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## Structural Imaging Predictors of Ketamine Response in Treatment-Resistant Depression: A Machine Learning Approach

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**Abstract**

Ketamine has demonstrated rapid antidepressant efficacy in treatment-resistant depression (TRD), but clinical decision-making is challenging due to variability in individual response. Current trial-and-error prescribing practices may expose patients to ineffective treatment and avoidable adverse effects, underscoring the need for reliable predictive tools to optimize treatment selection and support personalized, evidence-based care. We developed a machine-learning model (support vector classifier) to predict antidepressant response to ketamine using pre-treatment structural MRI data. The model was trained on 99 adults with TRD given a single intravenous ketamine infusion (0.5 mg/kg). Clinical response was defined as a  $\geq 50\%$  reduction in MADRS scores 24 hours post-infusion. Internal validation used repeated nested cross-validation, and generalizability was tested in two independent ketamine-treated cohorts ( $n = 51$ ) and a saline-treated control group ( $n = 49$ ). Among ketamine-treated participants, 52 (52.5%) responded to treatment. The model achieved a balanced accuracy of 72.2% (sensitivity = 72.3%, specificity = 73.1%, AUC = 0.72) in the discovery sample and 60.0% ( $p = .01$ , AUC = 0.65) in external validation. Greater gray matter volume in frontal regions predicted response, whereas greater cerebellar volume predicted non-response. Performance dropped to chance in the saline cohort (BAC = 41.1%, AUC = 0.45), supporting pharmacologic specificity. These findings present the first machine-learning model for the prediction of ketamine response in TRD using structural neuroimaging and highlight its potential utility for stratified treatment planning and biomarker-informed interventions while providing mechanistic insight into neuroanatomical predictors of antidepressant response.

## INTRODUCTION

Treatment-resistant depression (TRD), commonly defined as major depressive disorder (MDD) unresponsive to at least two adequate antidepressant trials, presents a significant clinical challenge[1–4]. Up to 60% of MDD patients fail to respond to initial pharmacotherapy, and many continue to experience persistent symptoms despite multiple subsequent interventions[1,5,6]. Current treatment approaches rely heavily on sequential medication trials; a trial-and-error process that delays effective intervention and increases the risk of chronicity and suicide[1,6–10].

In recent years, ketamine, an N-methyl-D-aspartate (NMDA) receptor antagonist, has emerged as a novel intervention for TRD, producing rapid antidepressant effects through proposed mechanisms including modulation of glutamatergic signaling and enhancement of synaptic plasticity[11–14]. Following a single subanesthetic intravenous infusion, nearly half of patients demonstrate a significant clinical response within 24 hours[12,15,16]. However, response is heterogeneous: while some patients achieve near-complete remission, others show minimal or transient benefit, and a subset experience adverse effects such as dissociation or symptom worsening[11,17,18].

The factors underlying response heterogeneity remain unclear[15,16]. In the absence of reliable predictors, clinicians cannot determine in advance which patients are likely to benefit from ketamine—potentially exposing some individuals to a treatment that offers no therapeutic gain, may carry adverse effects, and incurs substantial cost[11,15,19]. This variability highlights the need for predictive tools capable of identifying likely responders prior to treatment. Accurate stratification could optimize patient selection, reduce unnecessary exposure, and guide the development of more targeted interventions. For example, early identification of likely

responders could help avoid unnecessary exposure to ketamine in non-responders while enabling more timely intervention in high-risk individuals, including those with acute suicidality[20,21]. Importantly, predicting short-term response to ketamine may offer the highest clinical utility. Ketamine's rapid antidepressant effects typically emerge within 24 hours and may inform decisions about continued treatment or alternative strategies[12,22,23].

Previous research has explored various biological and clinical predictors of ketamine response, including peripheral inflammatory markers, cognitive processing speed, and dissociative symptoms[24–26]. However, these variables generally yield low individual-level predictive power and have not translated into actionable models[24,26]. More recently, neuroimaging studies have highlighted potential structural and functional differences between responders and non-responders in regions such as the anterior cingulate cortex, hippocampus, and prefrontal cortex, suggesting a neuroanatomical basis for response heterogeneity[27–29]. Still, findings remain inconsistent—likely due to small samples, methodological heterogeneity, and the limitations of conventional statistical approaches in capturing complex brain-behavior relationships[27,30].

Structural magnetic resonance imaging (sMRI) offers a non-invasive opportunity for investigating treatment-relevant brain morphology. Unlike functional MRI (fMRI) or electroencephalography (EEG), sMRI yields stable, trait-like morphometrics that can be standardized across sites and integrated into clinical workflows[27,30]. Systematic reviews have reported baseline volumetric differences in the anterior cingulate cortex and hippocampus as candidate predictors of antidepressant response to ketamine, but effect directions vary across studies, and no reproducible sMRI biomarker has been established to date[31–34].

To address these gaps, machine learning (ML) offers a data-driven framework capable of identifying high-dimensional, multivariate patterns that may better predict clinical outcomes [30,35]. ML algorithms can identify latent patterns across multiple brain regions that may distinguish responders from non-responders, with reported accuracies exceeding those of conventional regression-based approaches [36,37]. Predictive modeling of treatment response across neuroimaging modalities has shown promise in depression [36,38], but in ketamine specifically, existing ML studies remain limited. Previous efforts typically relied on a narrow set of pre-selected predictors, were constrained by small sample sizes, and employed only internal cross-validation without external test sets [39,40]. To date, no study has applied ML to pre-treatment structural MRI to predict treatment response to ketamine in TRD specifically.

In this study, we aimed to develop a supervised ML model trained on regional gray matter volume (GMV) parcellations derived from pre-treatment sMRI to predict short-term binary response to ketamine in patients with TRD. We chose to predict binary classification of response, as it offers a clinically interpretable endpoint that aligns with ketamine's time course of action and treatment decision timeline. We hypothesized that the model would distinguish responders from non-responders with above chance accuracy, and demonstrate external generalizability in independent TRD samples. Our findings aim to advance individualized treatment planning and offer mechanistic insight into the neuroanatomical substrates of ketamine response.

## **MATERIALS AND METHODS**

### **Study Design and Participants**

Our discovery sample used data from a single-site, double-blind, placebo-controlled randomized clinical trial of intravenous ketamine in adults with TRD (NCT03237286) [41].

Participants were randomized to receive a single IV infusion of ketamine (0.5 mg/kg) or saline over 40 minutes. Only patients from the ketamine-treated arm were included in model development. Clinical and demographic information were recorded at baseline. A total of 103 participants underwent pre-treatment MRI scanning; after quality control of imaging data (described below), 4 participants were excluded for poor image quality, yielding a final neuroimaging sample of 99.

Participants were aged 18 to 65 years and met DSM-5 criteria for major depressive disorder with non-response to at least one adequate antidepressant trial. Further inclusion and exclusion criteria are available on ClinicalTrials.gov (NCT03237286) and summarized in the supplement.

The external validation sample used data from two independent U.S.-based trials. From NCT00088699, 22 participants who received a single ketamine infusion and had available baseline sMRI and pre-/post-infusion MADRS data were included[12]. From NCT00768430, an additional 29 patients meeting the same criteria were included[22]. This yielded a pooled external validation cohort of 51 ketamine-treated TRD participants. Protocol details and eligibility criteria for both trials are available on ClinicalTrials.gov; brief trial descriptions and registry links are provided in the supplement.

