

## **Determinants of the maternal 25-hydroxyvitamin D response to vitamin D supplementation during pregnancy**

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**Author Disclosures**

C. Cooper reports personal fees from ABBH, Amgen, Eli Lilly, GSK, Medtronic, Merck, Novartis, Pfizer, Roche, Servier and Takeda, outside the submitted work. N. Harvey reports personal fees, consultancy, lecture fees and honoraria from Alliance for Better Bone Health, AMGEN, MSD, Eli Lilly, Servier, Shire, Consilient Healthcare and Internis Pharma, outside the submitted work. N. Bishop reports remuneration from Internis Pharmaceuticals Ltd, outside the submitted work. S. Kennedy has nothing to disclose. A. Papageorghiou reports grants from Arthritis Research Council, during the conduct of the study. I. Schoenmakers has nothing to disclose. R. Fraser has nothing to disclose. S. Gandhi has nothing to disclose. A. Carr has nothing to disclose. S. D'Angelo has nothing to disclose. S. Crozier has nothing to disclose. R. Moon has nothing to disclose. N. Arden has received honoraria, held advisory board positions (which involved receipt of fees), and received consortium research grants, respectively, from: Merck, grants from Roche, personal fees from Smith & Nephew, Nicox, Flexion, grants from Bioiberica, Novartis, and personal fees from Bioventus and Freshfields, outside the submitted work. E. Dennison has nothing to disclose. K. Godfrey reports reimbursement for speaking at Nestle Nutrition Institute conferences, grants from Abbott Nutrition & Nestec, outside the submitted work; in addition, K. Godfrey has a patent Phenotype Prediction pending, a patent Predictive Use of CpG Methylation pending, and a patent Maternal Nutrition Composition pending, not directly related to this work. H. Inskip reports grants from Medical Research Council, Arthritis Research UK, European Union's Seventh Framework Programme, during the conduct of the study; and while not directly receiving funding from other bodies, members of her team have received funding from the following companies from other work: Danone, Nestec, Abbott Nutrition. A Prentice has nothing to disclose. M.Z. Mughal has nothing to disclose. R. Eastell reports grants and personal fees from Amgen, grants from Department of Health, grants from AstraZeneca, grants, personal fees and non-financial support from Immunodiagnostic Systems, grants from

ARUK/MRC Centre for Excellence in Musculoskeletal Ageing Research, grants from National Institute for Health Research, grants from MRC/AZ Mechanisms of Diseases Call, grants from MRC, grants and personal fees from Alexion, grants and other from National Osteoporosis Society, grants, personal fees and other from Roche, personal fees from Otsuka, Novartis, Merck, Bayer, Johnson & Johnson, Fonterra Brands, Janssen Research, personal fees and other from Eli Lilly, personal fees from Ono Pharma, Alere (Unipath), Chronos, personal fees and other from CL Biosystems, other from European Calcified Tissue Society, IOF CSA, personal fees from Teijin Pham Limited, other from ASBMR, personal fees from D-STAR, personal fees from GSK Nutrition, outside the submitted work. D. Reid has nothing to disclose. S. Robinson has nothing to disclose M.K. Javaid reports personal fees from Stirling Anglia, Consilient Health and Internis, outside the submitted work.

**Short title:** Response to vitamin D in pregnancy

**Word Count:** 3167

**Figures:** 2

**Tables:** 3

## **Abstract**

### *Context*

Current approaches to antenatal vitamin D supplementation do not account for inter-individual differences in 25-hydroxyvitamin D [25(OH)D] response.

### *Objective*

We assessed which maternal and environmental characteristics were associated with 25-hydroxyvitamin D [25(OH)D] following supplementation with cholecalciferol.

### *Design*

Within-randomisation-group analysis of participants in the MAVIDOS trial of vitamin D supplementation in pregnancy.

### *Setting*

Hospital antenatal clinics

### *Participants*

829 pregnant women (422 placebo, 407 cholecalciferol). At 14 and 34 weeks' gestation, maternal anthropometry, health and lifestyle were assessed and 25(OH)D measured. Compliance was determined using pill counts at 19 and 34 weeks.

### *Interventions*

1000IU/day cholecalciferol or matched placebo from 14 weeks' gestation until delivery.

### *Main outcome measure*

25-(OH)D at 34 weeks, measured in a single batch (Diasorin Liaison).

### *Results*

25(OH)D at 34 weeks' gestation was higher in the women randomised to vitamin D [mean (SD): 67.7 (21.3) nmol/l] compared with placebo [43.1 (22.5) nmol/l,  $p < 0.001$ ]. In women randomised to cholecalciferol, higher pregnancy weight gain from 14 to 34 weeks' gestation [kg] ( $\beta = -0.81$  (95% CI -1.39, -0.22)), lower compliance with study medication [%] ( $\beta = -0.28$  (-0.072, -0.48)), lower early pregnancy 25(OH)D [nmol/l] ( $\beta = 0.28$  (0.16, 0.40)) and delivery in the winter vs the summer ( $\beta = -10.5$  (-6.4, -14.6)) were independently associated with lower 25(OH)D at 34 weeks' gestation.

### *Conclusions*

Women who gained more weight during pregnancy, had lower 25(OH)D in early pregnancy and delivered in winter achieved a lower 25(OH)D in late pregnancy when supplemented with 1000 IU/day cholecalciferol. Future studies should aim to determine appropriate doses to enable consistent repletion of 25-hydroxyvitamin D during pregnancy.

