-like structural fold (Uren et al., 2000) known as the caspase-hemoglobinase fold (CHF, Aravind and Koonin, 2002). Like caspases, meta- and paracaspases are multifunctional proteins regulating diverse biological phenomena, such as aging, immunity, proteostasis and programmed cell death. The broad phylogenetic distribution of meta- and paracaspases across all kingdoms of life and large variation of their biochemical and structural features complicate classification and annotation of the rapidly growing number of identified homologs. Establishment of an adequate classification and unified nomenclature of meta- and paracaspases is especially important to avoid frequent confusion of these proteases with caspases - a tenacious misnomer that unfortunately does not appear to decline with time. This letter represents a consensus opinion of researchers studying different aspects of caspases, meta- and paracaspases in various organisms, ranging from microbes to plants and animals.

Classification of meta- and paracaspases

The current classification of proteases provided by the MEROPS database clusters caspases, meta- and paracaspases to the same family, C14, within the CD clan (<https://www.ebi.ac.uk/merops/>). All members of C14 family are annotated to possess aspartate P1 cleavage specificity, and the family is further split into two subfamilies: C14A (caspases) and C14B (meta- and paracaspases).

Importantly, the MEROPS approach of grouping proteases into families or subfamilies is based on statistically significant similarities of the amino acid sequence within the peptidase domain or part thereof, without considering their biochemical properties (Rawlings et al., 2018). Being valuable for high-throughput protease classification, this approach, however, has substantial drawbacks if implemented without further adjustment. Indeed, in contradiction with the MEROPS description, none of the meta- or paracaspases characterized so far cleave after an aspartate residue. Instead, paracaspases are arginine-specific (Coornaert et al., 2008; Hachmann et al., 2012; Rebeaud et al., 2008), whereas metacaspases can cleave after either arginine or lysine (**Fig 1A**; Sundström et al., 2009; Vercammen et al., 2004). Such fundamental differences in the proteolytic specificity between caspases, meta- and paracaspases imply distinct repertoires of new proteoforms that they generate and point to the complex diversification and coevolution of their substrates and downstream pathways. One unfortunate consequence of the current classification is the misuse of

caspase-specific probes for studying meta- and paracaspases that is commonly found in the literature and leads to false conclusions.

Apart from substrate specificity, caspases, meta- and paracaspases feature other fundamental differences (**Fig. 1A**). For example, active metacaspases are monomers and their activation usually requires millimolar concentrations of calcium (Hander et al., 2019; McLuskey et al., 2012; Wong et al., 2012). In contrast, active caspases and paracaspases are calcium-independent dimers (Hachmann et al., 2012; Weismann et al., 2012; Yu et al., 2011). This indicates that upstream pathways regulating activation of caspases, meta- and paracaspases are likewise different.

In the past two decades we have learned about important differences between caspases, meta- and paracaspases. Thus, simple extrapolation of features typical for caspases to all other members of the C14 family is not justified anymore. Instead caspases, meta- and paracaspases should be separated into three corresponding groups within the family and each group should be properly annotated by providing its key biochemical and structural characteristics. We kindly request curators of MEROPS database to make corresponding changes.

Since structure and substrate specificity of prokaryotic caspase-like proteases named “orthocaspases” remain largely unknown (Klemenčič et al., 2015) we leave their classification and nomenclature open until their structural and biochemical properties have been clarified.

Unified nomenclature of meta- and paracaspases

The name “caspase” stands for “cysteine-dependent aspartate-specific protease”. Thus, the names “metacaspase” and “paracaspase” imply the wrong substrate specificity for these proteases. However, since these names have been used for two decades we propose to keep them, provided that caspases, meta- and paracaspases are recognized as three separate groups within the C14 family.

Based on domain composition and arrangement, meta- and paracaspases are further sub-divided into three and two types, respectively (**Fig. 1A**). For the sake of consistency, we propose to maintain a common nomenclature for the different types of meta- and paracaspases using Latin

numerals (e.g. type I metacaspases). As for the conserved protein structures, they will be referred to as the p20-like region, the p10-like region, the linker region and the N-terminal pro-domain, matching the nomenclature of caspases (**Fig. 1A**; Alnemri et al., 1996). The p20, p10 and linker regions have been previously defined for the caspase group of the C14 family (Fuentes-Prior and Salvesen, 2004) and can be easily identified in meta- and paracaspase homologs based on a hidden Markov model (HMM) alignment with the C14 peptidase domain (PF00656) (**Fig. 1B**). Notably, although not always clearly stated in the literature, most known members of the C14 family contain the linker region. Furthermore, type II metacaspases are distinguished by a long linker between the p20 and p10 regions and an additional linker within the p10 region (**Fig. 1A**), which are frequently referred to as a single linker.