### **MRI Acquisition and Image Preprocessing**

Baseline structural T1-weighted MRI scans were acquired prior to infusion using one of three 3T Siemens PRISMA scanners located at a single facility. Preprocessing was performed using the Computational Anatomy Toolbox (CAT12, version r2170) implemented within Statistical Parametric Mapping (SPM12) in MATLAB (version R2018b) for both the discovery and validation samples. Preprocessing included probabilistic tissue segmentation into gray

matter, white matter, and cerebrospinal fluid using CAT12's adaptive MAP approach with partial-volume estimation; spatial normalization to MNI space with Jacobian modulation of tissue maps to preserve local volume; and smoothing with an 8 mm Gaussian kernel. Image quality was assessed using the CAT12 image quality rating (IQR) metric; scans with an IQR below C were excluded, yielding a final sample with an average IQR of B.

From each processed image, regional gray-matter volume (GMV) features were derived from the modulated, normalized gray-matter maps produced by CAT12 (standard Voxel-based Morphometry preprocessing). Regional GMV values were summarized within 136 regions of interest from the Neuromorphometrics atlas and used as input features for the machine-learning analyses[42]. Although CAT12 can also generate surface-based morphometry measures (projection-based cortical thickness and surface area), these were not included here; we focused on volumetric GMV to obtain whole-brain (cortical and subcortical) coverage with an appropriate feature dimensionality for our models.

We additionally derived each subject's total intracranial volume (TIV) to use as a covariate, since head size can affect raw brain volumes. We also included a site covariate corresponding to the MRI acquisition protocol version for each scan (a binary indicator distinguishing two slightly different scanner sequences used over the course of the study), as well as participants' age and sex, as additional covariates. By accounting for these variables in our analysis, we aimed to isolate neuroanatomical differences related to treatment outcome rather than confounds such as age or scanner parameters.

### **Outcome Definition**

Antidepressant response was defined as a  $\geq 50\%$  reduction in MADRS scores from pre-infusion to 24 hours post-infusion, consistent with prior ketamine studies[12,15,22,23,41].

MADRS percentage change was calculated as:

$$\Delta MADRS\% = \frac{MADRS_{post} - MADRS_{pre}}{MADRS_{pre}} \times 100$$

Patients were categorized into response or nonresponse groups and these served as outcome labels for classification. Of the 99 patients in the discovery sample, 52 met response criteria (responders), while 47 did not (non-responders), and these group labels were used as classification outcomes.

### Machine Learning Analysis

All machine learning analyses were performed using NeuroMiner (v1.3; GitHub [<https://github.com/neurominer-git/NeuroMiner-1>]). Model training, preprocessing and hyperparameter optimization were performed within a repeated nested cross-validation (NCV) framework to avoid overfitting and ensure generalizability. The outer loop (CV2) used 3-fold cross-validation repeated 5 times. Each CV2 training set was further subdivided into an inner loop (CV1) using 3-fold cross-validation repeated 5 times for hyperparameter tuning and feature optimization. Figure 1 illustrates the NCV structure and model development pipeline.

The GMV parcellations were preprocessed by first scaling to the range  $[-1, 1]$ , and removing non-informative features (e.g., zero variance, NaN). Partial correlations were used to regress out covariates (age, sex, scanner sequence, and TIV): and robust principal component analysis (RobPCA) reduced dimensionality to 13 components[43]. A second scaling step was performed post-PCA to ensure all principal components had equal range. Feature selection was

conducted using a forward greedy wrapper, iteratively retaining (in steps of 10%) the top 20% of features that contributed to model performance.

Following pre-processing, a linear support vector machine classifier (SVC) was trained to classify responders versus non-responders. The SVC employed a linear solver with L2-regularized L2-loss, dual formulation, no kernel, and a tolerance of 0.01, with hyperplane weighting to account for class imbalances. The regularization parameter (C) was optimized over 11 values (0.0156, 0.0312, 0.0625, 0.1250, 0.2500, 0.5000, 1, 2, 4, 8, and 16). To evaluate the statistical significance of model performance, we conducted label permutation testing with 1,000 iterations and assessing performance using BAC, sensitivity, specificity, and AUC[44].

### **Model Interpretability, Specificity Testing, and External Validation**

To assess the neuroanatomical features contributing most strongly to classification, feature weights were analyzed across folds. To interpret regional contributions, model weights were back-projected from the PCA-transformed space to the original GMV feature space, enabling identification of the most predictive brain regions. Consistency and robustness of each predictor were quantified using cross-validation ratios (CVRs), calculated as the mean weight divided by its standard error across folds, and permutation-derived sign-based consistency tests, controlling for false discovery rate (FDR). The CVR measures the stability, magnitude and effect of each feature[45].

To evaluate whether the model learned patterns specific to ketamine-related response, the trained model was applied to the independent saline-treated cohort (n = 49) from the same trial as the discovery cohort. A reduction in predictive performance in this cohort would support specificity to ketamine mechanisms. The model was then applied to the pooled external validation sample to determine model generalizability. Scanning protocols varied across datasets,

but no harmonization was applied, to emulate real clinical use of our model; instead, site/sequence effects were addressed earlier via covariate regression within the nested cross-validation framework.

### **Post Hoc Transcriptomic Annotation of Predictive Regions**

We leveraged the abagen toolbox<sup>[46]</sup> to extract transcriptomic data from the Allen Human Brain Atlas (AHBA), which includes normalized microarray expression profiles for approximately 20,000 genes sampled from six adult human donors. <sup>[47]</sup> Using the Neuromorphometrics atlas aligned to MNI space, we focused on the six brain regions that emerged as the top structural predictors of ketamine response in our machine learning model. Upstream QC steps included probe filtering, donor consolidation, and gene/sample normalization per default abagen settings. <sup>[48]</sup> We specifically extracted expression levels for seven receptor genes—NMDA receptor subunits (GRIN1, GRIN2A, GRIN2B), AMPA receptor subunits (GRIA1, GRIA2), and GABA<sub>A</sub> receptor subunits (GABRA1, GABRB2)—selected based on their established pharmacological relevance to ketamine’s mechanism of action. <sup>[49,50]</sup>

### **Ethics approval and consent to participate**

The data used in this study were obtained from previously conducted clinical trials (NCT03237286, NCT00088699, and NCT00768430), each of which received approval from the relevant institutional review boards/ethics committees. Written informed consent was obtained from all participants in the original studies.

## **RESULTS**

### **Sample Characteristics**

Demographic and clinical characteristics for all cohorts are summarized separately in Table 1. The discovery sample had a mean (SD) age of 34.81 (11.01) years, with 62 (62.6%) female and 37 male (37.4%) participants. 52 patients (52.5%) were classified as responders and 47 (47.5%) as non-responders. Responders and non-responders had no difference in age ( $34.8 \pm 11.7$  vs.  $34.8 \pm 10.3$  years) or sex distributions (65.4% vs. 59.6% female, respectively). Baseline MADRS scores were comparable across groups ( $\sim 32$ ), while post-treatment scores differed substantially (mean = 9.1 for responders vs. 24.9 for non-responders).

The saline-treated validation sample (mean (SD) age, 33.84 (9.78) years; 31 female (63.3%); 18 male (36.7%)) included 12 responders (24.5%) and 37 (75.5%) non-responders. The pooled external validation sample (mean (SD) age, 42.84 (14.70) years; 25 female (49.0%), 26 male (51.0%)) included 21 responders (41.2%) and 30 non-responders (58.8%).

## **Machine Learning Analyses**

### Classification Performance in Discovery Sample.

The SVC trained on pre-treatment GMV parcellations achieved a BAC of 72.2%, sensitivity of 72.3%, specificity of 73.1%, and an AUC of 0.72. Permutation testing confirmed statistical significance ( $p < .001$ ). Performance metrics are detailed in Table 2.