**Keywords:** Vitamin D; pregnancy; osteoporosis; epidemiology; supplementation

## **Background**

Maternal vitamin D insufficiency during pregnancy is common (1,2), and there is evidence that this might have detrimental effects on maternal health, fetal development (3,4) and the long-term skeletal health of children (1,3). Severe maternal vitamin D deficiency during pregnancy can result in symptomatic hypocalcaemia in the neonate (3). Associations have been reported between maternal 25-hydroxyvitamin D [25(OH)D] and obstetric complications, including pre-eclampsia, gestational diabetes, preterm birth and offspring anthropometry, although the findings are inconsistent (3,4), and require confirmation in randomised controlled trials. Nonetheless, the Institute of Medicine (IOM) has suggested that risk of vitamin D insufficiency, defined as a 25(OH)D < 50 nmol/l, should be avoided during pregnancy (5), and this is supported by the recent Global Consensus on the Prevention of Rickets (6). Indeed many national guidelines recommend universal antenatal vitamin D supplementation to prevent vitamin D insufficiency (7-9).

Risk factors for vitamin D insufficiency are well described, and include ethnicity, extensive skin covering and liberal use of sun protection, overweight/obesity, low dietary vitamin D intake and smoking (1,10,11), in addition to the seasonal variation that is observed at temperate latitudes (11,12). Although vitamin D supplementation can improve maternal 25(OH)D status (10), little is known about how maternal characteristics might influence the 25(OH)D achieved following supplementation. In non-pregnant adults, baseline 25(OH)D concentration, body weight/adiposity and age are important determinants of the incremental rise in 25(OH)D following vitamin D supplementation (13,14). During pregnancy, maternal haemodilution is accompanied by a number of physiological changes to both vitamin D metabolism (15) and maternal body composition (16); such adaptations might lead to differences in the determinants of response to vitamin D supplementation between pregnant and non-pregnant women. Clinically, understanding how individuals respond could lead to individualised antenatal counselling regarding vitamin D supplementation to ensure vitamin D repletion is achieved without increasing the risk of vitamin D toxicity. We therefore undertook this study to determine maternal characteristics associated with achieved 25(OH)D following antenatal vitamin D supplementation in the context of a double-blind, randomised, placebo-controlled trial.

## Methods

### *The Maternal Vitamin D Osteoporosis Study (MAVIDOS)*

The MAVIDOS study is a multicentre, double-blind, randomised, placebo-controlled trial of vitamin D supplementation in pregnancy. The primary outcome was neonatal bone mass. A detailed description of the study methods (17) and primary findings **relating to offspring and maternal outcomes** have been published previously (18). The study was approved by the Southampton and South West Hampshire Research Ethics Committee. MAVIDOS was registered prospectively (ISRCTN:82927713; EUDRACT:2007-001716-23); full approval from UK MHRA was granted, and written, informed consent was obtained from all participants.

Briefly, women attending one of three United Kingdom (UK) hospitals [University Hospital Southampton NHS Foundation Trust, Southampton, UK (latitude 50.9° North); Oxford University Hospitals NHS Foundation Trust, Oxford, UK (latitude 51.8° North); Sheffield Hospitals NHS Trust (University of Sheffield), Sheffield, UK (latitude 53.4° North)] for early pregnancy ultrasound screening (11-14 weeks' gestation) were invited to participate in the study. Inclusion criteria were: age over 18 years, singleton pregnancy, and gestation less than 17 weeks based on last menstrual period (LMP) and ultrasound measurements. Women with known metabolic bone disease, renal stones, hyperparathyroidism or hypercalciuria, those taking medication known to interfere with fetal growth, fetal anomalies on ultrasonography and women already using >400IU/day vitamin D supplementation were excluded. A screening blood sample was obtained and analysed on the local NHS platform [all three laboratories (Southampton, Oxford and Sheffield) participate in DEQAS vitamin D quality assurance system (<http://www.deqas.org/>)]. Women with 25(OH)D between 25 and 100nmol/l and serum calcium <2.75mmol/l were eligible to **enroll** fully in the study.

Participants were randomised to either cholecalciferol 1000 IU/day or matched placebo [Merck KGaA, (Darmstadt, Germany)/ Sharp Clinical Services (Crickhowell, UK; previously DHP-Bilcare)], which was commenced before 17 weeks' gestation. The study medication was provided in a blister pack in a single box containing all medication for the whole pregnancy. All participants received standard antenatal care, and could continue self-administration of dietary supplements containing up to 400IU/day vitamin D.

### *Maternal assessments during pregnancy*

Prior to commencing the study medication, and again at 34 weeks' gestation, the women attended the research centre for a detailed assessment of diet (including supplement use), lifestyle (smoking, physical activity participation, employment) and health (past medical history, current medication use) using interviewer-led questionnaires. Ethnicity was determined by participant self-report, and subsequently categorised as white or non-white.

Anthropometric measurements included height, measured to the nearest 0.1cm using a stadiometer, and weight, assessed to the nearest 0.1kg using calibrated electronic scales. Four site (triceps, biceps, subscapular and suprailiac) skinfold thicknesses were measured to the nearest 0.2mm using a Harpenden skinfold calliper. Pregnancy weight gain was calculated as the difference between the weights at commencing study medication and 34 weeks' gestation.

#### *Compliance with study medication*

Participants were asked to bring any remaining study medication to each assessment. The pills were counted and compliance calculated as number consumed/expected consumption based on number of days since medication was dispensed, and expressed as a percentage. The 34 week visit was used for the calculation of compliance, and the count at 19 weeks was used if a 34 week count was not available.

