

1 **Title:** Climatic controls of decomposition drive the global biogeography of forest tree
2 symbioses

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431 **Manuscript**

432 **The identity of the dominant microbial symbionts in a forest determines the ability**
433 **of trees to access limiting nutrients from atmospheric or soil pools^{1,2}, sequester**
434 **carbon^{3,4} and withstand the impacts of climate change¹⁻⁶. Characterizing the global**
435 **distribution of symbioses, and identifying the factors that control it, are thus integral to**
436 **understanding present and future forest ecosystem functioning. Here we generate the first**
437 **spatially explicit global map of forest symbiotic status using a database of over 1.1 million**
438 **forest inventory plots with over 28,000 tree species. Our analyses indicate that climatic**
439 **variables, and in particular climatically-controlled variation in decomposition rate, are the**
440 **primary drivers of the global distribution of major symbioses. We estimate that**
441 **ectomycorrhizal (EM) trees, which represent only 2% of all plant species⁷, constitute**

442 **approximately 60% of tree stems on Earth. EM symbiosis dominates forests where**
443 **seasonally cold and dry climates inhibit decomposition, and are the predominant symbiosis**
444 **at high latitudes and elevation. In contrast, arbuscular mycorrhizal (AM) trees dominate**
445 **aseasonally warm tropical forests and occur with EM trees in temperate biomes where**
446 **seasonally warm-and-wet climates enhance decomposition. Continental transitions between**
447 **AM and EM dominated forests occur relatively abruptly along climate driven**
448 **decomposition gradients, which is likely caused by positive plant-microbe**
449 **feedbacks. Symbiotic N-fixers, which are insensitive to climatic controls on decomposition**
450 **compared with mycorrhizal fungi, are most abundant in arid biomes with alkaline soils**
451 **and high maximum temperatures. The climatically driven global symbiosis gradient we**
452 **document represents the first spatially-explicit, quantitative understanding of microbial**
453 **symbioses at the global scale and demonstrates the critical role of microbial mutualisms in**
454 **shaping the distribution of plant species.**

455 Microbial symbionts strongly influence the functioning of forest ecosystems. They
456 exploit inorganic, organic² and/or atmospheric forms of nutrients that enable plant growth¹,
457 determine how trees respond to elevated CO₂⁶, regulate the respiratory activity of soil
458 microbes^{3,8}, and affect plant species diversity by altering the strength of conspecific negative
459 density dependence⁹. Despite growing recognition of the importance of root symbioses for forest
460 functioning^{1,6,10} and the potential to integrate symbiotic status into Earth system models that
461 predict functional changes to the terrestrial biosphere¹⁰, we lack spatially-explicit, quantitative
462 maps of the different root symbioses at the global scale. Generating these quantitative maps of
463 tree symbiotic states would link the biogeography of functional traits of belowground microbial

464 symbionts with their 3.1 trillion host trees¹¹, spread across Earth's forests, woodlands, and
465 savannas.

466 The dominant guilds of tree root symbionts, arbuscular mycorrhizal (AM) fungi,
467 ectomycorrhizal (EM) fungi, ericoid mycorrhizal (ErM) fungi, and nitrogen (N)-fixing bacteria
468 (N-fixer) are all based on the exchange of plant photosynthate for limiting macronutrients. The
469 AM symbiosis evolved nearly 500 million years ago, with EM, ErM and N-fixer plant taxa
470 evolving multiple times from an AM basal state. Plants that form the AM symbiosis comprise
471 nearly 80% of all terrestrial plant species, and principally rely on AM fungi for enhancing
472 mineral phosphorus (P) uptake¹². In contrast to AM fungi, EM fungi evolved from multiple
473 lineages of saprotrophic ancestors, and as a result some EM fungi are more capable of mobilizing
474 organic sources of soil nutrients (particularly nitrogen)². Association with EM fungi, but not AM
475 fungi, has been shown to allow trees to accelerate photosynthesis in response to increased
476 atmospheric CO₂ when soil nitrogen (N) is limiting⁶ and to inhibit soil respiration by decomposer
477 microbes^{3,8}. Because increased plant photosynthesis and decreased soil respiration both reduce
478 atmospheric CO₂ concentrations, the EM symbiosis is associated with buffering the Earth's
479 climate against anthropogenic changes.

480 In contrast to mycorrhizal fungi, which extract nutrients from the soil, symbiotic N-fixers
481 (Rhizobia and Actinobacteria) convert atmospheric N₂ to plant-usable forms. Symbiotic N-fixers
482 are responsible for a large fraction of biological soil-N inputs, which can increase N-availability
483 in forests where they are locally abundant¹³. Both N-fixing bacteria and EM fungi often demand
484 more plant photosynthate than does the AM symbiosis^{12,14,15}. Because tree growth and
485 reproduction are limited by access to inorganic, organic and atmospheric sources of N, the
486 distribution of root symbioses is likely to reflect both environmental conditions that maximize

487 the cost-benefit ratio of symbiotic exchange as well as physiological constraints on different
488 symbionts.

489 In one of the earliest efforts to understand the functional biogeography of plant root
490 symbioses, Sir David Read¹⁶ categorically classified biomes by their perceived dominant
491 mycorrhizal type and hypothesized that seasonal climates favor hosts associating with EM fungi
492 due to their ability to compete directly for organic N. In contrast, it has been proposed that
493 sensitivity to low temperatures has prevented N-fixers from dominating outside the tropics,
494 despite the potential for N-fixation to alleviate N-limitation in boreal forests^{15,17}. However,
495 global scale tests of these proposed biogeographic patterns and their climate drivers are lacking.
496 To address this research gap, we compiled the first global ground-sourced survey database to
497 reveal numerical abundances of each symbiosis across the globe, which is essential for
498 identifying the potential mechanisms underlying transitions in forest symbiotic state along
499 climatic gradients^{18,19}.

