



CASE REPORT

Case Report: Disseminated, rifampicin resistant *Mycobacterium bovis* (BCG) infection in an immunocompromised child [version 1; peer review: 2 approved]

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V1 First published: 15 Oct 2020, 5:242
<https://doi.org/10.12688/wellcomeopenres.16280.1>

Latest published: 15 Oct 2020, 5:242
<https://doi.org/10.12688/wellcomeopenres.16280.1>

Abstract

Background: *Bacillus Calmette–Guérin* (BCG) is a live-attenuated vaccine used world-wide for prevention of tuberculosis disease. In some immunocompromised hosts it has the potential to cause disease. As with other members of the *M. tuberculosis* complex it has the potential for acquiring drug resistance.

Methods: We reviewed 10 years of paediatric clinical BCG strains referred to our clinical microbiology laboratory in Oxford where they underwent whole genome sequencing. We present a case series comparing clinical, pathogen genetic and pathogen phenotypic data, and consider the clinical implications.

Results: We identified 15 BCG isolates from 8 children under 16 years old. Only one child had clinical disease with the other seven reported as local inoculation-site reactions. Case 1 suffered disseminated disease secondary to an undiagnosed IL-12/IFN γ receptor defect and the BCG isolates evolved two different rifampicin resistance mutations. Across all 15 isolates, phenotypic resistance to each first line drug was seen.

Conclusions: BCG is a safe and effective vaccine in children. Most clinical specimens in our series were not related to disease. However, in the context of rare pathogen-specific immunocompromise, BCG can cause pathology and acquire drug resistance under selection from therapy.

Open Peer Review

Reviewer Status

	Invited Reviewers	
	1	2
version 1		
15 Oct 2020	report	report
1. Elizabeth Whitaker , Imperial College London, London, UK		
2. Ben J. Marais , University of Sydney, Sydney, Australia		
Any reports and responses or comments on the article can be found at the end of the article.		

Keywords

BCG, drug resistance, whole genome sequencing, immunocompromised

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Author roles: **Drysdale SB:** Conceptualization, Data Curation, Methodology, Project Administration, Writing – Original Draft Preparation, Writing – Review & Editing; **Kelly DF:** Conceptualization, Data Curation, Project Administration, Supervision, Writing – Review & Editing; **Morgan M:** Data Curation, Methodology, Writing – Review & Editing; **Peto T:** Funding Acquisition, Resources, Writing – Review & Editing; **Crook D:** Funding Acquisition, Project Administration, Resources, Writing – Review & Editing; **Matthews PC:** Conceptualization, Data Curation, Formal Analysis, Investigation, Methodology, Project Administration, Writing – Original Draft Preparation, Writing – Review & Editing; **Walker TM:** Conceptualization, Data Curation, Formal Analysis, Investigation, Methodology, Project Administration, Visualization, Writing – Original Draft Preparation, Writing – Review & Editing

Competing interests: No competing interests were disclosed.

Grant information: This work was supported by the National Institute for Health Research (NIHR) Oxford Biomedical Research Centre (BRC) and Health Innovation Challenge Fund (UK Department of Health and the Wellcome Trust [grant T5-358]. DFK receives salary support from the NIHR Oxford Biomedical Research Centre. PCM is funded by a Wellcome Trust Intermediate Fellowship (ref 110110). TMW is Wellcome Trust Clinical Career Development Fellow (214560).

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

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How to cite this article: Drysdale SB, Kelly DF, Morgan M *et al.* **Case Report: Disseminated, rifampicin resistant *Mycobacterium bovis* (BCG) infection in an immunocompromised child [version 1; peer review: 2 approved]** Wellcome Open Research 2020, 5:242 <https://doi.org/10.12688/wellcomeopenres.16280.1>

First published: 15 Oct 2020, 5:242 <https://doi.org/10.12688/wellcomeopenres.16280.1>

Introduction

The *Bacillus Calmette–Guérin* (BCG) vaccine has been in use worldwide for many years as prophylaxis against *Mycobacterium tuberculosis* infection. It is a live-attenuated vaccine derived from many cell culture passages of *Mycobacterium bovis*. There are multiple different *M. bovis* BCG strains¹ but the Danish Statens Serum Institut (SSI) 1331 strain has been the most widely used in the United Kingdom (UK).