We suggest to consider the active form of meta- or paracaspases being a monomer if it is a cleaved or intact polypeptide chain derived from a single translational event, and a dimer if it comprises uncut or processed products of two translational events.

We propose to optimize nomenclature recommendations previously established for *Arabidopsis thaliana* metacaspases (Tsiatsiani et al., 2011) and vertebrate paracaspases (Hulpiau et al., 2016) in order (i) to have a unified nomenclature across meta- and paracaspase groups of the C14 family, (ii) to facilitate comparison of orthologs from different organisms, and (iii) to make it suitable for annotating homologs of species with partially sequenced genomes. Thus, we suggest using simple root symbols such as MCA for metacaspases and PCA for paracaspases. When naming individual family members, these root symbols will be preceded by the abbreviated Latin name of the species and followed by a hyphen, Latin number representing the type and then a small alpha character indicating in alphabetical order the number of the homolog of this type in a given genome (**Fig. 1C**). Proenzymes that require proteolytic processing for activation could be annotated with a prefix “pro-“, e.g. pro-AtMCA-Ia for the metacaspase 1 of type I from *A. thaliana*. Spliceoforms should be indicated by a decimal number (e.g. AtMCA-Ia.1). Please note that these conventions do not consider the letter case, which should conform to gene and protein nomenclature established for a given model organism or taxonomic group.

Importantly, this nomenclature should be used synonymously for meta- and paracaspase homologs with well-established names, e.g. human MALT1/HsPCA-Ia or *A. thaliana* AtMC1/AtMCA-Ia. We encourage all researchers to adopt these recommendations. The new classification and unified nomenclature of meta- and paracaspases will facilitate a more comprehensive exchange of relevant findings within the scientific community and help to bridge already existing knowledge with newly discovered homologs, thus promoting mechanistic understanding of these ancient, evolutionarily conserved proteases.

Acknowledgements. This work was supported by Knut and Alice Wallenberg Foundation. We apologize to colleagues whose work has not been cited due to space limitation.

References

- Alnemri, E., Livingston, D., Nicholson, D., Salvesen, G., Thornberry, N., Wong, W., and Yuan, J. (1996). Human ICE/CED-3 Protease Nomenclature. *Cell* 87, 171.
- Aravind, L., and Koonin, E. V. (2002). Classification of the caspase-hemoglobinase fold: Detection of new families and implications for the origin of the eukaryotic separins. *Proteins Struct. Funct. Genet.* 46, 355–367.
- Coornaert, B., Baens, M., Heyninck, K., Bekaert, T., Haegman, M., Staal, J., Sun, L., Chen, Z.J., Marynen, P., and Beyaert, R. (2008). T cell antigen receptor stimulation induces MALT1 paracaspase - Mediated cleavage of the NF- κ B inhibitor A20. *Nat. Immunol.* 9, 263–271.
- Fuentes-Prior, P., and Salvesen, G.S. (2004). The protein structures that shape caspase activity, specificity, activation and inhibition. *Biochem. J.* 384, 201–232.
- Hachmann, J., Snipas, S.J., Van Raam, B.J., Cancino, E.M., Houlihan, E.J., Poreba, M., Kasperkiewicz, P., Drag, M., and Salvesen, G.S. (2012). Mechanism and specificity of the human paracaspase MALT1. *Biochem. J.* 443, 287–295.
- Hander, T., Fernández-Fernández, Á.D., Kumpf, R.P., Willems, P., Schatowitz, H., Rombaut, D., Staes, A., Nolf, J., Pottie, R., Yao, P., et al. (2019). Damage on plants activates Ca²⁺-dependent metacaspases for release of immunomodulatory peptides. *Science* 363, 1–10.
- Hulpiau, P., Driege, Y., Staal, J., and Beyaert, R. (2016). MALT1 is not alone after all: Identification of novel paracaspases. *Cell. Mol. Life Sci.* 73, 1103–1116.
- Klemenčič, M., and Funk, C. (2018). Type III metacaspases: calcium-dependent activity proposes new function for the p10 domain. *New Phytol.* 218, 1179–1191.
- Klemenčič, M., Novinec, M., and Dolinar, M. (2015). Orthocaspases are proteolytically active prokaryotic caspase homologues: The case of *Microcystis aeruginosa*. *Mol. Microbiol.* 98, 142–150.
- McLuskey, K., Rudolf, J., Proto, W.R., Isaacs, N.W., Coombs, G.H., Moss, C.X., and Mottram, J.C. (2012). Crystal structure of a *Trypanosoma brucei* metacaspase. *Proc. Natl. Acad. Sci. U. S. A.* 109, 7469–7474.
- Minina, E.A., Coll, N.S., Tuominen, H., and Bozhkov, P. V. (2017). Metacaspases versus caspases in development and cell fate regulation. *Cell Death Differ.* 24, 1314–1325.

