

# 1 **Methaemoglobin as a surrogate marker of primaquine antihypnozoite activity** 2 **in *Plasmodium vivax* malaria: a systematic review and individual patient data** 3 **meta-analysis**

## 4 5 **Methaemoglobin as a surrogate endpoint in vivax malaria**

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## 46 Abstract

47

48 **Background** The 8-aminoquinolines, primaquine and tafenoquine, are the only available drugs  
49 for the radical cure of *Plasmodium vivax* hypnozoites. Prior evidence suggests that there is  
50 dose-dependent 8-aminoquinoline induced methaemoglobinaemia and that higher  
51 methaemoglobin concentrations are associated with a lower risk of *P. vivax* recurrence. We  
52 undertook a systematic review and individual patient data meta-analysis to examine the utility  
53 of methaemoglobin as a surrogate endpoint for 8-aminoquinoline antihypnozoite activity to  
54 prevent *P. vivax* recurrence.

55

56 **Methods** We conducted a systematic search of Medline, Embase, Web of Science, and the  
57 Cochrane Library, from 1 January 2000 to 29 September 2022 inclusive, of prospective clinical  
58 efficacy studies of acute, uncomplicated *P. vivax* malaria mono-infections treated with radical  
59 curative doses of primaquine. The day 7 methaemoglobin concentration was the primary  
60 surrogate outcome of interest. The primary clinical outcome was the time to first *P. vivax*  
61 recurrence between day 7 and day 120 after enrolment. We used multivariable Cox  
62 proportional-hazards regression with site random-effects to characterise the time to first  
63 recurrence as a function of the day 7 methaemoglobin percentage ( $\log_2$  transformed), adjusted  
64 for the partner schizontocidal drug, the primaquine regimen duration as a proxy for the total  
65 primaquine dose (mg/kg), the daily primaquine dose (mg/kg), and other factors. The  
66 systematic review protocol was registered with PROSPERO (CRD42023345956).

67

68 **Findings** We identified 219 *P. vivax* efficacy studies, of which eight provided relevant  
69 individual-level data from patients treated with primaquine; all were randomised, parallel arm  
70 clinical trials assessed as having low or moderate risk of bias. In the primary analysis dataset,  
71 there were 1747 G6PD-normal patients enrolled from 24 study sites across 8 different  
72 countries (Indonesia, Brazil, Vietnam, Thailand, Peru, Colombia, Ethiopia, India). We observed  
73 an increasing dose-response relationship between the daily weight-adjusted primaquine dose  
74 and day 7 methaemoglobin level. For a given primaquine dose regimen, an observed doubling  
75 in day 7 methaemoglobin percentage was associated with an estimated 30% reduction in the  
76 risk of vivax recurrence (adjusted hazard ratio = 0.70; 95% CI = [0.57, 0.86];  $p = 0.0005$ ). These  
77 pooled estimates were largely consistent across the study sites. Using day 7 methaemoglobin  
78 as a surrogate endpoint for recurrence would reduce required sample sizes by approximately  
79 40%.

80

81 **Conclusions** For a given primaquine regimen, higher methaemoglobin on day 7 was associated  
82 with a reduced risk of *P. vivax* recurrence. Under our proposed causal model, this justifies the  
83 use of methaemoglobin as a surrogate endpoint for primaquine antihypnozoite activity in  
84 G6PD normal patients with *P. vivax* malaria.

85

86 **Word count** 408/500

## 87 Introduction

88 The human malaria parasites *Plasmodium vivax* and *Plasmodium ovale* are characterised by  
89 their ability to form dormant liver stage parasites called hypnozoites, which activate weeks to  
90 months later to cause relapsing bloodstream infection [1]. *P. vivax* is the most geographically  
91 widespread cause of human malaria and is a major challenge in malaria elimination. Relapse  
92 contributes substantially to the overall burden of symptomatic vivax malaria, causing over 75%  
93 of all symptomatic infections [2]. Preventing relapse is crucial for eliminating the burden of  
94 vivax malaria morbidity and mortality.

95  
96 The only drugs available for radical cure (killing latent hypnozoites) are primaquine and  
97 tafenoquine. Both are thought to be prodrugs [3, 4], necessitating metabolic activation to  
98 produce hypnozoicidal activity. The precise mechanism and active metabolites of  
99 primaquine and tafenoquine remain unknown [5]. Patients with vivax malaria, who receive an  
100 equivalent 8-aminoquinoline dose per body weight, may variably metabolise the drug leading  
101 to varying risks of later relapse. Some of this variation in biotransformation is due to  
102 polymorphisms in cytochrome *P450 (CYP) 2D6* [3] and poorer metabolisers are associated with  
103 higher relapse rates [6, 7].

104  
105 Both primaquine and tafenoquine cause predictable increases in blood methaemoglobin  
106 resulting from the action of their oxidative metabolites [8]. This involves a reversible increase  
107 in the conversion rate of intra-erythrocytic reduced ( $\text{Fe}^{2+}$ ) haem iron in haemoglobin to its  
108 oxidised ( $\text{Fe}^{3+}$ ) form [8]. Following daily administration of primaquine or single dose  
109 administration of tafenoquine, blood methaemoglobin gradually increases, reaching a peak  
110 concentration after approximately one week [9]. It has been hypothesised that the same  
111 oxidative metabolites responsible for methaemoglobinaemia are also responsible for inducing  
112 haemolysis in glucose-6-phosphate dehydrogenase (G6PD) deficiency, killing mature  
113 gametocytes of *P. falciparum*, and killing liver stage hypnozoites [8]. Early experiments with  
114 primaquine and its analogues indicated that there were greater increases in blood  
115 methaemoglobin for molecules which had improved radical-cure efficacy [8]. Two recent  
116 pharmacometric studies estimated an adjusted proportional reduction in the risk of vivax  
117 recurrence of approximately 10% for primaquine [10] and 20% for tafenoquine [4] for each  
118 additional percentage-point increase in day 7 methaemoglobin. Based on these results, it is  
119 hypothesised that increases in methaemoglobin may serve as a proxy for 8-aminoquinoline  
120 antihypnozoite activity and, as such, a potential surrogate endpoint for clinical trials to quantify  
121 the antirelapse efficacy of 8-aminoquinoline drugs in vivax malaria. A surrogate endpoint is a  
122 patient characteristic, such as a biomarker, intended to substitute for a clinical outcome [11]–  
123 specifically vivax recurrence in this context. To validate surrogacy, high-quality studies need to  
124 demonstrate that a putative biomarker is affected by the drug intervention and that drug-  
125 induced change in the biomarker level can predict the effect on the outcome of interest [12,  
126 13].

127  
128 We aimed to examine the utility of methaemoglobin as a surrogate endpoint for vivax  
129 recurrence using all available data in a pooled individual patient data meta-analysis involving  
130 *P. vivax* patients from multiple countries treated with primaquine.

## 131 Methods

### 132 Search strategy and selection criteria

133 We conducted a systematic search of Medline, Embase, Web of Science, and the Cochrane  
134 Library based on an existing living systematic review [14] of prospective clinical efficacy studies  
135 of acute, uncomplicated *P. vivax* malaria mono-infections treated with primaquine. We  
136 included randomised therapeutic trials and prospective cohort studies published between 1  
137 January 2000 and 29 September 2022 inclusive, in any language, with a minimum active follow-  
138 up of 42 days that recorded methaemoglobin data (at baseline and at least once in the first  
139 week of follow-up between day 5 and day 9) following daily primaquine administration given  
140 over multiple days. Only G6PD normal individuals (i.e.,  $\geq 30\%$  G6PD activity or a negative  
141 qualitative test) were included in the analysis. In G6PD deficient individuals, primaquine  
142 administration does not lead to the same methaemoglobin increases [15].

143  
144 Studies were included if the primaquine regimen was administered within the first three days  
145 of schizontocidal treatment. Search terms are provided in Supporting Information (List S1). This  
146 systematic review was conducted by two reviewers (IF and RJC), with discrepancies resolved  
147 through discussion. The review protocol was registered with PROSPERO (CRD42023345956).

### 148 149 Data pooling

150 The corresponding authors and/or principal investigators of eligible studies that met the study  
151 criteria were invited through direct email to contribute their individual patient data. Relevant  
152 data from unpublished studies were requested wherever possible. Shared data were uploaded  
153 to the Worldwide Antimalarial Resistance Network (WWARN) repository for curation and  
154 standardisation, utilising the IDDO SDTM Implementation Guide [16]. We excluded individual  
155 patients with missing information on age, sex, body weight, baseline parasite density,  
156 primaquine regimen, or schizontocidal treatment. Patients with severe malaria, pregnancy,  
157 mixed-species infection, or those who received adjunctive antimalarials after the initial  
158 schizontocidal treatment were also excluded.

159  
160 All studies included in our meta-analysis provided pseudonymised individual data and had  
161 obtained ethical approvals from the corresponding site of origin. Therefore, additional ethical  
162 approval was not required for the current analysis, as per the Oxford Tropical Research Ethics  
163 Committee. We adhered to the PRISMA-IPD guidelines [17] for reporting this systematic review  
164 and individual patient data meta-analysis (checklist available in Supporting Information, Table  
165 S1).

### 166 167 Outcomes

168 The primary clinical outcome was the time to first *P. vivax* recurrence (i.e., any episode of *P.*  
169 *vivax* parasitaemia irrespective of symptoms) between day 7 (the starting point for prediction  
170 and follow-up) and day 120 [4] after the initial primaquine administration.

171  
172 The secondary outcomes were the binary outcome of any *P. vivax* recurrence between day 7  
173 and day 120 after primaquine initiation and the maximum absolute change in haemoglobin  
174 concentration from day 0 to days 2–3 (expected days of the lowest haemoglobin level [18])  
175 following the start of any antimalarial treatment. In addition to being the main predictor of  
176 interest in the statistical model, we also specified the day 7 methaemoglobin concentration as

177 an outcome and modelled this biomarker as a function of daily mg (base) per kilogram  
178 primaquine dose.

179

## 180 Data analysis

181 We presented study-level summary statistics to highlight sample characteristics and potential  
182 heterogeneity across the included studies. The daily distributions of methaemoglobin levels,  
183 stratified by schizontocidal drug and primaquine regimen (low total dose 14-day, high total  
184 dose 7-day), were plotted to illustrate the temporal dynamics of primaquine-induced  
185 methaemoglobin production during radical cure treatment.

186

187 The primary predictor of interest (surrogate outcome) was the day 7 methaemoglobin  
188 concentration, expressed as a percentage of the total haemoglobin concentration. Day 7 was  
189 prespecified and typically, methaemoglobin concentrations peak after approximately a week  
190 of commencing the daily primaquine regimens. All studies measured methaemoglobin by  
191 transcutaneous pulse CO-oximetry. The day 7 methaemoglobin percentage ( $\log_2$  transformed)  
192 was included in the statistical model as a continuous variable. If the day 7 methaemoglobin  
193 percentage was recorded as zero, this remained untransformed (which assumed a  
194 methaemoglobin of 1%—approximately the physiological level). Zero recordings are likely to  
195 represent mis-readings of the analytical machine.

196

197 We proposed a causal directed acyclic graph for this analysis to guide model specification and  
198 aid interpretation of results (Fig 1). Missing day 7 methaemoglobin percentages were linearly  
199 imputed using levels measured within  $\pm 2$  days. If only one measurement was available, then  
200 the imputation assumed a constant (i.e., the single value observed was used). If no  
201 measurements were available within this timeframe (day 5 to day 9), the patient was excluded  
202 from the analysis.

203

204 **Figure 1. Directed acyclic graph showing our hypothesised causal relationships between**  
205 **primaquine-induced changes in blood methaemoglobin and *P. vivax* relapse.**

206

207 In the main analysis, patients were right censored at the time of the first recurrent vivax  
208 parasitaemia (outcome), any malaria parasitaemia, loss to follow up, blood smear gap of >60  
209 days, or the last day of study; whichever occurred first. We used multivariable, random-effects  
210 Cox proportional-hazards regression to model the time to first recurrence as a function of the  
211 day 7 methaemoglobin percentage ( $\log_2$  transformed) under a one-stage individual patient  
212 data meta-analysis framework. This model adjusted for daily mg/kg primaquine dose,  
213 primaquine duration (a proxy for total mg/kg primaquine dose), within-site and across-site  
214 linear interactions [19] between daily mg/kg primaquine dose and primaquine duration, age,  
215 sex, schizontocidal drug, and baseline parasite density (natural-log transformed). A random  
216 intercept and a random slope for day 7 methaemoglobin concentration were included to  
217 account for between-site effect-heterogeneity. Linearity and proportional-hazards  
218 assumptions were checked. The adjusted hazard ratio can be interpreted as the estimated  
219 predictive effect of each doubling in day 7 methaemoglobin percentage, over and above the  
220 adjustment factors.

221

222 To compare our estimates with those from previous studies [4, 10], we also specified the day  
223 7 methaemoglobin concentration on its original scale and refitted the main survival analysis  
224 model. Additionally, we separately (by study and primaquine regimen) fit a more parsimonious  
225 Cox proportional-hazards model that adjusted for daily mg/kg primaquine dose and pooled the  
226 estimates obtained from all the resulting clusters using a two-stage individual-patient data  
227 meta-analysis approach. A similar model specification to the one-stage approach that included  
228 a few more adjustment factors was not possible as the data were sparse (i.e., few recurrences).  
229 A forest plot was constructed to visualise the results under the common-effect and random-  
230 effects models.

