

Monkeypox virus isolation from longitudinal samples in 11 hospitalised patients.

Helen Callaby^{1,2}, Janie Olver¹, Kirsty Emery³, Kevin S Richards^{3,4}, Marian Killip³, Natalie Groves⁵, Mike BJ Beadsworth^{6,7}, D. Ashley Price⁸, Graham S Cooke^{9,10}, Paul Collini¹¹, Joby Cole¹¹, Jake Dunning^{13,14}, Malcolm G Semple^{15,16}, J Kenneth Baillie^{17,18}, ISARIC4C Investigators, Tommy Rampling^{1,19}, Catherine F Houlihan^{1,20}

1. Rare and Imported Pathogens Laboratory, UKHSA, Porton Down, UK
2. Institute of Medical Sciences, University of Aberdeen, UK
3. High Containment & Biological Services, UKHSA, Porton Down, UK
4. Faculty of Health and Life Sciences, Oxford Brookes University, Headington Campus, Oxford, UK
5. Genomic Public Health Analysis, UKHSA, Colindale, UK
6. Tropical and Infectious Disease Unit, Liverpool University Hospitals NHS Foundation Trust, Liverpool, UK
7. Department of Clinical Sciences, Liverpool School of Tropical Medicine, Liverpool, UK
8. The Newcastle upon Tyne NHS Hospitals Trust
9. Division of infections diseases, Imperial College London
10. Biomedical Research Centre Imperial College NHS Trust
11. Sheffield Teaching Hospitals NHS Foundation Trust
13. Pandemic Sciences Institute, University of Oxford, Oxford, UK
14. Department of Infectious Diseases, Royal Free London NHS Foundation Trust, Royal Free Hospital, London, UK
15. Health Protection Research Unit in Emerging and Zoonotic Infections, Institute of Infection, Veterinary and Ecological Sciences, Faculty of Health and Life Sciences, University of Liverpool, Liverpool, UK
16. Respiratory Department, Liverpool Institute for Child Health and Wellbeing, Alder Hey Children's Hospital, Liverpool, UK
17. Gifford Pandemic Science Hub, Centre for Inflammation Research, University of Edinburgh, Edinburgh, UK
18. Intensive Care Unit, Royal Infirmary of Edinburgh, Little France Crescent, Edinburgh, UK
19. Department of Infectious Diseases, University College London NHS Hospitals Trust
20. Department of Clinical Virology, University College London NHS Hospitals Trust

Monkeypox virus (MPXV) causes mpox, a disease largely confined to West and Central Africa until 2022, when the global spread of MPXV clade IIb was declared a public health emergency of international concern (PHEIC). Another PHEIC was declared in 2024 following the spread of MPXV clade I in Africa. Human transmission occurs mainly through contact with infectious lesions (1). While diagnostic swabs' PCR cycle threshold (Ct) is assumed to correlate with infectiousness, the evidence is weak (2,3). Prolonged MPXV shedding is widely reported, but the duration of infectiousness remains unclear (4–5).

We used the ISARIC WHO Clinical Characterisation Protocol UK to enrol patients admitted to UK hospitals for severe mpox. Seven patients were enrolled from 2018 to 2021 and four in 2022, with a median age of 35 years (range <1–48 years), seven male and four female. One patient was living with HIV. 169 samples were collected,

including lesion swabs, throat swabs, scabs, cerebrospinal fluid, endotracheal aspirates, plasma, and urine. The samples were taken from 13 days before symptom onset to 109 days after onset (supplementary Figure S1, patient 1).

The samples were inoculated into African Green Monkey Kidney (Vero E6) cells, incubated, and monitored for cytopathic effects (CPE). MPXV PCR of culture supernatant on days 0 and 7 confirmed CPE observations if there was a decrease in Ct. All MPXV detected were clade II. 99 samples were positive for MPXV by PCR on day 0 (Ct range 20.1-39.9), and virus was isolated from 47 samples (47.5%). The median Ct value for which MPXV could be isolated was 31.01 (range 22.3 – 38.0).