All *M. bovis* BCG strains are inherently resistant to pyrazinamide. The *in vitro* susceptibility of vaccine strains to other antibiotic agents has been investigated by previous studies^{1,2}. Unusually, the Danish BCG strain 1331 has only intermediate isoniazid susceptibility (sensitive at 0.4 µg/ml but resistant at 0.1 µg/ml)¹ and is resistant to ethionamide. *In vivo*, however, serum concentrations of isoniazid would often be >0.1 µg/ml³ and its low-grade resistance has not been considered a clinical problem. Alternative vaccine strains have occasionally been used during periods of interruption to the supply of the Danish 1331 preparation in the UK. These include the InterVax (Canada) BCG vaccine (Bulgarian substrain [Sofia] SL222)⁴ and the Connaught strain².

BCG adenitis following vaccination is relatively common, with an incidence of 1 in 1500–2800⁵. The history has been well described, and local infection usually regresses spontaneously⁶. In contrast, disseminated BCG infection is extremely rare (estimated to affect 3.4 children per million given BCG⁵) and is characteristically associated with primary or secondary immunodeficiency. There are no clear guidelines on how best to treat children with disseminated *M. bovis* BCG strain infection. However, treatment regimens generally include a prolonged course of rifampicin, isoniazid and ethambutol with adjustments made according to antibiotic susceptibility testing, led by an expert in mycobacterial infection, similar to recommendations for treating patients with multidrug-resistant (MDR) *M. tuberculosis*⁷.

In Oxfordshire, United Kingdom (UK), the annual incidence of TB is relatively low (approximately 4 per 100,000) and children are only given the BCG vaccine if they are in a high-risk group as defined by Public Health England (PHE) guidance⁸. This study was triggered by the isolation of a rifampicin-resistant BCG strain from a child in Oxfordshire who was vaccinated due to being in a high-risk epidemiological group, and went on to develop disseminated BCG disease. To better contextualise this case, we sought to assess the antibiotic susceptibilities of other *M. bovis* BCG isolates from children submitted to our clinical microbiology laboratory.

Methods

Study population

This study was conducted at Oxford University Hospitals NHS Foundation Trust, UK, a large tertiary referral teaching hospital for which specialist services serve a population of approximately 2.5 million people. Clinical isolates referred to the regional laboratory at Oxford University Hospital NHS Foundation Trust, UK, and identified as *Mycobacterium*

tuberculosis complex have been whole-genome-sequenced using Illumina technology since 2007⁹. Following the identification of an index case of a child with disseminated BCG infection ('Case one'), we used this sequence database to identify BCG isolates from children, and reviewed clinical notes and routine microbiology results of these cases.

Phenotypic susceptibility testing

Culture-based susceptibility testing was performed using liquid culture (Bactec MGIT 960) at the Public Health England National Mycobacterial Reference Laboratory, London.

Mycobacterial sequencing

For isolates identified as BCG, Illumina reads were mapped to a *Mycobacterium bovis* reference genome (AF2122/97 NC_002945.3) using Stampy (version 1.0.17), with repetitive regions masked. Variant calls were made using SAMtools mpileup (version 0.1.18), based on a minimum depth of 5X and at least one read on each strand. A mixed-call was assigned where the minority allele composed >10% of read depth¹⁰. Genotypic susceptibility predictions were based on a catalogue of resistance mutations previously described¹¹.

Ethics approval

Our retrospective review of laboratory data did not require formal ethical approval but the Caldicott guardian for Oxford University Hospitals NHS Foundation Trust approved the study and we obtained written informed consent from the parents of our index case ('Case one'). Mycobacterial sequencing was undertaken as part of service development within clinical microbiology.

Case report and results

Clinical context of *M. bovis* samples

We identified 15 clinical samples from eight children <16 years old (Table 1). Case one was a girl of Pakistani descent who first presented at 4 months of age. She had been vaccinated routinely with BCG (Danish strain 1331) soon after birth as she was from a high-risk epidemiological group. She developed disseminated BCG infection and was subsequently diagnosed with an Interleukin-12 receptor B1 (IL-12 RB1) gene mutation, which is known to increase susceptibility to mycobacterial disease¹². The samples from the remaining seven cases all arose from either injection site collections or local adenitis, and all arose in children who had received the same vaccine strain.