### Feature Importance and Regional Contributions

Increased GMV in frontal regions, including the left medial superior frontal gyrus, right superior frontal gyrus, and left frontal pole, was associated with a higher likelihood of response to ketamine. In contrast, greater volume in cerebellar regions, including cerebellar vermal lobules I–V, VI–VII, and the left cerebellar exterior, predicted non-response. These patterns are visualized in Figure 2.

### Mechanistic Specificity Assessment in Saline-Treated Cohort

To evaluate whether the model signature was specific to ketamine, the trained classifier was applied to the independent saline-treated TRD cohort. Model performance declined, yielding a balanced accuracy of 41.1% and an AUC of 0.45. These values did not exceed chance performance ( $p = .88$ , permutation test). This lack of generalization supports the specificity of the model for ketamine-induced antidepressant effects, rather than nonspecific symptom improvement or measurement artifacts.

### External Validation in Independent Ketamine Cohorts

The generalizability of the model was further tested in a pooled external validation cohort of ketamine-treated TRD patients drawn from geographically and methodologically distinct sites. Despite this heterogeneity, the model retained moderate performance predicting response in the external cohort (BAC 60.0%, sensitivity 66.7%, specificity 53.3%, AUC 0.65). Statistical significance was confirmed via permutation testing ( $p = .01$ ). These results support the external validity of the model across geographically and methodologically distinct TRD cohorts. Validation performance is summarized in Table 2.

### **Transcriptomic Characterization of Predictive Brain Regions**

Figure 4 displays normalized expression levels of ketamine-relevant receptor subunits across the six most predictive brain parcels identified by the ML model—three positively weighted (response-associated) frontal regions and three negatively weighted (non-response-associated) cerebellar regions. Frontal regions—where increased volume predicted treatment response—exhibited higher expression of NMDA subunits (GRIN1, GRIN2A, GRIN2B) and AMPA subunits (GRIA1, GRIA2) (range  $\approx 0.7$ – $0.9$ ) relative to cerebellar regions, which predicted non-response and showed lower expression (range  $\approx 0.2$ – $0.5$ ). Notably, GABA-A

subunit genes (GABRA1, GABRB2) were also elevated in frontal parcels, though differences were more modest.

## **DISCUSSION**

### **Prediction**

#### Main Findings

In this study, we developed an externally validated machine learning model that predicts short-term antidepressant response to IV ketamine in TRD patients using pre-treatment GMV data. We identified a multivariate neuroanatomical signature distinguishing ketamine responders from non-responders with a BAC of 72% in the original sample and 60% in an independent external cohort. This predictive performance substantially exceeds chance and is, to our knowledge, the first externally validated model for ketamine response in a TRD-only sample using structural imaging.

#### Relation to prior prediction work

Our work builds on prior studies that have sought biological predictors of ketamine's antidepressant effects[24–26]. Consistent with the literature, our results support the idea that neurobiological heterogeneity underlies differential ketamine responses[27,33]. Existing prediction efforts have largely relied on univariate associations or clinical markers, including symptom severity, inflammatory cytokines, or dissociative responses, which typically lack individual-level predictive power and fail to generalize across cohorts[24–26,39,51]. Even prior neuroimaging studies—often limited by small sample sizes or mixed MDD/TRD populations—report modest internal accuracy (~60–65%) and rarely attempt out-of-sample testing[40]. Moreover, most previous machine learning models do not focus on TRD specifically, despite this population carrying greater burden and clinical urgency[7,52]. In clinical practice, ketamine

remains administered largely by trial and error, with response rates showing high heterogeneity across patients and studies, including with repeated dosing, despite high costs, monitoring needs, and potential for dissociative or adverse effects[11,19,22,53]. Our model demonstrates the potential to address this unmet need by using pre-treatment structural imaging to identify a reproducible neuroanatomical signature of response, offering a pathway toward more personalized, evidence-informed treatment selection in TRD.

#### Generalizability and Mechanistic Specificity of the Model

External validation of our model demonstrated reasonable generalizability (60% BAC), despite inherent challenges such as heterogeneity in samples and MRI protocols, indicating the signature is not site-idiosyncratic. In contrast, the classifier failed to generalize to a saline/placebo cohort (BAC 41%), functioning as a negative control and supporting a ketamine-specific signal rather than a general prognostic marker. Together, these results increase confidence that the model captures pharmacologically relevant variance while illustrating the expected attenuation from discovery to external testing.

#### Clinical Implications

Our results carry potential practical implications for the clinical management of TRD. At present, the findings show proof-of-concept evidence that a reproducible neuroanatomical signal exists across heterogeneous ketamine datasets. If replicated and translated into a user-friendly tool, an MRI-based ketamine response predictor could substantially improve treatment planning. In current practice, clinicians have little objective basis for choosing ketamine for one patient vs. another aside from clinical impression and prior treatment history. Our MRI-based predictive model could prospectively identify likely responders, helping clinicians prioritize ketamine for appropriate candidates and spare likely non-responders unnecessary infusion, monitoring, and

side-effects. Many patients with TRD already undergo MRI during clinical work-ups or research participation; these routine scans could be repurposed to provide predictive information.

## **Mechanistic Insights**

### Main Findings

Two neuroanatomical features emerged as particularly predictive of ketamine response: gray matter volume in the frontal cortex and cerebellum. Greater frontal cortical volume at baseline was associated with response, whereas increased cerebellar volume was characteristic of non-responders. These structural features offer mechanistic insight into the neurobiological substrates modulating ketamine's antidepressant action.

### Frontal cortex

Frontal cortical morphology has consistently been implicated in ketamine's efficacy. The frontal regions identified as being important predictors of response overlap with dorsolateral and anterior prefrontal regions known to be important in the cognitive control of emotion; thought to be hypoactive in neurobiological models of depression; and used as targets in neuromodulation treatments such as transcranial magnetic stimulation, and direct current stimulation[54–57]. Mechanistically, ketamine induces a glutamate surge via NMDA receptor antagonism, which initiates synaptogenesis and restores connectivity in prefrontal circuits disrupted by chronic stress[58]. This cascade is thought to be driven in part by BDNF signaling and mTOR activation, facilitating rapid structural plasticity[13,14,49,50,59]. Patients with greater medial and superior frontal volume may have enhanced “neuroplastic reserve”—a denser or more intact neuronal architecture that better supports ketamine-induced remodeling. This interpretation aligns with prior findings associating higher pre-treatment activity or volume in frontal regions, including the rostral anterior cingulate, with superior clinical response to ketamine[31,60].

### Cerebellum

By contrast, the association of larger cerebellar volume with non-response presents a more novel and less intuitive observation. The cerebellum is increasingly being recognized for its role in mood, cognition, and emotion through bidirectional cerebello-cortical loops. Structural alterations in cerebellar lobules—particularly vermal and hemispheric subregions—have been reported in depression[61,62]. In our study, TRD patients with non-response exhibited relative hypertrophy in these same regions. This finding may reflect a hypothesis-generating observation consistent with a maladaptive compensatory phenotype in which aberrant cerebellar morphology contributes to persistent network dysfunction.

