

# SHORTROOT-Mediated Increase in Stomatal Density Has No Impact on Photosynthetic Efficiency<sup>1[CC-BY]</sup>

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The coordinated positioning of veins, mesophyll cells, and stomata across a leaf is crucial for efficient gas exchange and transpiration and, therefore, for overall function. In monocot leaves, stomatal cell files are positioned at the flanks of underlying longitudinal leaf veins, rather than directly above or below. This pattern suggests either that stomatal formation is inhibited in epidermal cells directly in contact with the vein or that specification is induced in cell files beyond the vein. The SHORTROOT pathway specifies distinct cell types around the vasculature in subepidermal layers of both root and shoots, with cell type identity determined by distance from the vein. To test whether the pathway has the potential to similarly pattern epidermal cell types, we expanded the expression domain of the rice (*Oryza sativa* ssp *japonica*) *OsSHR2* gene, which we show is restricted to developing leaf veins, to include bundle sheath cells encircling the vein. In transgenic lines, which were generated using the orthologous *ZmSHR1* gene to avoid potential silencing of *OsSHR2*, stomatal cell files were observed both in the normal position and in more distant positions from the vein. Contrary to theoretical predictions, and to phenotypes observed in eudicot leaves, the increase in stomatal density did not enhance photosynthetic capacity or increase mesophyll cell density. Collectively, these results suggest that the SHORTROOT pathway may coordinate the positioning of veins and stomata in monocot leaves and that distinct mechanisms may operate in monocot and eudicot leaves to coordinate stomatal patterning with the development of underlying mesophyll cells.

The coordinated differentiation of cell types within an organ is a crucial component of morphogenesis, necessary to ensure that the final form is appropriate for function. In this regard, photosynthetic function in plant leaves requires that chloroplast-containing cells in the middle leaf layers are interspersed with veins (to supply water and to redistribute metabolites) and are overlaid with stomatal pores through which carbon

dioxide can enter the leaf. In grass leaves, cellular arrangements are defined by parallel longitudinal veins that extend from the base of the leaf sheath to the tip of the leaf blade, with short transverse veins interconnecting the longitudinal network (Sharman, 1942; Esau, 1943; Nelson and Dengler, 1997). This vascular framework underpins linear files of stomata in the epidermis, with each vein being flanked by one to three rows of stomata on both the medial and lateral sides (Stebbins and Shah, 1960). The genetic mechanisms that ensure optimal photosynthetic capacity in grass leaves, by coordinating the development of veins, photosynthetic cell types, and stomata, are not known.

Grass leaves develop basipetally such that cellular differentiation proceeds from the tip of the leaf to the base, with cell divisions gradually becoming restricted to the base of the leaf blade (Sharman, 1942; Sylvester et al., 1990). In maize (*Zea mays*), the procambial centers that will form leaf veins are specified in subepidermal cell layers before cell divisions become restricted to the leaf base (Sharman, 1942; Russell and Evert, 1985; Sylvester et al., 1990), but the epidermal cell files that will form stomata only become distinguishable after that stage, when increased mitotic activity leads to smaller cells than in adjacent epidermal cell files (Stebbins and Shah, 1960). Although divisions in stomatal cell files normally increase cell numbers within

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rows rather than contribute to adjacent cell files, cell lineage analysis in maize revealed that cells within a stomatal row are not necessarily clonally related (Hernandez et al., 1999). As such, positional signals must trigger the specification of stomatal cell fate. Because stomatal cell files are specified in positions flanking procambial centers, it is conceivable that positional signals emanating from the developing vein either inhibit stomatal specification in epidermal cells directly above and below the procambium or induce specification in epidermal cell files at the flanks of the procambial center.

In rice (*Oryza sativa* ssp *japonica*), maize, and the model grass *Brachypodium distachyon*, stomatal identity and differentiation appear to be regulated by the same group of basic helix-loop-helix (bHLH) transcription factors that operate in the well-established pathway that was discovered in the eudicot *Arabidopsis thaliana*; Liu et al., 2009; Raissig et al., 2016, 2017). However, orthologous genes have been adopted into different roles in the two patterning networks. In *B. distachyon*, the combined activity of the bHLH proteins SPEECHLESS (BdSPCH1 and BdSPCH2) and INDUCER OF CBF EXPRESSION1 (BdICE) leads to the specification of stomatal fate, with loss-of-function mutants in either BdSPCH1/BdSPCH2 or BdICE lacking the characteristically spaced files of small epidermal cells at the leaf base that are the first visible indication of stomatal specification (Raissig et al., 2016). In considering what might induce stomatal specification in those cell files, we reflected on known patterning mechanisms that operate around the vasculature. Of note is the SHORTROOT (SHR) pathway, which acts to radially pattern cells around veins in *Arabidopsis* (Levesque et al., 2006; Cui et al., 2014). *AtSHR* is expressed specifically in the vasculature, but the protein, which is a member of the GRAS transcription factor family, moves outward to surrounding cell layers, where it interacts with a second GRAS family member, SCARECROW (SCR). SHR-SCR interactions specify cells as endodermis in the root (Helariutta et al., 2000) or bundle sheath in the shoot (Wysocka-Diller et al., 2000; Cui et al., 2014). Given that stomatal patterning is coordinated with vascular patterning, and that both *SHR* and *SCR* are expressed during stomatal development in rice (Kamiya et al., 2003), we speculated that the SHR pathway might act to induce the bHLH pathway in epidermal cell files that flank leaf veins and, thus, to regulate the spacing and localization of stomatal rows.

Two *SHR* genes are present in the rice genome (*OsSHR1* [Os07g39820] and *OsSHR2* [Os03g31880]; Cui et al., 2007). In situ hybridization analyses demonstrated that *OsSHR1* was expressed uniformly throughout the epidermis at the base of young leaf primordia, whereas the expression of *OsSHR2* was undetectable in either shoots or roots (Kamiya et al., 2003). This difference in expression level also was seen in recent transcriptomic data sets, where *OsSHR1* transcript levels were three times higher than those of *OsSHR2* (van Campen et al.,

2016). Notably, at the same stage that *OsSHR1* transcripts accumulate uniformly in the epidermis, *OsSCR1* transcripts are localized specifically in the files of small epidermal cells that will become stomatal rows (Kamiya et al., 2003). Later in development, once the asymmetric cell divisions associated with the specification of guard mother cells and subsidiary cells have occurred, both *OsSHR1* and *OsSCR1* transcripts are localized in individual stomatal complexes within stomatal cell files (Kamiya et al., 2003). This progressive spatial restriction is consistent with a role for *OsSHR1*-*OsSCR1* interactions in self-refining domains of activity during stomatal development, as seen during radial root patterning of *Arabidopsis* (Helariutta et al., 2000). However, there is no evidence that *OsSHR1* is expressed in the vasculature and, thus, no indication of a link between leaf veins and the specification of stomatal files in the epidermis. The expression domain of *OsSHR2* has yet to be resolved.

To dissect the mechanisms that coordinate vein formation, stomatal patterning, and photosynthetic capacity in grass leaves, we first tested whether SHR activity can influence stomatal development. We show that *OsSHR2* transcripts preferentially accumulate in developing vascular tissue of rice leaf primordia and, through a gain-of-function experiment, demonstrate that ectopic SHR activity in bundle sheath cells that surround leaf veins is sufficient to induce supernumerary stomatal rows in the epidermis. Despite theoretical predictions that the observed increase in stomatal density would improve photosynthetic capacity, no such gains were observed. Furthermore, in contrast to the phenotypes observed in *Arabidopsis* leaves with altered stomatal patterning (Dow et al., 2017), there was no increase in mesophyll cell density or in leaf thickness. These results illustrate the complexity of feedback mechanisms that coordinate the development of cell types in different layers of the leaf and point to distinct mechanisms operating in monocots and eudicots.

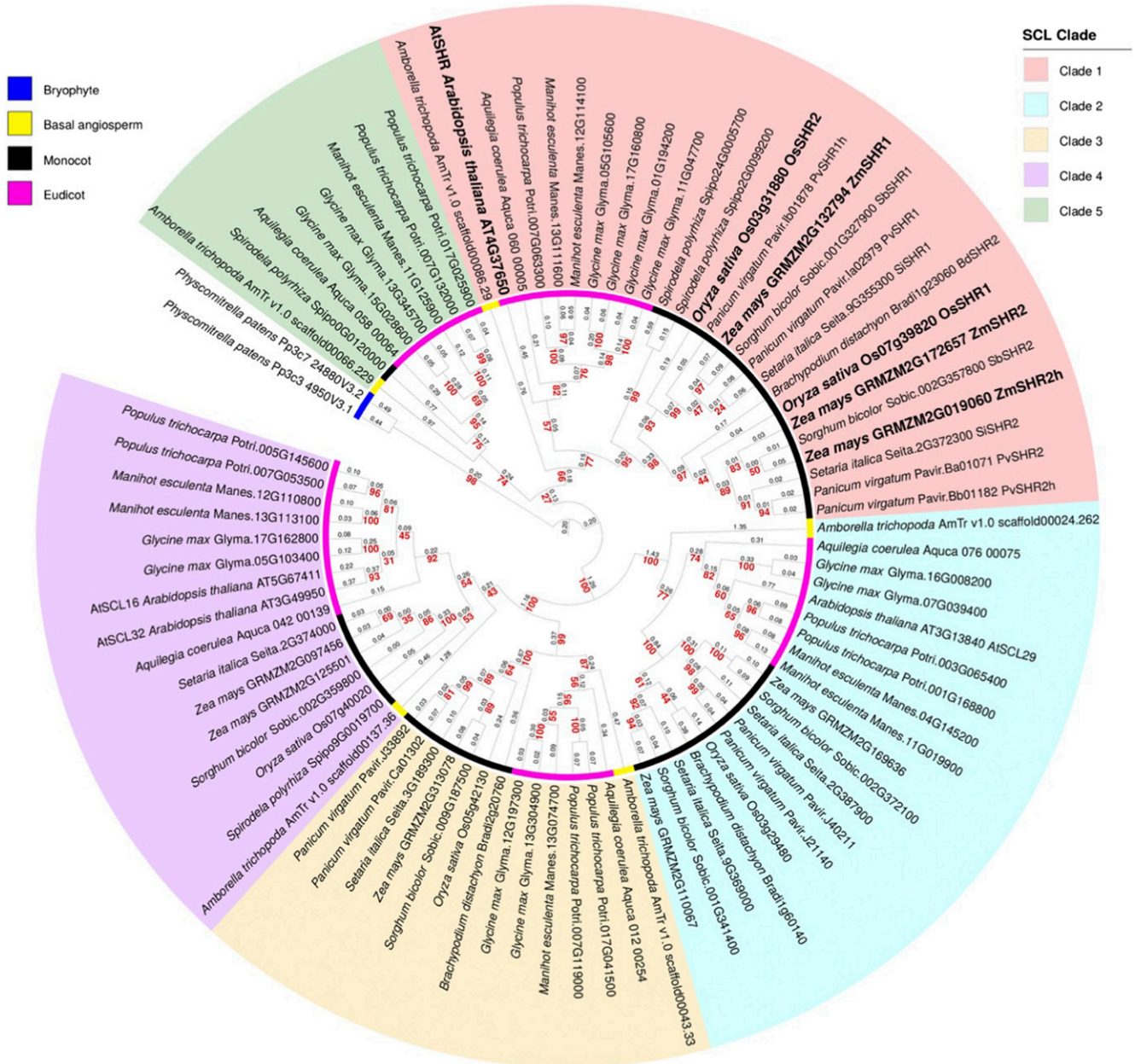
## RESULTS

### Duplicate *SHR* Genes in Grasses

Before testing the hypothesis that stomatal patterning in grass leaves is regulated by SHR activity in developing veins, the evolutionary relationships between *AtSHR* and related genes in the grasses were determined. Phylogenetic analysis was carried out with sequences from a broad range of land plant taxa, including five eudicot species: *Arabidopsis*, *Glycine max*, *Manihot esculenta*, *Populus trichocarpa*, and *Aquilegia coerulea*, the latter being a species that diverged early in the eudicot clade. Seven monocot species also were sampled: maize, rice, *Sorghum bicolor*, *Setaria italica*, *Panicum virgatum*, *B. distachyon*, and *Spirodela polyrrhiza*. To improve tree resolution, sequences were included from *Amborella trichopoda* (an angiosperm that diverged before the dicot-monocot split), and the tree was rooted

