



# Ultrahigh magnetic resonance contrast switching with water gated polymer–silica nanoparticles†

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**Very high  $T_1$  magnetic resonance imaging (MRI) switches can be obtained with pH-responsive polymer-coated paramagnetic mesoporous silica nanoparticles (MSNs), as the local environment traverses the  $pK_a$  of the polymer coat ( $\Delta r_1 \sim 50 \text{ mM}^{-1} \text{ s}^{-1}$  at 1.5 T and  $\Delta r_1 \sim 22 \text{ mM}^{-1} \text{ s}^{-1}$  at 3 T). We assign these characteristics to a strong peripheral hydration capping at the mesopores, impacting channel-confined water mobility such that outer sphere contributions to contrast are greatly enhanced.**

Magnetic resonance imaging (MRI) is a powerful non-invasive diagnostic technique with micron spatial resolution and deep tissue penetration that empowers clinicians to resolve and monitor a wide variety of potentially fatal internal pathological conditions.<sup>1,2</sup> Frequent low contrast-to-noise problems can be alleviated through the use of contrast agents (CAs), most notably chelated paramagnetic ions, such as gadolinium( $\text{Gd}^{3+}$ ).<sup>3</sup> The contrast generating efficiency for a  $\text{Gd}^{3+}$ -based CA can be defined by its longitudinal relaxivity, denoted  $r_1$  ( $r_1 = \Delta(1/T_1)/[\text{CA}]$ ; where  $T_1$  is the longitudinal relaxation time and  $[\text{CA}]$  is the concentration of the CA), with a high  $r_1$  correlating to a lower required  $\text{Gd}^{3+}$  dose, which is, of course, clinically desirable. To improve the natively low molecular  $r_1$  values a broad range of paramagnetically doped nanomaterials, with reduced tumbling rates ( $\tau_R$ ), have been reported.<sup>4</sup> Nanoparticulate CAs additionally present a route to controlled blood circulation times, additional imaging modality incorporation, and tumour accumulation.<sup>5</sup> In prior work, we, and other researchers, have shown that Gd-chelate modified mesoporous silica nanoparticles (Gd-MSNs) offer a synthetically tuneable, and biocompatible, platform with high associated image contrast.<sup>6–12</sup> It has been prior noted that restricted water mobility (elongated diffusional correlation times,  $\tau_D$ )

within the nanoconfinement of a porous reservoir significantly boosts  $r_1$ , in large part by increasing the role played by outer sphere effects.<sup>13,14</sup>

Stimuli-responsive “smart” nanoparticulate CAs, where contrast can be switched by an endogenous or exogenous stimulus, such as light, enzyme activity, redox environment, or local pH,<sup>15</sup> can provide specific information on the local physiological environment, facilitating the ability to distinguish between healthy tissues and lesions, for example.<sup>16,17</sup> In reference to proton relaxation theory,<sup>18,19</sup> tuneable contrast generation can be achieved by the modulation of the diffusive mobility of either inner-sphere (those bound to the metal ions at  $\sim 3.1 \text{ \AA}$ ) or outer-sphere (with water-to-Gd distances  $\geq 4.0 \text{ \AA}$ ) water molecules.<sup>20</sup> For example, a number of switchable nanoparticulate CAs have been designed where the inner-sphere (IS) water exchange rate ( $\tau_M$ ) can be modulated by the local environment.<sup>21–24</sup> Prior reported smart CAs possess, however, only moderate relaxometric switches (*i.e.*  $\Delta r_1 < 15 \text{ mM}^{-1} \text{ s}^{-1}$  and often much less).<sup>25,26</sup> It is also known that relaxivity unhelpfully decreases for typical Gd-based CAs at the higher imaging field strengths that are becoming increasingly common.<sup>20,27</sup> There is, therefore, substantial room to design a high field effective CA that exhibits a significant environmentally triggered switch. To date, no stimuli-responsive CA characteristics have been reported that operate through a modulation of OS contributions.

Herein, we report a versatile surface-initiated reversible addition-fragmentation chain-transfer (SI-RAFT) polymerisation approach to modify Gd-chelate doped MSNs with a stimuli-responsive, externally grafted, polymer shell (Fig. 1a). Poly(methacrylic acid) (pMAA) has been widely reported within pH-responsive drug delivery applications,<sup>28–30</sup> possessing an associated  $pK_a$  of 5.2, clinically relevant, for example, to mapping deviations in pH associated with chronic inflammation.<sup>31,32</sup> It is also known that in its charged state pMAA has a particularly strong association with water.<sup>33</sup> It was envisaged that such a capping would impact the mobility of particle internalised water, and hence optimise OS relaxivities ( $r_1^{\text{OS}}$ ).