Antibiotic management of *M. bovis* in an immunocompromised host

Case one initially received rifampicin (15 mg/kg daily), isoniazid (10 mg/kg daily), pyrazinamide (35 mg/kg daily) and ethambutol (20 mg/kg daily) for presumed *M. tuberculosis* infection. Following identification of the isolate as *M. bovis* by line-probe assay in the National Mycobacterial Reference Laboratory, London, approximately two months into therapy, pyrazinamide was stopped. A month later, phenotypic susceptibilities became available, suggesting resistance to ethambutol. This agent was therefore switched to moxifloxacin (10 mg/kg daily). She received 15 months of therapy and then

Table 1. Summary of 12 *M. bovis* BCG strain isolates from 8 children in a UK cohort. All children in this series had been immunised with the Danish 1331 *M. bovis* BCG strain.

	Site of BCG detection	Clinical diagnosis	Underlying condition	Outcome	Phenotypic drug sensitivity profile			
					INH	RIF	EMB	PZA
Case 1	Left axillary LN 2013	BCG adenitis	IL12Rb1 defect	Ongoing infection	S	S	R	R
	Left axillary LN 2013	BCG adenitis						
	Gastric aspirate 2014	Disseminated BCG			S	S	R	R
	Gastric aspirate 2014	Disseminated BCG						
	Right axillary LN 2014	Disseminated BCG			S	S	S	R
	Cervical LN 2015	Relapse disseminated BCG			S	R	S	R
	Pus, abdominal wall 2017	Relapse disseminated BCG			S	S	S	R
Case 2	Left axillary LN 2013	BCG adenitis	Metabolic disorder	Resolved infection	S	S	S	R
Case 3	Tissue L arm 2013	BCG adenitis	Nil	Resolved infection	S	S	R	R
	Axillary swab 2013	BCG adenitis	Nil	Resolved infection				
Case 4	Left axillary LN 2013	BCG adenitis	Nil	Resolved infection	S	S	S	R
Case 5	BCG site swab 2015	BCG site abscess	Nil	Resolved infection	R	S	S	R
Case 6	Left axillary LN 2007	BCG adenitis	HIV	Resolved infection	S	S	S	R
Case 7	Gastric aspirate/PEG swab 2009	Abscess over BCG scar	Nil	Resolved infection	S	S	S	R
Case 8	BCG scar 2007	ND	ND	ND	S	S	S	R

Key: LN, lymph node; RIF, Rifampicin; INH, isoniazid; PZA, pyrazinamide; EMB, ethambutol; ND, No data available. Blank indicates not tested.

commenced azithromycin (10 mg/kg daily, three days a week) for prophylaxis against non-tuberculous mycobacterial infection. She subsequently relapsed with disseminated BCG infection nine months after stopping initial antimycobacterial treatment, requiring multiple surgical procedures (drainage of abdominal collections) and prolonged antibiotic therapy, which are still ongoing three years later.

Antibiotic susceptibility of *M. bovis* isolates in Case one
Phenotypic susceptibility profiles are summarised in Table 1. All 5 of 7 isolates with phenotypic results from this child were reported as isoniazid susceptible. In total, two of five isolates were reported as phenotypically resistant to ethambutol. In the penultimate isolate (2015), *in vitro* rifampicin resistance was also reported. This corresponded to the identification of a rifampicin resistance mutation (*rpoB* D435V) in the 2015 isolate, a well characterised mutation in the rifampicin

resistance determining region that is the target of the widely-used MTB/RIF Xpert assay¹³. Interestingly, after rifampicin therapy was stopped and the patient suffered a further relapse in 2017, the organism was reported to be phenotypically susceptible and the D435V mutation was no longer present, although an alternative *rpoB* mutation (L430P) was present on WGS, and was also detected by Xpert MTB/RIF at the reference laboratory. The fact that the subsequent isolate had a different mutation is suggestive of a significant ongoing within-host selective pressure. That the rifampicin phenotype was susceptible despite the L430P resistance mutation is not unprecedented¹³.

Antibiotic susceptibility of other *M. bovis* isolates

Among the other seven children from whom BCG was isolated, one exhibited phenotypic resistance to ethambutol and one to isoniazid (Table 1). Despite resistance being reported to all first-line antibiotics within our set of isolates, no single isolate

was resistant to more than two drugs (i.e. more than one in addition to pyrazinamide). We were unable to identify any genetic basis for phenotypic resistance to isoniazid or ethambutol, despite searching through a list of known resistance-mechanisms with 94–98% sensitivity in *M. tuberculosis* infection¹³. As expected, based on derivation from a single vaccine strain, the paediatric isolates were all closely related, with a maximum of seven SNPs separating any two samples (Figure 1). Interestingly, we observed phenotypic variation in drug susceptibility even among isolates that are genomically indistinguishable.