231  
232 We estimated the adjusted predictive effect of the day 7 methaemoglobin percentage ( $\log_2$   
233 transformed) on the odds of vivax recurrence using multivariable, random-effects binary  
234 logistic regression. This model was limited to patients with at least 120 days of follow-up and  
235 adjusted for daily mg/kg primaquine dose, primaquine duration, within-site and across-site  
236 linear interactions between daily mg/kg primaquine dose and primaquine duration. A random  
237 intercept for study site was specified. The association between the maximum absolute change  
238 in haemoglobin concentration from day 0 to days 2–3 and the day 7 methaemoglobin  
239 percentage was estimated using multivariable, random-effects linear regression. This model  
240 included baseline haemoglobin concentration, daily mg/kg primaquine dose, age, sex,  
241 schizontocidal drug, baseline parasite density (natural-log transformed) as common-effect  
242 covariates, and a random intercept and slope for study site and daily mg/kg primaquine dose,  
243 respectively. This model was restricted to patients who started primaquine treatment on day  
244 0. If a haemoglobin measurement was missing, haematocrit was used to impute the  
245 haemoglobin concentration using the formula  $\text{haemoglobin} = (\text{haematocrit} - 5.62) \div 2.60$ ,  
246 where haematocrit was measured in percent and haemoglobin was measured in grams per  
247 decilitre [20]. If haematocrit remained missing, these patients were excluded. We also  
248 estimated the association of daily mg/kg primaquine dose and day 7 methaemoglobin  
249 percentage using a random-effects linear model, allowing for a random intercept and slope for  
250 study site and daily mg/kg primaquine dose, respectively.

251  
252 We provide illustrative sample-size calculations to demonstrate how our findings could  
253 contribute to making future studies of drug discovery or regimen optimisation in *P. vivax* more  
254 efficient by using blood methaemoglobin as a surrogate outcome. We estimated that a 0.5-  
255 mg/kg increase in daily primaquine dose results in a 0.39 increase in the  $\log_2$  day 7  
256 methaemoglobin (i.e. a 30% increase). We estimated the standard deviation of the  $\log_2$  day 7  
257 methaemoglobin level conditional on the daily mg/kg primaquine dose from the pooled data.  
258 Assuming a normal distribution for the  $\log_2$  day 7 methaemoglobin conditional on the daily  
259 dose allows for a simple calculation of the required sample size (based on a test for a difference  
260 between two normal distributions).

261  
262 Risk of bias related to individual studies was evaluated using the Quality in Prognosis Studies  
263 (QUIPS) tool [21] adapted to the current analysis (signalling questions are provided in  
264 Supporting Information, List S2). Statistical analysis followed a prespecified plan [22] and was  
265 conducted using R Statistical Software (version 4.3.0).

## 266 Findings

### 267 Study and patient selection

268 We identified 219 *P. vivax* efficacy studies published between 1 January 2000 and 29  
269 September 2022 (Fig 2). After review, 206 studies were excluded, leaving 13 studies eligible  
270 for the pooled analysis. Eight of these studies (8/13, 62%) provided individual level data from  
271 patients treated with primaquine; all were randomised, parallel arm clinical trials and assessed  
272 as having low risk of bias (Table S2). Individual level data were available for 4122 patients from  
273 these eight trials, of whom 1747 (42%) satisfied the inclusion and exclusion criteria for our  
274 analysis (primary dataset). These patients were enrolled from 24 study sites across 8 different  
275 countries (Indonesia, Brazil, Vietnam, Thailand, Peru, Colombia, Ethiopia, India; Fig S1). Most  
276 patients had a follow-up of at least 120 days since commencing primaquine radical cure  
277 treatment (1344, 77%—secondary sub-dataset) and had haemoglobin concentration (or  
278 haematocrit) measured on day 0 of antimalarial treatment (1360, 78%—haemolysis sub-  
279 dataset).

280

## 281 **Figure 2. Study and patient selection**

282

### 283 Patient characteristics

284 In the primary analysis (n = 1747 patients), the median age was 20 years (interquartile range  
285 [IQR]: 12 to 32), with 89 patients (5.1%) younger than 5 years. Overall, most patients were  
286 male (1116, 64%), resided in the Asia-Pacific region (1620, 93%), and were from locations with  
287 frequent relapse periodicity (i.e., a median interval from the first acute episode to relapse of  
288 less than 47 days [23]; 1614, 92%) and moderate transmission intensity (i.e., 1 to 9 cases per  
289 1000 person-years [24]; 1325, 76%). The majority of patients (1138, 65%) were treated with  
290 artemisinin-combination therapies (ACT), started primaquine on day of enrolment (day 0;  
291 1496, 86%), and took daily primaquine over 14 days (1194, 68%). The overall median daily-  
292 dose of primaquine was 0.52 mg/kg (IQR: 0.38 to 0.95, Fig S2 shows the weight-adjusted dose  
293 distribution by primaquine duration). Primaquine administration was fully supervised in most  
294 studies (i.e., all doses were directly observed; 1571, 90%). Further details on baseline patient  
295 characteristics are summarised in Table 1. Eligible studies that were not included [25-29] in the  
296 analysis tended to have a shorter duration of follow-up but were otherwise similar in other  
297 characteristics (Tables S3–5).

298

## 299 **Table 1. Demographic and patient characteristics at baseline**

300

### 301 Effect of primaquine on methaemoglobin concentrations

302 There were consistent increases in methaemoglobin concentration from baseline to day 7  
303 during primaquine treatment for both the 7-day and 14-day regimens. On average, the peak  
304 observed methaemoglobin concentrations were on day 7 following start of primaquine  
305 administration, with a median day 7 methaemoglobin level of 6.0% (IQR: 3.3 to 9.0).  
306 Subsequently, methaemoglobin levels tended to decrease slowly in the 7-day or plateau in the  
307 14-day regimen groups (Fig 3, Fig S3).

308

## 309 **Figure 3. Dynamics of primaquine-induced increases in blood methaemoglobin over time, 310 stratified by primaquine regimen and schizontocidal drug**

311

312 Among patients treated with a low-to-intermediate daily primaquine dose, day 7  
313 methaemoglobin was lower in the studies where primaquine was combined with chloroquine  
314 or quinine as a partner drug (Fig S4). However, there was no clear evidence suggesting a drug-

315 drug interaction at the patient level as there was insufficient within-site variation in the pooled  
316 data (within-site interaction  $p = 0.29$ ; across-site interaction  $p = 0.028$ ). Younger and older  
317 patients had lower primaquine-induced methaemoglobinaemia; with the highest levels  
318 observed among adolescent patients (Fig S5). Figure S6 shows the distribution of day 7  
319 methaemoglobin concentrations by primaquine regimen.

320

321 There was dose-dependent primaquine-induced methaemoglobin production. On average, for  
322 every additional 0.1 mg/kg increase in the daily primaquine dose, there was an associated 0.34  
323 percentage-point increase in the day-7 methaemoglobin concentration (95% confidence  
324 interval [CI] = [0.16, 0.52];  $p = 0.002$ ). This effect remained consistent across all sites (between-  
325 site standard deviation in mean difference = 0.20; this quantifies the variability in the estimated  
326 effect of the daily primaquine dose on the day-7 methaemoglobin concentration across  
327 different sites). For a particular mg/kg daily dose, patients who later developed vivax  
328 recurrences (largely attributable to relapses) during follow-up had lower day 7  
329 methaemoglobin values (Fig 4).

330

331 **Figure 4. Methaemoglobin levels (%) by daily mg/kg primaquine dose and *Plasmodium***  
332 ***vivax* recurrence status**

333

334 Association of day 7 methaemoglobin concentrations with the risk of vivax recurrence

335 After adjusting for the daily and total dose of primaquine and other covariates, a doubling in  
336 the observed or imputed day 7 methaemoglobin percentage was associated with an estimated  
337 30% reduction in the risk of vivax recurrence (adjusted hazard ratio [aHR] = 0.70; 95% CI =  
338 [0.57, 0.86];  $p = 0.0005$ ). These pooled estimates were largely consistent across the study sites  
339 (between-site standard deviation in aHR = 1.01; this is a multiplicative factor reflecting the  
340 variability in the estimated predictive-effect of the day-7 methaemoglobin concentration on  
341 the hazard across different study sites). There was no evidence of proportional hazards  
342 violations within the 120 days of follow-up. A sensitivity analysis restricted to patients with  
343 observed day 7 methaemoglobin values gave similar estimates (aHR = 0.66; 95% CI = [0.52,  
344 0.84];  $p = 0.0008$ ;  $n = 1502$ ). Controlling primaquine daily dose and duration, the relationship  
345 between day 7 methaemoglobin and the risk of vivax recurrence was generally consistent  
346 between the studies (Fig 5). A sensitivity analysis using a two-stage approach estimated a  
347 slightly smaller effect (dose-adjusted HR for vivax recurrence of 0.81 for each doubling in day  
348 7 methaemoglobin percentage; 95% CI = [0.67, 0.98];  $p = 0.037$ ). This numerically different  
349 value estimated by the two-stage approach was primarily a result of less flexibility in model  
350 specification (e.g., fewer adjustment factors allowed to avoid estimation issues) and sparsity  
351 of data.

352

353 **Figure 5. Forest plot**

354

355 On the absolute scale (i.e. percentage of the total haemoglobin), each additional percentage-  
356 point increase in day 7 methaemoglobin was associated with an estimated 10% reduction in  
357 the risk of vivax recurrence (aHR = 0.90; 95% CI = [0.84, 0.96];  $p = 0.003$ ). Improved model-fit  
358 ( $p < 0.0001$ ) was observed by specifying day 7 methaemoglobinaemia on the logarithmic scale  
359 (as in the main model), indicating linearity between multiplicative changes in day-7  
360 methaemoglobin level and the log-hazard ratio for vivax recurrence.

361

## 362 Additional analyses

363 Of the 1344 (77%) patients followed for at least 120 days, we observed a comparable  
364 relationship between day 7 methaemoglobin and the risk of any observed vivax recurrence  
365 (dose-adjusted odds ratio of 0.66 for each doubling in day 7 methaemoglobin; 95% CI = [0.52,  
366 0.83];  $p=0.0004$ ). Of the 1360 (78%) G6PD normal patients with haemoglobin (or haematocrit)  
367 measured at baseline, there was little or no evidence of an association between the maximum  
368 absolute decrease in haemoglobin concentration from day 0 to days 2–3 and the day-7  
369 methaemoglobinaemia (adjusted mean difference = 0.01; 95% CI = [−0.21, 0.23]  $p = 0.90$ ).

370

371 Assuming a comparison between two primaquine doses whereby the higher dose results in  
372 half the risk of vivax recurrence with recurrence rates of 16% versus 8% (corresponding to  
373 primaquine doses of 0.5 versus 1 mg/kg over 7 days), a two-arm randomised trial aiming to  
374 show superiority would require a sample size of 256 individuals per group to achieve 80%  
375 power with a 5% false positive rate. In contrast, using day 7 methaemoglobin as a surrogate  
376 endpoint for vivax recurrence, the required sample size would be reduced by approximately  
377 42% ( $n = 148$  individuals per group) for high daily dose primaquine and could be even more for  
378 lower daily doses (Fig S7).

## 379 Discussion

380 In this systematic review and individual patient data meta-analysis, we confirm that higher  
381 primaquine-induced methaemoglobin concentrations on day 7 are associated with lower rates  
382 of *P. vivax* recurrence. This finding remained consistent across diverse study sites with varying  
383 populations and levels of transmission intensity. Our analysis found no indication of a  
384 differential predictive effect of day 7 methaemoglobin between the 7-day and 14-day  
385 primaquine regimens. Additionally, we observed a positive dose-response relationship  
386 between the daily weight-adjusted primaquine dose and day 7 methaemoglobin level.

387

388 Our findings are in line with recent estimates derived from Chu 2021 [10] (a secondary analysis  
389 of primaquine trial [9] data from the endemic northwest Thailand-Myanmar border) and  
390 Watson 2022 [4] (an individual patient data meta-analysis of tafenoquine trials). These  
391 estimates suggested that an increase of one percentage-point of day 7 methaemoglobinaemia  
392 was associated with an estimated reduction of 10% for primaquine and 20% for tafenoquine  
393 in the risk of *P. vivax* recurrence. While our analysis of primaquine included data from these  
394 studies such that estimates are not completely uncorrelated, additional data from different  
395 studies contributed most (nearly 70%) of our pooled dataset. Watson 2022 analysed vivax  
396 patients who received tafenoquine only (not included in this analysis). According to the White  
397 2022 review [8] of early experiments with primaquine or its analogues conducted more than  
398 70 years ago [30–32], 8-aminoquinoline analogues which resulted in less than 6% of  
399 methaemoglobinaemia during treatment showed reduced efficacy. All these results point  
400 towards higher methaemoglobin levels following treatment as indicative of increased  
401 antihypnozoite activity.

402

403 In the current analysis of G6PD normal patients, we observed no evidence of an association  
404 between the extent of early haemolysis on days 2–3 from baseline and day 7  
405 methaemoglobinaemia. This suggests that during the early days of illness, early haemolysis was  
406 primarily attributable to the acute parasitaemia and rehydration, rather than iatrogenic  
407 haemolysis caused by the primaquine active metabolites. The lower day 7 methaemoglobin

408 observed among younger patients may be partly explained by age-related enzyme immaturity  
409 and lower drug exposures [10]. The reason for lower methaemoglobin in older patients is less  
410 clear.