#### *Assessment of 25(OH)D status*

On the day that the study medication was dispensed and at 34 weeks' gestation, a non-fasted venous blood sample was obtained, and serum stored at -80°C. 25(OH)D was assessed by radioimmunoassay (Liaison RIA automated platform, Diasorin, Minnesota, USA). All samples were analysed in a single batch at the end of the study at MRC Human Nutrition Research, Cambridge, UK. Details of assay performance and quality control through participation in DEQAS, NIST and NEQAS are given elsewhere (19,20).

#### *Statistical Analysis*

Women who had a measurement of 25(OH)D at both 14 and 34 weeks' gestation, and delivered a liveborn infant were included in the analysis (since pathology associated with fetal death might influence 25(OH)D concentrations). Maternal characteristics were compared between the women who did and did not complete the study using t-tests, Mann-Whitney U tests and  $\chi^2$  tests for normally distributed, non-normally distributed and categorical variables, respectively. Linear regression was used to assess the association between maternal characteristics and 25(OH)D at 34 weeks' gestation for each treatment

group separately. Multivariate linear regression was subsequently performed including all variables with a  $p < 0.2$  from the linear regression. Additionally, maternal factors associated with achieving a vitamin D replete status ( $>50\text{nmol/l}$ ) were determined using Poisson regression with robust standard errors(21). The cut-point of  $50\text{nmol/l}$  as the definition for vitamin D replete status was chosen to reflect the IOM guidelines (22). Additionally, we considered a  $25(\text{OH})\text{D} > 125\text{nmol/l}$  as indicating risk of toxicity, as suggested by the IOM (22). In the primary trial analysis, we classified season of birth according to the UK Meteorological office recommendations ([www.metoffice.gov.uk](http://www.metoffice.gov.uk)) with winter (December-February); spring (March-May); summer (June-August); and autumn (September-November). Since  $25(\text{OH})\text{D}$  concentrations are non-linearly associated with season, to facilitate ready comparison, we collapsed this classification into 2 groups with a notional “winter” (the months in which  $25(\text{OH})\text{D}$  concentrations tended to be lowest: December-May) and a “summer” (the months in which  $25(\text{OH})\text{D}$  concentrations tended to be highest: June-November). Finally, in sensitivity analysis, we excluded women who reported having taken any additional vitamin D-containing supplements within 90 days of the late pregnancy blood sampling. All analyses were performed in Stata v14 (Statacorp, College Station, Texas, USA). A p value of  $<0.05$  was considered statistically significant.

## Results

829 women, who delivered a live born infant and had measurements of  $25(\text{OH})\text{D}$  at both 14 and 34 weeks’ gestation, were included in the analysis (Figure 1). Women with missing  $25(\text{OH})\text{D}$  measurements at 34 weeks, who delivered a live born infant ( $n=136$ ) were of similar age, parity, height, ethnicity, educational achievement, early pregnancy BMI and smoking status to those included in this analysis ( $p > 0.05$  for all). There were no significant differences in baseline characteristics between women randomised to placebo and vitamin D supplementation (Table 1). Compliance with study medication was high in both treatment groups [placebo median 95% (IQR 88-98%), cholecalciferol median 96% (IQR 89-99%),  $p=0.11$ ].

### *Maternal 25(OH)D status at 34 weeks’ gestation by randomisation group*

Maternal  $25(\text{OH})\text{D}$  at 34 weeks’ gestation was greater in the women randomised to cholecalciferol [mean  $67.7\text{nmol/l}$  (SD  $21.3\text{nmol/l}$ )] compared with the placebo group (mean  $43.1\text{nmol/l}$  (SD  $22.5\text{nmol/l}$ ),  $p < 0.0001$ ). 83.3% of women randomised to cholecalciferol

achieved vitamin D replete status at 34 weeks' gestation ( $>50\text{nmol/l}$ ) compared with 35.6% in the placebo group ( $p<0.001$ ). Of the women who were not vitamin D replete at baseline ( $n=509$ ), 78.8% in the cholecalciferol group were replete at 34 weeks' gestation, compared with only 28.3% of the placebo group ( $p<0.001$ ). Similarly, only 48.4% of women who were vitamin D replete at baseline and received placebo remained vitamin D replete at 34 weeks' gestation, compared with 89.8% in the cholecalciferol group ( $p<0.001$ ). In both treatment groups, the proportion of women who were vitamin D replete at 34 weeks' gestation was lower in those who delivered in winter (Table 2). No participant reported symptoms suggestive of vitamin D toxicity. Two participants (0.5%) randomised to placebo and one to cholecalciferol (0.3%) ( $p$  difference=0.58) had a  $25(\text{OH})\text{D}\geq 125\text{nmol/l}$  at 34 weeks' gestation, with the maximum value being  $139\text{nmol/l}$ .

#### *Determinants of maternal 25(OH)D at 34 weeks' gestation*

In univariate analysis, maternal age, baseline 25(OH)D, season of delivery and compliance with study medication were significantly associated with 34 week 25(OH)D in both the placebo and vitamin D supplementation groups (Table 3). Additionally, women who reported smoking in late pregnancy had significantly lower 25(OH)D in the placebo group, but this association was not observed amongst women randomised to cholecalciferol. Conversely, markers of maternal weight and adiposity were significantly inversely associated with maternal 25(OH)D in the cholecalciferol group, but not the women randomised to placebo.