Sequence data

All raw sequence data have been uploaded to the ENA and are available under the project number [PRJNA548242](https://www.ebi.ac.uk/ena/browser/view/PRJNA548242).

Discussion

We have confirmed that isolation of *M. bovis* BCG strain from clinical samples is rare, with only eight positive cases recorded over an 11-year period in a large teaching hospital laboratory. The majority of these reflected local adenitis, which is a well-recognised complication of vaccination^{14,15}. There was only

one case of disseminated *M. bovis* BCG, which occurred in a child with an immunodeficiency known to increase susceptibility to severe mycobacterial infection¹².

This epidemiology reflects that which has been previously described; in a previous study in London 98% of BCG complications were local, with only 2% having systemic sequelae¹⁴. Likewise, National UK Medicines and Healthcare Products Regulatory Agency (MHRA) data from 2013/14 reported 41 adverse incidents related to the BCG vaccine, of which only two were disseminated BCG infection, the rest being local complications¹⁶. The Australian mycobacterial reference laboratory detected 24 *M. bovis* BCG isolates in 2009 (nine in children under six years old), with the majority of these being representing vaccine site abscesses (n=13)¹⁷. Similarly, in the USA between 2006–2013 there were only 73 cases of *M. bovis* BCG strain isolated compared with 91,985 cases of *M. tuberculosis*¹⁸.

In this study, we are able to draw on full-length sequence data for *M. bovis* to assess the presence of drug-resistance mutations and correlate these with phenotypic resistance testing. Rifampicin resistance in *M. bovis* (BCG) has been

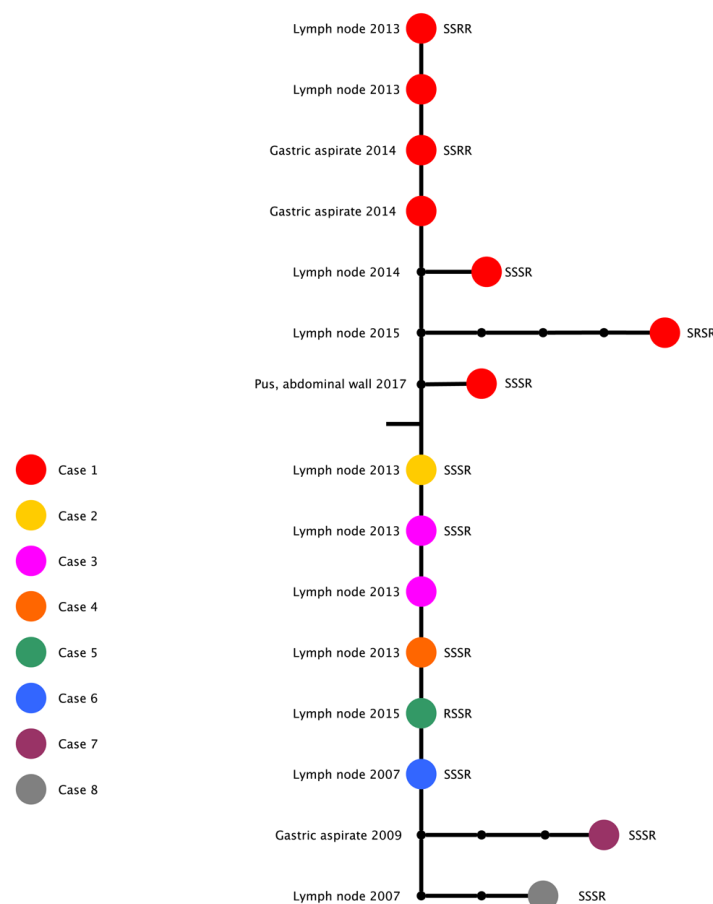


Figure 1. Maximum likelihood phylogenetic tree based on whole genome sequences of BCG isolates. Tip labels describe the case number, site of isolation, susceptibility profile ordered as isoniazid, rifampicin, ethambutol, pyrazinamide, and year of isolation. Scale bar shows a branch length of 0.5 SNPs.

reported previously, in each case in an immunocompromised child, including its emergence on treatment^{19–21}. Case one in our series had multiple isolates tested, with genotypic rifampicin resistance twice selected for, but only detected phenotypically once. This may be due to variations in the subpopulations of *M. bovis* organisms that are sampled in a patient with a high bacterial burden, or reflect antibiotic pressure (the rifampicin our patient had been receiving was stopped when the resistant isolate was detected). The presence of phenotypic resistance to isoniazid and ethambutol despite the absence of any identifiable genetic predictors of resistance is interesting: it might represent hitherto unrecognised mechanisms of resistance, or alternatively error in phenotypic determination of resistance. The latter is suggested by the fact that *in vitro* susceptibility reporting varies despite indistinguishable genomic sequences.