500 We determined the abundance of tree symbioses using GFBi, an extension from the plot-
501 based Global Forest Biodiversity (GFB) database, which contains over 1.1 million forest
502 inventory plots of individual-based measurement records from which we derive abundance
503 information for entire tree communities (Figure 1). Using published literature on the
504 evolutionary histories of mycorrhizal and N-fixer symbioses, we assigned plant species from the
505 GFBi to one of 5 symbiotic guilds: AM, EM, ErM, N-fixer, and non- or weakly-mycorrhizal
506 (NM). We then used the random forest algorithm with K-fold cross validation to determine the
507 importance and influence of variables related to climate, soil chemistry, vegetation, and
508 topography on the relative abundance of each tree-symbiotic guild (Figure 2). Because
509 decomposition is the dominant process by which soil nutrients become available to plants, we

510 calculated annual and quarterly decomposition coefficients according to the Yasso07 model²⁰,
511 which describes how temperature and precipitation gradients influence mass-loss rates of
512 different chemical pools of leaf litter (with parameters fit using a previous global study of leaf
513 decomposition, Figures 3, S5). Finally, we projected our predictive models across the globe over
514 the extent global biomes that fell within the multivariate distribution of our model training data
515 (Figures 4, S14-15, see Methods for full description).

516 Our analysis shows that the three most numerically abundant tree symbiotic guilds each
517 have reliable environmental signatures, with the four most important predictors accounting for
518 81, 79, and 52% of the total variability in EM, AM, and N-fixer relative basal area, respectively.
519 Models for ErM and NM lack strong predictive power given the relative rarity of these symbiotic
520 states amongst trees, although the raw data do identify some local abundance hotspots for ErM
521 (Figure S1). As a result, we focus the remainder of results and discussion on the three major tree
522 symbiotic states (EM, AM, N-fixer). Despite the fact that data from N. and S. America constitute
523 65% of the training data (at the 1 by 1 degree grid scale), our models accurately predict the
524 proportional abundances of the three major symbioses across all major geographic regions
525 (Figure S10). The high performance of our models, which is robust to both K-fold cross-
526 validation and rarefying samples so that all continents are represented with equal depth (Figures
527 S11-12), suggest that regional variations in climate (including indirect effects on decomposition)
528 and soil pH (for N-fixers) are the primary factors influencing the relative dominance of each
529 guild at the global scale (geographic origin only explained ~2-5% of the variability in residual
530 relative abundance) (Table S8, Figure S10).

531 Whereas a recent global analysis of root traits concluded that plant evolution has favored
532 reduced dependence on mycorrhizal fungi²¹, we find that trees associating with the relatively

533 more C-demanding and recently-derived EM fungi^{12,14} represent the dominant tree-symbiosis.
534 By taking the average proportion of EM trees, weighted by spatially-explicit global predictions
535 for tree stem density¹¹, we estimate that approximately 60% of trees on earth are EM, despite the
536 fact that only 2% of plant species associate with EM fungi (vs. 80% associating with AM fungi)⁷.
537 Outside of the tropics, the estimate for EM relative abundance increases to approximately 80%
538 of trees.

539 Turnover among the major symbiotic guilds results in a tri-modal latitudinal abundance
540 gradient, with the proportion of EM trees increasing (and AM trees decreasing) with distance
541 from the equator, while the upper-quantiles of N-fixing trees reach peak abundance in the arid
542 zone around 30 degrees (Figure 3A, Figure 4). These trends are driven by abrupt transitional
543 regions along continental climatic gradients (Figure 2), which skew the distribution of symbioses
544 among biomes (Figure 3A) and drive strong patterns across geographic and topographic features
545 that influence climate. Moving north or south from the equator, the first transitional zone
546 separates warm (aseasonal), AM-dominated, tropical broadleaf forests (>75% median basal area,
547 vs. 8% for EM trees) from the rest of the EM-dominated world forest system (Figure 2AB;
548 Figure 3A). The transition zone occurs across the globe around 25 degrees N and S latitude, just
549 beyond the dry tropical broadleaf forests (with 25% EM tree basal area; Figure 3A), where
550 average monthly temperature variation reaches 3-5°C (temperature seasonality, Figure 2AB).

551 Moving further N or S, the second transitional climate zone separates regions where
552 decomposition coefficients during the warmest quarter of the year are less than 2 (see Figure 3B
553 for the associated temperature and precipitation ranges). In N. America and China, this transition
554 zone occurs around 50 degrees N, separating the mixed AM / EM temperate forests from their
555 neighboring EM dominated boreal forests (75 vs 100% EM tree basal area, respectively; Figure

556 3A). This transitional decomposition zone bypasses W. Europe, which has temperature
557 seasonality $> 5^{\circ}\text{C}$, but lacks sufficiently wet summers to accelerate decomposition coefficients
558 beyond values associated with mixed AM/EM forests. The latitudinal transitions in symbiotic
559 state observed among biomes are mirrored by within-biome transitions along elevation gradients.
560 For example, in tropical Mexico, warm and wet quarter decomposition coefficients < 2 occur
561 along the slopes of the Sierra Madre, where mixed AM-exclusive and N-fixer woodlands in arid
562 climates transition to EM dominated tropical coniferous forests (75% basal area, Figure 3A,
563 Figure 4ABC, Figure S16-18). The southern hemisphere, which lacks the landmass to support
564 extensive boreal forests, experiences a similar latitudinal transition in decomposition rates along
565 the ecotone separating its tropical and temperate biomes, around 28 degrees S.

566 The abrupt transitions that we detected between forest symbiotic states along
567 environmental gradients suggest that positive feedbacks may exist between climatic and
568 biological controls of decomposition^{10,20}. In contrast to AM fungi, some EM fungi can use
569 oxidative enzymes to mineralize organic nutrients from leaf litter, converting nutrients to plant-
570 usable forms^{2,5}. Relative to AM trees, the leaf litter of EM trees is also chemically more resistant
571 to decomposition, with higher C:N ratios and higher concentrations of decomposition-inhibiting
572 secondary compounds¹⁰. Thus, EM leaf litter can exacerbate climatic barriers to decomposition,
573 promoting conditions where EM fungi have superior nutrient-acquiring abilities to AM-fungi^{5,10}.
574 A recent game theoretical model has shown that positive plant-soil-nutrient feedbacks can lead to
575 local bistability in mycorrhizal symbiosis²². Such positive-feedbacks are also known to cause
576 abrupt ecosystem transitions along smooth environmental gradients between woodlands and
577 grasses: trees suppress fires, which promotes seedling recruitment, while grass fuels fires, which
578 kill tree seedlings²³. The existence of abrupt transitions also suggests that forests in transitional

579 regions along decomposition gradients should be susceptible to drastic turnover in symbiotic
580 state with future environmental changes²³.