with the bryophyte *Physcomitrella patens* as the out-group. Figure 1 reveals five angiosperm gene clades, each of which contains genes from monocots, eudicots, and *A. trichopoda*. Within the *AtSHR* clade (Fig. 1, clade 1), there is evidence of a gene duplication prior to speciation in the grasses and further within-species duplications in maize and *P. virgatum* that are consistent with known whole-genome duplications (Schnable et al.,

2009; Okada et al., 2010). In maize, *ZmSHR2* and its homeolog *ZmSHR2h* have both been retained in the genome, whereas only one copy of *ZmSHR1* is present. The published nomenclature for these genes is somewhat confusing, because the *ZmSHR1* sequence (GRMZM2G132794; Slewinski et al., 2014) is orthologous to *OsSHR2* (Os03g31880), and the homeologs *ZmSHR2* (GRMZM2G172657) and *ZmSHR2h* (GRMZM2G019060)



**Figure 1.** Phylogenetic tree of SHR-like proteins. Maximum-likelihood inference was determined for phylogenetic relationships between SHR proteins in a basal angiosperm, six monocot, and five eudicot species, rooted with sequence from the bryophyte *P. patens*. Five distinct SCL clades are resolved. Branch lengths measured in number of amino acid substitutions per site are indicated in black, and bootstrap values for each node are shown in red. Homeologous relationships in monocot SHR proteins are indicated with h. Arabidopsis (*AtSHR*), rice (*OsSHR1* and *OsSHR2*), and maize (*ZmSHR1*, *ZmSHR2*, and homeolog *ZmSHR2h*) proteins are highlighted in boldface.



are orthologous to *OsSHR1* (Os07g39820). All five maize and rice *SHR* genes in clade 1 are equally related to *AtSHR*.

### *OsSHR2* Is Expressed in Developing Leaf Veins at the Time When Stomata Are Specified in the Overlying Epidermis

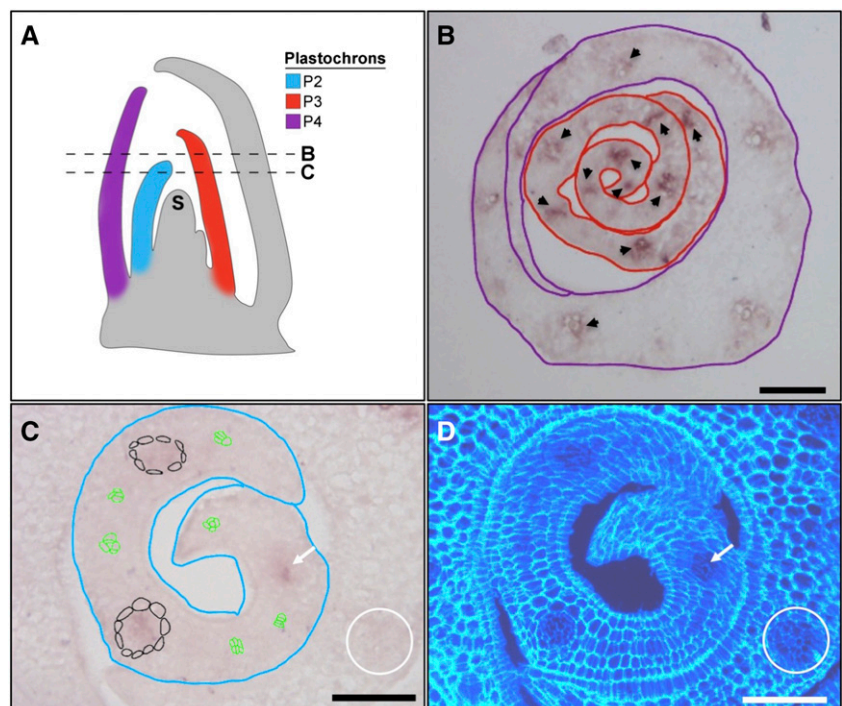
Although the *SHR* pathway is known to pattern cell types around both shoot and root vascular tissue in Arabidopsis, *OsSHR1* is not expressed in developing leaf veins of rice (Kamiya et al., 2003). To determine the spatial expression domain of *OsSHR2*, a gene-specific fragment was hybridized to developing leaf primordia. Figure 2 shows that *OsSHR2* transcripts accumulate in developing veins of plastochron P2 to P4 rice leaf primordia (where a plastochron is the time interval between the initiation of leaf primordia, and P1 is the youngest and closest to the shoot meristem from which the leaves are derived). In contrast to Arabidopsis, where *AtSHR* transcripts accumulate in procambium and persist throughout vascular development (Dhondt et al., 2010; Gardiner et al., 2011; Cui et al., 2014), *OsSHR2* transcripts are detectable only in veins after the surrounding bundle sheath cells have differentiated, and then expression is relatively short-lived (Fig. 2C). Notably, the time window of *OsSHR2* expression encompasses P3, which is when files of small cells that will become stomata are first visible in the epidermis (van Campen et al., 2016). The timing of *OsSHR2* transcript accumulation in leaf veins, therefore, precludes a role in procambium formation but is consistent with a subsequent role in cellular differentiation, including the specification of stomatal cell files in the overlying epidermis.

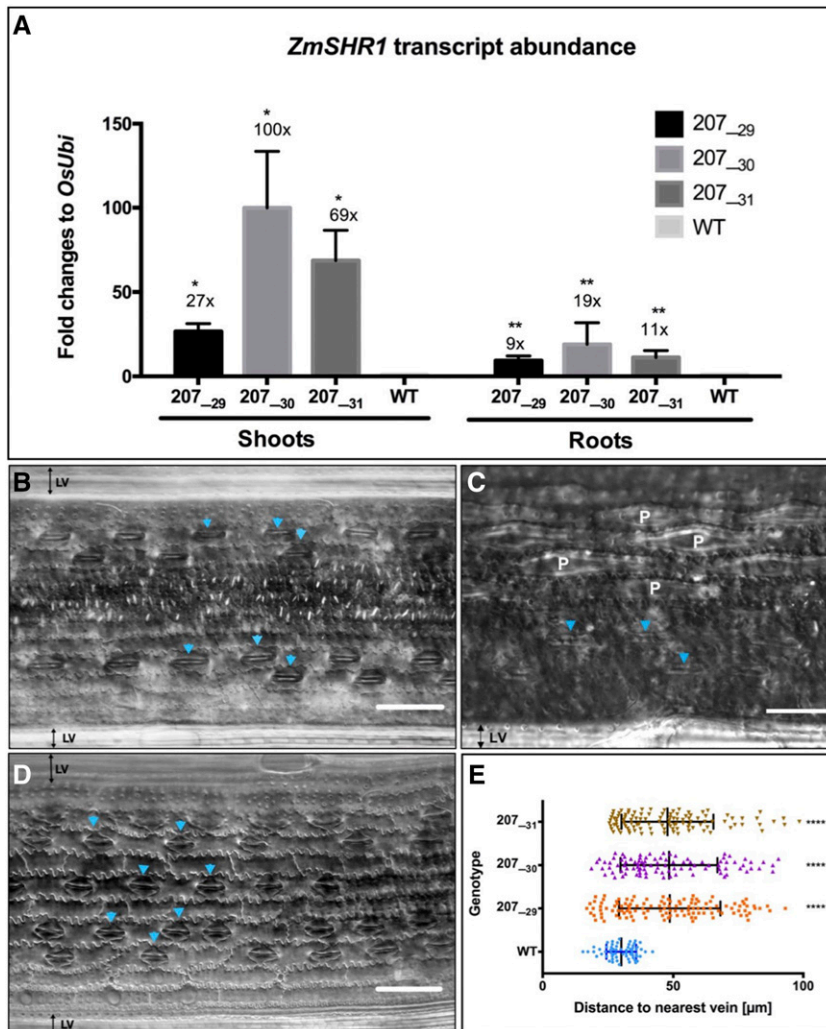
### Ectopic Expression of *ZmSHR1* Leads to the Formation of Extra Stomatal Rows in the Rice Epidermis

To determine whether *SHR* activity in subepidermal leaf layers has the potential to pattern stomata in the epidermis, transgenic rice lines were generated in which the temporal and spatial domains of *SHR* activity were expanded. Protein alignments gave no indication of substantial differences between rice and maize proteins in known functional domains (Supplemental Data Set S1); therefore, the orthologous *ZmSHR1* gene was used as the transgene to avoid potential silencing of *OsSHR2*. *ZmSHR1* was expressed either from the *ZmSHR1* promoter or from a promoter that is preferentially active in bundle sheath cells surrounding the vasculature. The rationale behind this approach was that if endogenous *OsSHR2* function in the developing vein specifies the position of stomatal rows on the flanks of the vein, expanding activity to cells encircling the vein should add extra rows of stomata in the epidermis.