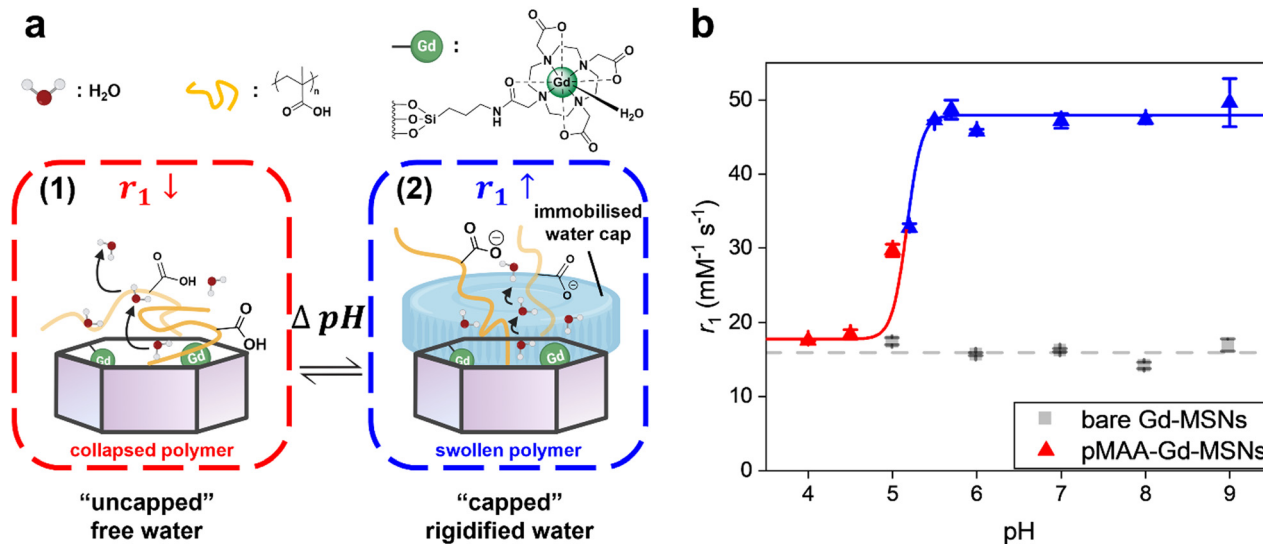
Initially, paramagnetic Gd-MSNs (with gadolinium (iii) 1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid, Gd-DO TA, modified pore channels) were synthesised according to

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**Fig. 1** (a) An illustration highlighting the pH-responsive  $T_1$  switch for pMAA-Gd-MSNs. For an uncharged/collapsed conformation (1), the peripheral pMAA shell possesses limited hydration, and moderate  $r_1$  values. As the polymer shell charges and swells, its hydration increases dramatically, and a “water cap” is formed (2). (b) A plot showing the longitudinal relaxivity values (at 1.41 T) for the bare Gd-MSNs and pMAA-Gd-MSNs. The latter exhibits a  $\Delta r_1\%$  > 182%, with the  $r_1$  trend fitted using a Boltzmann equation to give an associated estimated  $pK_a = 5.2$  ( $R^2 = 0.99$ ) as expected.