In multiple linear regression analysis, maternal factors significantly associated with greater 25(OH)D at 34 weeks' gestation in the vitamin D supplementation group were lower pregnancy weight gain [kg] ( $\beta=-0.81$ ; 95%CI: -1.39, -0.22;  $p=0.007$ ), higher compliance [%] ( $\beta=0.28$ ; 95%CI: 0.072, 0.48;  $p=0.008$ ), higher early pregnancy 25(OH)D [nmol/l] ( $\beta=0.28$ ; 95%CI: 0.16, 0.40;  $p<0.001$ ) and summer delivery [summer vs winter] ( $\beta=10.51$ ; 95%CI: 6.40, 14.63;  $p<0.001$ ) (Figure 2a). In the placebo group (Figure 2b), higher early pregnancy 25(OH)D [nmol/l] ( $\beta=0.59$ ; 95%CI: 0.49, 0.68;  $p<0.001$ ), summer delivery [summer vs winter] ( $\beta=24.97$ ; 95%CI: 21.77, 28.17;  $p<0.001$ ), and greater maternal age (years) ( $\beta=0.32$ ; 95%CI: 0.022, 0.62;  $p=0.04$ ) remained significantly associated with greater 25(OH)D at 34 weeks' gestation. When achievement of vitamin D replete status at 34 weeks' gestation was considered instead of absolute 25(OH)D concentration, in multivariate analyses, delivery in summer (RR=1.20; 95%CI: 1.09, 1.33;  $p<0.001$ ), white ethnicity (RR=1.267; 95%CI: 1.17, 1.37;  $p<0.001$ ), greater compliance with medication [%] (RR=1.01; 95%CI: 1.00, 1.02;

p=0.03), and greater early pregnancy 25(OH)D concentration [nmol/l] (RR=1.003; 95%CI: 1.001, 1.006; p=0.007) were significantly associated with achieving 25(OH)D>50nmol/l in the women randomised to cholecalciferol.

#### *Interaction between baseline 25(OH)D and randomisation group*

When comparing achieved 25(OH)D at 34 weeks' gestation between placebo and cholecalciferol groups, it was apparent that there was a statistically significant interaction between baseline 25(OH)D and randomisation group (p<0.001). Thus there was a smaller difference in 25(OH)D concentrations at 34 weeks' gestation between the placebo and treatment arms with increasing 25(OH)D at 14 weeks' gestation.

#### *Sensitivity analyses*

As participants were permitted to continue taking daily vitamin D supplements containing up to 400IU, in the sensitivity analysis we excluded 229 women (n=117 randomised to cholecalciferol) who reported taking other vitamin D containing dietary supplements at the late pregnancy interview. Similarly, 81.0% of women randomised to cholecalciferol were vitamin D replete at 34 weeks' gestation, compared with 29.4% of women randomised to placebo (p<0.001). The maternal characteristics associated with 25(OH)D at 34 weeks' gestation and achieving vitamin D replete status were similar to those observed in the whole cohort.

## **Discussion**

We have assessed anthropometric and demographic factors associated with the response to antenatal supplementation with 1000 IU/day cholecalciferol. This dose achieved vitamin D repletion in over 80% of women, without leading to 25(OH)D levels potentially associated with vitamin D toxicity (at least within the included baseline of 25-100nmol/l 25(OH)D). However, gaining less weight during pregnancy, having a higher 25(OH)D in early pregnancy, delivering in summer and having higher compliance with supplementation were independently associated with achieving a greater 25(OH)D concentration in late pregnancy amongst women randomised to vitamin D supplementation. Thus, those women who are at risk of vitamin D insufficiency in early pregnancy, gain greater weight, and deliver in winter might need supplementation with a higher dose of cholecalciferol to achieve similar 25(OH)D concentrations. However, when vitamin D replete status was considered as the outcome, only non-white maternal ethnicity and delivery in winter were significant predictors of vitamin D non-replete status following supplementation.

To our knowledge, the factors which determine the response to vitamin D supplementation in pregnancy have not previously been assessed. However, our findings are consistent with those in non-pregnant adults (13,14). It is well recognised that individuals who are overweight or obese are at higher risk of vitamin D insufficiency, and this is similarly observed in pregnancy (2,23). Studies in non-pregnant adults have also shown that obese individuals achieve a lower 25(OH)D with the same dose of supplementation as non-obese individuals (14). Meta-analysis of vitamin D supplementation studies has suggested that over 50% of the variance in 25(OH)D increment in response to supplementation is explained by body weight (13). Although the relationship between body weight and 25(OH)D increment following supplementation could reflect sequestration in adipose tissue, we found that in multivariate analysis, pre-pregnancy BMI and late pregnancy triceps skinfold thickness (as a marker of adiposity), were not associated with 25(OH)D after supplementation, but that pregnancy weight gain was positively associated. Similarly, we have previously demonstrated that greater gestational weight gain is associated with a decline in 25(OH)D status during pregnancy, independent of supplement use (12). Weight gain in pregnancy represents not only increased fat mass but also feto-placental tissues and haemodilution (16); our data, therefore, suggest that overall volume of dilution, and not just adiposity, may be important for response to vitamin D supplementation. However, importantly, when using a 25(OH)D > 50 nmol/l as a cut-point for repletion, pregnancy weight gain was not an independent predictor of achieving vitamin D repletion.