Transgenic lines were generated by fusing the coding sequence of *ZmSHR1* either to the native maize promoter (*ZmSHR1<sub>pro</sub>*) or to the *Zoysia japonica* phosphoenolpyruvate carboxykinase promoter (*ZjPCK<sub>pro</sub>*) and transforming the constructs into the rice cv Kitaake. Although *ZmSHR1* accumulates preferentially in bundle sheath cells of maize leaves (Tausta et al., 2014), the spatial domain of *ZmSHR1<sub>pro</sub>* activity in rice has not been determined, whereas *ZjPCK<sub>pro</sub>* has been shown previously to direct gene expression in veins and mature bundle sheath cells of rice leaves and in the vasculature of rice roots (Nomura et al., 2005). Transgenic lines were recovered from transformations with three different constructs (*ZmSHR1<sub>pro</sub>::ZmSHR1*, a translational reporter fusion [*ZmSHR1<sub>pro</sub>::mTurquoise::ZmSHR1*], and

**Figure 2.** *OsSHR2* transcripts accumulate in developing leaf veins. A, Schematic of a monocot shoot apex in longitudinal cross section showing the shoot apical meristem (S) and plastochron (P) leaf primordia. P2, P3, and P4 are color coded as indicated. Relative positions of the cross sections shown in B and C are indicated with dashed lines. B, Transverse cross section showing P3 (red outline) and P4 (purple outline) primordia hybridized in situ with a gene-specific fragment of *OsSHR2*. Transcripts are visible in developing veins of the P3 primordium (arrowheads) but are much less abundant by P4. C, Transverse cross section showing P2 primordium (blue outline) hybridized in situ with a gene-specific fragment of *OsSHR2*. Transcripts are detected in well-developed veins that are surrounded by bundle sheath cells (outlined in black or indicated by the white arrow) but not in procambium (outlined in green) or in older veins in the P3 primordium (white circle). D, Same section as in C counterstained with Calcofluor White and photographed under UV light to view cellular structure. Bars = 50  $\mu$ m.





**Figure 3.** Ectopic *ZmSHR1* expression perturbs stomatal patterning in rice. A, Bar charts representing *ZmSHR1* transcript abundance in shoots and roots of wild-type (WT) cv Kitaake plus three T2 transgenic lines (207<sub>-29Hv</sub>, 207<sub>-30Kv</sub>, and 207<sub>-31J</sub>). Expression was quantified by quantitative reverse transcription-PCR. Transcript levels were normalized to endogenous rice ubiquitin (*OsUbi*) transcripts and are presented as fold changes above background (wild-type levels are set to 1). Values represent means of two biological replicates, and error bars represent SE. \*,  $P < 0.02$  and \*\*,  $P < 0.004$  as determined by a two-tailed Student's *t* test. B to D, Paradermal views of the abaxial epidermis of cleared wild-type (B) and *ZjPCK<sub>pro</sub>:ZmSHR1* (D) leaves visualized by DIC microscopy. A magnified view (C) illustrates papillae cells (denoted as P) in the wild type. Arrowheads point to stomata. LV, Lateral vein. Bars = 50  $\mu\text{m}$  (B and D) and 75  $\mu\text{m}$  (C). E, Box plot representing measured distances between stomata and the nearest vein in leaves of the wild type and three *ZjPCK<sub>pro</sub>:ZmSHR1* T2 lines. Each data point represents a single stomatal complex. *P* values were determined by one-way ANOVA where transgenics were individually compared with the wild type: \*\*\*\*,  $P = 0.0001$ .

*ZjPCK<sub>pro</sub>:ZmSHR1*). In all three cases, supernumerary stomatal files were observed in the leaves of T0 regenerants (Supplemental Fig. S1), demonstrating that ectopic SHR activity is sufficient to perturb stomatal patterning in the rice epidermis. Due to very low fertility in *ZmSHR1<sub>pro</sub>:ZmSHR1* and *ZmSHR1<sub>pro</sub>:mTurquoise::ZmSHR1* lines and given the known fidelity of *ZjPCK<sub>pro</sub>* for bundle sheath preferential activity in rice (Nomura et al., 2005), the *ZjPCK<sub>pro</sub>:ZmSHR1* line was advanced for further phenotypic analysis. Three T1 individuals (207<sub>-29</sub>, 207<sub>-30</sub>, and 207<sub>-31</sub>) were propagated into the T2 generation, and then homozygous T2 lines (207<sub>-29Hv</sub>, 207<sub>-30Kv</sub>, and 207<sub>-31J</sub>) that encompassed a range of transgene expression levels (from 27- to 100-fold above background in the shoot) were selected for all subsequent analyses. Figure 3A shows that *ZmSHR1* transcripts accumulated in both shoot and root tissues of all three T2 lines, with levels relative to endogenous ubiquitin being highest in line 207<sub>-30</sub> and levels in all lines being higher in shoots than in roots.

To determine the extent to which the expression of the *ZjPCK<sub>pro</sub>:ZmSHR1* transgene perturbs stomatal patterning, both adaxial and abaxial epidermal layers

of transgenic and wild-type plants were cleared and analyzed by differential interference contrast (DIC) microscopy. In wild-type leaves, stomata are organized in one or two linear cell files close to both large and small veins. Given the distance between veins, this pattern leads to three or four stomatal cell files plus at least three or four nonstomatal files between each vein pair (Fig. 3, B and C). In leaves of the *ZjPCK<sub>pro</sub>:ZmSHR1* plants, stomatal rows were more evenly distributed between veins in both the adaxial and abaxial (Fig. 3D) epidermis. In the abaxial epidermis, the extra stomatal files replaced the epidermal papillae that are usually present on the wild-type epidermis (Fig. 3, B–D). To quantify the observed changes, the distance between individual stomata and the nearest vein was measured. Figure 3E illustrates how stomatal position varies from 20 to 100  $\mu\text{m}$  from a vein in the transgenic lines, as opposed to a range of just 20 to 40  $\mu\text{m}$  in the wild type.

To assess whether the ectopic positioning of stomata also reflects an increase in overall stomatal number across the leaf, stomatal density (stomata per  $\text{mm}^2$ )

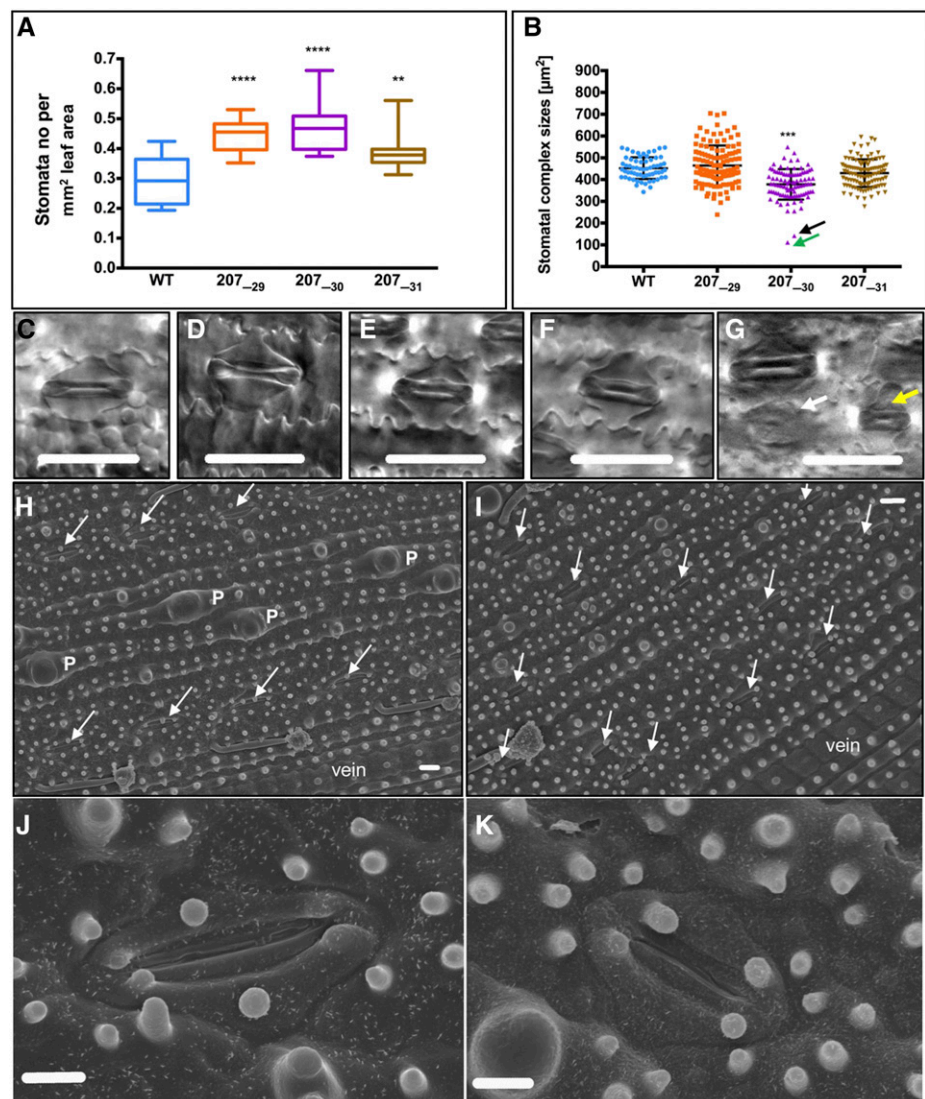


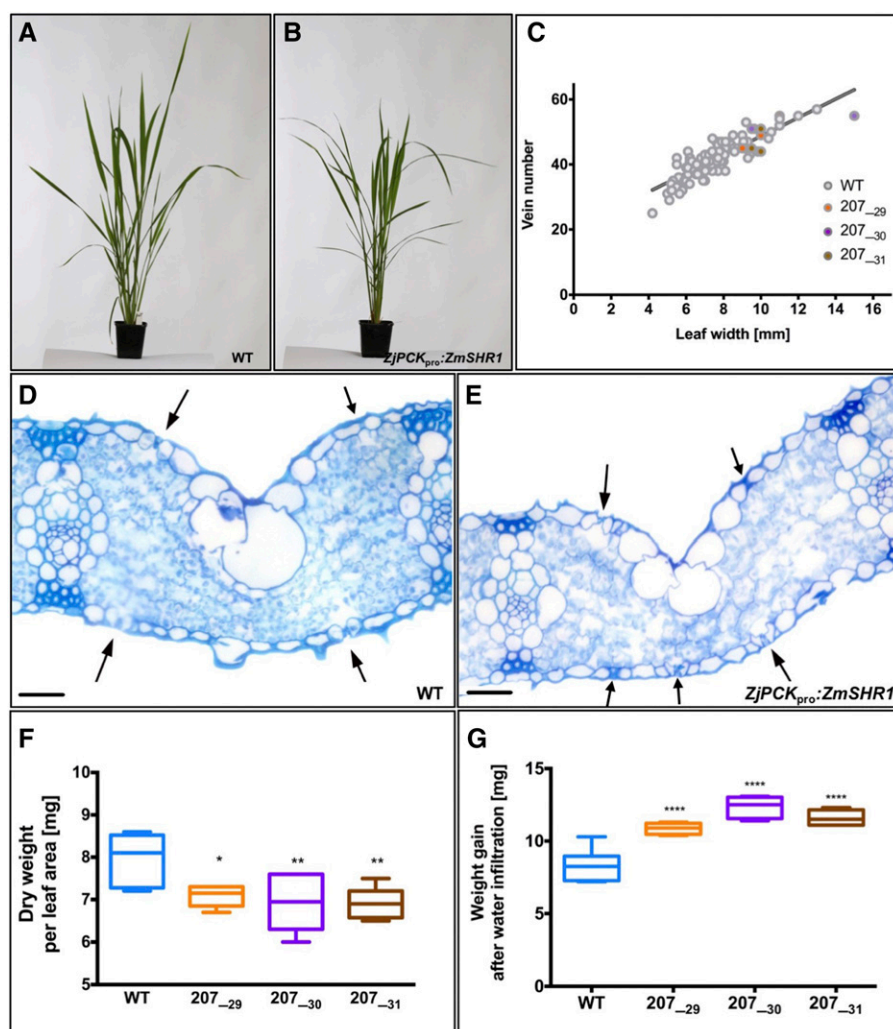
was measured. Figure 4A shows that stomatal density was increased significantly in all three *ZjPCK<sub>pro</sub>:ZmSHR1* lines as compared with the wild type. Because stomatal density has been shown previously to be negatively correlated with stomatal size (Ohsumi et al., 2007; Franks et al., 2009; Franks and Beerling, 2009), measurements of stomatal size also were taken. A significant decrease in size was observed only for line 207<sub>-30</sub> (Fig. 4, B and G), the line that had the highest levels of transgene expression (Fig. 3A) and the highest stomatal density (Fig. 4A). Other than the size, stomatal anatomy in all three transgenic lines was essentially indistinguishable from that of the wild type (Fig. 4, C–F and H–K). No consistent difference in the number of stomata per row could be detected. Together, these results demonstrate that the additional stomatal rows result from the formation of supernumerary stomata in the epidermis and that, to some extent, this increase in number is associated with a decrease in stomatal size.