### Transcriptomic bridge

To explore potential molecular correlates of these structural findings, we leveraged transcriptomic data from the Allen Human Brain Atlas using the abagen toolbox.[46–48] Gene expression analysis revealed that frontal regions—those associated with response—exhibited higher expression of NMDA receptor subunits (GRIN1, GRIN2A, GRIN2B), AMPA subunits (GRIA1, GRIA2), and GABA-A subunits (GABRA1, GABRB2) relative to negatively weighted cerebellar predictor regions identified by the model- patterns consistent with ketamine's glutamate–AMPA–BDNF/mTOR cascade[49,50,63]. These results provide biological plausibility for our structural findings: frontal regions may serve as optimal substrates for ketamine's pharmacodynamic effects, while enlarged cerebellar regions, with comparatively sparse receptor expression, may represent structurally dominant but functionally inert nodes within the antidepressant response network.

We therefore propose that cerebellar hypertrophy in non-responders may signify a morphologically entrenched and molecularly refractory circuit phenotype—potentially a high-

gain, low-plasticity feedback loop that exerts sustained influence over medial prefrontal systems. In this model, ketamine's cortical effects are counteracted by maladaptive cerebellar output, undermining antidepressant response. Meanwhile, patients with preserved frontal volume and higher receptor expression are structurally and molecularly positioned to benefit from ketamine-induced plasticity.

This dual-anatomical and transcriptomic framework extends current models of ketamine response and highlights the importance of considering cerebellar morphology not simply as a peripheral correlate, but as a potential determinant of treatment resistance in TRD.

### **Strengths and Limitations**

This study's strengths include, to our knowledge, being the first to demonstrate that structural MRI alone can support individual-level prediction of short-term ketamine response in a TRD-only cohort, with repeated nested cross-validation and independent external validation—addressing a gap noted in recent ketamine neuroimaging reviews and MRI prediction meta-analyses[27,33,64]. Methodologically, we combined whole-brain GMV parcellations, repeated nested cross-validation with permutation testing, external validation, and a saline negative-control generalization test—design choices aligned with best-practice recommendations for clinical prediction modelling[35,65]. Finally, using pretreatment sMRI, a stable and broadly available modality with demonstrated multi-centre reproducibility, increases translational potential.

Limitations include modest sample sizes, particularly in the external and saline cohorts, widening uncertainty around point estimates. We tested 24-hour outcomes only; whether baseline sMRI predicts durability of benefit remains unknown. Features were restricted to regional GMV; adding thickness, surface area, connectomic measures, and clinical/cognitive

variables may improve performance. We did not apply inter-site harmonization to the external ketamine cohorts, as healthy control data were unavailable to support conventional group-based methods such as ComBat. While this may have contributed to variability, the model's significant performance under these conditions supports its robustness and real-world transportability. Furthermore, participant-level psychiatric comorbidity information was not available across cohorts. We were, therefore, unable to evaluate whether comorbid conditions such as anxiety disorders influenced model predictions. Future studies with harmonized clinical phenotyping will be important to determine whether the observed neuroanatomical signature remains robust across diagnostic heterogeneity. Lastly, the post hoc imaging-transcriptomic analysis should also be interpreted cautiously, as the AHBA is based on six adult postmortem donors with incomplete regional sampling and does not reflect patient-specific gene expression.

### **Conclusions and Future Directions**

In this study, we present the first externally validated AI-based model of response to ketamine using machine learning models trained on pretreatment structural neuroimaging data. The model was specific to ketamine and implicated the superior frontal gyri, the left frontal pole, and the cerebellar vermal and lateral volumes in ketamine's neurobiological mechanism of action. These findings provide initial evidence that readily obtainable baseline sMRI scans could inform individualized decisions about ketamine treatment, potentially shortening the protracted trial-and-error trajectory that typifies care in TRD. Our results show promise that an sMRI-guided decision-support tool could help clinicians prioritize likely responders, spare non-responders unnecessary exposure, and shorten time-to-benefit in TRD pathways. Future work should extend validation to additional, independent datasets, evaluate predictors of longer-term outcomes, and test whether incorporating complementary data types such as clinical,

functional, or molecular measures further enhances prediction, and determine whether such models can inform treatment selection relative to other intervention.

### **Author contributions**

LB performed all analyses and drafted the manuscript. RP contributed the imaging and associated clinical data for the discovery cohort. LA provided the imaging data for the Maryland external validation cohort. JWM, YJ, JJ, ML, PTN, and LSM contributed the imaging data for the New York (Mount Sinai) external validation cohort and supported data processing and validation analyses. All other authors provided critical revisions to the manuscript. FC and PAL supervised the study and provided critical revisions. All authors reviewed and approved the final manuscript.

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### **Data availability**

Data were taken from previously conducted clinical trials (NCT03237286, NCT00088699, and NCT00768430). For details on the data and access requests, please contact the original study investigators, subject to appropriate data sharing agreements.

### **Competing Interests**

In the past 24 months, Dr. Murrough has provided consultation services for Autobahn Therapeutics, Inc., Biohaven Pharmaceuticals, Inc., Cliniclabs, Inc., Clexio Biosciences, Ltd., Compass Pathfinder, Plc., Dr Jay, Frontier Pharma, LLC, HMP Collective, Janssen Pharmaceuticals, LivaNova, Plc., Merck & Co., Inc., Otsuka Pharmaceutical, Ltd, WCG Clinical, Inc., and Xenon Pharmaceuticals, Inc. The Icahn School of Medicine (employer of Dr. Murrough) is named on a patent and has entered into a licensing agreement and will receive payments related to the use of ketamine or esketamine for the treatment of depression. The Icahn School of Medicine is also named on a patent related to the use of ketamine for the treatment of PTSD. Dr. Murrough is not named on these patents and will not receive any payments. Prof. Mehta has provided consultation services for Neurocentrx, Curie.bio and Nxera. Prof. Price is the named inventor on a University of Pittsburgh-owned patent filing optioned to TaJa Health, Inc., and has received consulting fees from Lightstone Ventures. Dr. Lalousis has received honoraria for talks presented at educational meetings organized by Boehringer Ingelheim outside the submitted work. All other authors report no potential conflicts of interest.

### **REFERENCES**

1. Thase M, Rush J. When at First You Don't Succeed: Sequential Strategies for Antidepressant Non responders. *J Clin Psychiatry* [Internet]. 1997 [cited 2024 Jul 31];58:23–9. [chrome-extension://efaidnbmnnnibpcajpcglclefindmkaj/https://www.psychiatrist.com/wp-content/uploads/2021/02/12682\\_dont-succeed-sequential-strategies-antidepressant.pdf](https://www.psychiatrist.com/wp-content/uploads/2021/02/12682_dont-succeed-sequential-strategies-antidepressant.pdf). Accessed 31 Jul 2024
2. Gaynes BN, Lux L, Gartlehner G, Asher G, Forman-Hoffman V, Green J, et al. Defining treatment-resistant depression. *Depress Anxiety* [Internet]. Blackwell Publishing Inc.; 2020 [cited 2025 Aug 21];37:134–45. <https://doi.org/10.1002/DA.22968>,