Despite receiving 1000IU cholecalciferol per day, 25% of mothers delivering in winter had a 25(OH)D less than 50 nmol/l. This is a higher non-repletion rate than that reported in other recent pregnancy supplement studies (24-26). However, it is notable that there were marked differences in baseline 25(OH)D concentrations between these investigations, and we observed that initial 25(OH)D status was positively associated with both the likelihood of achieving vitamin D replete status and absolute 25(OH)D status at 34 weeks' gestation. Importantly, the difference between the 25(OH)D achieved at 34 weeks' gestation in women randomised to placebo compared with cholecalciferol decreased with increasing baseline 25(OH)D. This is consistent with previous studies in adults, which have shown that the incremental response to vitamin D supplementation is higher in vitamin D insufficient than replete subjects (13,14), and that the increase in 25(OH)D relative to supplementation dose is negatively associated with dose of vitamin D supplement (27). This suggests that physiological processes such as saturation of the hepatic 25-hydroxylase involved in the

conversion of cholecalciferol to 25(OH)D or conversion to 24- or 4-OH metabolites, together with renal catabolism, limit attainment of very high 25(OH)D concentrations (28). This mechanism might be important in preventing hypervitaminosis D. However, studies comparing the effectiveness of differing doses of vitamin D in pregnancy have shown that 4000IU/day can achieve a higher 25(OH)D than 400IU/day (10,29), but whether these higher doses are of clinical benefit is yet to be demonstrated (30,31) and at the general population level, lower doses would be compatible with keeping 25(OH)D below a concentration which might be concerning.

It is evident from our findings that 1000 IU/day cholecalciferol in pregnancy does not eliminate the seasonal variation in 25(OH)D status observed in pregnant women in the UK (11,12). Similarly, non-white ethnicity was associated with a higher risk of not achieving vitamin D replete status in the supplemented women. Hollis et al. similarly found that even with 4000IU/day vitamin D during pregnancy, African-American women had lower 25(OH)D in late pregnancy than Caucasian or Hispanic women (10). Thus, future studies should aim to determine the dose required to achieve optimal 25(OH)D status amongst women of non-white ethnicity and amongst those who deliver in winter months. Maternal age was also positively associated in univariate analyses with 25(OH)D achieved at 34 weeks' gestation in women who received vitamin D supplementation. It has previously been shown in pregnant women that age is positively associated with 25(OH)D status (32,33). Whilst lower uptake of supplementation in younger women (12) could partly explain this observation, our finding would additionally suggest that even in younger women who do use supplements, the achieved 25(OH)D is lower. Data from healthy and hospitalised adults have similarly shown that older individuals achieve a higher 25(OH)D following vitamin D supplementation (13,34). As such, young pregnant women might particularly require advice on the need for, and compliance with, vitamin D supplementation.

Although our findings are novel, and may enable the development of individualised advice for antenatal vitamin D supplementation, there are a number of limitations which should be considered in the interpretation of this study. Firstly, we could not, as a result of stipulations made during the ethics approval process, include participants with 25(OH)D concentrations <25nmol/l or >100nmol/l. As baseline 25(OH)D was associated with the likelihood of achieving vitamin D replete status, it is likely that women with very low levels of 25(OH)D at baseline will require a higher supplementation dose to achieve vitamin D repletion. However, this needs to be confirmed in future studies. Secondly, only a small

proportion of the women included in this study were of non-white ethnicity. This reflects the local populations and care should be taken in translating these findings to a more ethnically diverse population. Thirdly, we did not examine genetic determinants of the response to vitamin D supplementation. It has been demonstrated previously that the incremental rise in 25(OH)D following supplementation differs by single nucleotide polymorphisms in vitamin D binding protein (35,36) and 25-hydroxylase genes (35). Although this genetic information can enable a more comprehensive understanding of the biochemical response to vitamin D supplementation, the current inability to undertake genotyping on a widespread population basis means this additional information would not allow for alterations to current clinical practice regarding vitamin D supplementation in pregnancy. Finally, during pregnancy, a number of physiological changes occur to vitamin D metabolism, including an increase in vitamin D binding protein and 1,25-dihydroxyvitamin D (15), indices that we were not able to include in our analysis.

In conclusion, we have demonstrated that women who gain more weight during pregnancy, have lower 25(OH)D in early pregnancy or deliver in winter tend to achieve a lower 25(OH)D in late pregnancy when supplemented with 1000IU/day cholecalciferol than do women with the converse attributes. Future studies should aim to determine appropriate doses to enable consistent repletion of 25-hydroxyvitamin D during pregnancy, and our findings support the notion that clinical approaches to vitamin D repletion may be informed by individual characteristics. **As such, personalised vitamin D supplementation advice might become part of future antenatal care.**

## **Acknowledgements**

This work was supported by grants from the Arthritis Research UK (17702), Medical Research Council (4050502589), Bupa Foundation, National Institute for Health Research (NIHR) Southampton Biomedical Research Centre, University of Southampton and University Hospital Southampton NHS Foundation Trust, and NIHR Musculoskeletal Biomedical Research Unit, University of Oxford. IS and AP were funded by the Medical Research Council (MRC) (programme code U105960371). The work leading to these results was supported by the European Union's Seventh Framework Programme (FP7/2007-2013), projects EarlyNutrition and ODIN under grant agreements numbers 289346 and 613977. <sup>†</sup>RJM, NCH and CC are joint first author. We are extremely grateful to Merck GmbH for the

kind provision of the Vigantoletten supplement. Merck GmbH had no role in the trial execution, data collection, analysis or manuscript preparation. The authors had full access to all study data.