411  
412 Methaemoglobinaemia is a simple and readily measured surrogate endpoint that can be  
413 quantified within a week after starting administration of an 8-aminoquinoline and has potential  
414 to improve the efficiency of exploratory trials (drug-drug interactions, regimen optimisation,  
415 drug screening, dose optimisation). Current antirelapse clinical trials generally require patient  
416 follow up for many months to observe recurrent infections in order to have sufficient power  
417 to determine comparative efficacy of different treatment arms. Quantifying  
418 methaemoglobinaemia also has potential to be a useful approach for monitoring of trials and  
419 clinical practices at an individual level, serving as a surrogate marker for patient adherence.  
420 Although methaemoglobin is a surrogate endpoint for adherence to treatment, these results  
421 are not driven by adherence (90% of patients had fully supervised treatment). Early-phase  
422 studies, preceding more definitive efficacy trials, may stand to benefit the most from this novel  
423 endpoint [8]. Minimum required sample sizes could also be substantially reduced by using day  
424 7 methaemoglobin as an endpoint to improve cost-efficiency in conducting *P. vivax* trials. An  
425 ongoing study (NCT05788094) is currently using day 7 methaemoglobin as a secondary  
426 outcome to address the question of whether there is important drug-drug interaction between  
427 tafenoquine and chloroquine, DHA-piperaquine, or artemether-lumefantrine [33]. The recent  
428 INSPECTOR trial in Indonesia used DHA-piperaquine as the partner schizonticidal drug and  
429 lower than expected efficacy was observed [34] compared to trials with chloroquine [35-37].  
430 In the INSPECTOR study, tafenoquine plus DHA-piperaquine resulted in a median  
431 methaemoglobin of 1.3% (range = 0.7% to 3.7%), which was similar to DHA-piperaquine alone  
432 (median = 1.0%, range = 0.5% to 1.8%). In contrast, primaquine plus DHA-piperaquine  
433 produced more than twice as much methaemoglobin (median = 2.9%, range = 0.9% to 7.9%)  
434 and superior efficacy than the tafenoquine plus DHA-piperaquine arm.

435  
436 Our study has several limitations. Some patients missed measurements and some studies by  
437 design intentionally did not measure methaemoglobin on day 7. We addressed such missing  
438 data through linear interpolation, since methaemoglobin was recorded within two days before  
439 and/or after day 7. It is important to note that methaemoglobinaemia itself is an inherently  
440 noisy measure with substantial variation between patients, especially in patients receiving  
441 higher doses of 8-aminoquinoline treatment (Fig S7 shows higher methaemoglobin variability  
442 in higher dose groups). Improved reliability may be achieved by measuring methaemoglobin  
443 on both hands, incorporating multiple or repeated measurements, waiting longer, and blocking  
444 fluorescent light (e.g., turning off lights or covering hand when checking). In an upcoming  
445 analysis, we plan to explore alternative summary metrics for methaemoglobin levels using  
446 pharmacometric modelling [22]. This approach aims to capture exposures to primaquine's  
447 active metabolites and improve the overall robustness of our findings. There is also a possibility  
448 of less accurate methaemoglobin measurements by using CO-oximetry for patients with dark  
449 skin pigment [38].

450  
451 Only two studies [39, 40] were conducted in locations with negligible reinfection. Hence  
452 another limitation of our analysis is the uncertainty regarding the aetiology of recurrences for  
453 most of the patients, since there is currently no standardised method to differentiate between  
454 recrudescence, reinfection, and relapse [1]. It is important to note that the use of highly

455 efficacious schizonticides across the study sites makes recrudescence unlikely. Another  
456 potential source of error includes participants who might not have had hypnozoites in the liver  
457 to start with, thus having zero risk of relapse, despite primaquine inducing  
458 methaemoglobinaemia. However, this circumstance should bias the effect estimates towards  
459 the null (i.e., no association between day 7 methaemoglobinaemia and the risk of vivax  
460 recurrence), making our estimates conservative.

461  
462 The generalisability of our findings is constrained to the large majority of vivax malaria patients  
463 with normal G6PD activity ( $\geq 30\%$  G6PD activity or a negative qualitative test). For individuals  
464 with G6PD deficiency, blood methaemoglobin will not serve as a valid surrogate endpoint for  
465 antihypnozoite activity [8, 10]. In G6PD deficiency, primaquine does not induce clear  
466 methaemoglobinaemia [15, 41]. We currently lack sufficient data to explore how *CYP2D6*  
467 polymorphism impacts primaquine biotransformation to its active metabolites, as most  
468 patients were not genotyped [10]. The previous analysis by Chu et al suggested lower  
469 methaemoglobin in null *CYP2D6* metabolisers (activity score of 0) [8]. Intermediate  
470 metabolisers may also have lower active metabolites and higher relapse rates [6, 7]. Further  
471 research specifically targeted at these vulnerable populations is imperative.

472  
473 Our pooled dataset includes patients from various countries with different antimalarial  
474 treatment policies for their first-line schizonticides, each having different elimination kinetics  
475 and thus different durations of suppressive post-treatment prophylaxis. The use of time-to-  
476 event models in such contexts may introduce bias in favour of drugs with longer half-lives, such  
477 as chloroquine. However, our sensitivity analysis, employing logistic regression that included  
478 partner drug as a covariate, yielded similar predictive-effect estimates.

479  
480 In conclusion, in primaquine-treated G6PD normal individuals, the day 7 methaemoglobin can  
481 serve as a pharmacodynamic proxy for exposure to the biologically active metabolites of  
482 primaquine, making it a valid surrogate endpoint in G6PD normal patients with *P. vivax* malaria.  
483 The consistency of results observed in tafenoquine studies suggests a common drug-class  
484 phenomenon for 8-aminoquinolines. Direct comparisons of 8-aminoquinoline induced  
485 methaemoglobinaemia between primaquine and tafenoquine in future studies could prove  
486 useful. These findings collectively enhance our understanding of the causal mechanisms by  
487 which 8-aminoquinoline drugs exert their effects, facilitate drug discovery and regimen  
488 optimisation, and influence clinical practices.

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502 submission.

503

#### 504 **Author contributions**

505 IF, RJC, RNP, NJW, JKB, and JAW conceived the study, analysed and interpreted the data, as  
506 well as drafted the manuscript. IF, RJC, and JAW accessed and verified the data. NHC, NPJD,  
507 JAG, GCKWK, MVGL, AL, EJN, FN, APP, IS, WRJT, KT, RNP, NJW, JKB conceived and undertook  
508 the individual studies and enrolled the patients. All authors critically reviewed the study for  
509 intellectual content, revised the manuscript, and were responsible for the decision to submit  
510 for publication.

511

#### 512 **Data sharing**

513 Pseudonymised participant data used in this study can be accessed via the WorldWide  
514 Antimalarial Resistance Network ([wwarn.org](http://wwarn.org)). Requests for access will be reviewed by a data  
515 access committee to ensure that use of data protects the interests of the participants and  
516 researchers according to the terms of ethics approval and principles of equitable data sharing.  
517 Requests can be submitted by email to [malariaDAC@iddo.org](mailto:malariaDAC@iddo.org) via the data access form  
518 available at [https://www.wwarn.org/working-together/sharing-accessing-data/accessing-](https://www.wwarn.org/working-together/sharing-accessing-data/accessing-data)  
519 [data](https://www.wwarn.org/working-together/sharing-accessing-data/accessing-data). WWARN is registered with the Registry of Research Data Repositories  
520 (<https://www.re3data.org/>). Code for data analysis and visualisation is available at  
521 <https://github.com/ihsanfakil/methb7>.

522

#### 523 **Declaration of interests**

524 JAG and GCKWK are former employees of GSK and hold shares in GSK and AstraZeneca. GCKWK  
525 reports travel support from AstraZeneca. JKB and KT receive institutional research funding  
526 from Medicines for Malaria Venture. JKB reports GSK, Wellcome Trust, and Sanaria;  
527 participation on the US National Institutes of Health data safety monitoring board; and  
528 membership of the editorial board of *Travel Medicine and Infectious Disease* and the guidelines  
529 development group for malaria control and elimination, Global Malaria Programme, WHO. RJC,  
530 JKB, and RNP report contributions to Up-to-Date. All other authors declare no competing  
531 interests.

532

#### 533 **References**

534

- 535 1. White NJ. Determinants of relapse periodicity in *Plasmodium vivax* malaria. *Malaria*  
536 *Journal*. 2011;10(1):297.
- 537 2. Commons RJ, Simpson JA, Watson J, White NJ, Price RN. Estimating the Proportion of  
538 *Plasmodium vivax* Recurrences Caused by Relapse: A Systematic Review and Meta-  
539 Analysis. *The American Journal of Tropical Medicine and Hygiene*. 2020;103(3):1094-9.
- 540 3. Baird JK, Louisa M, Noviyanti R, Ekawati L, Elyazar I, Subekti D, et al. Association of Impaired  
541 Cytochrome P450 2D6 Activity Genotype and Phenotype With Therapeutic Efficacy of  
542 Primaquine Treatment for Latent *Plasmodium vivax* Malaria. *JAMA Network Open*.  
543 2018;1(4):e181449.

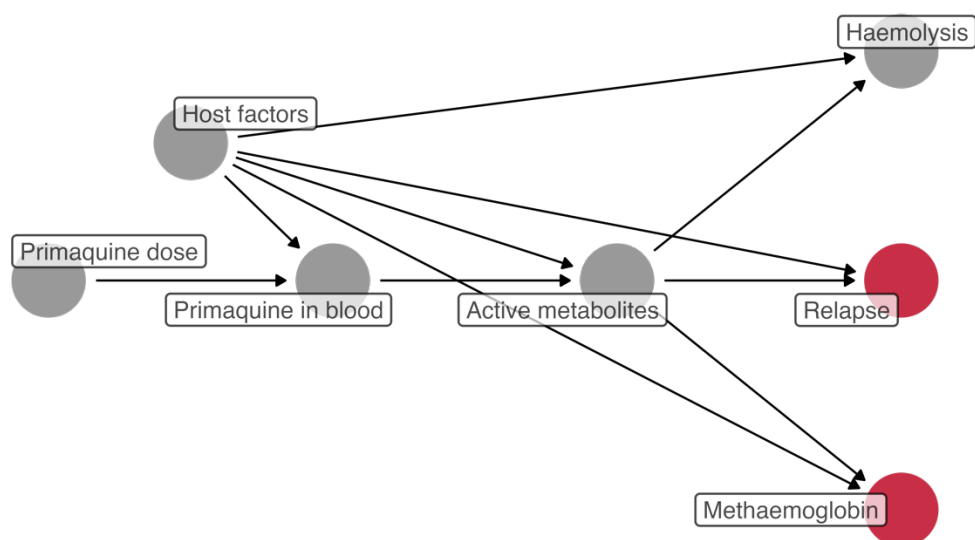
- 544 4. Watson JA, Commons RJ, Tarning J, Simpson JA, Llanos Cuentas A, Lacerda MV, et al. The  
545 clinical pharmacology of tafenoquine in the radical cure of *Plasmodium vivax* malaria: An  
546 individual patient data meta-analysis. *eLife*. 2022;11.
- 547 5. Baird JK. 8-Aminoquinoline Therapy for Latent Malaria. *Clinical Microbiology Reviews*.  
548 2019;32(4).
- 549 6. Pookmanee W, Thongthip S, Mungthin M, Sukasem C, Tankanitlert J, Chariyavilaskul P, et  
550 al. An increase in urinary primaquine and a reduction in urinary primaquine-5,6-  
551 orthoquinone in the Thai population with CYP2D6 reduced enzyme function. *Heliyon*.  
552 2024;10(2):e24351.
- 553 7. Bennett JW, Pybus BS, Yadava A, Tosh D, Sousa JC, McCarthy WF, et al. Primaquine Failure  
554 and Cytochrome P-450 2D6 in *Plasmodium vivax* Malaria. *New England Journal of*  
555 *Medicine*. 2013;369(14):1381-2.
- 556 8. White NJ, Watson JA, Baird JK. Methaemoglobinaemia and the radical curative efficacy of  
557 8-aminoquinoline antimalarials. *British Journal of Clinical Pharmacology*. 2022;88(6):2657-  
558 64.
- 559 9. Chu CS, Phyo AP, Turner C, Win HH, Poe NP, Yotyingaphiram W, et al. Chloroquine Versus  
560 Dihydroartemisinin-Piperaquine With Standard High-dose Primaquine Given Either for 7  
561 Days or 14 Days in *Plasmodium vivax* Malaria. *Clinical Infectious Diseases*.  
562 2019;68(8):1311-9.
- 563 10. Chu CS, Watson JA, Phyo AP, Win HH, Yotyingaphiram W, Thinraow S, et al. Determinants  
564 of Primaquine and Carboxyprimaquine Exposures in Children and Adults with *Plasmodium*  
565 *vivax* Malaria. *Antimicrobial Agents and Chemotherapy*. 2021;65(11):e0130221.
- 566 11. Ciani O, Manyara AM, Davies P, Stewart D, Weir CJ, Young AE, et al. A framework for the  
567 definition and interpretation of the use of surrogate endpoints in interventional trials.  
568 *eClinicalMedicine*. 2023;65:102283.
- 569 12. Fleming TR, DeMets DL. Surrogate end points in clinical trials: are we being misled? *Annals*  
570 *of internal medicine*. 1996;125(7):605-13.
- 571 13. Biomarkers and surrogate endpoints: Preferred definitions and conceptual framework.  
572 *Clinical Pharmacology & Therapeutics*. 2001;69(3):89-95.
- 573 14. Commons RJ, Thriemer K, Humphreys G, Suay I, Sibley CH, Guerin PJ, et al. The Vivax  
574 Surveyor: Online mapping database for *Plasmodium vivax* clinical trials. *The International*  
575 *Journal for Parasitology – Drugs and Drug Resistance*. 2017;7(2):181-90.
- 576 15. Pukrittayakamee S, Jittamala P, Watson JA, Hanboonkunupakarn B, Leungsinsiri P,  
577 Poovorawan K, et al. Pharmacometric assessment of primaquine induced haemolysis in  
578 glucose-6-phosphate dehydrogenase deficiency. *eLife*. 2023.
- 579 16. Infectious Diseases Data Observatory (IDDO). IDDO SDTM implementation manual 2023  
580 [Available from: <https://www.iddo.org/tools-and-resources/data-tools.>]