In our small series of cases we have observed *in vitro* resistance to each first-line anti-tuberculosis drug, despite BCG being a laboratory strain that is administered to patients. Despite known low-level isoniazid resistance in the Danish SSI 1331 BCG strain¹⁹, to our knowledge no underlying mechanism for this has been identified. Whatever the explanation, as far as we know the presence of resistance to any drug has only been clinically relevant in Case one in our cohort, whose primary problem was one of immunodeficiency predisposing to severe mycobacterial infection. Since local infection commonly either resolves spontaneously, or can be managed by aspiration or surgical excision^{14,15}, the antibiotic resistance profile may not be clinically relevant to developing a safe management plan.

Although the BCG vaccine is contraindicated in patients with immunodeficiencies, in the UK infants in high-risk groups for developing tuberculosis are usually vaccinated soon after birth, before any immunodeficiency will have become clinically apparent. In the cases with local vaccine site collections or adenitis, it could be argued that laboratory diagnosis did not inform the management, and thus in only one of the seven cases with clinical information available was the mycobacterial culture and/or sequence data clinically useful.

It is not known how many doses of BCG are given to UK children per year, but according to Public Health England

(PHE) data for high-risk areas, it will be considerably more than 30,000 doses²². A recent systematic review and meta-analysis²³ of the effect of BCG vaccination in children demonstrated a 19% protective efficacy against tuberculosis infection among vaccinated children after exposure compared with unvaccinated children, and protection by BCG against progression from infection to active disease of 58%. Thus, that only one case had invasive disease from all the children receiving the BCG in our large population catchment area, suggests BCG is safe, and that it is the host response that is important in the development of disease rather than any specific attribute of the vaccine strain.

In view of the dramatic changes in the development of whole genome sequencing of mycobacteria in the last few years, routine phenotypic antibiotic resistance testing has been discontinued in England in favour of genotypic testing. On the one hand this will provide more reproducible results, but on the other hand it will not detect resistance due to unknown mechanisms, such as that reported for isoniazid in the Danish BCG strain. However, work to enrich our knowledgebase of molecular resistance determinants is accelerating¹³ and the clinical relevance of this low-level isoniazid resistance that is only rarely detected even by phenotypic methods is questionable. Phenotyping only detected this purported isoniazid resistance in one of our clinical isolates.

In summary, we have shown isolation of *M. bovis* BCG strain is rare, and that the BCG vaccine is safe and well-tolerated in immunocompetent children. Only in the setting of profound immunocompromise do we see molecular and phenotypic rifampicin resistance emerging, most likely facilitated by a large bacillary burden. Because invasive disseminated BCG disease is rare, reporting and data sharing is important to allow assimilation of experience, particularly with respect to unusual cases of antibiotic resistance.

Data availability

Underlying data

European Nucleotide Archive: Clinical Mycobacterium bovis (BCG) isolates and antibiograms: a case series from children in Oxfordshire, 2007–2017. Accession number [PRJNA548242](https://identifiers.org/ebi/ena.embl:PRJNA548242); <https://identifiers.org/ebi/ena.embl:PRJNA548242>.