## References

1. Javaid MK, Crozier SR, Harvey NC, Gale CR, Dennison EM, Boucher BJ, Arden NK, Godfrey KM, Cooper C. Maternal vitamin D status during pregnancy and childhood bone mass at age 9 years: a longitudinal study. *Lancet* 2006; 367:36-43
2. McAree T, Jacobs B, Manickavasagar T, Sivalokanathan S, Brennan L, Bassett P, Rainbow S, Blair M. Vitamin D deficiency in pregnancy - still a public health issue. *Matern Child Nutr* 2013; 9:23-30
3. Harvey NC, Holroyd C, Ntani G, Javaid K, Cooper P, Moon R, Cole Z, Tinati T, Godfrey K, Dennison E, Bishop NJ, Baird J, Cooper C. Vitamin D supplementation in pregnancy: a systematic review. *Health technology assessment (Winchester, England)* 2014; 18:1-190
4. Moon RJ, Harvey NC, Cooper C. ENDOCRINOLOGY IN PREGNANCY: Influence of maternal vitamin D status on obstetric outcomes and the fetal skeleton. *Eur J Endocrinol* 2015; 173:R69-83
5. Ross AC, Manson JE, Abrams SA, Aloia JF, Brannon PM, Clinton SK, Durazo-Arvizu RA, Gallagher JC, Gallo RL, Jones G, Kovacs CS, Mayne ST, Rosen CJ, Shapses SA. The 2011 report on dietary reference intakes for calcium and vitamin D from the Institute of Medicine: what clinicians need to know. *J Clin Endocrinol Metab* 2011; 96:53-58
6. Munns CF, Shaw N, Kiely M, Specker BL, Thacher TD, Ozono K, Michigami T, Tiosano D, Mughal MZ, Makitie O, Ramos-Abad L, Ward L, DiMeglio LA, Atapattu N, Cassinelli H, Braegger C, Pettifor JM, Seth A, Idris HW, Bhatia V, Fu J, Goldberg G, Savendahl L, Khadgawat R, Pludowski P, Maddock J, Hypponen E, Oduwole A, Frew E, Aguiar M, Tulchinsky T, Butler G, Hogler W. Global Consensus Recommendations on Prevention and Management of Nutritional Rickets. *J Clin Endocrinol Metab* 2016; 101:394-415
7. National Institute for Health and Clinical Excellence. Antenatal care (NICE Clinical Guideline 62). [www.guidance.nice.org.uk/cg622010](http://www.guidance.nice.org.uk/cg622010).
8. Paxton GA, Teale GR, Nowson CA, Mason RS, McGrath JJ, Thompson MJ, Siafarikas A, Rodda CP, Munns CF. Vitamin D and health in pregnancy, infants, children and adolescents in Australia and New Zealand: a position statement. *Med J Aust* 2013; 198:142-143

9. Holick MF, Binkley NC, Bischoff-Ferrari HA, Gordon CM, Hanley DA, Heaney RP, Murad MH, Weaver CM. Evaluation, treatment, and prevention of vitamin D deficiency: an Endocrine Society clinical practice guideline. *The Journal of clinical endocrinology and metabolism* 2011; 96:1911-1930
10. Hollis BW, Johnson D, Hulsey TC, Ebeling M, Wagner CL. Vitamin D supplementation during pregnancy: double-blind, randomized clinical trial of safety and effectiveness. *Journal of bone and mineral research : the official journal of the American Society for Bone and Mineral Research* 2011; 26:2341-2357
11. Crozier SR, Harvey NC, Inskip HM, Godfrey KM, Cooper C, Robinson SM, Group SWSS. Maternal vitamin D status in pregnancy is associated with adiposity in the offspring: findings from the Southampton Women's Survey. *Am J Clin Nutr* 2012; 96:57-63
12. Moon RJ, Crozier SR, Dennison EM, Davies JH, Robinson SM, Inskip HM, Godfrey KM, Cooper C, Harvey NC. Tracking of 25-hydroxyvitamin D status during pregnancy: the importance of vitamin D supplementation. *Am J Clin Nutr* 2015; 102:1081-1087
13. Zittermann A, Ernst JB, Gummert JF, Borgermann J. Vitamin D supplementation, body weight and human serum 25-hydroxyvitamin D response: a systematic review. *Eur J Nutr* 2014; 53:367-374
14. Mazahery H, von Hurst PR. Factors Affecting 25-Hydroxyvitamin D Concentration in Response to Vitamin D Supplementation. *Nutrients* 2015; 7:5111-5142
15. Bouillon R, Van Assche FA, Van Baelen H, Heyns W, De Moor P. Influence of the vitamin D-binding protein on the serum concentration of 1,25-dihydroxyvitamin D<sub>3</sub>. Significance of the free 1,25-dihydroxyvitamin D<sub>3</sub> concentration. *The Journal of clinical investigation* 1981; 67:589-596
16. Widen EM, Gallagher D. Body composition changes in pregnancy: measurement, predictors and outcomes. *European journal of clinical nutrition* 2014; 68:643-652
17. Harvey NC, Javaid K, Bishop N, Kennedy S, Papageorgiou AT, Fraser R, Gandhi SV, Schoenmakers I, Prentice A, Cooper C. MAVIDOS Maternal Vitamin D Osteoporosis Study: study protocol for a randomized controlled trial. The MAVIDOS Study Group. *Trials* 2012; 13:13
18. Cooper C, Harvey NC, Bishop NJ, Kennedy S, Papageorgiou AT, Schoenmakers I, Fraser R, Gandhi SV, Carr A, D'Angelo S, Crozier SR, Moon RJ, Arden NK, Dennison EM, Godfrey