### The Development of Supernumerary Stomata Has No Significant Impact on General Shoot Morphology or Overall Leaf Anatomy

To determine whether the presence of extra stomata on the leaf surface had any impact on general shoot development in rice, overall plant morphology was examined. Plant stature, tiller number, growth rate, and flowering time were similar in both wild-type (Fig. 5A) and *ZjPCK<sub>pro</sub>:ZmSHR1* transgenic (Fig. 5B) lines, suggesting that neither ectopic *ZmSHR1* activity nor the presence of supernumerary stomatal files caused any gross alterations in shoot anatomy. To confirm this observation, particularly given that loss-of-function mutations in *shr1* in maize are reported to perturb vein spacing and the specification of bundle sheath cells (Slewisinski et al., 2014), a range of leaf anatomy traits also were quantified. Table I shows measurements of leaf width, leaf length, total number of longitudinal veins, bundle sheath cell size, cell number, and distance

**Figure 4.** Increased stomatal density is not associated with altered stomatal anatomy. A, Box plot showing stomata number measured per mm<sup>2</sup> of leaf area of the wild type (WT) and three *ZjPCK<sub>pro</sub>:ZmSHR1* T2 lines. B, Box plot of stomatal complex sizes calculated from stomatal height multiplied by stomatal length. Error bars = 50  $\mu$ m. All *P* values were determined by one-way ANOVA where transgenics were individually compared with the wild type: \*\*\*\*, *P* = 0.0001; \*\*\*, *P* < 0.001 and \*\*, *P* < 0.01. C to F, Representative examples of stomatal morphology in the wild type (C), 207<sub>-29</sub> (D), 207<sub>-30</sub> (E), and 207<sub>-31</sub> (F) lines photographed by DIC microscopy. G, Morphology of the smallest complexes in line 207<sub>-30</sub>. The white arrow corresponds to the data point indicated by the black arrow in B, and the yellow arrow corresponds to the green arrow in B. H and I, Positive imprints of the abaxial epidermis of wild-type (H) and *ZjPCK<sub>pro</sub>:ZmSHR1* line 207<sub>-31</sub> (I) leaves visualized by scanning electron microscopy. Papillae cells are indicated as P, arrows point to stomata, and veins are indicated. J and K, Representative examples of stomatal morphology in wild-type (J) and 207<sub>-31</sub> (K) lines photographed by scanning electron microscopy. Bars = 30  $\mu$ m (C–G), 10  $\mu$ m (H and I), and 5  $\mu$ m (J and K).





**Figure 5.** Expression of *ZmSHR1* in rice does not perturb overall shoot morphology or leaf anatomy. A and B, Representative phenotypes of 6-week-old wild-type (WT; A) and transgenic (B) soil-grown plants. C, Regression plot showing vein number versus leaf width for four individuals from three T2 lines. Gray dots represent wild-type measurements from Table I combined with measurements of wild-type leaves at various developmental stages (Supplemental Data Set S2). D and E, Transverse sections of penultimate leaves from representative wild-type (D) and *ZjPCK<sub>pro</sub>::ZmSHR1* (E) plants. Arrows indicate stomata. Bars = 500  $\mu$ m. F and G, Dry weight (F) and weight gain after water infiltration (G) for leaves of the wild type and three *ZjPCK<sub>pro</sub>::ZmSHR1* T2 lines. All  $P$  values were determined by one-way ANOVA where transgenics were individually compared with the wild type: \*\*\*\*,  $P = 0.0001$ ; \*\*,  $P < 0.01$ ; and \*,  $P < 0.1$ .

between veins for four individuals of each T2 line and the wild type. ANOVA revealed no significant differences between wild-type and transgenic lines for any of these traits. In addition, the ratio of leaf width to vein number was normal in transgenic lines (Fig. 5C). Histological examination of leaf sections similarly demonstrated normal patterning of subepidermal layers (Fig. 5, D and E), with no significant difference observed between wild-type and transgenic lines in mesophyll cell size, mesophyll cell number between veins, or leaf thickness (Table II). In lines 207<sub>-30</sub> and 207<sub>-31</sub>, a significant difference was observed in the number of cell layers present between abaxial and adaxial epidermal layers. However, given equivalent leaf thickness in the wild type and all three transgenic lines, the reduced number of cell layers must have been compensated for by subtle differences in cell size, cell number, and orientation (Table II). Notably, lines 207<sub>-30</sub> and 207<sub>-31</sub> did not have significantly higher stomatal densities than line 207<sub>-29</sub> (Fig. 4A), but they did have much higher transgene expression levels (Fig. 3A). As such, the effect on mesophyll cell layers could be a direct consequence

of ectopic SHR activity rather than an effect of stomatal density. Extra stomata in all of the transgenic lines were generally associated with extra substomatal air spaces, as determined by loss of leaf weight after drying and gain of leaf weight after vacuum infiltration of water (Byott, 1976). When compared with the wild type, transgenic leaves were clearly lighter after drying (Fig. 5F) and heavier after water infiltration (Fig. 5G), suggesting the presence of more air spaces in leaves of the transgenic lines. Together, these data suggest that *ZmSHR1*-mediated induction of supernumerary stomata in rice does not perturb general shoot morphology or the overall leaf anatomy.

#### Root Architecture Is Modified in Rice Lines That Express *ZmSHR1*

In all flowering plant species examined thus far, loss of SHR function in the root perturbs radial patterning of endodermal and cortical cells around the vasculature and leads to shorter roots overall. Consistent with this

**Table 1.** Quantification of leaf anatomy traits

Measurements were taken from the sixth fully expanded adult leaf on the first tiller for four individuals from each line. For distance and cell number between veins, measurements were taken from three or four pairs of intermediate veins for each leaf. Bundle sheath cell size measurements were taken from six cells in each leaf. Values represent means  $\pm$  SE. Significance was tested using a one-way ANOVA ( $P < 0.05$ ). There were no significant differences between the three transgenic lines and the wild type (WT) for any of the measurements.

Line	Leaf Width	Leaf Length	Total No. of Veins	Average Distances between Veins ( $n = 3$ )	Average Cell No. between Veins ( $n = 3-4$ )	Average Bundle Sheath Cell Size ( $n = 6$ )	Vein No. per mm of Leaf Area
	mm			$\mu\text{m}$		$\mu\text{m}$	
WT <sub>1a</sub>	11	465	52	212 $\pm$ 6.7	6.5 $\pm$ 0.6	9.9 $\pm$ 1.7	4.7
WT <sub>1b</sub>	11	435	54	185 $\pm$ 2.9	6.3 $\pm$ 0.6	10.2 $\pm$ 3.1	4.7
WT <sub>1c</sub>	12	550	55	219 $\pm$ 4.2	6.7 $\pm$ 0.6	9.8 $\pm$ 2.8	4.6
WT <sub>1d</sub>	13	565	57	213 $\pm$ 43.2	6 $\pm$ 0	10 $\pm$ 2.4	4.4
WT total	11.9 $\pm$ 0.4	503.8 $\pm$ 31.8	54.5 $\pm$ 1.0	207.3 $\pm$ 7.6	6.4 $\pm$ 0.1	10.0 $\pm$ 0.1	4.6 $\pm$ 0.1
207 <sub>-29-1a</sub>	10	455	44	150 $\pm$ 8.4	8.4 $\pm$ 0.5	8.6 $\pm$ 0.2	4.4
207 <sub>-29-1b</sub>	11	478	55	203 $\pm$ 6.3	5.2 $\pm$ 0.5	11.2 $\pm$ 3.8	5.0
207 <sub>-29-1c</sub>	10	490	49	194 $\pm$ 8.3	6.8 $\pm$ 0.5	8.3 $\pm$ 1.6	4.9
207 <sub>-29-1d</sub>	9	405	45	199 $\pm$ 16.2	6.25 $\pm$ 0.5	9.2 $\pm$ 0.9	5.0
207 <sub>-29</sub> total	10.0 $\pm$ 0.4	457.0 $\pm$ 18.8	48.4 $\pm$ 2.5	186.4 $\pm$ 12.4	6.7 $\pm$ 0.7	9.3 $\pm$ 0.7	4.8 $\pm$ 0.1
207 <sub>-30-1a</sub>	10	512	44	215 $\pm$ 13.3	7.4 $\pm$ 1.1	8.7 $\pm$ 0.3	4.4
207 <sub>-30-1b</sub>	15	595	55	202 $\pm$ 4.2	7.3 $\pm$ 0.6	9.0 $\pm$ 1.7	3.7
207 <sub>-30-1c</sub>	9.5	435	51	194 $\pm$ 3.5	7.3 $\pm$ 0.6	10.4 $\pm$ 1.7	5.4
207 <sub>-30-1d</sub>	10	443	51	205 $\pm$ 8.1	7.6 $\pm$ 0.6	8 $\pm$ 2.2	5.1
207 <sub>-30</sub> total	11.1 $\pm$ 1.3	496.3 $\pm$ 37.2	50.3 $\pm$ 2.3	204.1 $\pm$ 4.4	7.4 $\pm$ 0.1	9.0 $\pm$ 0.5	4.6 $\pm$ 0.4
207 <sub>-31-1a</sub>	10	420	44	187 $\pm$ 16.6	8 $\pm$ 1.0	9.4 $\pm$ 0.5	4.4
207 <sub>-31-1b</sub>	9.5	579	45	219 $\pm$ 2.5	8.6 $\pm$ 0.5	8.6 $\pm$ 1.7	4.7
207 <sub>-31-1c</sub>	11	560	52	211 $\pm$ 5.1	7.5 $\pm$ 0.5	10.1 $\pm$ 2.2	4.7
207 <sub>-31-1d</sub>	10	513	51	180 $\pm$ 10.4	6.5 $\pm$ 0.5	9.0 $\pm$ 1.9	5.1
207 <sub>-31</sub> total	10.1 $\pm$ 0.3	518 $\pm$ 35.5	48.0 $\pm$ 2.0	199.1 $\pm$ 9.3	7.7 $\pm$ 0.4	9.2 $\pm$ 0.3	4.7 $\pm$ 0.1

observation, gain of SHR function in *ZjPCK<sub>pro</sub>:ZmSHR1* transgenic rice plants led to longer roots with finer tips and an increased number of lateral roots compared with the wild type (Fig. 6, A–D). The formation of extra lateral roots is analogous to the role of *AtSHR* in lateral root formation in *Arabidopsis* (Lucas et al., 2011). However, the long-root phenotype contrasts with previous gain-of-function experiments in which the constitutive expression of *AtSHR* or *OsSHR2* led to short roots with perturbed radial patterning that was manifest by the presence of extra cortical cell layers (Henry et al., 2017). Transverse sections through zones of wild-type roots (Fig. 6, E, G, and I) and *ZjPCK<sub>pro</sub>:ZmSHR1* transgenic roots (Fig. 6, F, H, and J) revealed that the central vasculature, endodermal layer, and number of cortical layers were the same in both. Collectively, these data demonstrate that *ZjPCK<sub>pro</sub>*-driven expression of *ZmSHR1* in rice leads to root elongation and the production of more lateral roots, but these perturbations are not accompanied by patterning defects and do not impact shoot development in any obvious way.