3. McIntyre RS, Alsuwaidan M, Baune BT, Berk M, Demyttenaere K, Goldberg JF, et al. Treatment-resistant depression: definition, prevalence, detection, management, and investigational interventions. *World Psychiatry* [Internet]. John Wiley and Sons Inc; 2023 [cited 2025 Aug 21];22:394. <https://doi.org/10.1002/WPS.21120>
4. Sforzini L, Worrell C, Kose M, Anderson IM, Aouizerate B, Arolt V, et al. A Delphi-method-based consensus guideline for definition of treatment-resistant depression for clinical trials. *Mol Psychiatry* [Internet]. Springer Nature; 2022 [cited 2025 Aug 21];27:1286–99. <https://doi.org/10.1038/S41380-021-01381-X>;SUBJMETA=1414,2421,476,53,692,699;KWRD=DEPRESSION,DIAGNOSTIC+MARKER S
5. Henssler J, Alexander D, Schwarzer G, Bschor T, Baethge C. Combining Antidepressants vs Antidepressant Monotherapy for Treatment of Patients With Acute Depression: A Systematic Review and Meta-analysis. *JAMA Psychiatry* [Internet]. American Medical Association; 2022 [cited 2025 Aug 21];79:300–12. <https://doi.org/10.1001/JAMAPSYCHIATRY.2021.4313>
6. Rush AJ, Trivedi MH, Wisniewski SR, Nierenberg AA, Stewart JW, Warden D, et al. Acute and longer-term outcomes in depressed outpatients requiring one or several treatment steps: A STAR\*D report. *American Journal of Psychiatry* [Internet]. American Psychiatric Association; 2006 [cited 2025 Aug 21];163:1905–17. <https://doi.org/10.1176/AJP.2006.163.11.1905/ASSET/IMAGES/R114F4.JPEG>
7. Sussman M, Menzin J, O’Sullivan AK, Shah A, Olfson M. Economic burden of treatment-resistant depression on the U.S. Health care system. *J Manag Care Spec Pharm* [Internet]. Academy of Managed Care Pharmacy (AMCP); 2019 [cited 2025 Aug 21];25:823–35. <https://doi.org/10.18553/JMCP.2019.25.7.823>,
8. Kern DM, Canuso CM, Daly E, Johnson JC, Fu DJ, Doherty T, et al. Suicide-specific mortality among patients with treatment-resistant major depressive disorder, major depressive disorder with prior suicidal ideation or suicide attempts, or major depressive disorder alone. *Brain Behav* [Internet]. John Wiley and Sons Ltd; 2023 [cited 2025 Aug 21];13:e3171. <https://doi.org/10.1002/BRB3.3171>;REQUESTEDJOURNAL:JOURNAL:21579032;WGROU: STRING:PUBLICATION
9. Mann JJ, Michel CA, Auerbach RP. Improving Suicide Prevention Through Evidence-Based Strategies: A Systematic Review. *Am J Psychiatry* [Internet]. Am J Psychiatry; 2021 [cited 2023 Dec 13];178:611–24. <https://doi.org/10.1176/APPI.AJP.2020.20060864>
10. Lundberg J, Cars T, Lööv SÅ, Söderling J, Sundström J, Tiihonen J, et al. Association of Treatment-Resistant Depression With Patient Outcomes and Health Care Resource Utilization in a Population-Wide Study. *JAMA Psychiatry* [Internet]. American Medical Association; 2023 [cited 2025 Aug 21];80:167–75. <https://doi.org/10.1001/JAMAPSYCHIATRY.2022.3860>
11. Sanacora G, Frye MA, McDonald W, Mathew SJ, Turner MS, Schatzberg AF, et al. A Consensus Statement on the Use of Ketamine in the Treatment of Mood Disorders. *JAMA Psychiatry* [Internet]. American Medical Association; 2017 [cited 2024 Jan 16];74:399–405. <https://doi.org/10.1001/JAMAPSYCHIATRY.2017.0080>
12. Zarate CA, Singh JB, Carlson PJ, Brutsche NE, Ameli R, Luckenbaugh DA, et al. A Randomized Trial of an N-methyl-D-aspartate Antagonist in Treatment-Resistant Major Depression. *Arch Gen Psychiatry* [Internet]. American Medical Association; 2006 [cited 2025 Aug 21];63:856–64. <https://doi.org/10.1001/ARCHPSYC.63.8.856>
13. Maeng S, Zarate CA. The role of glutamate in mood disorders: results from the ketamine in major depression study and the presumed cellular mechanism underlying its antidepressant

- effects. *Curr Psychiatry Rep* [Internet]. *Curr Psychiatry Rep*; 2007 [cited 2024 Aug 21];9:467–74. <https://doi.org/10.1007/S11920-007-0063-1>
14. Duman RS, Aghajanian GK, Sanacora G, Krystal JH. Synaptic plasticity and depression: new insights from stress and rapid-acting antidepressants. *Nature Medicine* 2016 22:3 [Internet]. Nature Publishing Group; 2016 [cited 2025 Aug 21];22:238–49. <https://doi.org/10.1038/nm.4050>
15. Price RB, Kissel N, Baumeister A, Rohac R, Woody ML, Ballard ED, et al. International pooled patient-level meta-analysis of ketamine infusion for depression: In search of clinical moderators. *Mol Psychiatry* [Internet]. Springer Nature; 2022 [cited 2025 Aug 21];27:5096–112. <https://doi.org/10.1038/S41380-022-01757-7>;SUBJMETA=1414,476,477,631,692,699;KWRD=DEPRESSION,PSYCHOLOGY
16. Alnefeesi Y, Chen-Li D, Krane E, Jawad MY, Rodrigues NB, Ceban F, et al. Real-world effectiveness of ketamine in treatment-resistant depression: A systematic review & meta-analysis. *J Psychiatr Res*. Pergamon; 2022;151:693–709. <https://doi.org/10.1016/J.JPSYCHIRES.2022.04.037>
17. Williamson D, Turkoz I, Wajs E, Singh JB, Borentain S, Drevets WC. Adverse Events and Measurement of Dissociation After the First Dose of Esketamine in Patients With TRD. *International Journal of Neuropsychopharmacology* [Internet]. Oxford Academic; 2023 [cited 2025 Aug 21];26:198–206. <https://doi.org/10.1093/IJNP/PYAC081>
18. Serafini G, Howland RH, Rovedi F, Girardi P, Amore M. The Role of Ketamine in Treatment-Resistant Depression: A Systematic Review. *Curr Neuropharmacol* [Internet]. Bentham Science Publishers; 2014 [cited 2024 Aug 21];12:444. <https://doi.org/10.2174/1570159X12666140619204251>
19. Parikh S V., Lopez D, Vande Voort JL, Rico J, Achtyes E, Coryell W, et al. Developing an IV Ketamine Clinic for Treatment-Resistant Depression: a Primer. *Psychopharmacol Bull* [Internet]. NLM (Medline); 2021 [cited 2025 Aug 21];51:109. <https://pubmed.ncbi.nlm.nih.gov/articles/PMC8374924/>. Accessed 21 Aug 2025
20. Wilkinson ST, Ballard ED, Bloch MH, Mathew SJ, Murrough JW, Feder A, et al. The effect of a single dose of intravenous ketamine on suicidal ideation: a systematic review and individual participant data meta-analysis. *Am J Psychiatry* [Internet]. American Psychiatric Association; 2017 [cited 2025 Aug 21];175:150. <https://doi.org/10.1176/APPI.AJP.2017.17040472>
21. Grunebaum MF, Galfalvy HC, Choo TH, Keilp JG, Moitra VK, Parris MS, et al. Ketamine for rapid reduction of suicidal thoughts in major depression: a midazolam-controlled randomized clinical trial. *Am J Psychiatry* [Internet]. American Psychiatric Association; 2017 [cited 2025 Aug 21];175:327. <https://doi.org/10.1176/APPI.AJP.2017.17060647>
22. Murrough JW, Iosifescu D V., Chang LC, Al Jurdi RK, Green CE, Perez AM, et al. Antidepressant Efficacy of Ketamine in Treatment-Resistant Major Depression: A Two-Site Randomized Controlled Trial. *Am J Psychiatry* [Internet]. American Psychiatric Association; 2013 [cited 2025 Aug 21];170:1134. <https://doi.org/10.1176/APPI.AJP.2013.13030392>
23. Murrough JW, Perez AM, Pillemer S, Stern J, Parides MK, Aan Het Rot M, et al. Rapid and longer-term antidepressant effects of repeated ketamine infusions in treatment-resistant major depression. *Biol Psychiatry* [Internet]. Biol Psychiatry; 2013 [cited 2025 Oct 1];74:250–6. <https://doi.org/10.1016/j.biopsych.2012.06.022>
24. Rong C, Park C, Rosenblat JD, Subramaniapillai M, Zuckerman H, Fus D, et al. Predictors of response to ketamine in treatment resistant major depressive disorder and bipolar disorder. *Int J Environ Res Public Health* [Internet]. MDPI; 2018 [cited 2025 Aug 21];15. <https://doi.org/10.3390/IJERPH15040771>,