The longest duration of culture positivity was 109 days, observed in a throat swab with an MPXV Ct of 33.7 from a man with poorly controlled HIV (CD4+ count of 57 cells/mm³ and an HIV viral load of 52,800 copies/mL). He presented with an ulcerating limb rash and a necrotic tongue lesion, was treated with tecovirimat, and was discharged, only to return two weeks later with worsening symptoms (6).

Sequencing confirmed a prolonged infection rather than reinfection (see supplementary material). Since his disease was influenced by immune response and antiviral treatment, his data was included in Figure 1A but removed from 1B.

Our data showed MPXV isolation from lesion swabs, throat swabs, tracheal aspirates, scabs, and urine samples from day 2 to 109 days after illness onset. MPXV was isolated at higher Ct values (Ct=37) than previously reported, adding uncertainty to the assumption that patients with high Ct values on PCR are not infectious (2,3).

The duration of infectiousness for MPXV is still unclear, particularly in hospitalised patients (7). A Canadian cohort study found viral DNA detectable for 3-4 weeks,

though this was not linked to infectiousness (8). A Spanish study of outpatients with mpox, including culture data, found the median time to viral clearance to be 25 days, likely due to milder disease (9). Similarly, a 2023 review showed culture data varied by site, with testing ceasing after 19 days (10). The median time of isolation in our study, 12.0 days, fits within these thresholds but was far exceeded by the immunocompromised patient at 109 days.

A limitation of our study is that patients were sampled for clinical diagnosis with inconsistent sampling frequency. Additional freeze-thaw cycles may have impacted culture success. Despite this, we observed the longest virus culture period from a throat sample at over three months post-diagnosis. This study only included MPXV clade II samples, while clade I, which may be associated with a higher case fatality rate, may have different characteristics.

Immunocompromised patients may remain contagious with MPXV for extended periods, particularly through respiratory and oral routes, especially when oropharyngeal lesions are present. Infection prevention guidance should recommend close virological follow-up for these patients who may need extended isolation. High Ct values from oropharyngeal PCRs may not reliably indicate the end of infectiousness.