222 Rawlings, N.D., Barrett, A.J., Thomas, P.D., Huang, X., Bateman, A., and Finn, R.D. (2018). The
 223 MEROPS database of proteolytic enzymes, their substrates and inhibitors in 2017 and a
 224 comparison with peptidases in the PANTHER database. *Nucleic Acids Res.* *46*, D624–D632.
 225 Rebeaud, F., Hailfinger, S., Posevitz-Fejfar, A., Tapernoux, M., Moser, R., Rueda, D., Gaide, O.,
 226 Guzzardi, M., Iancu, E.M., Rufer, N., et al. (2008). The proteolytic activity of the paracaspase
 227 MALT1 is key in T cell activation. *Nat. Immunol.* *9*, 272–281.
 228 Suarez, M.F., Filonova, L.H., Smertenko, A., Savenkov, E.I., Clapham, D.H., Von Arnold, S.,
 229 Zhivotovsky, B., and Bozhkov, P. V. (2004). Metacaspase-dependent programmed cell death is
 230 essential for plant embryogenesis. *Curr. Biol.* *14*, 339–340.
 231 Sundström, J.F., Vaculova, A., Smertenko, A.P., Savenkov, E.I., Golovko, A., Minina, E., Tiwari,
 232 B.S., Rodriguez-Nieto, S., Zamyatnin Jr., A.A., Välineva, T., et al. (2009). Tudor staphylococcal
 233 nuclease is an evolutionarily conserved component of the programmed cell death degradome. *Nat.*
 234 *Cell Biol.* *11*, 1347–1354.
 235 Tsiatsiani, L., Van Breusegem, F., Gallois, P., Zavalov, A., Lam, E., and Bozhkov, P. V. (2011).
 236 Metacaspases. *Cell Death Differ.* *18*, 1279–1288.
 237 Uren, A., O’Rourke, K., Aravind, L., Pisabarro, M.T., Seshagiri, S., Koonin, E. V, and Dixit, V.M.
 238 (2000). Identification of paracaspases and metacaspases: two ancient families of caspase-like
 239 proteins, one of which plays a key role in MALT lymphoma. *Mol. Cell* *6*, 961–967.
 240 van Creveld, S.G., Ben-Dor, S., Mizrahi, A., Alcolombri, U., Hopes, A., Mock, T., Rosenwasser,
 241 S., and Vardi, A. (2018). A redox-regulated type III metacaspase controls cell death in a marine
 242 diatom. *bioRxiv* doi: <https://doi.org/10.1101/444109>
 243 Vercammen, D., De Cotte, B. Van, De Jaeger, G., Eeckhout, D., Casteels, P., Vandepoele, K.,
 244 Vandenberghe, I., Van Beeumen, J., Inzé, D., and Van Breusegem, F. (2004). Type II
 245 metacaspases Atmc4 and Atmc9 of *Arabidopsis thaliana* cleave substrates after arginine and
 246 lysine. *J. Biol. Chem.* *279*, 45329–45336.
 247 Wiesmann, C., Leder, L., Blank, J., Bernardi, A., Melkko, S., Decock, A., D’Arcy, A., Villard, F.,
 248 Erbel, P., Hughes, N., et al. (2012). Structural determinants of MALT1 protease activity. *J. Mol.*
 249 *Biol.* *419*, 4–21.
 250 Wong, A.H.H., Yan, C., and Shi, Y. (2012). Crystal structure of the yeast metacaspase Yca1. *J.*
 251 *Biol. Chem.* *287*, 29251–29259.

252 Yu, J.W., Jeffrey, P.D., Ha, J.Y., Yang, X., and Shi, Y. (2011). Crystal structure of the mucosa-
253 associated lymphoid tissue lymphoma translocation 1 (MALT1) paracaspase region. Proc. Natl.
254 Acad. Sci. U. S. A. *108*, 21004–21009.