#### Increased Stomatal Density Predicts a Higher Photosynthetic Capacity in *ZjPCK<sub>pro</sub>:ZmSHR1* Lines, But the Predicted Gains Are Not Realized

Because stomatal density and the size of the stomatal pore determine stomatal conductance to CO<sub>2</sub> and water, and thus strongly correlate with photosynthetic capacity (Fischer et al., 1998; Franks et al., 2009; Franks and Beerling,

2009), it has been hypothesized that increased stomatal density could increase photosynthetic capacity and, hence, crop yield (Jones, 1977, 1987). However, such an approach has had little success in conventional breeding programs. The increased stomatal density observed in transgenic *ZjPCK<sub>pro</sub>:ZmSHR1* lines enabled the impact of varying stomatal numbers to be evaluated both theoretically and experimentally. Measurements of stomatal density and of pore length ( $\mu\text{m}$ ) and depth ( $\mu\text{m}$ ; equivalent to guard cell width) can be used to calculate the theoretical maximum rate of photosynthesis (Fig. 7A; Brown and Escombe, 1900; Franks and Farquhar, 2001). Therefore, pore length and depth were measured in leaves of wild-type and *ZjPCK<sub>pro</sub>:ZmSHR1* transgenic lines (Table III). These values were then averaged and used to calculate the theoretical maximum rate of stomatal conductance to water vapor ( $g_{\text{max}}$ ; Fig. 7A) and, from that, the theoretical maximum stomatal conductance to CO<sub>2</sub> ( $g_{\text{cmx}}$  [by dividing  $g_{\text{max}}$  values by 1.6 to account for the differences in molecular diffusion between water vapor and CO<sub>2</sub>]; Farquhar and Sharkey, 1982). Table III shows that predicted  $g_{\text{max}}$  values of *ZjPCK<sub>pro</sub>:ZmSHR1* lines are 25% to 60% higher than those in the wild type and that  $g_{\text{cmx}}$  values are 20% to 40% higher than those in the wild type. Together, these data suggest the potential for a higher stomatal conductance and, therefore, greater photosynthetic capacity in *ZjPCK<sub>pro</sub>:ZmSHR1* lines as compared with the wild type.

To investigate whether the theoretically predicted increase in photosynthetic capacity was realized in *ZjPCK<sub>pro</sub>:ZmSHR1* transgenic lines, leaf chlorophyll



**Table II.** Quantification of mesophyll cell size and layers

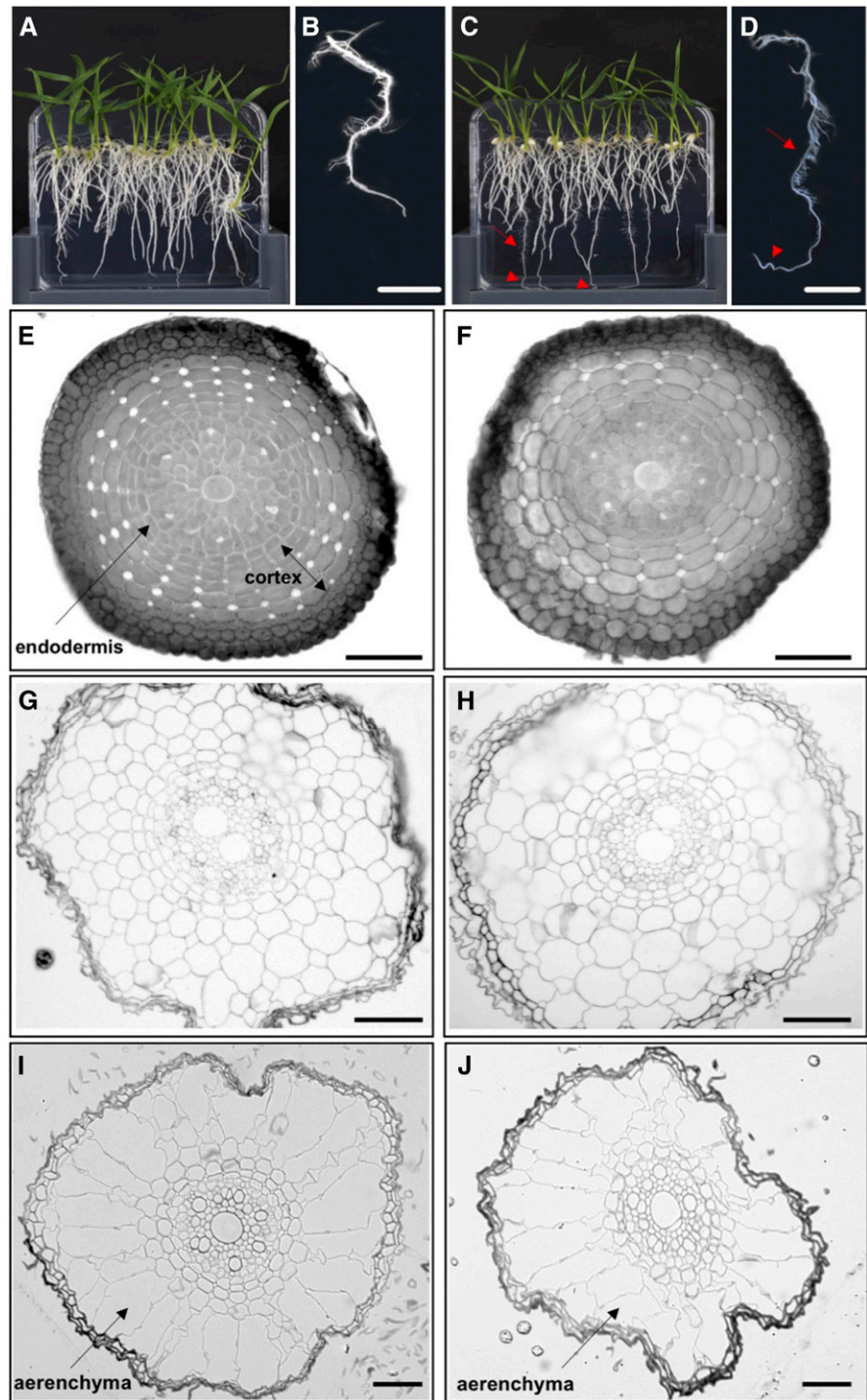
Measurements were taken from hand-cut cross sections of fully expanded adult leaves of different leaf widths. Leaf thickness represents the mean of three measurements taken at three points across the mediolateral axis: at a vein, directly adjacent to a vein, and midway between two veins. To cover the normal range seen in the wild type (WT), leaves with widths between 9 and 13 mm were chosen for measurements. Mesophyll cell sizes represent means of six cell size measurements in one leaf. For cell number between veins, measurements were taken between three or four pairs of intermediate veins for each leaf. Values represent means  $\pm$  SE. Significance was tested using a one-way ANOVA. There was a significantly lower number of mesophyll cell layers between line 207<sub>30</sub> and the wild type (\*\*\*,  $P < 0.0005$ ) and between line 207<sub>31</sub> and the wild type (\*,  $P < 0.05$ ). Leaf thickness, mesophyll cell size, and mesophyll cell number between veins were not significantly different between transgenic lines and the wild type.