581 To illustrate the sensitivity of global patterns of tree symbiosis to climate change, we use
582 the relationships we developed for current climate to project potential changes in forest
583 symbiotic status in the future. Relative to our global predictions using the most recent climate
584 data, model predictions using the projected climates for 2070 suggest the abundance of EM trees
585 will decline by as much as 10% (using a relative concentration pathway of 8.5 W/m²; Figure
586 S24). Due to their position along decomposition gradients relative to the abrupt shift from EM to
587 AM forests (Figure 2AB), our models predict the largest declines in EM abundance will occur
588 along the boreal-temperate ecotone, although this model does not estimate the time lags between
589 climate change and forest community responses. The predicted decline in EM trees corroborates
590 the results of common garden transfer and simulated warming experiments, which demonstrate
591 that some important EM hosts will decline at the boreal-temperate ecotone in altered climates²⁴.

592 The change in dominant nutrient exchange symbioses along climate gradients highlights
593 the interconnection between atmospheric and soil compartments of the biosphere. The transition
594 from AM to EM dominance corresponds with a shift from P to N limitation of plant growth with
595 increasing latitude^{25,26}. Including published global projections of total soil N or P, microbial N,
596 or soil P fractions (labile, occluded, organic, and apatite) did not increase the amount of variation
597 explained by the model or alter the variables identified as most important, and thus were dropped
598 from our analysis. However, our finding that climatic controls of decomposition best predict the
599 dominant mycorrhizal associations mechanistically links symbiont physiology with climatic
600 controls of soil nutrient release from leaf litter. These findings are consistent with Read's
601 hypothesis¹⁶ that slow decomposition at high latitudes favors EM fungi due to their increased

602 capacity to liberate organic nutrients². Thus, while more experiments are necessary to understand
603 the specific mechanism by which nutrient competition favors dominance of AM or EM
604 symbioses¹⁸, we propose that the latitudinal and elevational transitions from AM to EM
605 dominated forests be called Read's Rule.

606 While our analyses focus on prediction at large spatial scales appropriate to the available
607 data, our findings with respect to Read's Rule also provide insight into how soil factors structure
608 the fine-scale distributions of tree symbioses within our grid cells. For example, while at a coarse
609 scale we find that EM trees are relatively rare in many wet tropical forests, individual tropical
610 sites in our raw data span the full range from 0 – 100 % EM basal area. In much of the wet
611 tropics, these EM dominated sites exist as outliers within a matrix of predominantly AM trees. In
612 an apparent exception that proves Read's Rule, in aseasonal warm neotropical climates, which
613 accelerate leaf-decomposition and promote regional AM dominance (Figure 3), EM dominated
614 tree stands can develop in sites where poor soils and recalcitrant litter slow decomposition and N
615 mineralization^{18,27}. Landscape-scale variation in the relative abundance of symbiotic states also
616 changes along climate gradients, with variability highest in xeric and temperate biomes (Figure
617 S3-4), suggesting that the potential of local nutrient variability to favor particular symbioses is
618 contingent on climate.

619 Whereas EM trees are associated with ecosystems where plant growth is thought to be
620 primarily N-limited, N-fixer trees are not. Our results highlight the global extent of the “N-
621 cycling paradox,” wherein some metrics suggest that N-limitation is greater in the temperate
622 zone^{25,26}, yet N-fixing trees are relatively more common in the tropics^{15,28} (Figure 3A). We find
623 that N-fixers, which we estimate represent 7% of all trees, dominate forests with annual max
624 temperatures >35°C and alkaline soils (particularly in North America and Africa, Figure 2C).

625 They have the highest relative abundance in xeric shrublands (24%), tropical savannas (21%),
626 and dry broadleaf forest biomes (20%), but are nearly absent from boreal forests (<1%) (Figure
627 3A, Figure 4). The decline in N-fixer tree abundance we observed with increasing latitude is also
628 associated with a previously documented latitudinal shift in the identity of N-fixing microbes,
629 from facultative N-fixing rhizobial bacteria in tropical forests to obligate N-fixing actinorhizal
630 bacteria in temperate forests²⁸. Our data are not capable of fully disentangling the several
631 hypotheses that have been proposed to reconcile the N-cycling paradox¹⁵. However, our results
632 are consistent with the model prediction¹⁷ and regional empirical evidence^{19,29,30} that N-fixing
633 trees are particularly important in arid biomes. Based primarily on the observed positive,
634 nonlinear association of N-fixer relative abundance with the mean temperature of the hottest
635 month (Figure 2C), our models predict a two-fold increase in N-fixer relative abundance when
636 transitioning from humid to dry tropical forest biomes (Figure 3A).

637 Although soil microbes are a dominant component of forests, both in terms of diversity
638 and ecosystem functioning^{5,6,10}, identifying global-scale microbial biogeographic patterns
639 remains an ongoing research priority. Our analyses confirm that Read's Rule, which is one of the
640 first proposed biogeographic rules specific to microbial symbioses, successfully describes global
641 transitions between mycorrhizal guilds. More generally, climate driven turnover among the
642 major plant-microbe symbioses represents a fundamental biological pattern in the Earth system,
643 as forests transition from low-latitude arbuscular mycorrhizal, to N-fixer, to high-latitude
644 ectomycorrhizal ecosystems. The predictions of our model (which we make available as a global
645 raster layer) can now be used to represent these critical ecosystem variations in global
646 biogeochemical models used to predict climate-biogeochemical feedbacks within and between
647 trees, soils, and the atmosphere. Additionally, the layer containing the proportion of N-fixing

648 trees can be used to map potential symbiotic N-fixation, which links together atmospheric pools
649 of C and N. Future work can extend our findings to incorporate multiple plant growth forms and
650 non-forested biomes, where similar patterns likely exist, to generate a complete global
651 perspective. Our predictive maps leverage the most comprehensive global forest dataset to
652 generate the first quantitative global map of forest tree symbioses, demonstrating how nutritional
653 mutualisms are coupled with the global distribution of plant communities.