prior reports.<sup>34,35</sup> The particles exhibited high colloidal uniformity with an associated size of  $49.5 \pm 4.2$  nm (resolved by transmission electron microscopy, TEM, ESI 1a†) and a corresponding pore diameter of  $3.2 \pm 0.2$  nm (Barrett-Joyner-Halenda pore size analysis, ESI 2†). A time delayed co-condensation method (with 0.15 mol% of an aminated silane added), biases the localisation of amino anchor groups at either the internal or external pore channel, as prior reported.<sup>36</sup> Chemical modification with an activated DOTA-NHS ester and subsequent metalation leads to the generation of the desired paramagnetic MSNs. The outer surface of Gd-MSNs was exclusively modified with a 2-(dodecylthiocarbonothioylthio)-2-methylpropionic acid modified silane (DDMAT-silane) chain transfer agent (CTA), ( $\sim 2.9$  nm in size, too large to enter the mesopores, ESI 3†). Particle modification was confirmed by ultraviolet-visible spectroscopy (UV-Vis, ESI 4†), thermogravimetric analysis (TGA, ESI 5†) and attenuated total reflectance infrared spectroscopy (ATR-IR, ESI 6†), with a calculated CTA grafting density of *ca.* 4.5 groups  $\text{nm}^{-2}$  (ESI 7†). This density facilitates the formation of a dense, thickness tuneable (based on RAFT conditions, ESI 8†), polymer brush coating from a wide variety of potential polymerizable monomers.<sup>37–39</sup> The generated polymer coated Gd-MSNs show an observed (and expected) increase in particle diameter after the SI-RAFT polymerisation (TEM, ESI 1b, c, and DLS ESI 8†). The living-character of the polymerisation process was confirmed by proton nuclear magnetic resonance spectroscopy ( $^1\text{H}$  NMR, ESI 9†), where monomer consumption was shown to follow pseudo-first-order kinetics (ESI 9 insets†).<sup>40,41</sup> The generated pMAA-Gd-MSNs possess high colloidal stability, across a broad pH range (in 10 mM Britton-Robinson buffer solution, pH 4.0–9.0), with extremely low polydispersity variation over 30 days (ESI 10,† polydispersity indices < 0.08). An expected pH-responsive switch in hydrodynamic diameter as the  $pK_a$  of pMAA<sup>42</sup> is traversed (ESI 8a†). The particles

exhibit no toxic effects during a 48 h exposure to HeLa cells or HEK-293T cells (ESI 11†).

Relaxivities of pMAA-Gd-MSNs were first assessed by NMR (1.4 T), where dramatic enhancements in  $r_1$  were observed as the  $pK_a$  of pMAA is traversed (Fig. 1b,  $\Delta r_1 = 30.3 \pm 3.2$   $\text{mM}^{-1} \text{s}^{-1}$  across 1.0 pH-unit, from pH 4.0 to pH 7.0). This “switch-on” response is not observed in the absence of a pMAA polymer coating; native Gd-MSNs exhibit constant relaxivities across the full pH range. In analogous poly(dimethylaminoethyl methacrylate) (pDMAEMA) brush coated particles, where a much weaker H-bond association with bulk water is expected,<sup>43,44</sup> (ESI 12†) the determined relaxivities overlap with those of bare Gd-MSNs and are non-responsive (ESI 13†). For the pMAA particles, associated image contrast enhancements were confirmed through spatially-resolved  $T_1$ -mapping experiments on clinical imaging scanners (1.5 T and 3 T; Fig. 2a), with associated  $T_1$  and  $R_1$  values shown in ESI 14.† These switches in  $r_1$  are the highest reported at both magnetic field strengths (Fig. 2b and c,  $\Delta r_1 = 50.5$   $\text{mM}^{-1} \text{s}^{-1}$  at 1.5 T and at  $\Delta r_1 = 21.8$   $\text{mM}^{-1} \text{s}^{-1}$  at 3 T).<sup>25,26</sup> It is also notable that the relaxivities, at both fields, exceed the theoretical maxima for solely IS contributions (optimised  $r_1^{IS} \sim 40$   $\text{mM}^{-1} \text{s}^{-1}$  at 1.5 T and  $\sim 20$   $\text{mM}^{-1} \text{s}^{-1}$  at 3 T),<sup>20,27</sup> suggesting the presence of a composite IS/OS contribution. In fact,<sup>6,45</sup> IS contributions are expected to be much lower than that here (*ca.* 16  $\text{mM}^{-1} \text{s}^{-1}$  at 1.5 T, seen ESI 16†). OS contributions are, then, both substantial and responsible for the observed  $\Delta r_1$  switch (more analyses detailed below).