- 581 17. Stewart LA, Clarke M, Rovers M, Riley RD, Simmonds M, Stewart G, et al. Preferred  
582 Reporting Items for Systematic Review and Meta-Analyses of individual participant data:  
583 the PRISMA-IPD Statement. *JAMA*. 2015;313(16):1657-65.
- 584 18. Rajasekhar M, Simpson JA, Ley B, Edler P, Chu CS, Abreha T, et al. Primaquine dose and  
585 the risk of haemolysis in patients with uncomplicated *Plasmodium vivax* malaria: a  
586 systematic review and individual patient data meta-analysis. *The Lancet Infectious*  
587 *Diseases*. 2023.
- 588 19. Riley RD, Debray TPA, Fisher D, Hattle M, Marlin N, Hoogland J, et al. Individual participant  
589 data meta-analysis to examine interactions between treatment effect and participant-  
590 level covariates: Statistical recommendations for conduct and planning. *Statistics in*  
591 *Medicine*. 2020;39(15):2115-37.
- 592 20. Lee SJ, Stepniewska K, Anstey N, Ashley E, Barnes K, Binh TQ, et al. The relationship  
593 between the haemoglobin concentration and the haematocrit in *Plasmodium falciparum*  
594 malaria. *Malaria Journal*. 2008;7(1):149.
- 595 21. Hayden JA, van der Windt DA, Cartwright JL, Côté P, Bombardier C. Assessing bias in  
596 studies of prognostic factors. *Annals of Internal Medicine*. 2013;158(4):280-6.
- 597 22. WorldWide Antimalarial Resistance Network (WWARN). Primaquine methaemoglobin  
598 study group 2023 [Available from: [https://www.wwarn.org/working-together/study-](https://www.wwarn.org/working-together/study-groups/primaquine-methaemoglobin-study-group)  
599 [groups/primaquine-methaemoglobin-study-group.](https://www.wwarn.org/working-together/study-groups/primaquine-methaemoglobin-study-group)]
- 600 23. Battle KE, Karhunen MS, Bhatt S, Gething PW, Howes RE, Golding N, et al. Geographical  
601 variation in *Plasmodium vivax* relapse. *Malaria Journal*. 2014;13(1):144.
- 602 24. Battle KE, Lucas TCD, Nguyen M, Howes RE, Nandi AK, Twohig KA, et al. Mapping the global  
603 endemicity and clinical burden of *Plasmodium vivax*, 2000–17: a spatial and temporal  
604 modelling study. *The Lancet*. 2019;394(10195):332-43.
- 605 25. Carmona-Fonseca J. Vivax malaria in children: recurrences with standard total dose of  
606 primaquine administered in 3 vs. 7 days. *Iatreia*. 2010;23(1):10-20.
- 607 26. Chu CS, Phyo AP, Lwin KM, Win HH, San T, Aung AA, et al. Comparison of the cumulative  
608 efficacy and safety of chloroquine, artesunate, and chloroquine-primaquine in  
609 *Plasmodium vivax* malaria. *Clinical Infectious Diseases*. 2018;67(10):1543-9.
- 610 27. Fukuda MM, Krudsood S, Mohamed K, Green JA, Warrasak S, Noedl H, et al. A randomized,  
611 double-blind, active-control trial to evaluate the efficacy and safety of a three day course  
612 of tafenoquine monotherapy for the treatment of *Plasmodium vivax* malaria. *PLoS One*.  
613 2017;12(11):e0187376.
- 614 28. Ley B, Alam MS, Thriemer K, Hossain MS, Kibria MG, Auburn S, et al. G6PD deficiency and  
615 antimalarial efficacy for uncomplicated malaria in Bangladesh: a prospective observational  
616 study. *PloS One*. 2016;11(4):e0154015.

- 617 29. Solari-Soto L, Soto-Tarazona A, Mendoza-Requena D, Llanos-Cuentas A. Ensayo clínico del  
618 tratamiento de la malaria vivax con esquema acertado de primaquina comparado con el  
619 esquema tradicional. *Revista de la Sociedad Peruana de Medicina Interna*. 2002;15:196-9.
- 620 30. Alving AS, Pullman TN, Craige B, Jones R, Whorton CM, Eichelberger L. The clinical trial of  
621 eighteen analogues of pamaquin (plasmochin) in vivax malaria (Chesson strain). *The*  
622 *Journal of Clinical Investigation*. 1948;27(3):34-45.
- 623 31. Cooper WC, Myatt AV, Hernandez T, Jeffery GM, Coatney G. Studies in Human Malaria.  
624 XXXI. Comparison of Primaquine, Isopentaquine, SN-3883, and Pamaquine as Curative  
625 Agents against Ghesson Strain vivax Malaria. *Airier J Trap Med & Hyg*. 1953;2(6):949-57.
- 626 32. Edgcomb JH, Arnold J, Yount Jr E, Alving AS, Eichelberger L, Jeffery G, et al. Primaquine, SN  
627 13272, a new curative agent in vivax malaria: a preliminary report. *Journal of the National*  
628 *Malaria Society*. 1950;9(4):285-92.
- 629 33. ACT vs CQ With Tafenoquine for *P. vivax* Mono-infection (ACTQ) 2023 [Available from:  
630 [https://www.clinicaltrials.gov/study/NCT05788094?cond=NCT05788094&rank=1#collab](https://www.clinicaltrials.gov/study/NCT05788094?cond=NCT05788094&rank=1#collaborators-and-investigators)  
631 [orators-and-investigators.](https://www.clinicaltrials.gov/study/NCT05788094?cond=NCT05788094&rank=1#collaborators-and-investigators)]
- 632 34. Sutanto I, Soebandrio A, Ekawati LL, Chand K, Noviyanti R, Satyagraha AW, et al.  
633 Tafenoquine co-administered with dihydroartemisinin–piperaquine for the radical cure of  
634 *Plasmodium vivax* malaria (INSPECTOR): a randomised, placebo-controlled, efficacy and  
635 safety study. *The Lancet Infectious Diseases*. 2023.
- 636 35. Lacerda MVG, Llanos-Cuentas A, Krudsood S, Lon C, Saunders DL, Mohammed R, et al.  
637 Single-Dose Tafenoquine to Prevent Relapse of *Plasmodium vivax* Malaria. *New England*  
638 *Journal of Medicine*. 2019;380(3):215-28.
- 639 36. Llanos-Cuentas A, Lacerda MV, Rueangweerayut R, Krudsood S, Gupta SK, Kochar SK, et al.  
640 Tafenoquine plus chloroquine for the treatment and relapse prevention of *Plasmodium*  
641 *vivax* malaria (DETECTIVE): a multicentre, double-blind, randomised, phase 2b dose-  
642 selection study. *The Lancet*. 2014;383(9922):1049-58.
- 643 37. Llanos-Cuentas A, Lacerda MVG, Hien TT, Vélez ID, Namaik-Larp C, Chu CS, et al.  
644 Tafenoquine versus Primaquine to Prevent Relapse of *Plasmodium vivax* Malaria. *New*  
645 *England Journal of Medicine*. 2019;380(3):229-41.
- 646 38. Patterson S, Sandercock N, Verhovsek M. Understanding pulse oximetry in hematology  
647 patients: Hemoglobinopathies, racial differences, and beyond. *American Journal of*  
648 *Hematology*. 2022;97(12):1659-63.
- 649 39. Sutanto I, Tjahjono B, Basri H, Taylor WR, Putri FA, Meilia RA, et al. Randomized, Open-  
650 Label Trial of Primaquine against Vivax Malaria Relapse in Indonesia. *Antimicrobial Agents*  
651 *and Chemotherapy*. 2013;57(3):1128-35.
- 652 40. Nelwan EJ, Ekawati LL, Tjahjono B, Setiabudy R, Sutanto I, Chand K, et al. Randomized trial  
653 of primaquine hypnozoitocidal efficacy when administered with artemisinin-combined

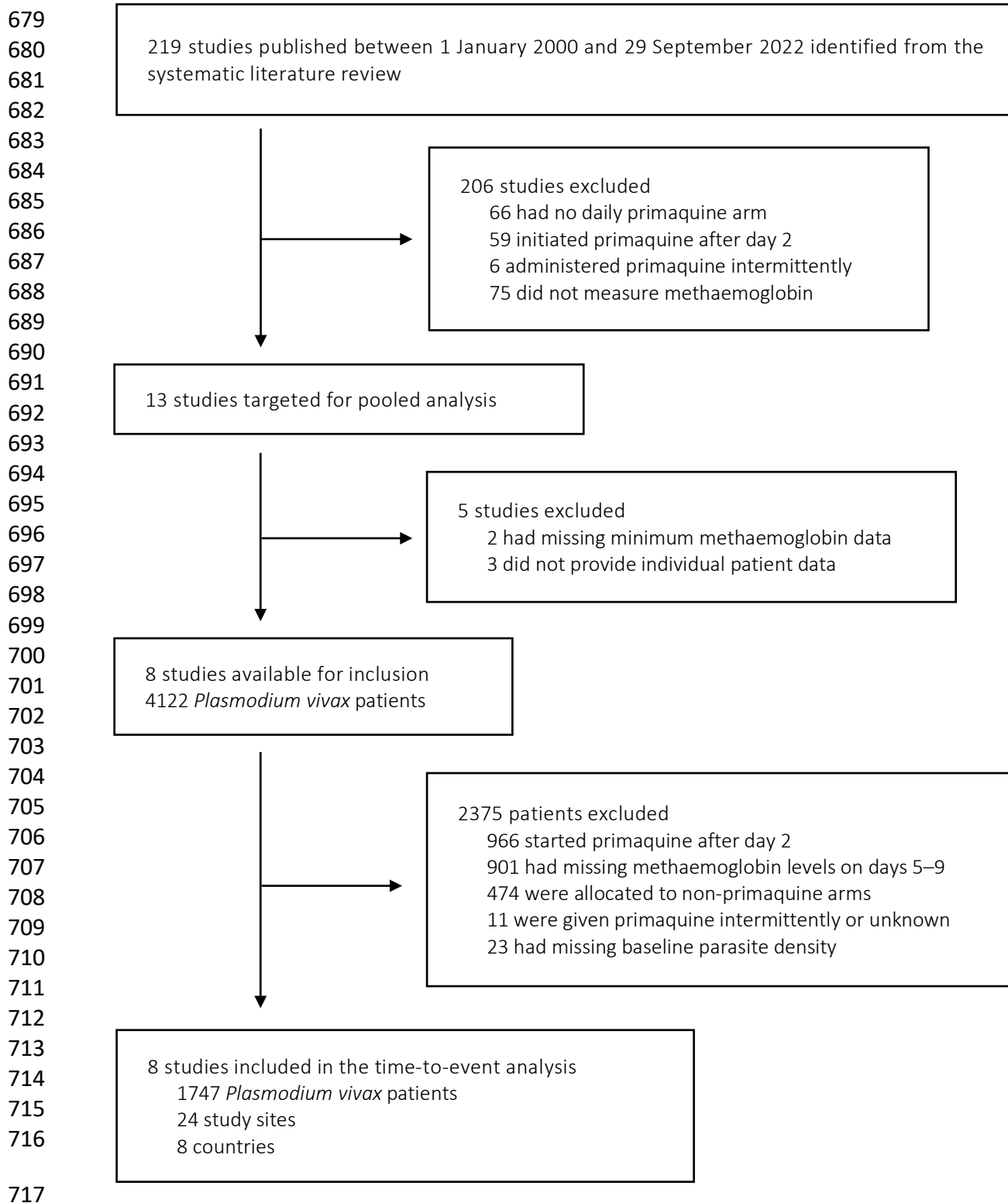
- 654 blood schizontocides for radical cure of *Plasmodium vivax* in Indonesia. BMC Medicine.  
655 2015;13:294.
- 656 41. Brewer GJ, Tarlov AR, Kellermeyer RW, Alving AS. The hemolytic effect of primaquine. XV.  
657 Role of methemoglobin. The Journal of Laboratory and Clinical Medicine. 1962;59(6):905-  
658 17.
- 659 42. Pasaribu AP, Chokejindachai W, Sirivichayakul C, Tanomsing N, Chavez I, Tjitra E, et al. A  
660 randomized comparison of dihydroartemisinin-piperaquine and artesunate-amodiaquine  
661 combined with primaquine for radical treatment of vivax malaria in Sumatera, Indonesia.  
662 The Journal of Infectious Diseases. 2013;208(11):1906-13.
- 663 43. Taylor WRJ, Thriemer K, Von Seidlein L, Yuentrakul P, Assawariyathipat T, Assefa A, et al.  
664 Short-course primaquine for the radical cure of *Plasmodium vivax* malaria: a multicentre,  
665 randomised, placebo-controlled non-inferiority trial. The Lancet. 2019;394(10202):929-  
666 38.
- 667

668 Table and figures



669  
670

671 **Figure 1. Directed acyclic graph showing our hypothesised causal relationships between**  
672 **primaquine-induced changes in blood methaemoglobin and *P. vivax* relapse.** Red nodes  
673 represent the outcomes of interest: relapse and blood methaemoglobin (measured on day 7,  
674 for example), between which the association was estimated. Under this causal model, blood  
675 methaemoglobin is a proxy measurement for the hypnozoontocidal activity of primaquine (but  
676 not on the causal pathway mediating the effect of primaquine on relapse). Host factors include  
677 but not limited to patient's genetics (e.g., those related to *CYP2D6* and *G6PD*), behaviours, age,  
678 immunity to *P. vivax*, and geographical location.