Line	Leaf Width	Leaf Thickness	No. of Mesophyll Cell Layers	Mesophyll Cell Size	Mesophyll Cell No. between Veins
	mm	$\mu\text{m}$		$\mu\text{m}^2$	
WT <sub>1a</sub>	11	91.7 $\pm$ 4.3	6	240.5 $\pm$ 35.3	8–9
WT <sub>1b</sub>	11	94.3 $\pm$ 5.0	5	186.33 $\pm$ 45.8	8–9
WT <sub>1c</sub>	11	88.3 $\pm$ 2.8	5	172.7 $\pm$ 46	8
WT <sub>2a</sub>	12	88.7 $\pm$ 8.8	6	218.3 $\pm$ 20.5	7–8
WT <sub>2b</sub>	12	91 $\pm$ 1.0	6	205.7 $\pm$ 30.2	7–9
WT <sub>2c</sub>	12	82 $\pm$ 2.6	5	210.5 $\pm$ 25.1	7
WT <sub>3a</sub>	13	85.6 $\pm$ 5.6	6	251.8 $\pm$ 33.9	7–9
WT <sub>3b</sub>	13	82.6 $\pm$ 2.6	5	241.8 $\pm$ 32.9	7–9
WT <sub>3c</sub>	13	76 $\pm$ 5.1	5	195.3 $\pm$ 30.1	7–9
WT <sub>4a</sub>	10	79.7 $\pm$ 4.9	4	205.2 $\pm$ 12.4	8
WT <sub>4b</sub>	10	80.7 $\pm$ 4.1	4	229.8 $\pm$ 30.9	7
WT <sub>4c</sub>	10	80 $\pm$ 5.1	3	190.7 $\pm$ 20.2	8
WT <sub>5a</sub>	9	77.5 $\pm$ 4.3	4	205.8 $\pm$ 23.3	8
WT <sub>5b</sub>	9	82 $\pm$ 6.0	4	197.5 $\pm$ 37	8
WT <sub>5c</sub>	9	69.3 $\pm$ 9.2	3	172.8 $\pm$ 37.2	7
WT total		83.3 $\pm$ 1.7	4.7 $\pm$ 0.3	205.3 $\pm$ 6.1	7–9
207 <sub>–29</sub> –1a	9	71.7 $\pm$ 6.0	5	131.2 $\pm$ 34.3	11
207 <sub>–29</sub> –1b	9	67 $\pm$ 4.0	4	186.5 $\pm$ 43.2	8
207 <sub>–29</sub> –1c	9	78 $\pm$ 2.1	5	229.7 $\pm$ 63.7	7
207 <sub>–29</sub> –2a	10	91 $\pm$ 7.5	4	233.3 $\pm$ 58.8	7
207 <sub>–29</sub> –2b	10	77 $\pm$ 7.1	4	191.2 $\pm$ 16.5	7
207 <sub>–29</sub> –2c	10	74.3 $\pm$ 9.7	4	185.7 $\pm$ 41	9
207 <sub>–29</sub> –3a	12	84.7 $\pm$ 12.3	4	224.2 $\pm$ 29.9	8–9
207 <sub>–29</sub> –3b	12	90.3 $\pm$ 11.8	4	200.3 $\pm$ 40.3	8
207 <sub>–29</sub> –3c	12	84.7 $\pm$ 12.9	3	209 $\pm$ 27.9	7
207 <sub>–29</sub> total		79.9 $\pm$ 2.8	4.1 $\pm$ 0.2	199 $\pm$ 10.4	7–11
207 <sub>–30</sub> –1a	10	78 $\pm$ 4.2	3	261 $\pm$ 44.9	6–8
207 <sub>–30</sub> –1b	10	81.3 $\pm$ 3.7	3	188.8 $\pm$ 26.3	8
207 <sub>–30</sub> –1c	10	85 $\pm$ 5.7	3	220 $\pm$ 48.5	7
207 <sub>–30</sub> –2a	11	79 $\pm$ 3.1	3	200.4 $\pm$ 30.6	6
207 <sub>–30</sub> –2b	11	88.3 $\pm$ 7.5	3	210.5 $\pm$ 51.8	7
207 <sub>–30</sub> –2c	11	78.7 $\pm$ 9.9	3	251.3 $\pm$ 39.8	6
207 <sub>–30</sub> total		81.7 $\pm$ 1.7	3 $\pm$ 0***	222 $\pm$ 11.7	6–8
207 <sub>–31</sub> –1a	10	80 $\pm$ 9.1	4	158 $\pm$ 40.3	9
207 <sub>–31</sub> –1b	10	81.7 $\pm$ 4.1	4	227.3 $\pm$ 51.7	7
207 <sub>–31</sub> –1c	10	81.7 $\pm$ 3.5	3	147 $\pm$ 10.9	8
207 <sub>–31</sub> –2a	11.5	84.3 $\pm$ 3.2	4	271.7 $\pm$ 27.8	8
207 <sub>–31</sub> –2b	11.5	79.7 $\pm$ 3.9	4	230.3 $\pm$ 22.5	7
207 <sub>–31</sub> –2c	11.5	74.3 $\pm$ 5.2	3	198.2 $\pm$ 28.8	8
207 <sub>–31</sub> total		80.3 $\pm$ 1.4	3.6 $\pm$ 0.2*	205.4 $\pm$ 19.3	7–9

content was quantified, and gas-exchange experiments were carried out to measure both stomatal conductivity to water vapor ( $g_s$ ) and photosynthetic carbon assimilation rates ( $A$ ). No significant difference in chlorophyll content was observed between wild-type and transgenic lines (Fig. 7B), suggesting that mesophyll cell chloroplast capacity is normal in the transgenic lines. For gas-exchange experiments, a light intensity of 1,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$  (photosynthetic photon flux density [PPFD]) was first identified as the lowest PPFD at which the saturation point of  $\text{CO}_2$  assimilation was reached in one representative *ZjPCK<sub>pro</sub>:ZmSHR1* line

and the wild type (Supplemental Fig. S2). To resolve the impact of stomatal distribution on stomatal opening and photosynthetic induction, net gas exchange was measured in leaves during a low (100  $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) to high (1,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) light transition. There was a rapid and immediate increase in  $g_s$  (Fig. 7C) and  $A$  (Fig. 7D) in response to high light in all measured lines. However, despite the increased  $g_{\text{max}}$  values (Table III), the three transgenic lines showed no increase in either  $g_s$  (steady-state values or rate of induction; Fig. 7C) or  $A$  (Fig. 7D). These results indicate that alterations to stomatal density are physiologically buffered to protect

**Figure 6.** *ZjPCK<sub>pro</sub>:ZmSHR1* transgenic seedlings develop longer roots than the wild type. A to D, Two-week-old wild-type (A and B) and *ZjPCK<sub>pro</sub>:ZmSHR1* (C and D) plants grown on one-half-strength Murashige and Skoog plates, with magnified images of detached wild-type (B) and *ZjPCK<sub>pro</sub>:ZmSHR1* (D) roots. Red arrows indicate fine lateral roots, and red arrowheads point to elongated root tips. E to J, Transverse sections through the root apex (E and F), elongation zone (G and H), and maturation zone (I and J) of 2-week-old wild-type (E, G, and I) and *ZjPCK<sub>pro</sub>:ZmSHR1* (F, H, and J) roots. Bars = 1 cm (B and D) and 50  $\mu$ m (E–J).

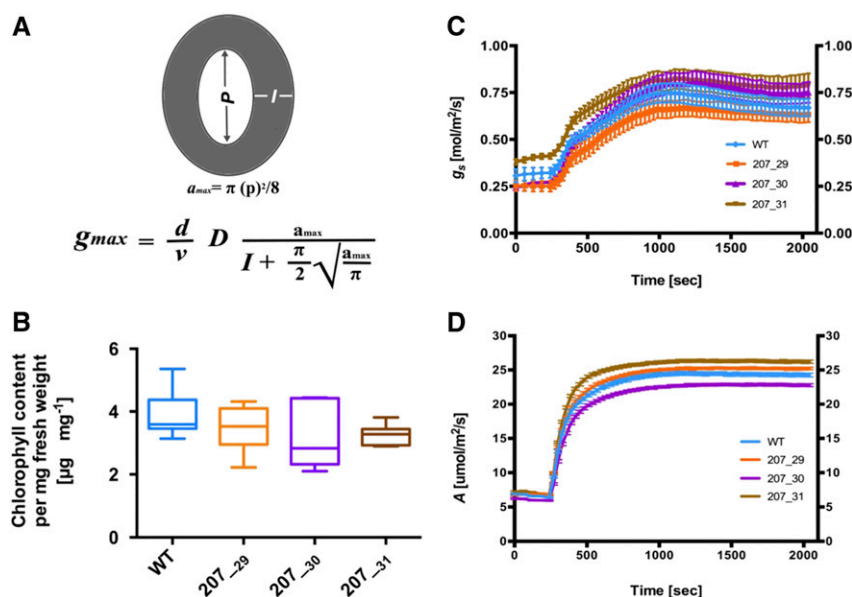


the leaf from water loss and, presumably, to coordinate capacity between veins, photosynthetic cell types, and stomata.

## DISCUSSION

The SHR pathway has been characterized primarily in *Arabidopsis*, where a single gene encodes a mobile

protein that moves outward from its expression domain in the vasculature to specify cell fate in surrounding cell layers (Helariutta et al., 2000; Nakajima et al., 2001; Gallagher et al., 2004; Levesque et al., 2006; Cui et al., 2007). Phylogenetic analysis revealed that the presence of a single *SHR* gene is typical of eudicot genomes, except where within-species genome duplications have occurred (Fig. 1). This contrasts with grasses, where duplicated *SHR1* and *SHR2* genes are present in



**Figure 7.** Increased stomatal number in *ZjPCK<sub>pro</sub>:ZmSHR1* lines does not lead to significant increases in chlorophyll, photosynthetic assimilation rates, or stomatal conductivity. **A**, Schematic representation of calculations used to determine photosynthetic capacity. Pore length ( $P$ ;  $\mu\text{m}$ ), pore depth ( $l$ ;  $\mu\text{m}$ ), and stomatal density ( $D$ ) measurements were used to calculate  $g_{max}$ , which is an integrated measurement of  $D$  and the stomatal geometry embedded in a physical gas-diffusion model.  $P$  was used to first calculate the mean maximum stomatal pore area ( $a_{max}$ ;  $\mu\text{m}^2$ ).  $g_{max}$  was then calculated from  $a_{max}$ ,  $D$ , and  $l$ . The equation constants are  $\pi$ , approximated to 3.142,  $d$  represents the diffusivity of water ( $24.6 \times 10^{-6} \text{ m}^2 \text{ s}^{-1}$  at  $25^\circ\text{C}$ ), and  $\nu$  represents the molar volume of air ( $24.4 \times 10^{-3} \text{ m}^3 \text{ mol}^{-1}$  at  $25^\circ\text{C}$  and 101.3 kPa). **B**, Box plot representing chlorophyll content in each line ( $n = 6$ ). **C**, Temporal response of net  $g_s$  to an increase in irradiance from 100 to 1,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$ . Data points are means  $\pm$  SE ( $n = 5-7$ ). **D**, Temporal response of net  $A$  to an increase in irradiance from 100 to 1,000  $\mu\text{mol m}^{-2} \text{s}^{-1}$ . Data points are means  $\pm$  SE ( $n = 5-7$ ).  $g_s$  and  $A$  were both measured under ambient  $\text{CO}_2$  levels of  $400 \mu\text{L L}^{-1}$ . Steady-state levels of  $g_s$  were achieved within 30 to 60 min from an initial PPFD of  $100 \mu\text{mol m}^{-2} \text{s}^{-1}$ . WT, Wild type.

all species examined except *B. distachyon*, where only one member of the orthogroup has been retained (Fig. 1). Transgenic Arabidopsis lines that express *OsSHR1*, *OsSHR2*, or *BdSHR* in the expression domain of *AtSHR* exhibit supernumerary cortical cell layers in the roots of a wild-type background and a phenotype in the loss-of-function *Atshr* background that indicates rescue of the mutant phenotype in addition to the supernumerary cortical cell layers (Wu et al., 2014). Together with data showing that all three of the monocot proteins are mobile in Arabidopsis (Wu et al., 2014), these observations suggest that the role of the SHR pathway in patterning cell layers around the root vasculature is conserved in Arabidopsis, rice, and *B. distachyon*, with the monocot proteins able to move farther from the vasculature than *AtSHR*.

As in roots, the SHR pathway regulates radial patterning around vascular tissue in the hypocotyl of Arabidopsis (Fukaki et al., 1998; Tasaka et al., 1999), but roles in shoot development have not been well described in monocots, with the only functional insight provided by the *Zmshr1* loss-of-function mutant in maize, in which vascular patterning in the leaf is perturbed (Slewisinski et al., 2014). Vascular perturbations in *Zmshr1* plants are subtle, however, as might be expected given the potential for redundancy with

*ZmSHR2* and *ZmSHR2h*. The transgenic rice lines reported here express *ZmSHR1* in the bundle sheath cells and mature veins of the leaf in addition to expression of the endogenous ortholog *OsSHR2* in developing leaf veins. Despite the spatial and temporal expansion of the expression domain, no defects in vascular patterning were observed, and the specification of cell types around the veins was normal (Fig. 5; Table I). As such, *ZmSHR1* expression in bundle sheath cells is not sufficient to alter vascular patterning in rice. There are three possible explanations for this observation. Most likely, by the time the transgene is expressed, the developmental time window for procambium initiation in neighboring cells may have passed. Alternatively, downstream targets and/or interaction partners of *ZmSHR1* may not be recognized in rice. Or *ZmSHR1* plays no role in leaf venation patterning per se. Although *OsSHR2* is expressed in developing veins, the timing of expression (Fig. 2) is consistent with the suggestion that the *ZmSHR1/OsSHR2* orthologs are not involved in the initiation of new veins in the leaf but, instead, play a role in specifying cell types either within or around the developing vascular bundle.