- KM, Inskip HM, Prentice A, Mughal MZ, Eastell R, Reid DM, Javaid MK. Maternal gestational vitamin D supplementation and offspring bone health (MAVIDOS): a multicentre, double-blind, randomised placebo-controlled trial. *The lancet Diabetes & endocrinology* 2016; 4:393-402
19. Jones KS, Assar S, Harnpanich D, Bouillon R, Lambrechts D, Prentice A, Schoenmakers I. 25(OH)D<sub>2</sub> half-life is shorter than 25(OH)D<sub>3</sub> half-life and is influenced by DBP concentration and genotype. *J Clin Endocrinol Metab* 2014; 99:3373-3381
  20. Sempos CT, Vesper HW, Phinney KW, Thienpont LM, Coates PM. Vitamin D status as an international issue: national surveys and the problem of standardization. *Scand J Clin Lab Invest Suppl* 2012; 243:32-40
  21. Barros AJ, Hirakata VN. Alternatives for logistic regression in cross-sectional studies: an empirical comparison of models that directly estimate the prevalence ratio. *BMC medical research methodology* 2003; 3:21
  22. Ross AC, Taylor CL, Yaktine AL, Del Valle HB. *Dietary Reference Intakes for Calcium and Vitamin D* 2011.
  23. Bodnar LM, Catov JM, Roberts JM, Simhan HN. Prepregnancy obesity predicts poor vitamin D status in mothers and their neonates. *The Journal of nutrition* 2007; 137:2437-2442
  24. Litonjua AA, Carey VJ, Laranjo N, Harshfield BJ, McElrath TF, O'Connor GT, Sandel M, Iverson RE, Jr., Lee-Paritz A, Strunk RC, Bacharier LB, Macones GA, Zeiger RS, Schatz M, Hollis BW, Hornsby E, Hawrylowicz C, Wu AC, Weiss ST. Effect of Prenatal Supplementation With Vitamin D on Asthma or Recurrent Wheezing in Offspring by Age 3 Years: The VDAART Randomized Clinical Trial. *JAMA* 2016; 315:362-370
  25. Chawes BL, Bonnelykke K, Stokholm J, Vissing NH, Bjarnadottir E, Schoos AM, Wolsk HM, Pedersen TM, Vinding RK, Thorsteinsdottir S, Arianto L, Hallas HW, Heickendorff L, Brix S, Rasmussen MA, Bisgaard H. Effect of Vitamin D<sub>3</sub> Supplementation During Pregnancy on Risk of Persistent Wheeze in the Offspring: A Randomized Clinical Trial. *JAMA* 2016; 315:353-361
  26. March KM, Chen NN, Karakochuk CD, Shand AW, Innis SM, von Dadelszen P, Barr SI, Lyon MR, Whiting SJ, Weiler HA, Green TJ. Maternal vitamin D<sub>3</sub> supplementation at 50 mug/d protects against low serum 25-hydroxyvitamin D in infants at 8 wk of age: a randomized controlled trial of 3 doses of vitamin D beginning in gestation and continued in lactation. *Am J Clin Nutr* 2015; 102:402-410

27. Barger-Lux MJ, Heaney RP, Dowell S, Chen TC, Holick MF. Vitamin D and its major metabolites: serum levels after graded oral dosing in healthy men. *Osteoporosis international : a journal established as result of cooperation between the European Foundation for Osteoporosis and the National Osteoporosis Foundation of the USA* 1998; 8:222-230
28. Heaney RP, Armas LA, Shary JR, Bell NH, Binkley N, Hollis BW. 25-Hydroxylation of vitamin D3: relation to circulating vitamin D3 under various input conditions. *The American journal of clinical nutrition* 2008; 87:1738-1742
29. Dawodu A, Saadi HF, Bekdache G, Javed Y, Altaye M, Hollis BW. Randomized controlled trial (RCT) of vitamin D supplementation in pregnancy in a population with endemic vitamin D deficiency. *The Journal of clinical endocrinology and metabolism* 2013; 98:2337-2346
30. Moon R, Harvey N, Cooper C. ENDOCRINOLOGY IN PREGNANCY: Influence of maternal vitamin D status on obstetric outcomes and the foetal skeleton. *European Journal of Endocrinology* 2015;
31. Wagner CL, McNeil RB, Johnson DD, Hulsey TC, Ebeling M, Robinson C, Hamilton SA, Hollis BW. Health characteristics and outcomes of two randomized vitamin D supplementation trials during pregnancy: a combined analysis. *J Steroid Biochem Mol Biol* 2013; 136:313-320
32. Davies-Tuck M, Yim C, Knight M, Hodges R, Doery JC, Wallace E. Vitamin D testing in pregnancy: Does one size fit all? *The Australian & New Zealand journal of obstetrics & gynaecology* 2015; 55:149-155
33. Xiao JP, Zang J, Pei JJ, Xu F, Zhu Y, Liao XP. Low maternal vitamin D status during the second trimester of pregnancy: a cross-sectional study in Wuxi, China. *PloS one* 2015; 10:e0117748
34. Singh G, Bonham AJ. A predictive equation to guide vitamin d replacement dose in patients. *J Am Board Fam Med* 2014; 27:495-509
35. Didriksen A, Grimnes G, Hutchinson MS, Kjaergaard M, Svartberg J, Joakimsen RM, Jorde R. The serum 25-hydroxyvitamin D response to vitamin D supplementation is related to genetic factors, BMI, and baseline levels. *European journal of endocrinology / European Federation of Endocrine Societies* 2013; 169:559-567