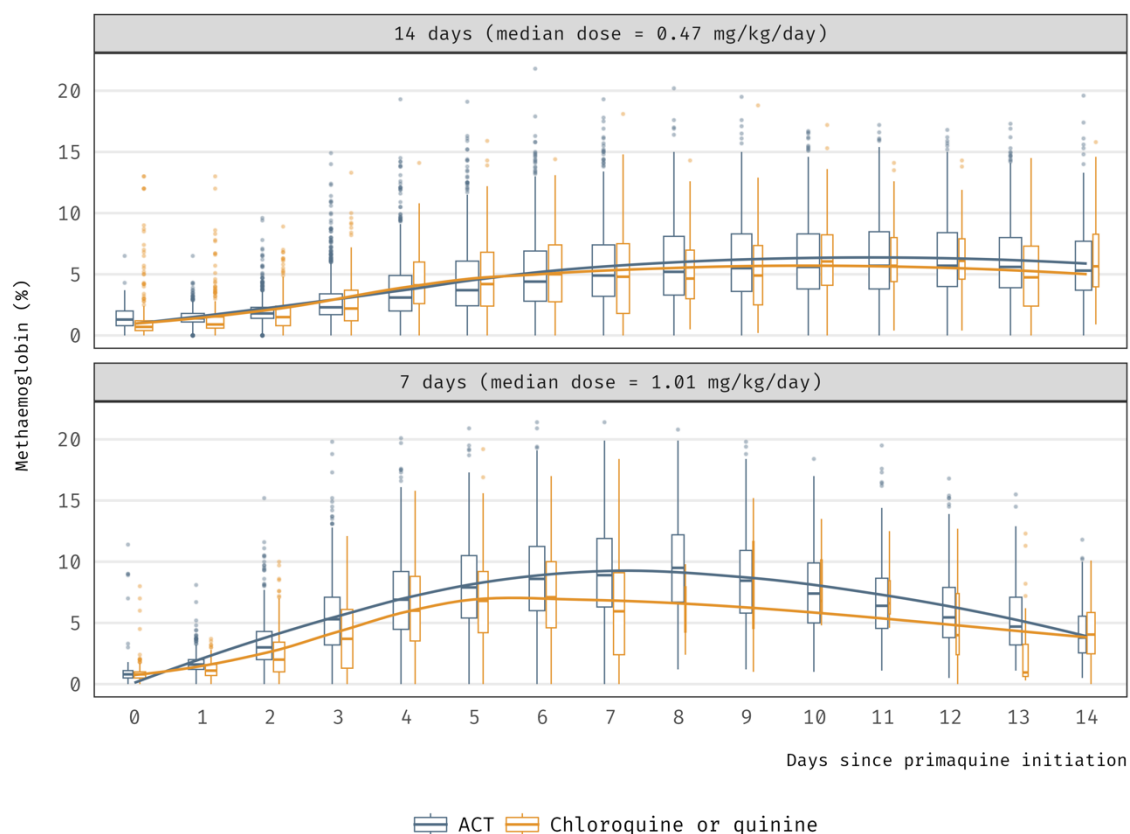
718 **Figure 2. Study and patient selection.** Databases systematically searched were from Medline,  
719 Embase, Web of Science, and the Cochrane Library. Patients included in the secondary analyses  
720 were subsets of the patients in the primary, time-to-event analysis.

Table 1. Demographic and patient characteristics at baseline.

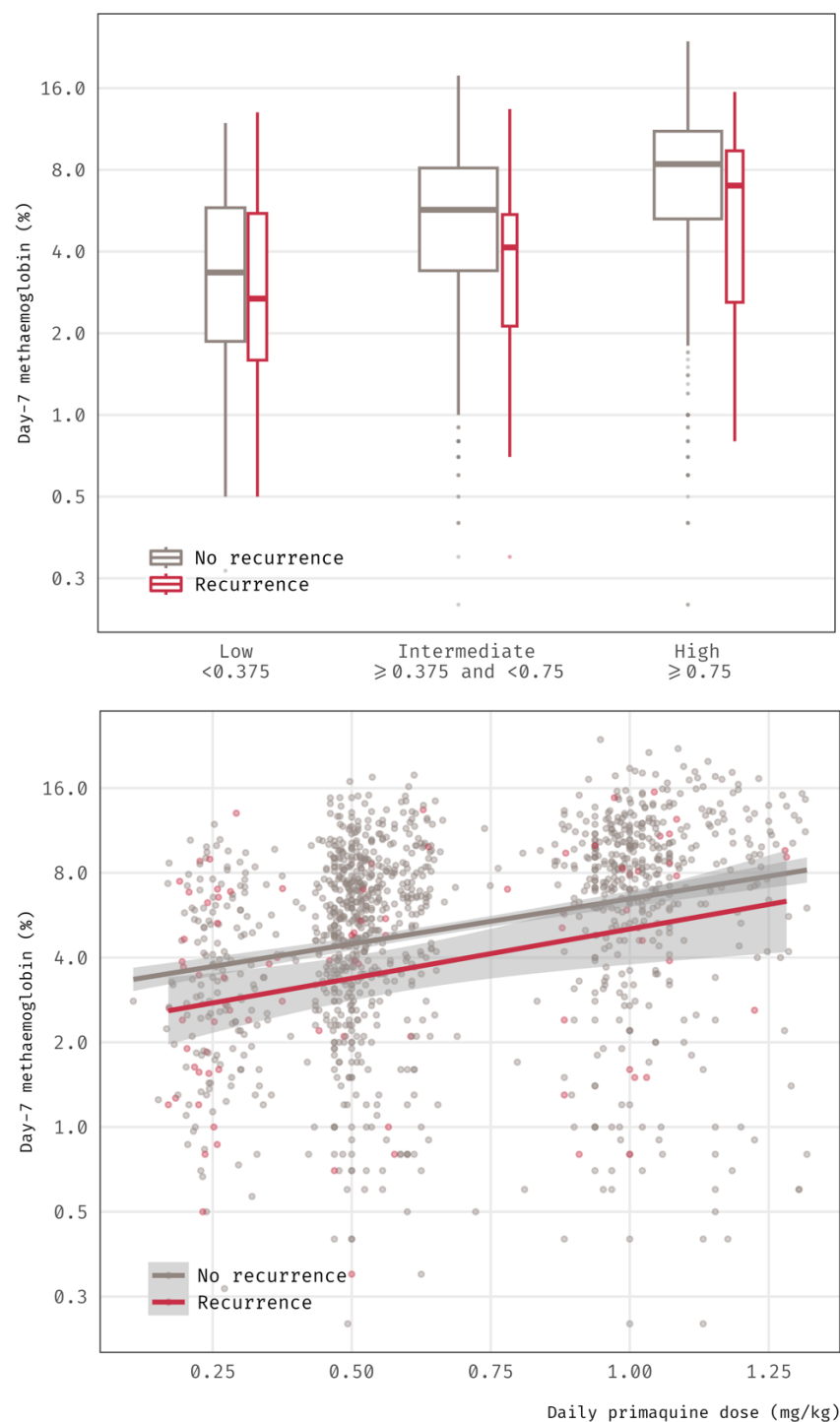
Characteristic	Overall	Study							
		Pasaribu 2013 [42]	Sutanto 2013 [39]	Llanos-Cuentas 2014 [36]	Nelwan 2015 [40]	Chu 2019 [9]	Lacerda 2019 [35]	Llanos-Cuentas 2019 [37]	Taylor 2019 [43]
<b>Number of patients</b>	1,747	303	38	50	120	578	42	84	532
<b>Region</b>									
Asia-Pacific	1,620 (93%)	303 (100%)	38 (100%)	22 (44%)	120 (100%)	578 (100%)	4 (9.5%)	23 (27%)	532 (100%)
Americas	124 (7.1%)	0 (0%)	0 (0%)	28 (56%)	0 (0%)	0 (0%)	35 (83%)	61 (73%)	0 (0%)
Africa	3 (0.2%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)	3 (7.1%)	0 (0%)	0 (0%)
<b>Relapse periodicity<sup>§</sup></b>									
Low	133 (7.6%)	0 (0%)	0 (0%)	34 (68%)	0 (0%)	0 (0%)	38 (90%)	61 (73%)	0 (0%)
High	1,614 (92%)	303 (100%)	38 (100%)	16 (32%)	120 (100%)	578 (100%)	4 (9.5%)	23 (27%)	532 (100%)
<b>Transmission intensity<sup>#</sup></b>									
Low	262 (15%)	0 (0%)	0 (0%)	16 (32%)	120 (100%)	0 (0%)	4 (9.5%)	17 (20%)	105 (20%)
Moderate	1,325 (76%)	303 (100%)	0 (0%)	6 (12%)	0 (0%)	578 (100%)	0 (0%)	11 (13%)	427 (80%)
High	160 (9.2%)	0 (0%)	38 (100%)	28 (56%)	0 (0%)	0 (0%)	38 (90%)	56 (67%)	0 (0%)
<b>Age (years)</b>	20 (12, 32)	13 (9, 25)	27 (25, 29)	34 (26, 46)	28 (25, 31)	20 (13, 32)	38 (24, 47)	36 (25, 50)	15 (10, 29)
<b>Age (years)</b>									
<5	89 (5.1%)	26 (8.6%)	0 (0%)	0 (0%)	0 (0%)	24 (4.2%)	0 (0%)	0 (0%)	39 (7.3%)
≥5 and <15	534 (31%)	144 (48%)	0 (0%)	0 (0%)	0 (0%)	168 (29%)	0 (0%)	0 (0%)	222 (42%)
≥15	1,124 (64%)	133 (44%)	38 (100%)	50 (100%)	120 (100%)	386 (67%)	42 (100%)	84 (100%)	271 (51%)
<b>Male</b>	1,116 (64%)	169 (56%)	38 (100%)	35 (70%)	120 (100%)	363 (63%)	30 (71%)	52 (62%)	309 (58%)
<b>Body weight (kg)</b>	48 (30, 59)	35 (21, 50)	65 (59, 72)	59 (49, 68)	69 (63, 74)	47 (33, 54)	64 (55, 72)	63 (55, 70)	43 (24, 54)
<b>Schizontocidal drug</b>									
Artesunate/Amodiaquine	146 (8.4%)	146 (48%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)
Artesunate/Pyronaridine	60 (3.4%)	0 (0%)	0 (0%)	0 (0%)	60 (50%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)
Dihydroartemisinin/Piperaquine	932 (53%)	157 (52%)	0 (0%)	0 (0%)	60 (50%)	290 (50%)	0 (0%)	0 (0%)	425 (80%)
Quinine	38 (2.2%)	0 (0%)	38 (100%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)
Chloroquine	571 (33%)	0 (0%)	0 (0%)	50 (100%)	0 (0%)	288 (50%)	42 (100%)	84 (100%)	107 (20%)

<b>Primaquine duration</b>									
7 days	553 (32%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)	289 (50%)	0 (0%)	0 (0%)	264 (50%)
14 days	1,194 (68%)	303 (100%)	38 (100%)	50 (100%)	120 (100%)	289 (50%)	42 (100%)	84 (100%)	268 (50%)
<b>Primaquine start</b>									
Day 0	1,496 (86%)	303 (100%)	27 (71%)	0 (0%)	120 (100%)	511 (88%)	3 (7.1%)	0 (0%)	532 (100%)
Day 1	181 (10%)	0 (0%)	9 (24%)	49 (98%)	0 (0%)	0 (0%)	39 (93%)	84 (100%)	0 (0%)
Day 2	70 (4.0%)	0 (0%)	2 (5.3%)	1 (2.0%)	0 (0%)	67 (12%)	0 (0%)	0 (0%)	0 (0%)
<b>Primaquine daily dose (mg/kg)</b>	0.52 (0.38, 0.95)	0.31 (0.27, 0.34)	0.51 (0.47, 0.59)	0.25 (0.22, 0.30)	0.53 (0.47, 0.60)	0.87 (0.50, 1.00)	0.23 (0.21, 0.27)	0.25 (0.23, 0.29)	0.66 (0.53, 1.03)
<b>Primaquine daily dose (mg/kg)</b>									
<0.375	436 (25%)	253 (83%)	0 (0%)	49 (98%)	0 (0%)	4 (0.7%)	42 (100%)	83 (99%)	5 (0.9%)
≥0.375 and <0.75	760 (44%)	50 (17%)	38 (100%)	1 (2.0%)	120 (100%)	282 (49%)	0 (0%)	1 (1.2%)	268 (50%)
≥0.75	551 (32%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)	292 (51%)	0 (0%)	0 (0%)	259 (49%)
<b>Primaquine dose calculation</b>									
Actual dosing	1,324 (76%)	0 (0%)	38 (100%)	50 (100%)	0 (0%)	578 (100%)	42 (100%)	84 (100%)	532 (100%)
Protocol dosing	423 (24%)	303 (100%)	0 (0%)	0 (0%)	120 (100%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)
<b>Primaquine supervision</b>									
Fully supervised	1,571 (90%)	303 (100%)	38 (100%)	0 (0%)	120 (100%)	578 (100%)	0 (0%)	0 (0%)	532 (100%)
Partially supervised	176 (10%)	0 (0%)	0 (0%)	50 (100%)	0 (0%)	0 (0%)	42 (100%)	84 (100%)	0 (0%)
<b>Parasite density (asexual parasites/μl)</b>	2,515 (617, 7,515)	760 (320, 2,980)	2,928 (784, 4,496)	4,631 (1,993, 8,580)	872 (144, 2,464)	4,517 (1,440, 13,565)	3,471 (1,589, 8,348)	5,079 (1,913, 12,700)	2,215 (551, 7,500)
<b>Haemoglobin (g/dl)</b>	12.40 (11.30, 13.70)	11.90 (10.80, 12.80)	14.20 (13.10, 14.67)	NA (NA, NA)	NA (NA, NA)	12.50 (11.40, 13.70)	NA (NA, NA)	NA (NA, NA)	12.65 (11.50, 13.90)
Number of missing data	212	0	38	28	120	0	11	15	0
<b>Day-7 methaemoglobin (%)</b>	6.0 (3.3, 9.0)	3.8 (2.6, 5.6)	5.6 (4.1, 7.5)	2.5 (1.5, 6.0)	5.5 (3.5, 8.2)	6.5 (4.0, 8.9)	2.4 (1.5, 6.1)	1.4 (1.2, 7.3)	7.1 (3.6, 11.0)
Number of missing data	245	9	3	38	6	68	37	77	7
<b>Manufacturer of primaquine tablets</b>	–	Phapros	Shin Poon	Sanofi	Sanofi	Thai Government	Sanofi	Sanofi	Centurion Laboratories