25. Kiraly DD, Horn SR, Van Dam NT, Costi S, Schwartz J, Kim-Schulze S, et al. Altered peripheral immune profiles in treatment-resistant depression: Response to ketamine and prediction of treatment outcome. *Transl Psychiatry* [Internet]. Nature Publishing Group; 2017 [cited 2025 Aug 21];7. <https://doi.org/10.1038/TP.2017.31>,
26. Niciu MJ, Luckenbaugh DA, Ionescu DF, Guevara S, Machado-Vieira R, Richards EM, et al. Clinical predictors of ketamine response in treatment-resistant major depression. *Journal of Clinical Psychiatry* [Internet]. Physicians Postgraduate Press Inc.; 2014 [cited 2025 Aug 21];75. <https://doi.org/10.4088/JCP.13M08698>,
27. Zavaliangos-Petropulu A, Al-Sharif NB, Taraku B, Leaver AM, Sahib AK, Espinoza RT, et al. Neuroimaging-Derived Biomarkers of the Antidepressant Effects of Ketamine. *Biol Psychiatry Cogn Neurosci Neuroimaging* [Internet]. Elsevier Inc.; 2023 [cited 2025 Aug 21];8:361–86. <https://doi.org/10.1016/j.bpsc.2022.11.005>
28. Zhou YL, Wu FC, Liu WJ, Zheng W, Wang CY, Zhan YN, et al. Volumetric changes in subcortical structures following repeated ketamine treatment in patients with major depressive disorder: a longitudinal analysis. *Transl Psychiatry* [Internet]. Springer Nature; 2020 [cited 2025 Aug 21];10. <https://doi.org/10.1038/S41398-020-00945-9>,
29. Abdallah CG, Jackowski A, Salas R, Gupta S, Sato JR, Mao X, et al. The Nucleus Accumbens and Ketamine Treatment in Major Depressive Disorder. *Neuropsychopharmacology* [Internet]. Nature Publishing Group; 2017 [cited 2025 Aug 21];42:1739–46. <https://doi.org/10.1038/NPP.2017.49>,
30. Woo CW, Chang LJ, Lindquist MA, Wager TD. Building better biomarkers: Brain models in translational neuroimaging. *Nat Neurosci* [Internet]. Nature Publishing Group; 2017 [cited 2025 Aug 21];20:365–77. <https://doi.org/10.1038/NN.4478>,
31. Herrera-Melendez A, Stippl A, Aust S, Scheidegger M, Seifritz E, Heuser-Collier I, et al. Gray matter volume of rostral anterior cingulate cortex predicts rapid antidepressant response to ketamine. *European Neuropsychopharmacology* [Internet]. Elsevier B.V.; 2021 [cited 2025 Aug 29];43:63–70. <https://doi.org/10.1016/j.euroneuro.2020.11.017>
32. Abdallah CG, Salas R, Jackowski A, Baldwin P, Sato JR, Mathew SJ. Hippocampal Volume And The Rapid Antidepressant Effect Of Ketamine. *J Psychopharmacol* [Internet]. SAGE Publications Ltd; 2014 [cited 2025 Aug 29];29:591. <https://doi.org/10.1177/0269881114544776>
33. Yun JY, Kim YK. Neural correlates of treatment response to ketamine for treatment-resistant depression: A systematic review of MRI-based studies. *Psychiatry Res* [Internet]. Elsevier Ireland Ltd; 2024 [cited 2025 Aug 29];340. <https://doi.org/10.1016/j.psychres.2024.116092>
34. Medeiros GC, Matheson M, Demo I, Reid MJ, Matheson S, Twose C, et al. Brain-based correlates of antidepressant response to ketamine: a comprehensive systematic review of neuroimaging studies. *Lancet Psychiatry* [Internet]. Elsevier Ltd; 2023 [cited 2025 Aug 29];10:790–800. [https://doi.org/10.1016/S2215-0366\(23\)00183-9](https://doi.org/10.1016/S2215-0366(23)00183-9)
35. Dwyer DB, Falkai P, Koutsouleris N. Machine Learning Approaches for Clinical Psychology and Psychiatry. *Annu Rev Clin Psychol* [Internet]. Annual Reviews Inc.; 2018 [cited 2025 Aug 29];14:91–118. <https://doi.org/10.1146/ANNUREV-CLINPSY-032816-045037>,
36. Chekroud AM, Zotti RJ, Shehzad Z, Gueorguieva R, Johnson MK, Trivedi MH, et al. Cross-trial prediction of treatment outcome in depression: A machine learning approach. *Lancet Psychiatry* [Internet]. Elsevier Ltd; 2016 [cited 2025 Aug 29];3:243–50. [https://doi.org/10.1016/S2215-0366\(15\)00471-X](https://doi.org/10.1016/S2215-0366(15)00471-X)
37. Lin E, Kuo PH, Liu YL, Yu YWY, Yang AC, Tsai SJ. A deep learning approach for predicting antidepressant response in major depression using clinical and genetic biomarkers.