654 **References**

- 655 1 Batterman, S. A. *et al.* Key role of symbiotic dinitrogen fixation in tropical forest
656 secondary succession. *Nature* **502**, 224-227, doi:10.1038/nature12525 (2013).
- 657 2 Shah, F. *et al.* Ectomycorrhizal fungi decompose soil organic matter using oxidative
658 mechanisms adapted from saprotrophic ancestors. *New Phytol* **209**, 1705-1719,
659 doi:10.1111/nph.13722 (2016).
- 660 3 Averill, C., Turner, B. L. & Finzi, A. C. Mycorrhiza-mediated competition between
661 plants and decomposers drives soil carbon storage. *Nature* **505**, 543-+,
662 doi:10.1038/nature12901 (2014).
- 663 4 Clemmensen, K. E. *et al.* Roots and associated fungi drive long-term carbon
664 sequestration in boreal forest. *Science* **339**, 1615-1618, doi:10.1126/science.1231923
665 (2013).
- 666 5 Cheeke, T. E. *et al.* Dominant mycorrhizal association of trees alters carbon and nutrient
667 cycling by selecting for microbial groups with distinct enzyme function. *New Phytol.*
668 **214**, 432-442, doi:10.1111/nph.14343 (2017).
- 669 6 Terrer, C., Vicca, S., Hungate, B. A., Phillips, R. P. & Prentice, I. C. Mycorrhizal
670 association as a primary control of the CO₂ fertilization effect. *Science* **353**, 72-74,
671 doi:10.1126/science.aaf4610 (2016).
- 672 7 Brundrett, M. C. in *Biogeography of Mycorrhizal Symbiosis* 533-556 (Springer, 2017).
- 673 8 Averill, C. & Hawkes, C. V. Ectomycorrhizal fungi slow soil carbon cycling. *Ecol Lett*
674 **19**, 937-947, doi:10.1111/ele.12631 (2016).
- 675 9 Bennett, J. A. *et al.* Plant-soil feedbacks and mycorrhizal type influence temperate forest
676 population dynamics. *Science* **355**, 181-184 (2017).
- 677 10 Phillips, R. P., Brzostek, E. & Midgley, M. G. The mycorrhizal-associated nutrient
678 economy: a new framework for predicting carbon-nutrient couplings in temperate forests.
679 *New Phytol.* **199**, 41-51, doi:10.1111/nph.12221 (2013).
- 680 11 Crowther, T. W. *et al.* Mapping tree density at a global scale. *Nature* **525**, 201 (2015).
- 681 12 Heijden, M. G., Martin, F. M., Selosse, M. A. & Sanders, I. R. Mycorrhizal ecology and
682 evolution: the past, the present, and the future. *New Phytol.* **205**, 1406-1423 (2015).
- 683 13 Binkley, D., Sollins, P., Bell, R., Sachs, D. & Myrold, D. Biogeochemistry of adjacent
684 conifer and alder-conifer stands. *Ecology* **73**, 2022-2033 (1992).

685 14 Leake, J. *et al.* Networks of power and influence: the role of mycorrhizal mycelium in
686 controlling plant communities and agroecosystem functioning. *Canadian Journal of*
687 *Botany* **82**, 1016-1045 (2004).

688 15 Hedin, L. O., Brookshire, E. N. J., Menge, D. N. L. & Barron, A. R. in *Annual Review of*
689 *Ecology Evolution and Systematics* Vol. 40 *Annual Review of Ecology Evolution and*
690 *Systematics* 613-635 (Annual Reviews, 2009).

691 16 Read, D. J. Mycorrhizas in Ecosystems. *Experientia* **47**, 376-391, doi:Doi
692 10.1007/Bf01972080 (1991).

693 17 Houlton, B. Z., Wang, Y.-P., Vitousek, P. M. & Field, C. B. A unifying framework for
694 dinitrogen fixation in the terrestrial biosphere. *Nature* **454**, 327 (2008).

695 18 Peay, K. G. The mutualistic niche: mycorrhizal symbiosis and community dynamics.
696 *Annual Review of Ecology, Evolution, and Systematics* **47**, 143-164 (2016).

697 19 Pellegrini, A. F., Staver, A. C., Hedin, L. O., Charles-Dominique, T. & Tourgee, A.
698 Aridity, not fire, favors nitrogen-fixing plants across tropical savanna and forest biomes.
699 *Ecology* **97**, 2177-2183 (2016).

700 20 Tuomi, M. *et al.* Leaf litter decomposition—estimates of global variability based on
701 Yasso07 model. *Ecological Modelling* **220**, 3362-3371 (2009).

702 21 Ma, Z. *et al.* Evolutionary history resolves global organization of root functional traits.
703 *Nature* (2018).

704 22 Lu, M. & Hedin, L. O. Global plant-symbiont organization and emergence of
705 biogeochemical cycles resolved by evolution-based trait modelling. *Nature ecology &*
706 *evolution*, 1 (2019).

707 23 Scheffer, M., Carpenter, S., Foley, J. A., Folke, C. & Walker, B. Catastrophic shifts in
708 ecosystems. *Nature* **413**, 591 (2001).

709 24 Reich, P. B. *et al.* Geographic range predicts photosynthetic and growth response to
710 warming in co-occurring tree species. *Nature Climate Change* **5**, 148 (2015).

711 25 McGroddy, M. E., Daufresne, T. & Hedin, L. O. Scaling of C: N: P stoichiometry in
712 forests worldwide: Implications of terrestrial redfield-type ratios. *Ecology* **85**, 2390-
713 2401 (2004).

714 26 Reich, P. B. & Oleksyn, J. Global patterns of plant leaf N and P in relation to temperature
715 and latitude. *Proceedings of the National Academy of Sciences of the United States of*
716 *America* **101**, 11001-11006 (2004).

717 27 Corrales, A., Mangan, S. A., Turner, B. L. & Dalling, J. W. An ectomycorrhizal nitrogen
718 economy facilitates monodominance in a neotropical forest. *Ecol Lett* **19**, 383-392,
719 doi:10.1111/ele.12570 (2016).

720 28 Menge, D. N., Lichstein, J. W. & Ángeles-Pérez, G. Nitrogen fixation strategies can
721 explain the latitudinal shift in nitrogen-fixing tree abundance. *Ecology* **95**, 2236-2245
722 (2014).