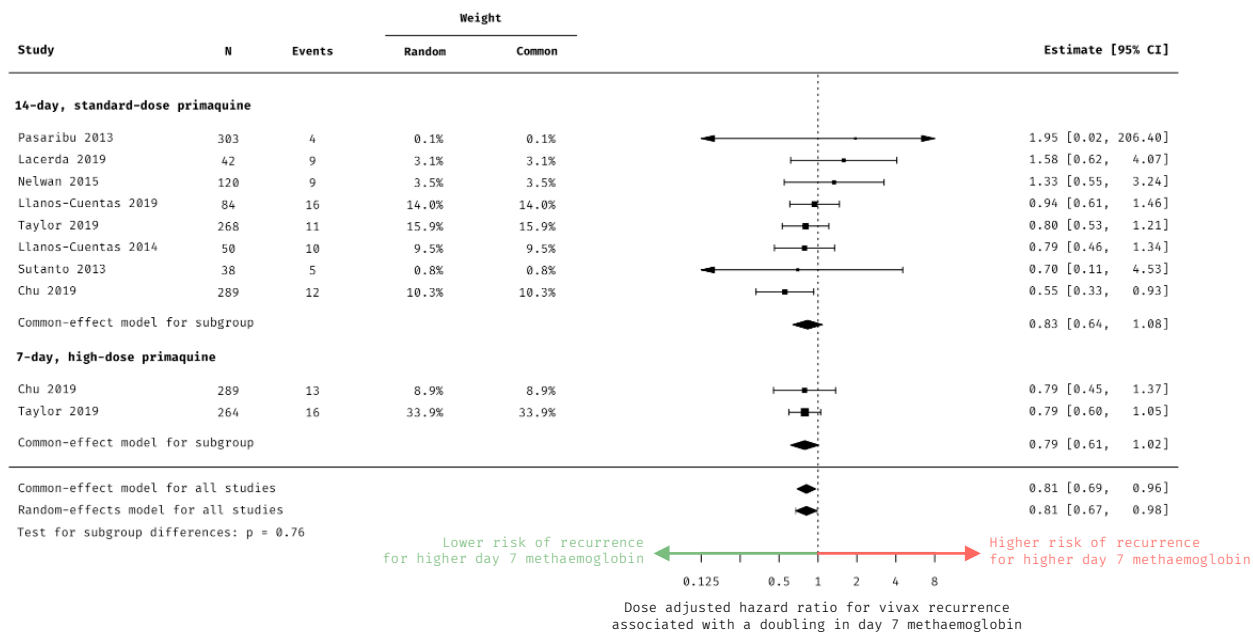
Numbers are in median (first quartile, third quartile) or frequency (percentage). NA not available, ACT artemisinin-based combination therapy, g gram, dl decilitre, mg milligram, μl microlitre, kg kilogram. § Relapse periodicity was categorised as high (median relapse periodicity of 47 days or less) and low (median relapse periodicity of more than 47) [23]; # Transmission intensity was categorised as low (<1 case per 1000 person-years), moderate (1 case to <10 cases per 1000 person-years), and high (≥10 cases per 1000 person-years) according to subnational malaria incidence estimates for the median year of study enrolment [24]. Missing day 7 methaemoglobin were linearly imputed with methaemoglobin data on ±2 days. Patients with missing haemoglobin or haematocrit data were excluded for the haemolysis sub-dataset.



**Figure 3. Dynamics of primaquine-induced increases in blood methaemoglobin over time, stratified by primaquine regimen and schizontocidal drug.** Methaemoglobin levels increased after starting primaquine in both regimens, usually reaching a maximum after about a week. Methaemoglobin increased at a faster rate among the 7-day regimen reflecting the higher daily dose taken and generally methaemoglobin started to decrease during the second week, when primaquine was no longer administered. Meanwhile, after peaking at also day 7 for the 14-day regimen, methaemoglobin appears to be at a more constant level during the second week. Box width is proportional to the square root of the number of patients. ACT artemisinin-based combination therapy (artesunate/amodiaquine, artesunate/pyronaridine, dihydroartemisinin/piperaquine).



**Figure 4. Day 7 blood methaemoglobin (%) as a function of daily mg/kg primaquine dose and *Plasmodium vivax* recurrence status.** There was an increasing trend of day 7 methaemoglobin as daily primaquine dose increased among patients with at least 120 days of follow up. Patients who developed *P. vivax* recurrences typically had lower day 7 methaemoglobin levels. Vertical axis is show on the logarithmic scale. Box width is proportional to the square root of the number of patients.



**Figure 5. Forest plot.** Common-effect and random-effects models converged to comparable estimates. Horizontal axis for dose-adjusted hazard ratios is shown on the logarithmic scale.

## Supporting Information

### List S1. Systematic search terms for the databases

Vivax AND (artefenomel OR arterolane OR amodiaquine OR atovaquone OR artemisinin OR arteether OR artesunate OR artemether OR artemotil OR azithromycin OR artekin OR chloroquine OR chlorproguanil OR cycloguanil OR clindamycin OR coartem OR dapsone OR dihydroartemisinin OR duo-cotecxin OR doxycycline OR halofantrine OR lumefantrine OR lariam OR malarone OR mefloquine OR naphthoquine OR naphthoquinone OR piperaquine OR primaquine OR proguanil OR pyrimethamine OR pyronaridine OR proguanil OR quinidine OR quinine OR riamet OR sulphadoxine OR tetracycline OR tafenoquine)

Table S1. PRISMA-IPD checklist

PRISMA-IPD Section/topic	Item No	Checklist item	Reported on page
<b>Title</b>			
Title	1	Identify the report as a systematic review and meta-analysis of individual participant data.	1
<b>Abstract</b>			
Structured summary	2	Provide a structured summary including as applicable:	2
		<b>Background:</b> state research question and main objectives, with information on participants, interventions, comparators and outcomes.	
		<b>Methods:</b> report eligibility criteria; data sources including dates of last bibliographic search or elicitation, noting that IPD were sought; methods of assessing risk of bias.	
		<b>Results:</b> provide number and type of studies and participants identified and number (%) obtained; summary effect estimates for main outcomes (benefits and harms) with confidence intervals and measures of statistical heterogeneity. Describe the direction and size of summary effects in terms meaningful to those who would put findings into practice.	
		<b>Discussion:</b> state main strengths and limitations of the evidence, general interpretation of the results and any important implications.	
<b>Other:</b> report primary funding source, registration number and registry name for the systematic review and IPD meta-analysis.			
<b>Introduction</b>			
Rationale	3	Describe the rationale for the review in the context of what is already known.	3
Objectives	4	Provide an explicit statement of the questions being addressed with reference, as applicable, to participants, interventions, comparisons, outcomes and study design (PICOS). Include any hypotheses that relate to particular types of participant-level subgroups.	3
<b>Methods</b>			
Protocol and registration	5	Indicate if a protocol exists and where it can be accessed. If available, provide registration information including registration number and registry name. Provide publication details, if applicable.	4
Eligibility criteria	6	Specify inclusion and exclusion criteria including those relating to participants, interventions, comparisons, outcomes, study design and characteristics (e.g. years when conducted, required minimum follow-up). Note whether these were applied at the study or individual level i.e. whether eligible participants were included (and ineligible participants excluded) from a study that included a wider population than specified by the review inclusion criteria. The rationale for criteria should be stated.	4
Identifying studies -	7	Describe all methods of identifying published and unpublished studies including, as applicable: which bibliographic databases were searched with dates of coverage; details of any hand searching including of conference proceedings; use of study registers	4

information sources		and agency or company databases; contact with the original research team and experts in the field; open adverts and surveys. Give the date of last search or elicitation.	
Identifying studies - search	8	Present the full electronic search strategy for at least one database, including any limits used, such that it could be repeated.	24
Study selection processes	9	State the process for determining which studies were eligible for inclusion.	4
Data collection processes	10	Describe how IPD were requested, collected and managed, including any processes for querying and confirming data with investigators. If IPD were not sought from any eligible study, the reason for this should be stated (for each such study). If applicable, describe how any studies for which IPD were not available were dealt with. This should include whether, how and what aggregate data were sought or extracted from study reports and publications (such as extracting data independently in duplicate) and any processes for obtaining and confirming these data with investigators.	4
Data items	11	Describe how the information and variables to be collected were chosen. List and define all study level and participant level data that were sought, including baseline and follow-up information. If applicable, describe methods of standardising or translating variables within the IPD datasets to ensure common scales or measurements across studies.	4
IPD integrity	A1	Describe what aspects of IPD were subject to data checking (such as sequence generation, data consistency and completeness, baseline imbalance) and how this was done.	4–6
Risk of bias assessment in individual studies.	12	Describe methods used to assess risk of bias in the individual studies and whether this was applied separately for each outcome. If applicable, describe how findings of IPD checking were used to inform the assessment. Report if and how risk of bias assessment was used in any data synthesis.	6
Specification of outcomes and effect measures	13	State all treatment comparisons of interests. State all outcomes addressed and define them in detail. State whether they were pre-specified for the review and, if applicable, whether they were primary/main or secondary/additional outcomes. Give the principal measures of effect (such as risk ratio, hazard ratio, difference in means) used for each outcome.	4–6
Synthesis methods	14	Describe the meta-analysis methods used to synthesise IPD. Specify any statistical methods and models used. Issues should include (but are not restricted to): <ul style="list-style-type: none"> <li>• Use of a one-stage or two-stage approach.</li> <li>• How effect estimates were generated separately within each study and combined across studies (where applicable).</li> <li>• Specification of one-stage models (where applicable) including how clustering of patients within studies was accounted for.</li> <li>• Use of fixed or random effects models and any other model assumptions, such as proportional hazards.</li> <li>• How (summary) survival curves were generated (where applicable).</li> <li>• Methods for quantifying statistical heterogeneity (such as <math>I^2</math> and <math>t^2</math>).</li> </ul>	4–6

		<ul style="list-style-type: none"> <li>How studies providing IPD and not providing IPD were analysed together (where applicable).</li> <li>How missing data within the IPD were dealt with (where applicable).</li> </ul>	
Exploration of variation in effects	A2	If applicable, describe any methods used to explore variation in effects by study or participant level characteristics (such as estimation of interactions between effect and covariates). State all participant-level characteristics that were analysed as potential effect modifiers, and whether these were pre-specified.	5–6
Risk of bias across studies	15	Specify any assessment of risk of bias relating to the accumulated body of evidence, including any pertaining to not obtaining IPD for particular studies, outcomes or other variables.	6
Additional analyses	16	Describe methods of any additional analyses, including sensitivity analyses. State which of these were pre-specified.	5–6
<b>Results</b>			
Study selection and IPD obtained	17	Give numbers of studies screened, assessed for eligibility, and included in the systematic review with reasons for exclusions at each stage. Indicate the number of studies and participants for which IPD were sought and for which IPD were obtained. For those studies where IPD were not available, give the numbers of studies and participants for which aggregate data were available. Report reasons for non-availability of IPD. Include a flow diagram.	6, 7, 18
Study characteristics	18	For each study, present information on key study and participant characteristics (such as description of interventions, numbers of participants, demographic data, unavailability of outcomes, funding source, and if applicable duration of follow-up). Provide (main) citations for each study. Where applicable, also report similar study characteristics for any studies not providing IPD.	6, 7, 19, 20, 32–35
IPD integrity	A3	Report any important issues identified in checking IPD or state that there were none.	7, 34
Risk of bias within studies	19	Present data on risk of bias assessments. If applicable, describe whether data checking led to the up-weighting or down-weighting of these assessments. Consider how any potential bias impacts on the robustness of meta-analysis conclusions.	6, 7, 30
Results of individual studies	20	For each comparison and for each main outcome (benefit or harm), for each individual study report the number of eligible participants for which data were obtained and show simple summary data for each intervention group (including, where applicable, the number of events), effect estimates and confidence intervals. These may be tabulated or included on a forest plot.	23
Results of syntheses	21	Present summary effects for each meta-analysis undertaken, including confidence intervals and measures of statistical heterogeneity. State whether the analysis was pre-specified, and report the numbers of studies and participants and, where applicable, the number of events on which it is based.	8, 9
		When exploring variation in effects due to patient or study characteristics, present summary interaction estimates for each characteristic examined, including confidence intervals and measures of statistical heterogeneity. State whether the analysis was pre-specified. State whether any interaction is consistent across trials.	