In examining the effect of polymer thickness and Gd-localisation within the particle mesopores, we note that a larger magnitude switch is observed with thicker polymer shells (at 1.4 T, ESI 17a†) but that switching magnitudes are largely



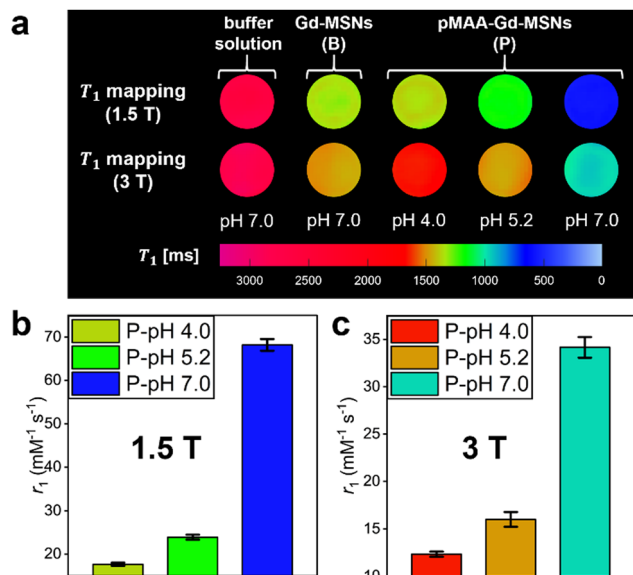


Fig. 2 (a)  $T_1$  maps, recoded at pH 4.0, pH 5.2 and pH 7.0 (1.5 T and 3 T clinical MRI scanners), for the pMAA-Gd-MSNs (denoted as "P-"). The  $T_1$  maps for the bare-Gd-MSNs are reported at pH 7.0 (denoted as "B-"). The MR derived relaxivities for the associated nanoparticles, across a range of pH, are included at both 1.5 T (b) and 3 T (c).

insensitive to Gd-depth (at 1.4 T, ESI 17b<sup>†</sup>). This is further confirmatory of a polymer-mediated origin.

To further examine the origin of this large magnitude relaxivity switch we refer to Solomon-Bloembergen-Morgan (SBM) theory (ESI 15<sup>†</sup>). Contributions from polymer-swelling induced changes in global particle rotation are negligible (ESI 16a<sup>†</sup>). An analysis of Eu-analogues confirms metal hydration to

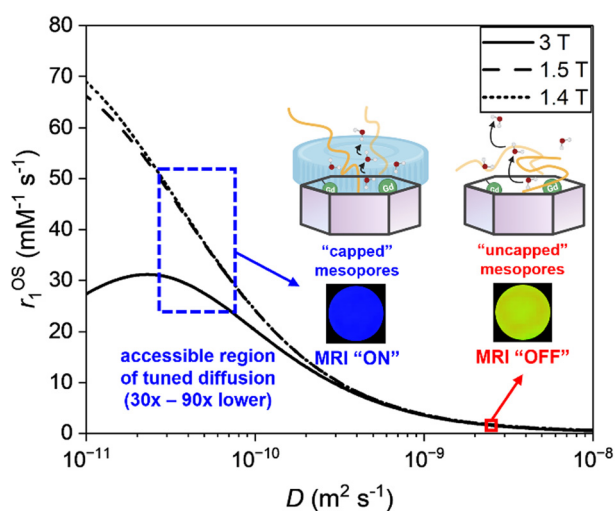


Fig. 3 The effect of the local diffusion coefficient of water on the outer-sphere relaxivity. This analytical treatment employs a modified Freed equation (at 1.4, 1.5 and 3 T), and considers the diffusion of water at  $r_{GdH} = 4.0$  Å from the paramagnetic centres (ref. 49). The diffusion coefficient of confined H<sub>2</sub>O with "uncapped" mesopores is taken from ref. 46. The diffusion coefficient of confined H<sub>2</sub>O with capped mesopores is deduced from the experimental relaxivity data, where  $D$  is observed to be reduced by 30–90x.

be the same (ESI 18<sup>†</sup>) in polymer modified particles at both pH 4.0 and pH 7.0. Additionally, the modulation of the water exchange rate ( $\tau_M$ ) through changes in the conformational state of the polymer cannot fully account for such a high observed switch in the acquired  $r_1$  value. Specifically, the modelled nuclear magnetic relaxation dispersion (NMRD) profile (ESI 16b<sup>†</sup>) for the IS contribution to  $r_1$  highlights that the role of  $\tau_M$  is much less significant than the expected influence of  $\tau_D$ , accounting only for (at best) ~20% of the switch in relaxivity. In recent work the presence of an MSN peripheral immobile water layer has been reported to have a substantial effect on the diffusion coefficient ( $D$ ) of pore-internal water.<sup>46</sup> It is known that a charged polyacid brush has an unusually strong association with water, dramatically reducing its mobility.<sup>47,48</sup> Within a modified SBM model (ESI 15<sup>†</sup>), an entirely realistic (30–90 fold) reduction in internal water diffusion,  $D$ , can account for the enhancement in relaxivity (Fig. 3). We propose, then, that the triggered polymer charging generates a peripheral particle water "cap" that dramatically increases OS relaxivity by virtue of its impact on particle internalised water.