References

- Ritz N, Tebruegge M, Connell TG, et al.: **Susceptibility of Mycobacterium bovis BCG vaccine strains to antituberculous antibiotics.** *Antimicrob Agents Chemother.* 2009; **53**(1): 316–318.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Durek C, Rüscho-Gerdes S, Jocham D, et al.: **Sensitivity of BCG to modern antibiotics.** *Eur Urol.* 2000; **37** Suppl 1: 21–5.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Mount FW, Anastasiades AA, Schnack G: **Control study of biologically active isoniazid in serum of children with primary tuberculosis.** *Am Rev Respir Dis.* 1961; **83**: 173–183.
[PubMed Abstract](#)
- Cernuschi T, Malvolti S, Nickels E, et al.: **Bacillus Calmette-Guérin (BCG) vaccine: A global assessment of demand and supply balance.** *Vaccine.* 2018; **36**(4): 498–506.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Riordan A, Cole T, Broomfield C: **Fifteen-minute consultation: Bacillus Calmette-Guérin abscess and lymphadenitis.** *Arch Dis Child Educ Pract Ed.* 2014; **99**(3): 87–9.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Brummaier T, Yen SPY, Keerecharoen D, et al.: **The natural course of a localized BCG vaccine abscess.** *Vaccine.* 2019; **37**(20): 2757.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Talbot EA: **Mycobacterium bovis.** In: UpToDate.com. 2017; Accessed 29 Oct 2017.
[Reference Source](#)
- Public Health England: **Immunisation against infectious disease “The Green**

book". 2015.

[Reference Source](#)

9. Walker TM, Lalor MK, Broda A, *et al.*: **Assessment of *Mycobacterium tuberculosis* transmission in Oxfordshire, UK, 2007-12, with whole pathogen genome sequences: An observational study.** *Lancet Respir Med.* 2014; **2**(4): 285–292.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
10. Walker TM, Ip CLC, Harrell RH, *et al.*: **Whole-genome sequencing to delineate *Mycobacterium tuberculosis* outbreaks: a retrospective observational study.** *Lancet Infect Dis.* 2013; **13**(2): 137–146.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
11. Walker TM, Kohl TA, Omar SV, *et al.*: **Whole-genome sequencing for prediction of *Mycobacterium tuberculosis* drug susceptibility and resistance: A retrospective cohort study.** *Lancet Infect Dis.* 2015; **15**(10): 1193–1202.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
12. Wu U-I, Holland SM: **Host susceptibility to non-tuberculous mycobacterial infections.** *Lancet Infect Dis.* 2015; **15**(8): 968–980.
[PubMed Abstract](#) | [Publisher Full Text](#)
13. The CRYPTIC Consortium and 100 000 Genomes Project: **Prediction of Susceptibility to First-Line Tuberculosis Drugs by DNA Sequencing.** *N Engl J Med.* 2018; **379**(15): 1403–15.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
14. Venkataraman A, Yusuff M, Liebeschuetz S, *et al.*: **Management and outcome of Bacille Calmette-Guérin vaccine adverse reactions.** *Vaccine.* 2015; **33**(41): 5470–5474.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
15. Cuello-García C, Pérez-Gaxiola G, Gutiérrez CJ: **Treating BCG-induced disease in children (Review).** *Cochrane Database Syst Rev.* 2013; **2013**(1): CD008300.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
16. MHRA: **Paper provided by MHRA for Joint Committee on Vaccination and Immunisation October 2014: Vaccine-associated suspected adverse reactions reported via the yellow card scheme during 2013/14.** 2014.
[Reference Source](#)
17. Lumb R, Bastian I, Carter R, *et al.*: **Tuberculosis in Australia: bacteriologically confirmed cases and drug resistance, 2008 and 2009. A report of the Australian Mycobacterium Reference Laboratory Network.** *Commun Dis Intell Q Rep.* 2011; **35**(2): 154–61.
[PubMed Abstract](#)
18. Scott C, Cavanaugh JS, Pratt R, *et al.*: **Human Tuberculosis Caused by *Mycobacterium bovis* in the United States, 2006–2013.** *Clin Infect Dis.* 2016; **63**(5): 594–601.
[PubMed Abstract](#) | [Publisher Full Text](#)
19. Serum Staten Institute: **Summary of Product Characteristics (SPC)-BCG.** 2015.
20. Hesselting AC, Schaaf HS, Victor T, *et al.*: **Resistant *Mycobacterium bovis* Bacillus Calmette-Guérin Disease: Implications for Management of Bacillus Calmette-Guérin Disease in Human Immunodeficiency Virus-Infected Children.** *Pediatr Infect Dis J.* 2004; **23**(5): 476–479.
[PubMed Abstract](#) | [Publisher Full Text](#)
21. Hesselting AC, Schaaf HS, Hanekom WA, *et al.*: **Danish Bacille Calmette-Guerin Vaccine-Induced Disease in Human Immunodeficiency Virus-Infected Children.** *Clin Infect Dis.* 2003; **37**(9): 1226–1233.
[PubMed Abstract](#) | [Publisher Full Text](#)
22. Public Health England: **Quarterly vaccination coverage statistics for children aged up to five years in the UK (COVER programme): April to June 2017.** 2017.
23. Roy A, Eisenhut M, Harris RJ, *et al.*: **Effect of BCG vaccination against *Mycobacterium tuberculosis* infection in children: Systematic review and meta-analysis.** *BMJ.* 2014; **349**: g4643.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)