723 29 Liao, W., Menge, D. N., Lichstein, J. W. & Ángeles-Pérez, G. Global climate change
724 will increase the abundance of symbiotic nitrogen-fixing trees in much of North
725 America. *Global Change Biol* (2017).

726 30 Gei, M. *et al.* Legume abundance along successional and rainfall gradients in Neotropical
727 forests. *Nature ecology & evolution*, 1 (2018).

728

729 **Figure Legends**

730 **Figure 1. The global distribution of GFBi training data. The global map has n=2,768 grid**
731 **cells at a 1 x 1 degree latitude/longitude resolution. Cells are colored in the red, green and**
732 **blue spectrum according to the % of total tree basal area occupied by N-fixer, AM, and**
733 **EM tree symbiotic guilds, as indicated by the ternary plot. Grey cells show the global**
734 **distribution of forests where we make model projections.**

735

736 **Figure 2. A small number of environmental variables predict the majority of global**
737 **turnover in forest symbiotic status. Panels show the partial feature contributions of**
738 **different environmental variables on forest symbiotic state. Each row plots the shape of the**
739 **contribution of the four most important predictors of the proportion of tree basal area**
740 **belonging to the (a) ectomycorrhizal (EM), (b) arbuscular mycorrhizal (AM), and (c) N-**
741 **fixer symbiotic guilds (n=2,768). Variables are listed in declining importance from left to**
742 **right, as determined by inc node purity, with points colored with a red-green-blue gradient**
743 **according to their position on the x-axis of the most important variable (left-most panels**
744 **for each guild), allowing cross visualization between predictors. Each panel lists two**
745 **measures of variable importance, inc node purity (used for sorting) and %IncMSE (see**
746 **Supplemental Information for description). The abundance of each symbiont type**
747 **transitions sharply along climatic gradients, suggesting that sites near the threshold are**
748 **particularly vulnerable to switching their dominant symbiont guild with climate changes.**

749

750 **Figure 3. The distribution of forest symbiotic status across biomes is related to climatic**
751 **controls over decomposition. (a) Biome level summaries of the median +/- 1 quartile of the**
752 **predicted % tree basal area per biome for ectomycorrhizal (EM), arbuscular mycorrhizal**

753 (AM), and N-fixer symbiotic guilds (n=100 random samples per biome). (b) The
754 dependency of decomposition coefficients (k, solid and dotted lines) on temperature and
755 precipitation during the warmest quarter with respect to predicted dominance of
756 mycorrhizal symbiosis. The transition from AM forests to EM forests between k=1 and 2 is
757 abrupt, which is consistent with positive feedback between climatic and biological controls
758 of decomposition.

759

760 **Figure 4. Global maps of predicted forest tree symbiotic state. Maps (left) and latitudinal**
761 **gradients (right, with solid line indicating the median and colored ribbon spanning the**
762 **range from the 5% and 95% quantiles) of the % of tree basal area for (a) ectomycorrhizal**
763 **(EM), (b) arbuscular mycorrhizal (AM), and (c) N-fixer symbiotic guilds. All projections**
764 **are displayed a 0.5 by 0.5 degree lat/long scale with n=28,454.**

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767 work of over 200 independent investigators and their public and private funding agencies (see
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769 **Author Contributions**

770 KGP & TWC conceived the study; TWC, JL, PBR, GN, SdM, MZ, NP, BH, XZ, & CZ
771 conceived and organized the GFBi database; KGP, BSS, GDAW, & MVN compiled the
772 symbiosis database; BSS carried out the primary data analysis; MVN & DR contributed to data
773 compilation and analysis; BSS, TWC, MVN & KGP wrote the initial manuscript; BSS, TWC,
774 JL, MVN, GDAW, PBR, GN, SdM, MZ, NP, BH, XZ, CZ & KGP made substantial revisions to

775 all versions of the manuscript; all other named authors provided forest inventory data and
776 commented on the manuscript.

777 **Data Availability**

778 The GFBi database is available upon written request at <https://www.gfbinitiative.org/datarequest>.
779 Additionally, the symbiotic state assigned to tree species as a supplementary file, as are global
780 rasters of our model projections for EM, AM, and N-fixer proportion of tree basal area.

781 **Conflict of Interest**

782 The authors declare we have no conflict of interests.

783 **Supplementary Information**

784 For more information on symbiotic guild assignments, model selection, and supplementary
785 analyses, refer to SupplementaryInfo_Steidinger_etal2019.pdf. For the full suite of Supplemental
786 Files, including symbiotic guild assignments and rasters of model projections, refer to
787 SupplementalFiles_Steidinger_etal2019.zip

788

789 **Methods**

790 We quantified the relative abundance of tree symbiotic guilds across >1.1 million forest
791 census plots combined in the GFBi database, an extension from the plot-based Global Forest
792 Biodiversity (GFB³¹) database. The GFBi database consists of individual-based data that we
793 compiled from all the regional and national GFBi forest inventory data sets. The standardized
794 GFBi data frame, i.e. tree list, comprises tree ID, a unique number assigned to each individual
795 tree; plot ID, a unique string assigned to each plot; plot coordinates, in decimal degrees of
796 WGS84 datum; tree size, in diameter-at-breast-height; trees-per-hectare expansion factor; year of

797 measurement; data set name, a unique number assigned to each forest inventory data set; and
798 binomial scientific tree species names.

799 We error checked all species names from different forest inventory data sets in three
800 steps. First, we extracted scientific names from original data sets, keeping only the names of
801 genus and species (authority names are removed). Next, we compiled all the species names into
802 five general species lists, one for each continent. Finally, we verified individual species names
803 against 23 online taxonomic databases using the ‘taxize’ package of R programming language³².
804 We assigned each morphospecies a unique name comprising the genus, the string “spp”,
805 followed by the data set name and a unique number for that species. For example, “Picea
806 sppCNI1” and “Picea sppCNI2” represent two different species under the genus “Picea”,
807 observed in the first Chinese data set (CNI).

808 We derived plot-level abundance information in terms of species abundance matrices.
809 Each species abundance matrix consisted of the number of individuals by species (column
810 vectors) within individual sample plots (row vectors). In addition, key plot-level information was
811 also added to the matrices, including plot ID, data set name, plot coordinates, the year of
812 measurement, and basal area, i.e. the total cross-sectional areas (m²) of living trees per one
813 hectare of ground area.