To summarise, we present here paramagnetic inorganic-organic hybrid nanoparticles that exhibit a pH-mediated contrast switch that is sharp (across < 1.0 pH-unit), of an unprecedented magnitude, clinically relevant, and mechanistically new.

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## Conflicts of interest

There are no conflicts of interest to declare.

## References

- 1 D. Ni, W. Bu, E. B. Ehlerting, W. Cai and J. Shi, *Chem. Soc. Rev.*, 2017, **46**, 7438–7468.
- 2 J. Wahsner, E. M. Gale, A. Rodríguez-Rodríguez and P. Caravan, *Chem. Rev.*, 2019, **119**, 957–1057.
- 3 T. J. Clough, L. Jiang, K.-L. Wong and N. J. Long, *Nat. Commun.*, 2019, **10**, 1420.
- 4 J. Pellico, C. M. Ellis and J. J. Davis, *Contrast Media Mol. Imaging*, 2019, 1845637.
- 5 N. Zhao, L. Yan, X. Zhao, X. Chen, A. Li, D. Zheng, X. Zhou, X. Dai and F.-J. Xu, *Chem. Rev.*, 2019, **119**, 1666–1762.
- 6 F. Carniato, L. Tei, M. Cossi, L. Marchese and M. Botta, *Chem. – Eur. J.*, 2010, **16**, 10727–10734.
- 7 F. Carniato, L. Tei, A. Arrais, L. Marchese and M. Botta, *Chem. – Eur. J.*, 2013, **19**, 1421–1428.
- 8 K. M. L. Taylor, J. S. Kim, W. J. Rieter, H. An, W. Lin and W. Lin, *J. Am. Chem. Soc.*, 2008, **130**, 2154–2155.
- 9 F. Carniato, L. Tei and M. Botta, *Eur. J. Inorg. Chem.*, 2018, 4936–4954.
- 10 D. Yuan, C. M. Ellis and J. J. Davis, *Materials*, 2020, **13**, 3795.
- 11 F. Tang, L. Li and D. Chen, *Adv. Mater.*, 2012, **24**, 1504–1534.
- 12 W.-Y. Huang, G.-L. Davies and J. J. Davis, *Chem. Commun.*, 2013, **49**, 60–62.