Although subepidermal patterning was normal in rice leaves expressing *ZmSHR1*, stomatal patterning was perturbed (Figs. 3 and 4; Table III). Specifically,



**Table III.** Stomatal pore measurements

Mean stomatal measurements  $D$ ,  $P$ , and  $I$  (see Fig. 7A) were used to calculate the maximum theoretical rate of  $g_{\max}$  and  $g_{c\max}$  in the wild type and three independent transgenic lines. Values represent means  $\pm$  SE of stomatal measurements taken from the middle of the penultimate leaf of the first tiller. Asterisks indicate significant differences from the wild type as determined by a one-way ANOVA test: \*\*,  $P > 0.01$ ; \*\*\*,  $P > 0.001$ ; and \*\*\*\*,  $P > 0.0001$ .

Genotype	$D$ ( $n = 12$ )	$P$ ( $n = 40-48$ )	$I$ ( $n = 40-48$ )	$g_{\max}$	$g_{c\max}$
	$\text{mm}^{-2}$	$\mu\text{m}$		$\text{mol water m}^{-2} \text{s}^{-1}$	$\text{mol CO}_2 \text{m}^{-2} \text{s}^{-1}$
Wild type	$0.29 \pm 0.1$	$13.11 \pm 0.3$	$2.53 \pm 0.10$	1.03	0.65
207 <sub>-29</sub>	$0.46 \pm 0.1$ ****	$11.22 \pm 0.2$ ****	$1.89 \pm 0.04$ ****	1.62	1.01
207 <sub>-30</sub>	$0.48 \pm 0.1$ ****	$11.00 \pm 0.2$ ****	$1.99 \pm 0.10$ ****	1.68	1.05
207 <sub>-31</sub>	$0.39 \pm 0.1$ **	$10.90 \pm 0.2$ ****	$2.11 \pm 0.10$ ***	1.28	0.80

expanding the effective expression domain of *OsSHR2*, through transgenic manipulation of the maize ortholog *ZmSHR1*, led to the formation of additional stomatal rows at a greater distance from the vein than in wild-type leaves. The normally consistent position of stomatal cell files on the flanks of veins in monocot leaves has been proposed previously to result either from an inhibitory signal transmitted from the developing vein to directly overlying epidermal cells or from an inductive signal transmitted to epidermal cells at a specific distance from the vein (Hernandez et al., 1999). The stomatal phenotype reported here supports an inductive role for the SHR pathway, either through direct action of the mobile SHR protein or indirectly via SHR target molecules (but does not preclude the possibility of an antagonistic inhibitory signal also acting directly over the veins). In an analogous mechanism to that observed in the radial patterning of roots, limited movement of the SHR protein from the site of vascular expression could specify stomatal cell files in the epidermis at the flanks of the vein. In this scenario, expansion of the SHR expression domain to bundle sheath cells surrounding the vein would lead to extra stomatal rows positioned farther away from the vein than normal.

A number of genes act downstream of SHR during root development, with SCR being the best characterized (Helariutta et al., 2000; Nakajima et al., 2001; Levesque et al., 2006). The distinct expression pattern of *OsSCR* during guard cell and subsidiary cell formation in the rice epidermis predicts a further role for SCR in stomatal formation (Kamiya et al., 2003). However, ectopic expression of *ZmSCR1* in rice does not perturb stomatal density or patterning, at least when expressed constitutively and at high levels (Wang et al., 2017). Therefore, stomatal specification must be mediated by interactions with alternative downstream targets, the most likely candidates being homologs of BdSPCH and BdICE, which have been shown to act in the earliest stages of specification in *B. distachyon* (Raissig et al., 2016). The role of *OsSCR* may be limited to later stages in the process, where asymmetric cell divisions are needed to form the stomatal complex (Kamiya et al., 2003). Clearly, further analyses are needed to determine exactly how the SHR pathway integrates with the canonical bHLH pathway that regulates stomatal differentiation.

Many aspects of the mechanisms that coordinate stomatal patterning with leaf venation and leaf gas exchange

remain unresolved, particularly in crop plants. In part, this paucity of information is because appropriate lines for phenotypic analyses have been unavailable. The transgenic rice lines generated in this work allowed an initial investigation into the interrelationship between stomatal patterning and photosynthetic capacity in a monocot. Our results showed that, despite predicted increases in photosynthetic capacity through increased  $g_{\max}$  (Table III), actual rates of  $g_s$  and photosynthesis were indistinguishable between the transgenic lines and the wild type (Fig. 7). Studies with stomatal density and distribution (*sdd1-1*) mutants in Arabidopsis also showed that stomatal density cannot be correlated directly with  $g_s$ , because increased numbers can be compensated for by decreased functional stomatal aperture (Büßis et al., 2006; Lawson and Blatt, 2014). This compensation mechanism appears to be environmentally dependent, in that no difference in  $g_s$  or assimilation rates were detected when the wild type, *sdd1* mutants with increased stomatal density, and *SDD1* overexpression lines with a 40% reduction in stomatal number were compared under light conditions of  $180 \mu\text{mol m}^{-2} \text{s}^{-1}$  PPFD (Berger and Altmann, 2000; Von Groll et al., 2002). When measured at high light, however, the increase in stomatal density in *sdd1* mutants translated into 30% higher photosynthetic rates in Arabidopsis (Schlüter et al., 2003). Future analyses of *ZjPCK<sub>pro</sub>:ZmSHR1* leaves under different light intensities may reveal that a similar compensatory mechanism operates in grasses to balance the tradeoff between carbon gain and water loss via transpiration. A second compensatory mechanism also operates in Arabidopsis, whereby altered stomatal patterning is accompanied by altered development of underlying mesophyll cells, ensuring that both gas exchange and water use efficiency are optimized across the leaf (Dow et al., 2017). Given that increased stomatal density did not lead to increased mesophyll cell density in the rice leaves examined here, it is possible that different compensatory mechanisms operate in eudicot and monocot leaves to coordinate gas exchange, carbon fixation, and water loss.

## CONCLUSION

Collectively, the work reported here has revealed that the spatial expression domain of *ZmSHR1/OsSHR2*

orthologs in the subepidermal layers of rice leaves influences the positioning of stomatal cell files in the overlying epidermis. By expanding the vascular-specific expression domain to include bundle sheath cells surrounding the vein at later stages of development, extra stomatal rows were formed. The resulting increased stomatal density did not increase mesophyll cell density in underlying tissue and did not enhance photosynthetic efficiency under the limited conditions tested, but the transgenic lines provide a vehicle for more rigorous testing of the relationship between stomatal density and photosynthetic capacity in the future.

## MATERIALS AND METHODS

### Plant Growth Conditions

Rice (*Oryza sativa* ssp *japonica*) 'Kitaake' lines were grown in soil (John Innes Compost No. 2) in a transgenic greenhouse in either Oxford, United Kingdom, or Düsseldorf, Germany. Day/night temperature was maintained at 28°C/22°C  $\pm$  3°C with a diurnal light regime of 16 h of light (supplemented to ~300  $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) and 8 h of dark. For fixation prior to in situ hybridization, seedlings were grown in water-saturated medium-grade vermiculite (Sinclair) for 17 d in a glasshouse at 28°C (day) and 23°C (night) with up to 16 h of supplemental light (100  $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) per day.

### Generation of Transformation Constructs

*ZmSHR1* (GRMZM2G132794) coding sequence was amplified from genomic DNA by PCR using Phusion High-Fidelity Polymerase (Thermo Scientific) and Gateway-compatible primers (MS5F, 5'-GGGGACAAGTTTGTACAAAAAAGCAGGCTCCGTTGGTTAGATGGACACACTGTTTAGA-3', and MS5R, 5'-GGGGACCACTTGTACAAAGAAAGCTGGGTTCATGGCCTCCACGCGCTCGT-3'). The amplified sequence was subcloned into the Gateway donor vector pDONR207 in a BP reaction. The resultant entry clones were sequenced, and the target sequences were then cloned in an LR reaction downstream of *Zoysia japonica* *PKK<sub>pro</sub>* (Nomura et al., 2005) into a destination vector modified from pVec8-GFP (Kim and Dolan, 2016) or into the binary destination vector pSC210. pSC210 vector was created by Sarah Covshoff and kindly gifted to us by Julian Hibberd (University of Cambridge). *ZmSHR1<sub>pro</sub>*: *mTurquoise*::*ZmSHR1* and *ZmSHR1<sub>pro</sub>*::*ZmSHR1* constructs were generated using the Golden Gate cloning system (Weber et al., 2011) with synthesized promoter and coding sequence modules.

### Rice Transformation

Callus induced from mature rice seeds was used for the transformation of cv Kitaake with *Agrobacterium tumefaciens* strain EHA105 carrying the construct of interest. Callus induction, transformant selection, and seedling regeneration were performed at 32°C under continuous light according to a protocol modified from Toki et al. (2006; available to download from [https://langdalelab.files.wordpress.com/2015/07/kitaake\\_transformation\\_2015.pdf](https://langdalelab.files.wordpress.com/2015/07/kitaake_transformation_2015.pdf)). Hygromycin-resistant T0 seedlings confirmed positive for transgene presence by PCR screening (see below) were transplanted into soil in 0.73-L pots. T1 lines with single transgene insertion events were selected on the basis of a 3:1 segregation pattern on hygromycin-containing selective medium. Homozygous plants were identified by quantification of transgene copy number in hygromycin-resistant T1 plants, using the gCount service of iDNA genetics, and propagated into the T2 generation. Homozygous T2 lines 207<sub>29H</sub>, 207<sub>30K</sub>, and 207<sub>31J</sub>, which represented a single transgene insertion event, were used for all phenotypic analyses except those shown in Supplemental Figure S1.

### DNA and RNA Extraction

Genomic DNA was isolated from cv Kitaake lines using a modified 2% (v/v) cetyl-trimethyl-ammonium bromide method (Murray and Thompson, 1980), and rice leaf RNA was extracted using TRIzol reagent.