		Provide a description of the direction and size of effect in terms meaningful to those who would put findings into practice.	
Risk of bias across studies	22	Present results of any assessment of risk of bias relating to the accumulated body of evidence, including any pertaining to the availability and representativeness of available studies, outcomes or other variables.	30
Additional analyses	23	Give results of any additional analyses (e.g. sensitivity analyses). If applicable, this should also include any analyses that incorporate aggregate data for studies that do not have IPD. If applicable, summarise the main meta-analysis results following the inclusion or exclusion of studies for which IPD were not available.	8, 9
<b>Discussion</b>			
Summary of evidence	24	Summarise the main findings, including the strength of evidence for each main outcome.	9
Strengths and limitations	25	Discuss any important strengths and limitations of the evidence including the benefits of access to IPD and any limitations arising from IPD that were not available.	10, 11
Conclusions	26	Provide a general interpretation of the findings in the context of other evidence.	9, 10
Implications	A4	Consider relevance to key groups (such as policy makers, service providers and service users). Consider implications for future research.	11
<b>Funding</b>			
Funding	27	Describe sources of funding and other support (such as supply of IPD), and the role in the systematic review of those providing such support.	11, 12

## List S2. Signalling questions for risk of bias assessment using the QUIPS tool adapted to the current analysis

Domain 1: The study sample represents the population of interest on key characteristics, sufficient to limit potential bias of the observed relationship between the predictive factor and outcome.

- The source population or population of interest is adequately described.
- The baseline study sample (i.e., individuals entering the study) is adequately described.
- The sampling frame and recruitment are adequately described, including methods to identify the sample sufficient to limit potential bias.
- Period of recruitment is adequately described.
- Place of recruitment (setting, level of endemicity, geographic location) are adequately described.
- Inclusion and exclusion criteria are adequately described).

Domain 2: Loss to follow up (from baseline sample to study population analysed) is not associated with certain characteristics (i.e., the study data adequately represent the sample) sufficient to limit potential bias to the observed relationship between predictive factor and outcome.

- Response rate (i.e., proportion of study sample completing the study and providing outcome data) is adequate.
- Attempts to collect information on participants who dropped out of the study are described.
- Reasons for loss to follow up are provided.
- Participants lost to follow up are adequately described.
- There are no important differences between participants who completed the study and those who did not.

Domain 3: Predictive factor and drug intervention are adequately measured in study participants to sufficiently limit potential bias.

- A clear definition or description of the primaquine regimen and measured methaemoglobin is provided (e.g., including dose, level, duration of exposure, and clear specification of the method of measurement).
- Adequately accurate and reliable measurement of primaquine doses and methaemoglobin concentrations to limit misclassification bias.
- Continuous variables are reported, or clinically relevant cut points (i.e., not data-dependent) are used.
- Method and setting of methaemoglobin measurement are the same for all study participants.
- Adequate proportion of the study sample has complete methaemoglobin data.
- Adequate adherence or supervision of primaquine administration.

Domain 4: Outcome of interest is adequately measured in study participants to sufficiently limit potential bias.

- A clear definition of the outcome is provided, including duration of follow up.
- Method of outcome measurement used is adequately accurate and reliable to limit misclassification bias.
- Method and setting of outcome measurement are the same for all study participants.

Table S2. Risk of bias assessment

Study	QUIPS domain			
	Study participant	Study attrition	Predictive factor measurement	Outcome measurement
Pasaribu 2013 [42]	Low	Low	Low	Low
Sutanto 2013 [39]	Low	Low	Low	Low
Llanos-Cuentas 2014 [36]	Low	Low	Moderate	Low
Nelwan 2015 [40]	Low	Low	Low	Low
Chu 2019 [9]	Low	Low	Low	Low
Lacerda 2019 [35]	Low	Moderate	Moderate	Low
Llanos-Cuentas 2019 [37]	Low	Low	Low	Low
Taylor 2019 [43]	Low	Moderate	Moderate	Low

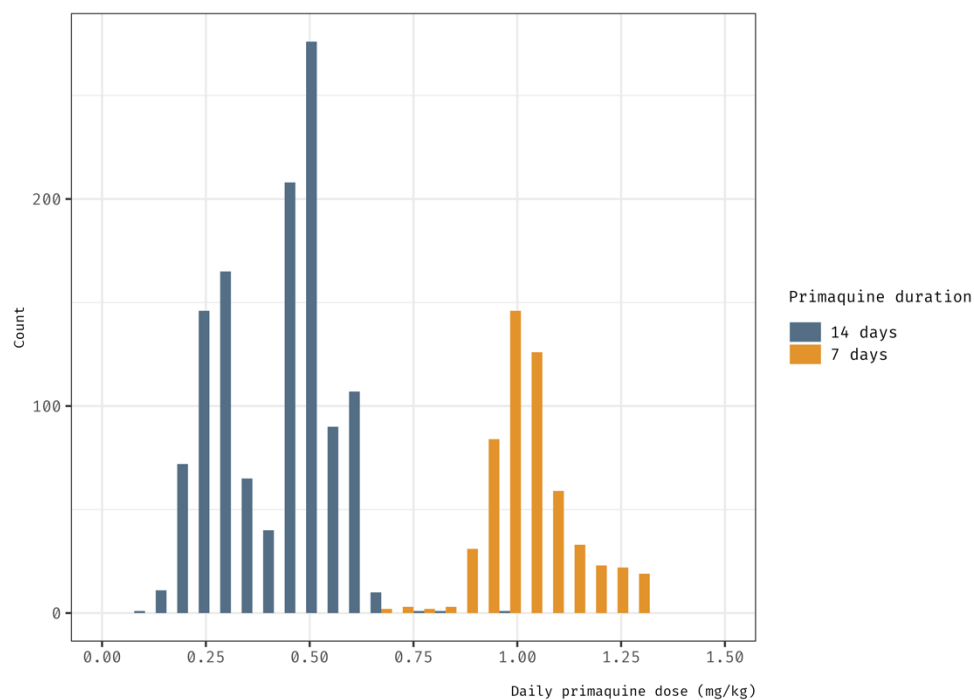


**Figure S1. Study sites that contributed to the pooled data in this individual patient data meta-analysis. Red bubble represents a study site with size proportional to the square root of the number of patients.**

Table S3. Studies included in analysis

Paper	Study site	Country	Region	Latitude	Longitude	Year start	Year end	MAP incidence rate (per 1000 persons)	Transmission intensity#	Relapse periodicity§
<b>Pasaribu 2013 [42]</b>	Tanjung Leidong	Indonesia	Asia-Pacific	2.77	99.98	2011	2011	2.75	Moderate	High
<b>Sutanto 2013 [39]</b>	Lumajang	Indonesia	Asia-Pacific	-8.13	113.22	2010	2011	36.88*	High	High
<b>Llanos-Cuentas 2014 [36]</b>	Bangkok	Thailand	Asia-Pacific	13.76	100.50	2011	2013	0.16	Low	High
<b>Llanos-Cuentas 2014 [36]</b>	Mae Sot	Thailand	Asia-Pacific	16.72	98.58	2011	2013	3.07	Moderate	High
<b>Llanos-Cuentas 2014 [36]</b>	Lucknow	India	Asia-Pacific	26.85	80.95	2011	2013	2.84	Moderate	Low
<b>Llanos-Cuentas 2014 [36]</b>	Bikaner	India	Asia-Pacific	28.02	73.31	2011	2013	2.85	Moderate	Low
<b>Llanos-Cuentas 2014 [36]</b>	Iquitos	Peru	Americas	-3.74	-73.25	2011	2013	40.49	High	Low
<b>Llanos-Cuentas 2014 [36]</b>	Manaus	Brazil	Americas	-3.12	-60.02	2011	2013	42.81	High	Low
<b>Nelwan 2015 [40]</b>	Sragen	Indonesia	Asia-Pacific	-7.42	111.02	2013	2013	42.44*	High	High
<b>Chu 2019 [9]</b>	Mae Sot	Thailand	Asia-Pacific	16.72	98.58	2012	2014	3.09	Moderate	High
<b>Lacerda 2019 [35]</b>	Manaus	Brazil	Americas	-3.12	-60.02	2013	2016	41.14	High	Low
<b>Lacerda 2019 [35]</b>	Porto Velho	Brazil	Americas	-8.76	-63.90	2013	2016	9.23	Moderate	Low
<b>Lacerda 2019 [35]</b>	Jimma	Ethiopia	Africa	7.67	36.84	2013	2016	40.53	High	Low
<b>Lacerda 2019 [35]</b>	Gondar	Ethiopia	Africa	12.60	37.45	2013	2016	6.72	Moderate	Low
<b>Lacerda 2019 [35]</b>	Mae Sot	Thailand	Asia-Pacific	16.72	98.58	2013	2016	0.64	Low	High
<b>Llanos-Cuentas 2019 [37]</b>	Manaus	Brazil	Americas	-3.12	-60.02	2015	2016	18.55	High	Low
<b>Llanos-Cuentas 2019 [37]</b>	Monteira	Colombia	Americas	8.75	-75.88	2015	2016	5.36	Moderate	Low
<b>Llanos-Cuentas 2019 [37]</b>	Cali	Colombia	Americas	3.45	-76.53	2015	2016	1.93	Moderate	Low
<b>Llanos-Cuentas 2019 [37]</b>	Iquitos	Peru	Americas	-3.74	-73.25	2015	2016	58.05	High	Low
<b>Llanos-Cuentas 2019 [37]</b>	Umphang	Thailand	Asia-Pacific	15.88	98.92	2015	2016	1.08	Moderate	High
<b>Llanos-Cuentas 2019 [37]</b>	Mae Sot	Thailand	Asia-Pacific	16.72	98.58	2015	2016	1.08	Moderate	High
<b>Llanos-Cuentas 2019 [37]</b>	Ho Chi Minh City	Vietnam	Asia-Pacific	10.82	106.63	2015	2016	0.01	Low	High
<b>Taylor 2019 [43]</b>	Dak O	Vietnam	Asia-Pacific	12.00	107.50	2015	2017	0.24	Low	High
<b>Taylor 2019 [43]</b>	Hanura	Indonesia	Asia-Pacific	-5.53	105.24	2015	2017	1.01	Moderate	High

MAP malaria atlas project. § Relapse periodicity was categorised as high (median relapse periodicity of 47 days or less) and low (median relapse periodicity of more than 47) [23]; # Transmission intensity was categorised as low (<1 case per 1000 person-years), moderate (1 case to <10 cases per 1000 person-years), and high (≥10 cases per 1000 person-years) according to subnational malaria incidence estimates for the median year of study enrolment [24]. \*Based on the location where patients were infected by *P. vivax* in Indonesian Papua.



**Figure S2. Distribution of weight-adjusted primaquine daily dose by primaquine regimen.** In the 14-day primaquine regimen, the observed two peaks reflect the targeted total primaquine dose of 3.5 and 7 mg (base) per kg body weight.

Table S4. Studies that were eligible for analysis but not included in the pooled data

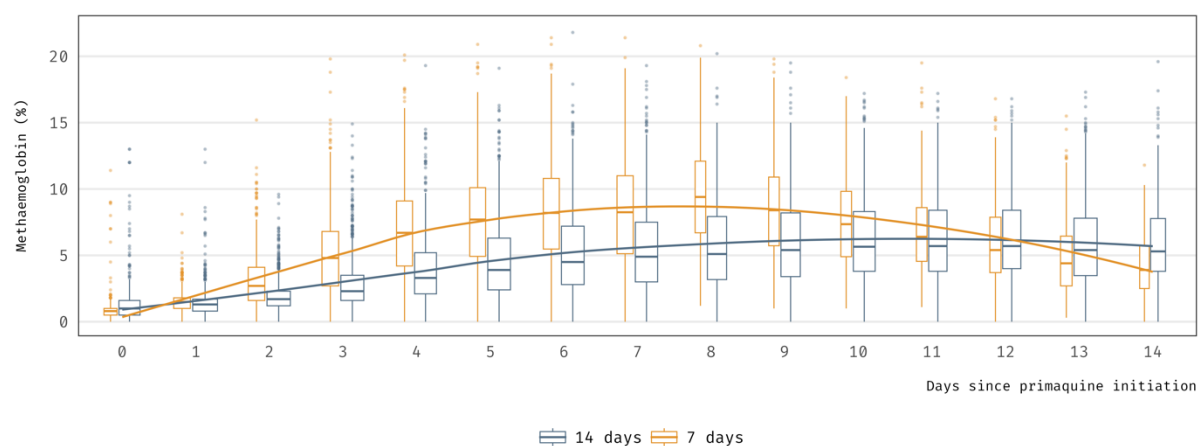
Characteristic	Study				
	Solari Soto [29]	Carmona-Fonseca [25]	Ley [28]	Fukuda [27]	Chu [26]
<b>Year published</b>	2002	2010	2016	2017	2018
<b>Number of treatment arms</b>	2	2	1	2	3
<b>Number of sites</b>	1	1	2	1	1
<b>Region</b>	Americas	Americas	Asia-Pacific	Asia-Pacific	Asia-Pacific
<b>Country</b>	Peru	Colombia	Bangladesh	Thailand	Thailand
<b>Follow-up (days)</b>	60	120	28	120	365
<b>Randomised</b>	Yes	Yes	No	Yes	Yes
<b>Recruitment period</b>	1998–1999	2005–2008	2014–2015	2003–2005	2010–2012
<b>Treatment arms</b>	(1) Chloroquine + 14-day, low-dose primaquine, (2) Chloroquine + 7-day, low-dose primaquine	(1) Chloroquine + 7-day, low-dose primaquine, (2) Chloroquine + 3-day, low-dose primaquine	Chloroquine + 14-day, low-dose primaquine	(1) Chloroquine + 14-day, low-dose primaquine, (2) Tafenoquine	(1) Artesunate, (2) Chloroquine, (3) Chloroquine + 14-day, high-dose primaquine
<b><i>P. vivax</i> patients enrolled</b>	60	79	66	70	644
<b>Treated with primaquine</b>	60	79	66	24	198
<b>Supervision</b>	Yes	Yes	Yes	Yes	Yes
<b>Sex (primaquine receiving arm)</b>	57% male	Mostly male	62% male	83% male	64% male
<b>Age (in years, primaquine receiving arm)</b>	Average = 26.5	Range = 10–17	Median = 18 (mono-infection), 14 (mixed)	Median = 30	Median = 18
<b>Reason for exclusion</b>	No response from investigators	Data not available	Missing minimum data	Data not provided	Missing minimum data

IPD individual patient data.