- Front Psychiatry [Internet]. Frontiers Media S.A.; 2018 [cited 2025 Aug 29];9:367995. <https://doi.org/10.3389/FPSYT.2018.00290/BIBTEX>
38. Drysdale AT, Grosenick L, Downar J, Dunlop K, Mansouri F, Meng Y, et al. Resting-state connectivity biomarkers define neurophysiological subtypes of depression. *Nature Medicine* 2016 23:1 [Internet]. Nature Publishing Group; 2016 [cited 2025 Aug 29];23:28–38. <https://doi.org/10.1038/nm.4246>
39. Bao Z, Zhao X, Li J, Zhang G, Wu H, Ning Y, et al. Prediction of repeated-dose intravenous ketamine response in major depressive disorder using the GWAS-based machine learning approach. *J Psychiatr Res* [Internet]. Elsevier Ltd; 2021 [cited 2025 Aug 29];138:284–90. <https://doi.org/10.1016/j.jpsychires.2021.04.014>
40. Cao Z, Lin CT, Ding W, Chen MH, Li CT, Su TP. Identifying Ketamine Responses in Treatment-Resistant Depression Using a Wearable Forehead EEG. *IEEE Trans Biomed Eng. IEEE Computer Society*; 2019;66:1668–79. <https://doi.org/10.1109/TBME.2018.2877651>
41. Price RB, Spotts C, Panny B, Griffio A, Degutis M, Cruz N, et al. A Novel, Brief, Fully Automated Intervention to Extend the Antidepressant Effect of a Single Ketamine Infusion: A Randomized Clinical Trial. <https://doi.org/10.1176/appi.ajp.2022.20216> [Internet]. American Psychiatric Association Washington, DC; 2022 [cited 2025 Oct 13];179:959–68. <https://doi.org/10.1176/APPI.AJP.2022.20216>
42. Neuromorphometrics, Inc. | Building a Model of the Living Human Brain [Internet]. [cited 2025 Oct 14]. <https://www.neuromorphometrics.com/>. Accessed 14 Oct 2025
43. Hubert M, Rousseeuw PJ, Vanden Branden K. ROBPCA: A New Approach to Robust Principal Component Analysis. *Technometrics* [Internet]. Taylor & Francis; 2005 [cited 2025 Oct 14];47:64–79. <https://doi.org/10.1198/004017004000000563>
44. Golland P, Fischl B. Permutation Tests for Classification: Towards Statistical Significance in Image-Based Studies. *Lecture Notes in Computer Science (including subseries Lecture Notes in Artificial Intelligence and Lecture Notes in Bioinformatics)* [Internet]. Springer, Berlin, Heidelberg; 2003 [cited 2025 Oct 14];2732:330–41. [https://doi.org/10.1007/978-3-540-45087-0\\_28](https://doi.org/10.1007/978-3-540-45087-0_28)
45. Gómez-Verdejo V, Parrado-Hernández E, Tohka J. Sign-Consistency Based Variable Importance for Machine Learning in Brain Imaging. *Neuroinformatics* [Internet]. Humana Press Inc.; 2019 [cited 2025 Oct 14];17:593–609. <https://doi.org/10.1007/S12021-019-9415-3/FIGURES/5>
46. Markello RD, Arnatkevičiūtė A, Poline JB, Fulcher BD, Fornito A, Misic B. Standardizing workflows in imaging transcriptomics with the Abagen toolbox. *Elife* [Internet]. eLife Sciences Publications Ltd; 2021 [cited 2025 Aug 31];10. <https://doi.org/10.7554/ELIFE.72129>,
47. Hawrylycz M, Miller JA, Menon V, Feng D, Dolbeare T, Guillozet-Bongaarts AL, et al. Canonical genetic signatures of the adult human brain. *Nat Neurosci* [Internet]. Nature Publishing Group; 2015 [cited 2025 Aug 31];18:1832–44. <https://doi.org/10.1038/NN.4171>,
48. Arnatkevičiūtė A, Fulcher BD, Fornito A. A practical guide to linking brain-wide gene expression and neuroimaging data. *Neuroimage* [Internet]. Academic Press Inc.; 2019 [cited 2025 Aug 31];189:353–67. <https://doi.org/10.1016/j.neuroimage.2019.01.011>
49. Li N, Lee B, Liu RJ, Banasr M, Dwyer JM, Iwata M, et al. mTOR-dependent synapse formation underlies the rapid antidepressant effects of NMDA antagonists. *Science (1979)* [Internet]. Science; 2010 [cited 2025 Aug 31];329:959–64. <https://doi.org/10.1126/SCIENCE.1190287>,

50. Duman RS, Aghajanian GK. Synaptic dysfunction in depression: Potential therapeutic targets. *Science* (1979) [Internet]. American Association for the Advancement of Science; 2012 [cited 2025 Aug 31];338:68–72. <https://doi.org/10.1126/SCIENCE.1222939>,
51. Medeiros GC, Gould TD, Prueitt WL, Nanavati J, Grunebaum MF, Farber NB, et al. Blood-based biomarkers of antidepressant response to ketamine and esketamine: A systematic review and meta-analysis. *Mol Psychiatry* [Internet]. Springer Nature; 2022 [cited 2025 Aug 29];27:3658–69. <https://doi.org/10.1038/S41380-022-01652-1>,
52. Klok MPC, van Eijndhoven PhilipF, Argyelan M, Schene AH, Tendolkar I. Structural brain characteristics in treatment-resistant depression: review of magnetic resonance imaging studies. *BJPsych Open* [Internet]. Royal College of Psychiatrists; 2019 [cited 2025 Sep 5];5. <https://doi.org/10.1192/BJO.2019.58>,
53. Nikolin S, Rodgers A, Schwaab A, Bahji A, Zarate C, Vazquez G, et al. Ketamine for the treatment of major depression: a systematic review and meta-analysis. *EClinicalMedicine* [Internet]. Elsevier Ltd; 2023 [cited 2025 Oct 21];62. <https://doi.org/10.1016/j.eclinm.2023.102127>
54. Dunlop K, Hanlon CA, Downar J. Noninvasive brain stimulation treatments for addiction and major depression. *Ann N Y Acad Sci* [Internet]. Ann N Y Acad Sci; 2017 [cited 2025 Oct 21];1394:31–54. <https://doi.org/10.1111/NYAS.12985>
55. Disner SG, Beevers CG, Haigh EAP, Beck AT. Neural mechanisms of the cognitive model of depression. *Nat Rev Neurosci* [Internet]. Nature Publishing Group; 2011 [cited 2025 Oct 21];12:467–77. <https://doi.org/10.1038/NRN3027;SUBJMETA>
56. Mayberg HS. Limbic-cortical dysregulation: a proposed model of depression. *J Neuropsychiatry Clin Neurosci* [Internet]. J Neuropsychiatry Clin Neurosci; 1997 [cited 2025 Oct 21];9:471–81. <https://doi.org/10.1176/JNP.9.3.471>
57. Friedman NP, Robbins TW. The role of prefrontal cortex in cognitive control and executive function. *Neuropsychopharmacology* [Internet]. Springer Nature; 2022 [cited 2025 Oct 21];47:72–89. <https://doi.org/10.1038/S41386-021-01132-0;SUBJMETA>
58. Arnsten AFT, Joyce MKP, Roberts AC. The Aversive Lens: Stress effects on the prefrontal-cingulate cortical pathways that regulate emotion. *Neurosci Biobehav Rev* [Internet]. Elsevier Ltd; 2023 [cited 2025 Oct 21];145. <https://doi.org/10.1016/j.neubiorev.2022.105000>
59. Alt A, Witkin J, Bleakman D. AMPA Receptor Potentiators as Novel Antidepressants. *Curr Pharm Des* [Internet]. Bentham Science Publishers Ltd.; 2005 [cited 2025 Sep 8];11:1511–27. <https://doi.org/10.2174/1381612053764814>,
60. Vasavada MM, Leaver AM, Espinoza RT, Joshi SH, Njau SN, Woods RP, et al. Structural connectivity and response to ketamine therapy in major depression: A preliminary study. *J Affect Disord* [Internet]. Elsevier; 2016 [cited 2025 Sep 8];190:836–41. <https://doi.org/10.1016/J.JAD.2015.11.018>
61. Depping MS, Schmitgen MM, Kubera KM, Wolf RC. Cerebellar Contributions to Major Depression. *Front Psychiatry* [Internet]. Frontiers Media S.A.; 2018 [cited 2025 Sep 8];9:634. <https://doi.org/10.3389/FPSYT.2018.00634>
62. Depping MS, Schmitgen MM, Bach C, Listunova L, Kienzle J, Kubera KM, et al. Abnormal Cerebellar Volume in Patients with Remitted Major Depression with Persistent Cognitive Deficits. *Cerebellum* [Internet]. Springer; 2020 [cited 2025 Sep 8];19:762–70. <https://doi.org/10.1007/S12311-020-01157-Z>,
63. Abdallah CG, Sanacora G, Duman RS, Krystal JH. Ketamine and rapid-acting antidepressants: A window into a new neurobiology for mood disorder therapeutics. *Annu Rev*

Med [Internet]. Annual Reviews Inc.; 2015 [cited 2025 Sep 8];66:509–23.