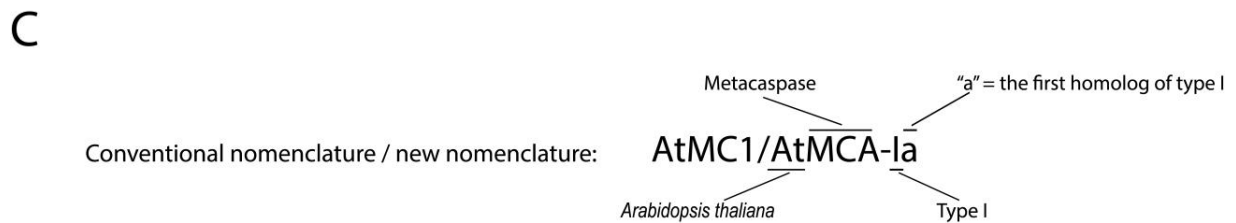
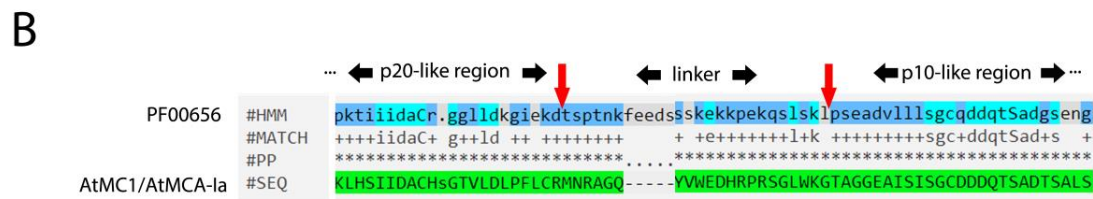
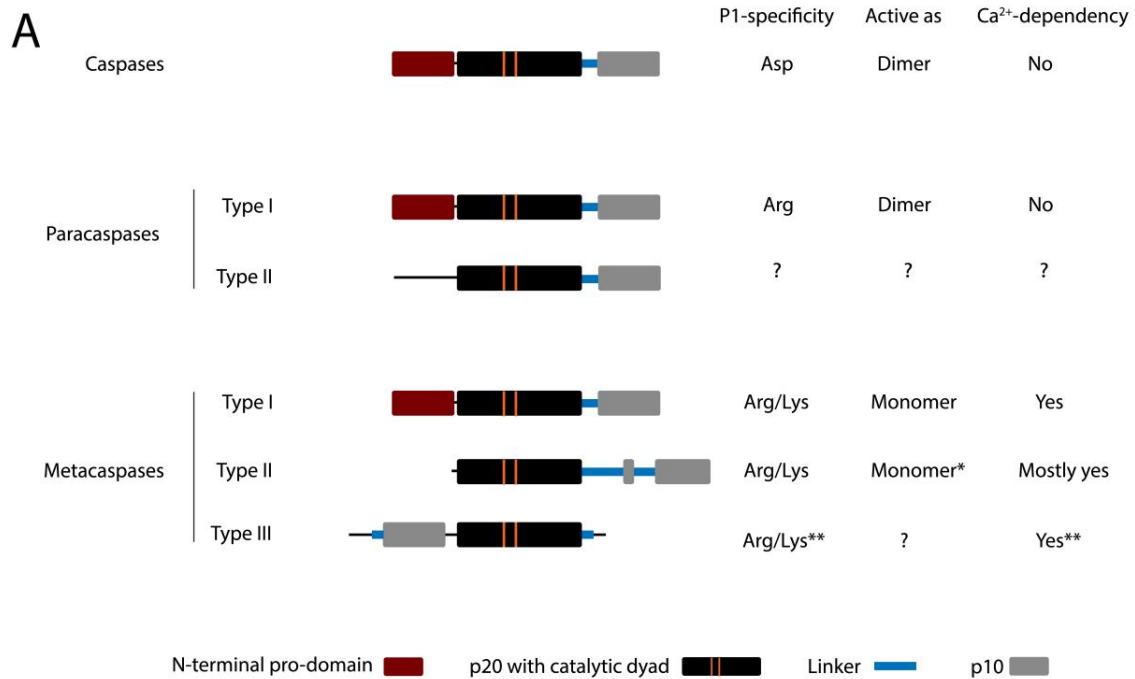


Figure 1. Classification and nomenclature of meta- and paracaspases.

(A) Comparison between caspases, metacaspases and paracaspases: domain composition and biochemical characteristics.

*Bozhkov and Smertenko, unpublished data for mcII-Pa/PaMC-IIb from *Picea abies* (Minina et al., 2017; Suarez et al., 2004).

262 **Only two orthologs of the type III metacaspases have been characterized so far,
263 GtMC2/GtMC-IIIa from *Guillardia theta* (Klemenčič and Funk, 2018) and PtMC5/PtMC-
264 IIIc from *Phaeodactylum tricornutum* (van Creveld et al., 2018).

265 **(B)** Part of the HMM alignment of the *Arabidopsis thaliana* metacaspase 1 with the C14
266 peptidase domain (PF00656), red arrows indicate borders between the p20-like region, linker
267 and the p10-like region.

268 **(C)** An example of the use of the new nomenclature for the *A. thaliana* type I metacaspase.
269 For homologs with well-established names we recommend to use the new nomenclature
270 synonymously; this will significantly ease comparison with orthologs from species with
271 partially sequenced genome.
272