814 Tree genera were assigned to a plant family using a plant taxonomy lookup table
815 generated by Will Cornwell (hosted on Github <https://github.com/traitecoevo/taxonlookup>),
816 which uses the accepted taxonomy from “The Plant List.” The majority (96.5%) of genera from
817 the GFBi species were successfully matched to family; for those that could not be assigned, we
818 manually checked the GFBi genus and species against synonyms from The Plant List. Of the
819 remaining 1,038 mismatches, an additional 440 were assigned to family either by updating older

820 genera and species names with their more recent synonyms or else by correcting obvious
821 misspellings. The remaining 598 entries that could not be matched to family were excluded from
822 analysis.

823 We used a taxonomically-informed approach to assign symbiotic states to plant species
824 from the GFBi. Plant species were assigned to one of 5 symbiotic guilds – ectomycorrhizal
825 (EM), arbuscular mycorrhizal (AM), ericoid mycorrhizal (ErM), weakly AM or non-mycorrhizal
826 (AMNM), or N-fixer (Table S1). Although we did not model the relative abundance of ErM
827 trees, due to their rarity, we have included a map of their relative abundance from our grid
828 (Figure S1). We also include as a supplementary file the full species list, which includes columns
829 used to assign species to guild. In addition, we include here a list of families and genera assigned
830 to all guilds except AM (Tables S2-5) with notes for cases of species from individual genera that
831 were either assigned to two guilds simultaneously (e.g., *Alnus* is an N-fixer and EM) or where
832 species from individual genera were split between two different guilds (e.g., some *Pisonia* sp. are
833 weakly AM and some are EM). An AM summary table is excluded for length considerations—
834 the same information is available in the Supplementary File “SymbioticGuildAssignment.csv”.

835 The taxonomy of species in our inventory was compared with recently published
836 literature on the evolutionary history of mycorrhizal symbiosis^{7,33,34} and N-fixation³⁵⁻³⁸. Most
837 species symbiotic status could be reliably assigned at the genus (e.g. *Dicymbe*) or family level
838 (e.g. Pinaceae). For the few groups where status was unreliable or variable within a genus (e.g.
839 *Pisonia*) we conducted additional literature searches.

840 We assigned species to the EM category in three stages. First, at the family level (e.g.,
841 Pinaceae); next, at the genus level (e.g., *Dicymbe*); and finally, using literature searches for
842 unclear genera. For example, for the genus *Pisonia*, some species are AM and others are EM. We

843 used a published list from Hayward & Hynson (2014)³⁹ to sort species into the appropriate guild.
844 For the genus *Acacia*, we followed Brundrett (2017)⁷ in assuming that only endemic Australian
845 species associate with EM, while all others are AM (we sorted *Acacia* species according to
846 provenance using <http://worldwidewattle.com/>).

847 The AMNM category lumped together all genera of terrestrial, non-epiphytic plants that
848 either lack arbuscular mycorrhizal fungi (AMF), or have low or inconsistent records of AMF
849 colonization of roots. For example, although there are some published records of AMF
850 colonization in the roots of Proteaceae, these records are inconsistent, and colonization is
851 generally low. Further, as Proteaceae are associated with a non-mycorrhizal root morphology
852 (the “cluster” or “proteoid root system”) that allows them to access otherwise unavailable forms
853 of soil nutrients⁴⁰, we placed the entire family within AMNM. The family Urticaceae, which we
854 also characterized as AMNM, was somewhat problematic – early-successional species from
855 tropical forests, such as those in the genus *Cecropia*, have records of both low and absent AMF
856 colonization⁴¹. Our approach was to use the most broadly inclusive AMNM categorization.

857 N-fixer status was assigned at the genus level, using previously compiled databases of
858 global symbiotic N₂-fixation³⁵⁻³⁸. Given that symbiotic N₂-fixation with rhizobial or *Frankia*
859 bacteria has only evolved in four orders (Rosales, Cucurbitales, Fabales and Fagales)⁴², all
860 species outside of this nitrogen-fixing clade were assigned non-fixing status. Some species could
861 not be assigned a N-fixer status because they were typed to a higher taxonomic level (e.g.
862 family) that is ambiguous from a N-fixer status perspective. We recorded when our assignment
863 of N-fixer status was based on phylogenetic criteria but where symbiotic N-fixation is
864 evolutionarily labile. Since these cases are more likely to be misassigned we excluded them from
865 the N-fixation category. The N-fixer group contains species that are colonized by AMF (e.g.,

866 most genera from Leguminosae) and others that are colonized by ectomycorrhizal fungi (e.g.,
867 *Alnus* sp.).

868 Most plant species form AM symbiosis, which is the basal symbiotic state to the later
869 derived EM and N-fixing symbioses. Further, many EM and N-fixing plants maintain the ability
870 to form AM symbiosis. Thus, a tree species is most likely AM if it does not form associations
871 with another symbiotic guild (or forgoes root symbiosis entirely), as evidenced by their inclusion
872 in exhaustive databases of plant symbiotic state^{7,33-38,41}. In keeping with other large-scale studies
873 in the field (e.g. ³⁴), we assigned tree species from the GFBi database to an AM-exclusive state if
874 they belonged to taxa that were not matched to EM, ErM, non-or-weakly mycorrhizal or N-fixer
875 symbioses. Thus, the AM and N-fixer groups in our dataset are non-overlapping despite the fact
876 that most N-fixers also associate with AM fungi.

877 The proportions of tree basal area and tree individuals were aggregated to a 1' by 1'
878 degree grid by taking the weighted average of the plot-level proportions (Table S6). This resulted
879 in a total of 2,768 grid cells, each with a score for the proportional abundance of EM, AM, N-
880 fixer, ErM, and AMNM trees. We calculated two measures of relative abundance for each
881 symbiotic guild: proportion of tree stems and proportion of tree basal area. Because the
882 measurements are highly correlated with one another (Figure S2) we chose to model only
883 proportion of total tree basal area, which should scale more approximately to proportion of tree
884 biomass as it accounts for differences in size among individual stems. Additionally, we
885 quantified variability among plots within each grid cell by calculating the weighted standard
886 deviation across the grid (Supplemental Information, Figure S3-4).