- 13 J. S. Ananta, B. Godin, R. Sethi, L. Moriggi, X. Liu, R. E. Serda, R. Krishnamurthy, R. Muthupillai, R. D. Bolskar, L. Helm, M. Ferrari, L. J. Wilson and P. Decuzzi, *Nat. Nanotechnol.*, 2010, **5**, 815–821.
- 14 P. H. Fries and E. Belorizky, *J. Chem. Phys.*, 2010, **133**, 024504.
- 15 C. M. Ellis, J. Pellico and J. J. Davis, *Materials*, 2019, **12**, 4096.
- 16 C. d I H. Alarcón, S. Pennadam and C. Alexander, *Chem. Soc. Rev.*, 2005, **34**, 276–285.
- 17 Y. Lu, A. A. Aimetti, R. Langer and Z. Gu, *Nat. Rev. Mater.*, 2016, **2**, 16075.
- 18 I. Solomon, *Phys. Rev.*, 1955, **99**, 559–565.
- 19 N. Bloembergen and L. O. Morgan, *J. Chem. Phys.*, 1961, **34**, 842–850.
- 20 P. Caravan, C. T. Farrar, L. Frullano and R. Uppal, *Contrast Media Mol. Imaging*, 2009, **4**, 89–100.
- 21 L. Zhu, Y. Yang, K. Farquhar, J. Wang, C. Tian, J. Ranville and S. G. Boyes, *ACS Appl. Mater. Interfaces*, 2016, **8**, 5040–5050.
- 22 J. Pellico, C. M. Ellis, J. Miller and J. J. Davis, *Chem. Commun.*, 2019, **55**, 8540–8543.
- 23 C. Caro, M. L. García-Martín and M. Pernia Leal, *Biomacromolecules*, 2017, **18**, 1617–1623.
- 24 Y. Li, Y. Qian, T. Liu, G. Zhang and S. Liu, *Biomacromolecules*, 2012, **13**, 3877–3886.
- 25 G.-L. Davies, I. Kramberger and J. J. Davis, *Chem. Commun.*, 2013, **49**, 9704–9721.
- 26 S. Fu, Z. Cai and H. Ai, *Adv. Healthcare Mater.*, 2021, **10**, 2001091.
- 27 L. M. De León-Rodríguez, A. F. Martins, M. C. Pinho, N. M. Rofsky and A. D. Sherry, *J. Magn. Reson. Imaging*, 2015, **42**, 545–565.
- 28 P. Yang, D. Li, S. Jin, J. Ding, J. Guo, W. Shi and C. Wang, *Biomaterials*, 2014, **35**, 2079–2088.
- 29 B. Tian, S. Liu, W. Lu, L. Jin, Q. Li, Y. Shi, C. Li, Z. Wang and Y. Du, *Sci. Rep.*, 2016, **6**, 21335.
- 30 Y. Q. Yang, X. D. Guo, W. J. Lin, L. J. Zhang, C. Y. Zhang and Y. Qian, *Soft Matter*, 2012, **8**, 454–464.
- 31 S. R. Bonam, F. Wang and S. Muller, *Nat. Rev. Drug Discovery*, 2019, **18**, 923–948.
- 32 F. Wang and S. Muller, *Front. Immunol.*, 2015, **6**, 252.
- 33 S.-P. Ju, W.-J. Lee, C.-I. Huang, W.-Z. Cheng and Y.-T. Chung, *J. Chem. Phys.*, 2007, **126**, 224901.
- 34 W. Stober, A. Fink and E. Bohn, *J. Colloid Interface Sci.*, 1968, **26**, 62–69.
- 35 J. J. Davis, W. Y. Huang and G. L. Davies, *J. Mater. Chem.*, 2012, **22**, 22848–22850.
- 36 M. J. Hollamby, D. Borisova, P. Brown, J. Eastoe, I. Grillo and D. Shchukin, *Langmuir*, 2012, **28**, 4425–4433.
- 37 J. O. Zoppe, N. C. Ataman, P. Mocny, J. Wang, J. Moraes and H.-A. Klok, *Chem. Rev.*, 2017, **117**, 1105–1318.
- 38 R. Ranjan and W. J. Brittain, *Macromol. Rapid Commun.*, 2008, **29**, 1104–1110.
- 39 K. Ohno, Y. Ma, Y. Huang, C. Mori, Y. Yahata, Y. Tsujii, T. Maschmeyer, J. Moraes and S. Perrier, *Macromolecules*, 2011, **44**, 8944–8953.
- 40 A. Goto, K. Sato, Y. Tsujii, T. Fukuda, G. Moad, E. Rizzardo and S. H. Thang, *Macromolecules*, 2001, **34**, 402–408.
- 41 M. J. Flanders and W. M. Gramlich, *Polym. Chem.*, 2018, **9**, 2328–2335.
- 42 X. Wang, X. Ye and G. Zhang, *Soft Matter*, 2015, **11**, 5381–5388.
- 43 S. H. Min, S. K. Kwak and B.-S. Kim, *Soft Matter*, 2015, **11**, 2423–2433.
- 44 R. Chockalingam and U. Natarajan, *Mol. Phys.*, 2015, **113**, 3370–3382.
- 45 L. Tei, G. Gugliotta, Z. Baranyai and M. Botta, *Dalton Trans.*, 2009, 9712–9714.
- 46 M. Weigler, E. Winter, B. Kresse, M. Brodrecht, G. Buntkowsky and M. Vogel, *Phys. Chem. Chem. Phys.*, 2020, **22**, 13989–13998.
- 47 H. S. Sachar, T. H. Pial, P. R. Desai, S. A. Etha, Y. Wang, P. W. Chung and S. Das, *Matter*, 2020, **2**, 1509–1521.
- 48 H. S. Sachar, B. S. Chava, T. H. Pial and S. Das, *Macromolecules*, 2021, **54**, 2011–2021.
- 49 M. Botta, *Eur. J. Inorg. Chem.*, 2000, 399–407.