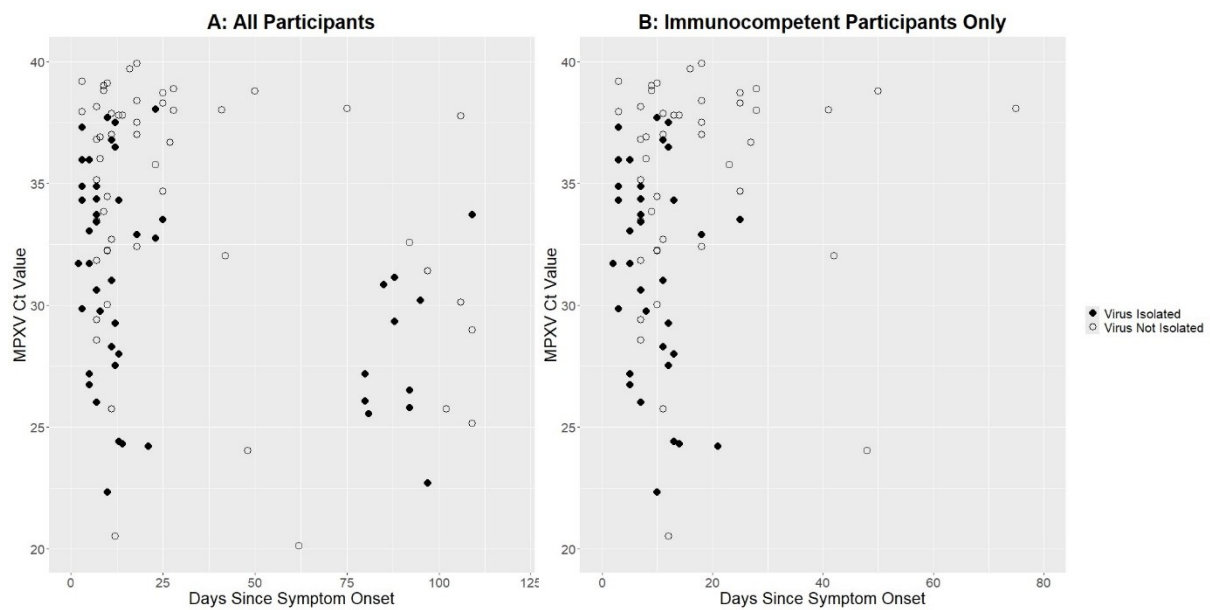


Figure 1. Participants' MPXV PCR Ct value against the number of days since symptom onset. Panel A displays all participants, panel B displays only immunocompetent participants (excluding participant 1). The x-axis limit changes in each figure depending on the maximum number of days since symptom onset. On the y-axis, a decrease in Ct value corresponds to an increase in MPXV DNA in the sample. The symbol used to denote each value is solid black if the virus was able to be isolated at day 7 of culture and hollow if the virus could not be isolated. Graphs were produced using the ggplot2 package (version 3.4.4) in R (version 4.3.2).

References

1. Vaughan A, Aarons E, Astbury J, Brooks T, Chand M, Flegg P, et al. Human-to-Human Transmission of Monkeypox Virus, United Kingdom, October 2018. *Emerg Infect Dis* [Internet]. 2020 Apr;26(4):782–5. Available from: http://wwwnc.cdc.gov/eid/article/26/4/19-1164_article.htm
2. Hernaez B, Muñoz-Gómez A, Sanchiz A, Orviz E, Valls-Carbo A, Sagastagoitia I, et al. Monitoring monkeypox virus in saliva and air samples in Spain: a cross-sectional study. *Lancet Microbe*. 2023 Jan;4(1):e21–8.

3. Lim CK, McKenzie C, Deerrain J, Chow EPF, Towns J, Chen MY, et al. Correlation between monkeypox viral load and infectious virus in clinical specimens. *Journal of Clinical Virology*. 2023 Apr;161:105421.
4. Nörz D, Brehm TT, Tang HT, Grewe I, Hermanussen L, Matthews H, et al. Clinical characteristics and comparison of longitudinal qPCR results from different specimen types in a cohort of ambulatory and hospitalized patients infected with monkeypox virus. *Journal of Clinical Virology*. 2022 Oct;155:105254.
5. Palich R, Burrell S, Monsel G, Nouchi A, Bleibtreu A, Seang S, et al. Viral loads in clinical samples of men with monkeypox virus infection: a French case series. *Lancet Infect Dis*. 2023 Jan;23(1):74–80.
6. Stafford A, Rimmer S, Gilchrist M, Sun K, Davies EP, Waddington CS, et al. Use of cidofovir in a patient with severe mpox and uncontrolled HIV infection. *Lancet Infect Dis*. 2023 Jun;23(6):e218–26.
7. European Centre for Disease Prevention and Control. <https://www.ecdc.europa.eu/en/all-topics-z/monkeypox/factsheet-health-professionals>. Factsheet for Health Professionals on Mpox.
8. Tan DHS, Pico Espinosa O, Matelski J, Khera SS, Qamar A, Persaud R, et al. Longitudinal Analysis of Mpox Virus DNA Detectability From Multiple Specimen Types During Acute Illness: A Cohort Study. *Open Forum Infect Dis*. 2024 Feb 1;11(2).
9. Suñer C, Ubals M, Tarín-Vicente EJ, Mendoza A, Alemany A, Hernández-Rodríguez Á, et al. Viral dynamics in patients with monkeypox infection: a prospective cohort study in Spain. *Lancet Infect Dis*. 2022;12(12):e2022.
10. Kim H, Kwon R, Lee H, Lee SW, Rahmati M, Koyanagi A, et al. Viral load dynamics and shedding kinetics of mpox infection: a systematic review and meta-analysis. *J Travel Med*. 2023 Sep 5;30(5).