887 To identify the key factors structuring symbiotic distributions we assembled 70 global
888 predictor layers: 19 climatic (annual, monthly, and quarterly temperature and precipitation

889 variables), 14 soil chemical (total soil N density, microbial N, C:N ratios and soil P fractions,
890 pH, cation exchange capacity), 5 soil physical (soil texture and bulk density), 26 vegetative
891 indices (leaf area index, total stem density, enhanced vegetation index means and variances), and
892 5 topographic variables (elevation, hillshade) (Table S7). Because decomposition is the dominant
893 process by which soil nutrients become available to plants, we generated 5 additional layers that
894 estimate the climatic control of decomposition. We parameterized decomposition coefficients
895 according to the Yasso07 model^{20,43} using the following equation:

$$896 \quad k = \text{Exp}(0.095T_i - 0.00014 T_i^2) (1 - \text{Exp}[-1.21 P_i]), \quad (1)$$

897 where P_i and T_i are precipitation and mean temperature, either quarterly or annually, and the
898 constants 0.0095 ($=\beta_1$) -0.00014 ($=\beta_2$), and -1.21 ($=\gamma$) are parameters fit using a previous global
899 study of leaf litter mass-loss²⁰. Although local decomposition rates can vary significantly based
900 on litter quality or microbial community composition⁴⁴, climate is the primary control at the
901 global scale²⁰. Decomposition coefficients describe how fast different chemical pools of leaf
902 litter lose mass over time relative to a parameter, α , that accounts for leaf-chemistry.
903 Decomposition coefficients (k) with values of 0.5 and 2 indicate a halving and doubling of
904 decomposition rates relative to α , respectively (Supplemental Information, Figure S5).

905 We implemented the random forest algorithm using the “randomForest” packaged in R.
906 Random forest models average over multiple regression trees, each of which uses a random
907 subset of all the model variables to predict a response. We first determined the influence and
908 relationship of all 75 predictor layers on forest symbiotic state and then optimized our models
909 using a stepwise reduction in variables, from least- to most-important. Variable importance was
910 measured in two ways: Inc Node Purity and %IncMSE (with values reported in each panel of
911 Figure 2). The inc node purity of variable x considers the decrease in the residual sum of squares

912 that results from splitting regression trees using variable x . %IncMSE (mean square error)
913 quantifies the increase in model error as a result of randomly shuffling the order of values in the
914 vector x . We chose to rank variables according to inc node purity because we found that higher
915 inc node purities were associated with larger effect sizes, whereas larger %IncMSE were
916 associated with more linear responses of smaller effect. Whereas our inspection of partial feature
917 contributions is derived from univariate random forest models, we additionally ran multivariate
918 random forests to predict the proportional abundance of EM, AM, and N-fixer trees for each
919 pixel. The multivariate models were run using 50-regression trees each, with the unique set of the
920 best 4 predictor variables for each symbiotic guild in the univariate models (Table S7, Figure 2).
921 Despite strong negative correlations between the proportions of EM and AM basal area (Figure
922 S22), the results from multivariate and univariate random forests are strongly correlated with one
923 another (Figure S23).

924 Using model selection based on eliminating variables with low Inc Node Purity, we
925 removed most soil nutrient, vegetative, and topographic variables from our models (Figure S6-
926 7). Our final models include the remaining 34 predictor layers with climate, decomposition, and
927 certain soil physical and chemical information (Figure S8). To determine the parsimony of our
928 models, we compared the coefficient of determination in models run with a stepwise reduction in
929 the number of variables (starting with those with the lowest Inc Node Purity). Based on
930 performance of the ratio of coefficient of determination in models with 4 vs 34 variables, we
931 determined that the 4 most important variables accounted for >85% of the explained variability
932 (Figure S9). We also compared model performance visually with plots of actual vs predicted
933 proportions of each tree symbiotic guild among continents and geographic subregions (Figure
934 S10). We used the “forestFloor” packaged in R to plot the partial variable response of tree

935 symbiotic guilds to each predictor variable (Figure 2ABC, see Figure S19-21 for partial plots of
936 the partial feature contributions of all 34 variables).

937 In order to test the sensitivity of model performance and predictions, we performed cross
938 validation in R using the “rfUtilities” package²⁴. K-fold cross validation tests the sensitivity of
939 model predictions to losing random subsets from the training data. For EM, AM, and N-fixer
940 models we ran 99 iterations that withheld 10% of the model training data. We assessed the drop
941 in model performance in the 99 iterations by manually calculating the coefficient of
942 determination, which uses the following formula: $1 - \frac{\sum (\text{actual \% basal area} - \text{predicted \% basal area})^2}{\sum (\text{actual \% basal area} - \text{mean actual \% basal area})^2}$. For all symbiotic guilds,
943 withholding 10% of the training data resulted in a mean loss in variance explained of less than
944 1% (Figure S11). This shows that our training data has sufficient redundancy to ensure that our
945 model conclusions are robust. Similarly, to determine whether our random forest models would
946 make similar predictions if data were equally distributed among continents, we rarefied our
947 aggregated grid of symbiotic states and predictor layers to an even depth. Specifically, we sub-
948 sampled all continents – N. America (including Central America and the Caribbean), S. America,
949 Europe, Asia, and Oceania – to match the number of grid-pixels from Africa (n=50). This is a
950 much more aggressive reduction of training data than is typically used in K-fold cross
951 validations, as it involves dropping ~90% of training data rather than retaining the same amount.
952 We performed 99 iterations of rarefaction each for the three symbiotic guilds. On average,
953 models run with the rarefied data explained about 10% less variance over the full training data
954 (the entire predictor / response grid) than did models run with all of the training data (Figure
955 S12-13).

957 To avoid projecting our random forest models outside the ranges of their training data
958 (e.g., grid cells with higher mean annual temperatures than the maximum used to fit the models),
959 we subset a global grid of predictor layers depending on whether (1) the grid cell fell within the
960 top 60% of land surface with respect to tree stem density¹¹ and either (2) fell within the
961 univariate distribution of all the predictor layers from our training data and/or (3) fell within an
962 8-dimensional hypervolume defined by the unique set of the 4-best predictors of the relative
963 abundance of each guild (Figure S14). We then projected our models across only those grid cells
964 that met these criteria, which constitutes 46% of the global land surface and 88% of global tree
965 stems (Figure 1; Figure S15). Model projections were made at two resolutions: both 1 by 1
966 degree and 0.5 by 0.5 degree resolution (Figure 4). While model validation indicates that our
967 projections are robust, additional ground truthing of predictions to identify any discrepancies
968 would be incredibly valuable. If such discrepancies exist they can help fine tune climate-
969 symbiosis models, or identify areas where climate might favour invasion by symbioses that have
970 not yet evolved or dispersed to a particular biogeographic region.