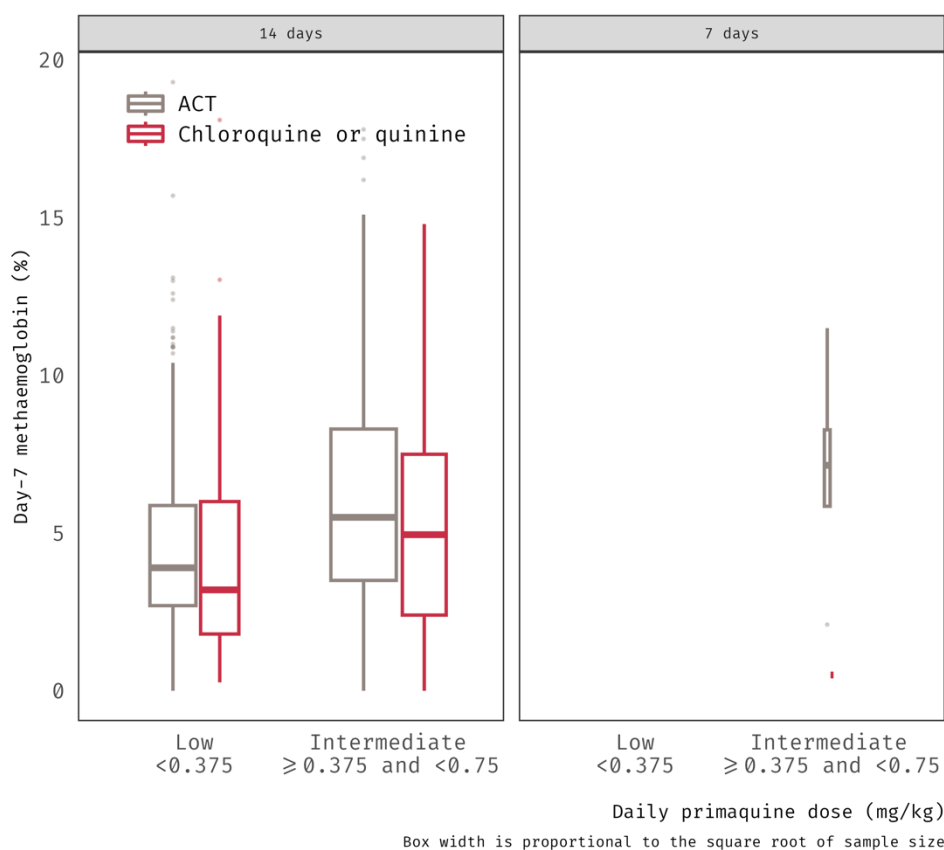
Table S5. Comparison of characteristics of patients (as originally reported) who received primaquine between included and eligible but not included studies

Characteristic	Included studies (n = 8)	Eligible but not included studies (n = 5)
<b>Region, studies (percentage)</b>		
Asia-Pacific	21 (58.3%) <sup>€</sup>	3 (60%)
Africa	4 (11.1%) <sup>€</sup>	0 (0%)
The Americas	11 (30.6%) <sup>€</sup>	2 (40%)
<b>Year of enrolment, studies (percentage)</b>		
Pre-2015	6 (75%)	5 (100%)
2015-2019	2 (25%)	0 (0%)
<b>Follow up duration in days, studies (percentage)</b>		
42	0 (0%)	1 (20%)
>42 to <120	0 (0%)	1 (20%)
120	0 (0%)	2 (40%)
>120	8 (100%)	1 (20%)
<b>Age (weighted-average years)<sup>#,§</sup></b>	19.1	20.0 <sup>§</sup>
<b>Male (weighted percentage)<sup>#</sup></b>	65.8%	63.7%

€ Included multinational studies. Number and percentage are derived from the number of study sites, not studies, within each region. # Weights approximated by the numbers of vivax patients treated with primaquine. § Study-specific average is the mean or median in vivax patients treated with primaquine. § Excluding one study for which the required summary statistics not available.



**Figure S3. Dynamics of primaquine-induced increases in blood methaemoglobin over time.** Box width is proportional to the square root of the number of patients.



**Figure S4. Day 7 methaemoglobin concentrations by primaquine regimen and dose group.** Among patients treated with a low-to-intermediate daily primaquine dose, day 7 methaemoglobin was lower when primaquine was combined with chloroquine or quinine as a partner drug.

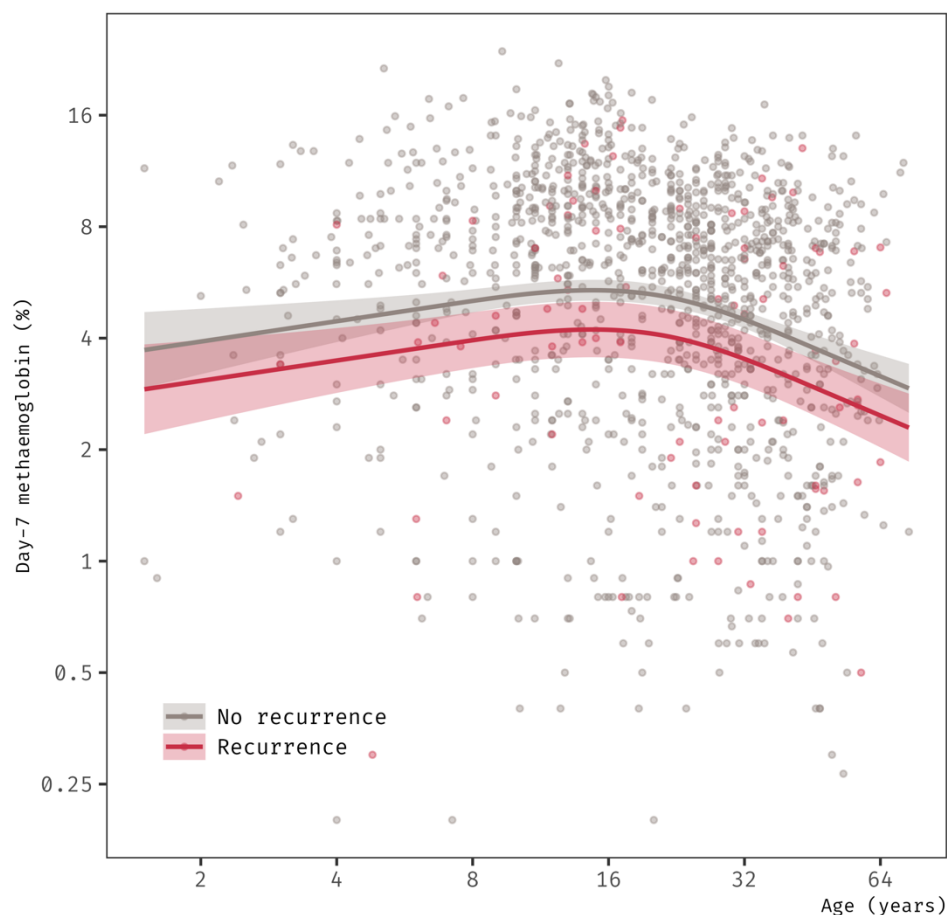


Figure S5. Inverse J-shaped association between patient age and day 7 methaemoglobin by recurrence status, after controlling for daily mg/kg primaquine dose. Horizontal and vertical axes are shown on the logarithmic scale.

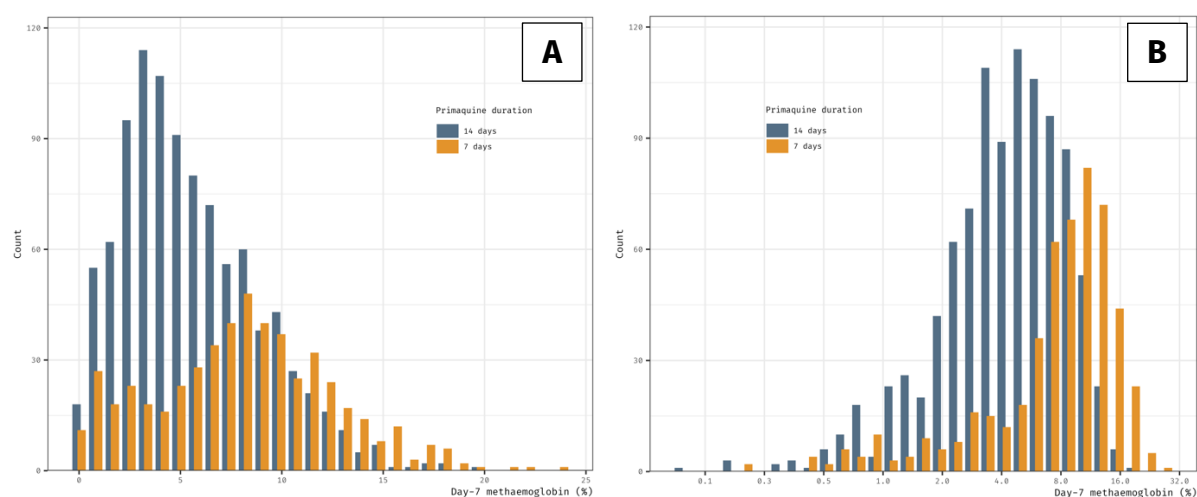
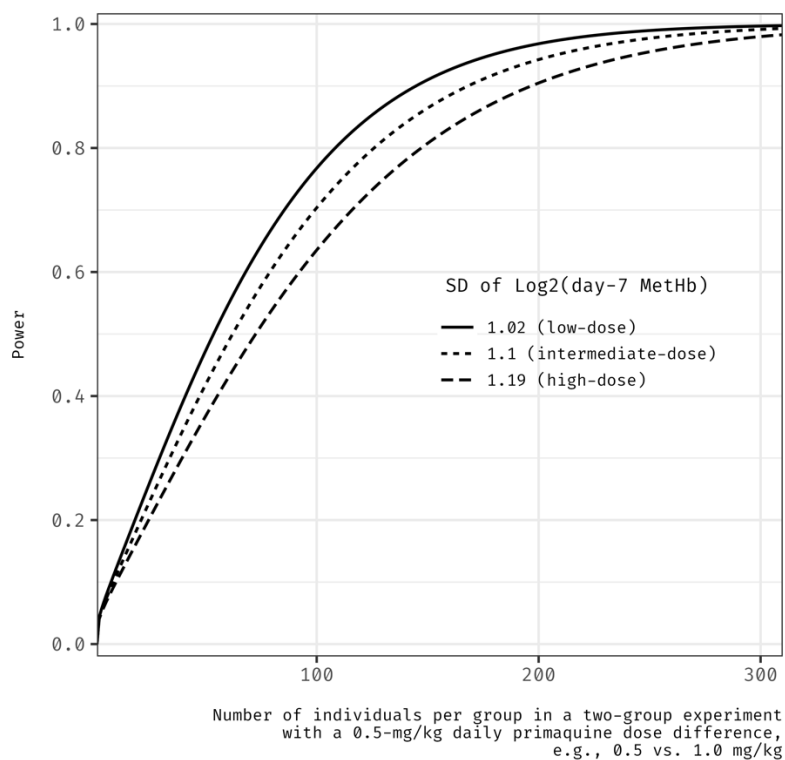


Figure S6. Distribution of day 7 methaemoglobin among the patients on (A) the original scale and (B) the logarithmic scale.



**Figure S7. Example of sample size calculations for future studies.** The assumed effect size is 0.39 (change on the  $\log_2$  day 7 methaemoglobin) which is equivalent to a 0.5-mg/kg increase in daily primaquine dose. The standard deviation (SD) of the  $\log_2$  day 7 methaemoglobin level were calculated for different categories of daily mg/kg primaquine dose based on pooled data. The false positive rate was set to 5%. The population distribution of the  $\log_2$  of day 7 methaemoglobin level conditional on the daily dose was assumed to follow a normal distribution.