- 36.** Fu L, Yun F, Oczak M, Wong BY, Vieth R, Cole DE. Common genetic variants of the vitamin D binding protein (DBP) predict differences in response of serum 25-hydroxyvitamin D [25(OH)D] to vitamin D supplementation. *Clinical biochemistry* 2009; 42:1174-1177

## **Figure Legends**

**Figure 1:** Consort diagram

**Figure 2:** Independent determinants of maternal 25(OH)D at 34 weeks gestation a) following supplementation with 1000IU cholecalciferol per day from 14 weeks' gestation until delivery; and b) receiving placebo from 14 weeks' gestation until delivery. Shown as standard deviation change in 25(OH)D per unit predictor. \* $p < 0.05$ , \*\* $p < 0.01$

**Table 1:** Maternal characteristics at baseline according to randomisation group.

	Placebo	1000IU/day cholecalciferol
N	422	407
Gestation (weeks), mean (SD)	15.9 (1.5)	15.9 (1.5)
Maternal age (years), mean (SD)	30.7 (5.4)	30.7 (5.0)
Nulliparous (%)	44.8	42.7
Current smoker (%)	7.7	7.7
BMI (kg/m <sup>2</sup> ), median (IQR)	25.4 (22.7-29.7)	24.6 (22.2-28.6)
Height (cm), mean (SD)	165.6 (6.6)	165.5 (6.3)
White ethnicity (%)	94.8	95.6
25(OH)D (nmol/l), median (IQR)	44.4 (33.2-57.0)	45.7 (34.3-57.8)

**Table 2:** Percentage of women achieving vitamin D replete status (>50nmol/l) according to randomisation group and season of delivery.

	Placebo	1000 IU/day cholecalciferol	p comparing randomisation groups
Season of delivery			
Winter (December-May)	13.9	75.0	<0.001
Summer (June-November)	54.2	90.1	<0.001
p comparing seasons	<0.001	<0.001	

**Table 3:** 25-hydroxyvitamin D status at 34 weeks gestation according to maternal characteristics in women randomised to placebo or vitamin D supplementation from 14 weeks gestation. Shown as nmol/l change in 25(OH)D per unit predictor.

	Placebo		1000 IU/day cholecalciferol	
	Beta (95% CI)	p	Beta (95% CI)	p
Maternal age (years)	<b>0.67 (0.28, 1.07)</b>	<b>0.001</b>	<b>0.70 (0.28, 1.11)</b>	<b>0.001</b>
Parity (yes vs no)	-1.25 (-5.69, 3.20)	0.581	-0.29 (-4.60, 4.03)	0.896
Smoking at 34 weeks gestation (yes vs no)	<b>-13.45 (-22.12, -4.78)</b>	<b>0.002</b>	-1.49 (-9.50, 6.52)	0.715
Ethnicity (other vs white)	-8.69 (-18.59, 1.21)	0.085	1.99 (-8.50, 12.48)	0.709
Height (cm)	0.15 (-0.19, 0.48)	0.389	-0.072 (-0.41, 0.27)	0.675
BMI at 14 weeks gestation (kg/m <sup>2</sup> )	-0.24 (-0.69, 0.20)	0.284	<b>-0.47 (-0.90, -0.048)</b>	<b>0.029</b>
Weight at 34 weeks gestation (kg)	-0.056 (-0.22, 0.11)	0.492	<b>-0.23 (-0.38, -0.085)</b>	<b>0.002</b>
Weight gain early to late pregnancy (kg)	-0.23 (-0.85, 0.40)	0.473	<b>-0.65 (-1.26, -0.039)</b>	<b>0.037</b>
Triceps SFT at 34 weeks gestation (mm)	-0.059 (-0.38, 0.26)	0.718	<b>-0.42 (-0.74, -0.10)</b>	<b>0.01</b>
Moderate/strenuous exercise in late pregnancy (hrs/week)	1.36 (-1.75, 4.47)	0.389	-0.76 (-3.63, 2.10)	0.60
25(OH)D at 14 weeks gestation (nmol/l)	<b>0.52 (0.40, 0.64)</b>	<b>&lt;0.001</b>	<b>0.21 (0.089, 0.33)</b>	<b>0.001</b>
Season of delivery (summer vs winter)	<b>22.77 (19.05, 26.50)</b>	<b>&lt;0.001</b>	<b>10.09 (6.039, 14.15)</b>	<b>&lt;0.001</b>
Compliance (%)	<b>0.23 (0.044, 0.42)</b>	<b>0.016</b>	<b>0.39 (0.19, 0.59)</b>	<b>&lt;0.001</b>

SFT: skinfold thickness

**Figure 1:** Consort diagram

**Figure 2:** Independent determinants of maternal 25(OH)D at 34 weeks gestation a) following supplementation with 1000IU cholecalciferol per day from 14 weeks' gestation until delivery; and b) receiving placebo from 14 weeks' gestation until delivery. Shown as standard deviation change in 25(OH)D per unit predictor. \*p<0.05, \*\*p<0.01

*a) Cholecalciferol 1000IU/day*

*b) Placebo*