Open Peer Review

Current Peer Review Status:  

Version 1

Reviewer Report 20 November 2020

<https://doi.org/10.21956/wellcomeopenres.17888.r41519>

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Ben J. Marais 

Westmead Children's Hospital and the Marie Bashir Institute for Infectious Diseases and Biosecurity, University of Sydney, Sydney, NSW, Australia

Well written and interesting case series. Informed discussion and balanced conclusion.

Minor comments for consideration:

1. Variability in DST and SNPs in Danish strain 1331 BCG (without any treatment selection) is interesting. DST variability may be explained by inaccurate phenotypic results - it would be good to know the level of resistance detected in these instances. It is also relevant for INH, given that resistance was VERY rarely detected phenotypically, which is unexpected if tested at 0.1ug/ml.
2. For SNP variability the question remains if this represents post administration diversity or diversity in the 'vaccine stock' itself. Have the authors tested 'vaccine stock' (straight from the vial) to assess possible SNP variability?
3. In the discussion it would be good to provide an estimated denominator for the statement that BCG adverse effects were VERY rare. Do we know how many BCGs (roughly) were administered to children in the study area over the study period?
4. Any suggestions on how the single case could/should have been dealt with differently to prevent the recurrent relapses observed? Should the initial treatment period have been extended in light of the immune abnormality and ineffectiveness of PZA?
5. Can we accept 'significant within host selective pressure' as a likely explanation in a child with significant immune compromise? It seems as if treatment induced selective pressure, differential drug penetration and selective sampling of bacterial sub-populations offer the most likely explanation in this instance.

Overall this is an informative and well-presented report that makes an important contribution to the existing literature. Consideration of minor comments above should help to refine it further.

Is the background of the case's history and progression described in sufficient detail?

Yes

Are enough details provided of any physical examination and diagnostic tests, treatment given and outcomes?

Yes

Is sufficient discussion included of the importance of the findings and their relevance to future understanding of disease processes, diagnosis or treatment?

Yes

Is the case presented with sufficient detail to be useful for other practitioners?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Tuberculosis

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 04 November 2020

<https://doi.org/10.21956/wellcomeopenres.17888.r40938>

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Elizabeth Whitaker 

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This is an interesting and well written case report highlighting the complexity of managing a rare case of disseminated BCG. It also showed the importance of phenotypic sensitivity testing to ensure the most appropriate anti-mycobacterial treatment course for this rare condition. It also explains why multiple drugs are included in regimens to treat patients such as these. The authors do not comment on toxicity/adverse drug reactions in this child. They also don't comment on drug levels - metabolism of rifampicin in children differs greatly to adults, and higher drug doses are required to achieve therapeutic drug levels - this child may have benefited from therapeutic drug monitoring - the authors could comment on its role in these complex cases. It may be that low levels of rifampicin contributed to the within-host selective pressure.

As it is a case report, I would be interested to see more details on the clinical case, but understand why the authors have concentrated on the drug sensitivity element of the presentation - as this has relevance for the readers, in particular if they have a similar case in the future. I wonder if the team have considered the role of phage therapy for this child, as she seems to be still be on an antibiotic regimen 3 years later.

Reporting rare cases such as these has value as clinical decisions are made on expert opinion,

rather than evidence based data. This case will ensure future similar cases are considered in the context of drug sensitivity/resistance.

It would be interesting for the authors to comment on screening for primary immune deficiencies and planned changes to BCG delivery in the UK. Of particular note, this child would not have been identified by TREC screening, so the case would not have been avoided.

Is the background of the case's history and progression described in sufficient detail?

Yes

Are enough details provided of any physical examination and diagnostic tests, treatment given and outcomes?

Partly

Is sufficient discussion included of the importance of the findings and their relevance to future understanding of disease processes, diagnosis or treatment?

Yes

Is the case presented with sufficient detail to be useful for other practitioners?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Tuberculosis, BCG, mycobacterial immunity immunodeficiency in children SARS-CoV-2

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.