971 We used the following equation to estimate the % of global tree stems that belong to
972 each tree symbiotic guild: $\sum_i (\text{predicted proportion of trees of guild } g \text{ in pixel } i) \times (\text{total number}$
973 $\text{of tree stems in pixel } i) / \sum_i (\text{total number of tree stems in pixel } i)$. The proportion of tree stems
974 and the proportion of tree basal area in each guild are highly correlated throughout the training
975 data (Figure S4). The figures cited in the main text for each guild were calculated using model
976 projections across all pixels, even those that did not meet the criteria for model projection
977 because they fell outside the multivariate distribution of the predictor layers or had insufficient
978 stem density. However, our estimates for the global % of trees occupied by each tree symbiotic
979 guild change by <1% when using only those pixels that met our criteria for model projection.

980 In the main manuscript we state that sharp transitions between dominant symbiotic states
981 with climate variables could lead to declines in EM trees, particularly in southern boreal forests.
982 To determine this, we projected our random forest models for each symbiotic guild using climate
983 change projections over our 19 bioclimatic variables (Table S7), including the decomposition
984 coefficients that use temperature and precipitation values. Specifically, we considered the 2070
985 scenario with a relative concentration pathway (RCP) of 8.5 (W/m²), which predicts an increase
986 of greenhouse gas emissions throughout the 21st century⁴⁵. We plot difference in the proportion
987 of forest basal area between the projections for 2070 and those using current climate data (Table
988 S7, Figure S24). We qualify this prediction with the note that vegetative changes to forests are
989 constrained by rates of mortality, recruitment, and growth.

990 After training and cross-validating our models with GFBi data exclusively, we
991 additionally tested whether our models accurately predicted the symbiotic state of Eurasian
992 forests previously published by Schepaschenko et al. (2017)⁴⁶. We assigned symbiotic status to
993 all trees in Schepaschenko et al. (2017) and aggregated plot level data to a 1 by 1 degree grid
994 using the same methods as with the GFBi dataset (Figure S25). We found that, on average, our
995 models predicted the symbiotic state in the regional dataset within 13.6% of the value of this
996 other dataset (Figure S26). For projected maps in Figure 4abc, we included the Schepaschenko et
997 al. (2017) data with the GFBi training data to increase geographic coverage throughout Eurasia.

998

999 **Methods References**

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1001 31 Liang, J. *et al.* Positive biodiversity-productivity relationship predominant in global
1002 forests. *Science* **354**, aaf8957 (2016).

1003 32 Chamberlain, S. A. & Szöcs, E. taxize: taxonomic search and retrieval in R.
1004 *F1000Research* **2** (2013).

1005 33 Brundrett, M. C. & Tedersoo, L. Evolutionary history of mycorrhizal symbioses and
1006 global host plant diversity. *New Phytol.* (2018).

1007 34 Brundrett, M. C. Mycorrhizal associations and other means of nutrition of vascular
1008 plants: understanding the global diversity of host plants by resolving conflicting
1009 information and developing reliable means of diagnosis. *Plant and Soil* **320**, 37-77,
1010 doi:10.1007/s11104-008-9877-9 (2009).

1011 35 Werner, G. D., Cornwell, W. K., Sprent, J. I., Kattge, J. & Kiers, E. T. A single
1012 evolutionary innovation drives the deep evolution of symbiotic N₂-fixation in
1013 angiosperms. *Nature Communications* **5**, 4087 (2014).

1014 36 Werner, G. D., Cornwell, W. K., Cornelissen, J. H. & Kiers, E. T. Evolutionary signals of
1015 symbiotic persistence in the legume–rhizobia mutualism. *Proceedings of the National
1016 Academy of Sciences* **112**, 10262-10269 (2015).

1017 37 Afkhami, M. E. *et al.* Symbioses with nitrogen-fixing bacteria: nodulation and
1018 phylogenetic data across legume genera. *Ecology* **99**, 502-502 (2018).

1019 38 Tedersoo, L. *et al.* Global database of plants with root-symbiotic nitrogen fixation: Nod
1020 DB. *Journal of Vegetation Science* (2018).

1021 39 Hayward, J. & Hynson, N. A. New evidence of ectomycorrhizal fungi in the Hawaiian
1022 Islands associated with the endemic host *Pisonia sandwicensis* (Nyctaginaceae). *Fungal
1023 Ecol* **12**, 62-69 (2014).

1024 40 Lambers, H., Martinoia, E. & Renton, M. Plant adaptations to severely phosphorus-
1025 impoverished soils. *Current opinion in plant biology* **25**, 23-31 (2015).

1026 41 Wang, B. & Qiu, Y.-L. Phylogenetic distribution and evolution of mycorrhizas in land
1027 plants. *Mycorrhiza* **16**, 299-363 (2006).

1028 42 Soltis, D. E. *et al.* Chloroplast gene sequence data suggest a single origin of the
1029 predisposition for symbiotic nitrogen fixation in angiosperms. *Proceedings of the
1030 National Academy of Sciences* **92**, 2647-2651 (1995).

1031 43 Palosuo, T., Liski, J., Trofymow, J. & Titus, B. Litter decomposition affected by climate
1032 and litter quality—testing the Yasso model with litterbag data from the Canadian intersite
1033 decomposition experiment. *Ecological Modelling* **189**, 183-198 (2005).

1034 44 Bradford, M. A. *et al.* Climate fails to predict wood decomposition at regional scales.
1035 *Nature Climate Change* **4**, 625 (2014).

1036 45 Allen, M. R. *et al.* IPCC fifth assessment synthesis report-climate change 2014 synthesis
1037 report. (2014).

1038 46 Schepaschenko, D. *et al.* A dataset of forest biomass structure for Eurasia. *Scientific data*
1039 **4**, 170070 (2017).

1040