### Genomic PCR Screening

Regenerated T0 plants and T1 seedlings (2–3 weeks after germination) were subjected to genomic PCR using primers specific to the cloning vector pSC210 (pSC210-F, 5'-CTGCTGCTGCTGCTCTCTC-3', located in *ZjPKK<sub>pro</sub>*, and pSC1/2/310R, 5'-AAGACCGGCAACAGGATTC-3', located in the *nos* terminator). PCR amplification was performed in a total reaction volume of 20  $\mu\text{L}$  containing 9.2  $\mu\text{L}$  of 2xGoTaq mix (Promega) and 1 M betaine. PCR conditions were as follows: 95°C for 5 min; 35 cycles of 95°C for 30 s, 58°C for 40 s, and 72°C for 2.5 min; and 72°C for 5 min. All PCR products were detected by agarose gel electrophoresis using standard protocols (Sambrook, 1982).

### Real-Time Quantitative Reverse Transcription-PCR

Gene expression analysis was carried out using RNA extracted from fully expanded fourth leaf tissue. Total RNA was treated with TURBO DNase (Thermo Scientific), and cDNA was synthesized using SuperScript III reverse transcriptase (Invitrogen) according to the manufacturer's instructions. Specific primers were used to amplify genes of interest (qRT5F, 5'-ATGAAGGTGTTACCGAGGG-3', and qRT5R, 5'-TCGTTGCTCGCCTTGGG-3') or an endogenous *UBIQUITIN* gene (Os03g13170; pW65F, 5'-CCAGGACAAGATGATCTGCC-3', and pW65R, 5'-AAGAAGCTGAAGCATCCAGC-3'). Reactions were carried out using KAPA SYBR FAST Master Mix (2x) universal (KAPA Biosystems) in a 20- $\mu\text{L}$  PCR volume. Transcript amplification and detection was performed on the 7500 Fast Real-Time PCR System (Applied Biosystems) using the following thermal profile: 95°C for 5 min; followed by 45 cycles of 95°C for 15 s, 60°C for 30 s, and 72°C for 30 s. Three technical replicates were carried out per sample, and transcript abundance was normalized to the endogenous rice *UBIQUITIN* gene. Relative expression fold changes were calculated using the Pfaffl method (Pfaffl, 2001).

### In Situ Hybridization

Shoot apices were harvested on ice, fixed in 4% (w/v) paraformaldehyde overnight, and then dehydrated using a graded ethanol series before storage in 70% (v/v) ethanol at 4°C. Following wax embedding in Paraplast Plus (Sigma-Aldrich), 8- $\mu\text{m}$ -thick mounted sections were pretreated as described by Javelle et al. (2011) with the following modifications: the 30-s 10% (v/v) ethanol and 1-min water washes were replaced with a 2-min 0.85% (w/v) sodium chloride wash, and the proteinase K incubation time was increased to 30 min. The slides were hybridized to a 141-bp digoxigenin-labeled RNA probe specific to *OsSHR2*. The probe sequence was PCR amplified using primers appended with T3 and T7 RNA polymerase-binding sites (5'-AATTAACCTCACTAAAGGGAGATCCCTTTGTCATCCAT-3' and 5'-TAATACGACTACTATAGGCTGGGTACCACCATACAGCA-3') and then labeled according to Langdale (1994), but with the following reaction mix: 1x RNA polymerase buffer, 0.5 mM ATP, 0.5 mM GTP, 0.5 mM CTP, 0.1 mM digoxigenin-UTP, 0.8 units  $\mu\text{L}^{-1}$  RNA polymerase (all Thermo Scientific), and 1.6 units  $\mu\text{L}^{-1}$  RNaseOUT (Invitrogen). The reaction was incubated for 1.5 h at 37°C and terminated by 10 min of incubation at 37°C with 3 volumes of DNase solution (10 mM Tris-HCl, pH 7.5, 10 mM magnesium chloride, 50 mM sodium chloride, 2.5  $\mu\text{g} \mu\text{L}^{-1}$  tRNA, and 0.02 units  $\mu\text{L}^{-1}$  DNase I). Hybridization was carried out at 55°C essentially according to Langdale (1994) with posthybridization washes and visualization as described by Javelle et al. (2011) using 0.05x SSC buffer. After dehydration through ethanol and counterstaining for 5 min with 0.1% (w/v) Calcofluor White (Fluorescent Brightener 28; Sigma-Aldrich), slides were mounted in Entellan (Merck Millipore) and imaged under bright-field and UV light using a Leica DMR8 microscope, QImaging MicroPublisher (QImaging) camera, and Image-Pro Insight (MediaCybernetics) software.

### Histological Analysis of Root Tissue

Roots were fixed in 3:1 ethanol:acetic acid for 30 min, dehydrated in an ethanol series, and embedded in Araldite epoxy resin (prepared from four single embedding components of Durcupan ACMA, B, C, D; Sigma-Aldrich). Transverse sections of 10  $\mu\text{m}$  were cut with a glass knife microtome, stained with a drop of Toluidine staining solution (1% (w/v) Toluidine Blue in 1% (w/v) borate) for 20 s, and rinsed with distilled water. Dried slides were then mounted with DePeX medium (Serva) for microscopic analysis.

### Scanning Electron Microscopy

Negative imprints of the abaxial leaf surface were taken from the middle of fully expanded adult leaves (11–12 mm diameter) using ESPE Express

two-component ultra-light body dental wax (3M). Silicone imprints were dried for 5 min, covered with epoxy resin HP E25KL (HP Textiles), and dried overnight at room temperature to produce positive imprints for scanning electron microscopy. Resin imprints were coated with gold particles in an argon sputter coater and visualized with a Zeiss Leo 1430 VP scanning electron microscope.

## Phenotypic Analysis of Leaf Anatomy

Anatomical measurements were carried out on the expanded sixth leaf of the first tiller. Leaf width and length were first measured, and then fresh segments were hand cut from the middle of the leaf blade. Sections were exposed to UV light and then viewed and photographed with a Leica DMRB microscope or a Zeiss Axio Imager M2m. The cell number between a pair of intermediate veins positioned between the second and third lateral veins from the leaf margin, as well as interveinal distances of three pairs of veins and cell size of six bundle sheath cells, were recorded for each sample. Two to five leaf samples were quantified for each line. Least-squares regression lines of leaf width versus vein number were plotted for the cv Kitaake wild type and *ZjPCK<sub>pro</sub>:ZmSHR1* data sets using GraphPad Prism 6.

## Quantification of Stomatal Phenotypes

The leaf clearing method used was adapted from Dow et al. (2014). Penultimate leaves were fixed and cleared with 7:1 ethanol:acetic acid solution overnight, softened for 30 min in 1 M potassium hydroxide, rinsed twice with water, and then mounted on microscope slides using a chloral hydrate-glycerol mounting solution (40 g of chloral hydrate in 16 g of glycerol). The abaxial epidermis was analyzed by differential interference contrast microscopy with Zeiss Axio ImagerM2m. Stomatal density ( $\text{mm}^{-2}$ ) was counted between two lateral veins in three different areas of four individual leaves per line. Stomatal sizes defined as stomatal height multiplied by stomatal width ( $\mu\text{m}^2$ ) were measured and averaged from 40 to 48 stomata counted from four to five leaves per individual line using ImageJ software (Abràmoff et al., 2004), available at <https://imagej.nih.gov/ij/>.

## Leaf Drying and Weight-Gain Experiments

Leaf discs were punched from freshly detached leaves of equivalent size and then dried at room temperature for 3 d. Dry weights were determined for three pooled leaf discs per leaf, from six leaves in total. For weight-gain experiments, four leaf discs were punched from six freshly detached leaves and weighed. The discs were then immediately vacuum infiltrated with distilled water, blotted dry on filter paper, and then weighed once more to determine weight gain.

## Quantification of Chlorophyll

Chlorophyll was extracted from freshly detached leaves of equivalent sizes as described previously (Lichtenthaler and Wellburn, 1983; Porra et al., 1989). Three leaf discs (10 mm each in diameter) were punched from the middle of mature fully expanded leaves and ground in liquid nitrogen. Chlorophyll was extracted by incubating the ground powder for 20 min in 80% (v/v) acetone in 50 mM Tris-HCl, pH 7.8. The absorption properties of 1 mL of extract were measured with an Eppendorf Biospectrometer, and chlorophyll content was calculated as follows: chlorophyll *a* ( $\mu\text{g mL}^{-1}$ ) =  $12.21 (A_{663}) - 2.81 (A_{646})$ ; chlorophyll *b* ( $\mu\text{g mL}^{-1}$ ) =  $20.13 (A_{646}) - 5.03 (A_{663})$ ; total chlorophyll content =  $17.76 (A_{646}) + 7.43 (A_{663})$ .

## Leaf Gas-Exchange Measurements

*A* and *g<sub>s</sub>* were measured on fully expanded leaves of 5- to 6-week-old plants using infrared gas analysis (Li-Cor 6400XT, Portable Photosynthesis System; LICOR Biosciences). Leaves were first equilibrated at a PPFD of  $100 \mu\text{mol m}^{-2} \text{s}^{-1}$  until both *A* and *g<sub>s</sub>* reached steady state, this being defined as a less than 10% change in rate during a 10-min period (after 30–60 min). Once steady state was reached, *A* and *g<sub>s</sub>* were recorded for an initial period of 5 min at  $100 \mu\text{mol m}^{-2} \text{s}^{-1}$  every 60 s. PPFD was then increased to  $1,000 \mu\text{mol m}^{-2} \text{s}^{-1}$  for 30 min, while *A* and *g<sub>s</sub>* were measured every 30 s. The leaf cuvette was maintained at  $400 \mu\text{mol mol}^{-1} \text{CO}_2$  concentration, leaf temperature of  $27^\circ\text{C} \pm 2^\circ\text{C}$ , and a vapor pressure deficit (VPD) of  $1.3 \pm 0.3 \text{ kPa}$ . All measurements were completed before 3 PM to avoid any unwanted diurnal or circadian effects on photosynthesis.

## Protein Alignment and Phylogenetic Analysis

BLASTP (protein) and BLASTN (nucleotide) search algorithms were used on proteomes and genomes from 13 angiosperm and one bryophyte species obtained from the Phytozome version 10 database (Goodstein et al., 2012). Protein and genomic sequences that were significantly similar (threshold,  $E < 1 \times 10^{-3}$ ) to the amino acid and transcript sequences from *ZmSHR1* were identified. The amino acid sequences were aligned using MergeAlign software (Collingridge and Kelly, 2012; Supplemental Data Set S1). A maximum-likelihood method was used to infer the phylogenetic tree using RAxML software (Stamatakis, 2014); 100 rapid bootstrap analysis runs were used to produce a best-scoring maximum-likelihood tree (the best-fitting protein substitution model used for inference was selected automatically by the software). The phylogenetic tree was visualized using Interactive Tree of Life software (Letunic and Bork, 2016).

## Supplemental Data

The following supplemental materials are available.

**Supplemental Figure S1.** Three independent T0 lines exhibit extra stomatal rows when *ZmSHR1* is ectopically expressed.

**Supplemental Figure S2.** Light saturation curve of 207<sub>-30</sub> versus the wild type.

**Supplemental Data Set S1.** Multiple sequence alignment of SHR-like proteins.

**Supplemental Data Set S2.** Data used for regression analysis of vein number versus leaf width.

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