<https://doi.org/10.1146/ANNUREV-MED-053013-062946>,

64. Cohen SE, Zantvoord JB, Wezenberg BN, Bockting CLH, van Wingen GA. Magnetic resonance imaging for individual prediction of treatment response in major depressive disorder: a systematic review and meta-analysis. *Transl Psychiatry* [Internet]. Springer Nature; 2021 [cited 2025 Sep 8];11. <https://doi.org/10.1038/S41398-021-01286-X>,

65. Jovicich J, Marizzoni M, Sala-Llonch R, Bosch B, Bartrés-Faz D, Arnold J, et al. Brain morphometry reproducibility in multi-center 3T MRI studies: A comparison of cross-sectional and longitudinal segmentations. *Neuroimage* [Internet]. Neuroimage; 2013 [cited 2025 Aug 29];83:472–84. <https://doi.org/10.1016/J.NEUROIMAGE.2013.05.007>,

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## Tables

Sample	Group	n	Female, n (%)	Male, n (%)	Age, mean (SD)	MADRS Pre, mean (SD)	MADRS Post, mean (SD)	MADRS Change, mean (SD)
Discovery	Responders	52	34 (65.4%)	18 (34.6%)	34.84 (11.72)	32.15 (5.56)	9.48 (5.54)	-22.67 (5.43)
	Non-responders	47	28 (59.6%)	19 (40.4%)	34.77 (10.29)	33.34 (5.13)	25.17 (6.72)	-8.00 (6.06)
Saline	Responders	12	7 (58.3%)	5 (41.7%)	29.37 (6.49)	33.92 (5.38)	12.42 (3.60)	-21.50 (6.30)
	Non-responders	37	24 (64.9%)	13 (35.1%)	35.44 (10.66)	31.97 (5.03)	27.24 (7.14)	-4.73 (5.06)
External validation – MD	Responders	5	2 (60.0%)	3 (40.0%)	39.60 (5.41)	31.40 (4.62)	7.20 (6.34)	-24.20 (7.16)
	Non-responders	17	11 (64.7%)	6 (35.3%)	34.59 (9.64)	32.76 (5.02)	27.47 (6.86)	-5.29 (7.67)
External validation – NY	Responders	16	7 (43.8%)	9 (56.3%)	46.34 (16.39)	30.56 (5.40)	8.50 (5.16)	-22.06 (5.81)
	Non-responders	13	5 (38.5%)	8 (61.5%)	41.44 (15.32)	31.46 (5.82)	25.46 (5.65)	-6.00 (4.16)

**Table 1. Demographic and Clinical Characteristics Across samples.** Each cohort is stratified by treatment response status (responders vs non-responders). Values are presented as mean (SD) for continuous variables and number (%) for categorical variables. MADRS = Montgomery–Åsberg Depression Rating Scale. Clinical response was defined as  $\geq 50\%$  reduction in MADRS scores 24 hours post-infusion. Change score calculated as post-ketamine – pre-ketamine. Discovery sample from NCT03237286; Saline sample from the same trial; External validation – MD from NCT00088699; External validation – NY from NCT00768430.

	TP, n	TN, n	FP, n	FN, n	CCR, %	BAC, %	PV, %	AUC		
					Non- responders	Responders	PPV	NPV		
Discovery model	34	38	14	13	72.3	73.1	72.7	70.8	74.5	0.73
Saline validation	15	5	7	22	40.5	41.7	41.1	68.2	18.5	0.45
External validation	16	14	7	14	53.3	66.7	60.0	69.6	50.0	0.65

**Table 2. Classification performance metrics across discovery, saline validation, and external validation cohorts.** Performance is reported using confusion matrix counts—true positives (TP), true negatives (TN), false positives (FP), and false negatives (FN)—alongside correct classification rate (CCR), balanced accuracy (BAC) positive predictive value (PPV), negative predictive value (NPV), and area under the receiver operating characteristic curve (AUC).

## Figure Legends

**Figure 1. Machine learning analysis design.** The discovery model was trained on data from a single-site ketamine trial (NCT03237286) using nested, repeated cross-validation. The outer loop (CV2: 3-fold, 5 repetitions) provided validation data for unbiased performance estimation, while the inner loop (CV1: 3-fold, 5 repetitions) was used for hyperparameter tuning and feature selection. The preprocessing pipeline included scaling, pruning, partial correlation to control for age, sex, site, and TIV, dimensionality reduction via robust PCA, and forward feature selection. Model generalizability was assessed through external validation on two independent single-dose ketamine cohorts (NCT00088699 and NCT00768430; total n=51), and mechanistic specificity was tested on a saline-treated cohort (n=49) from the same trial as the discovery sample.

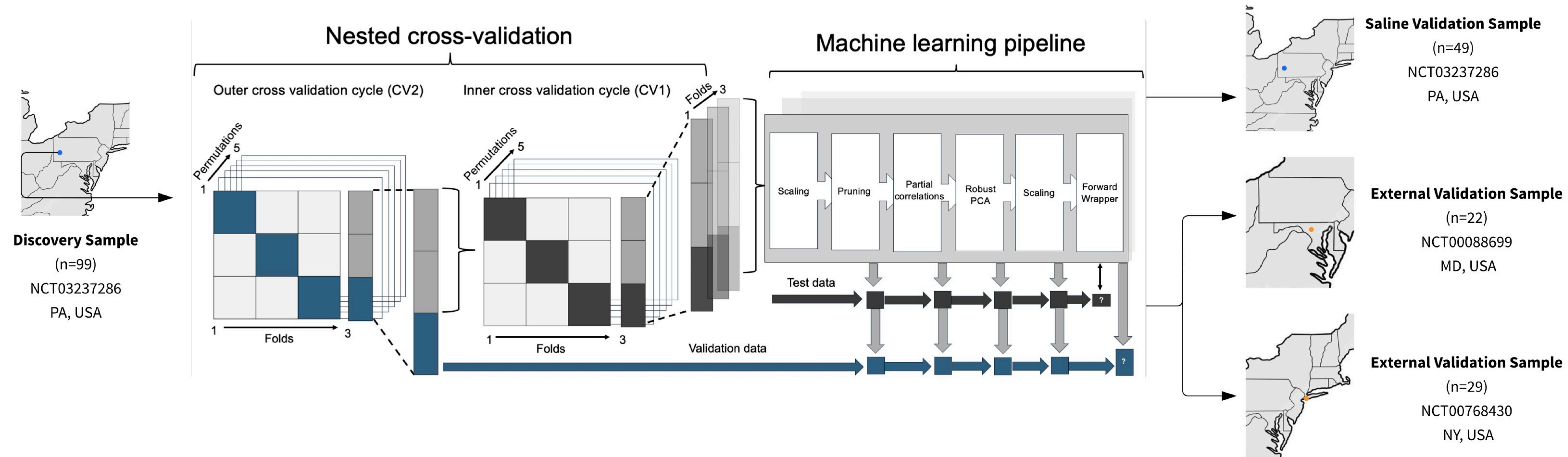
**Figure 2. Top 25 features contributing to classification of ketamine response.** Bars represent feature weights, with negative weights indicating features that contributed to classification as responders and positive weights indicating features that contributed to classification as non-responders, based on the model's class coding. Features are ordered by the absolute magnitude of their weight, with the most influential features shown at the bottom. Bar color reflects statistical significance of sign-based consistency across cross-validation folds: blue bars denote features with a consistent direction of contribution across folds at FDR-corrected  $p < 0.05$ , while grey bars did not meet this threshold. CVRs, annotated inside each bar (values rescaled  $\times 5$  for visualization only), quantify the robustness of each feature's contribution across the inner folds of the nested cross-validation framework. The CV ratio was calculated as the mean weight divided by the standard error of that weight across folds, serving as an interpretable measure of feature stability akin to a standardized effect size.

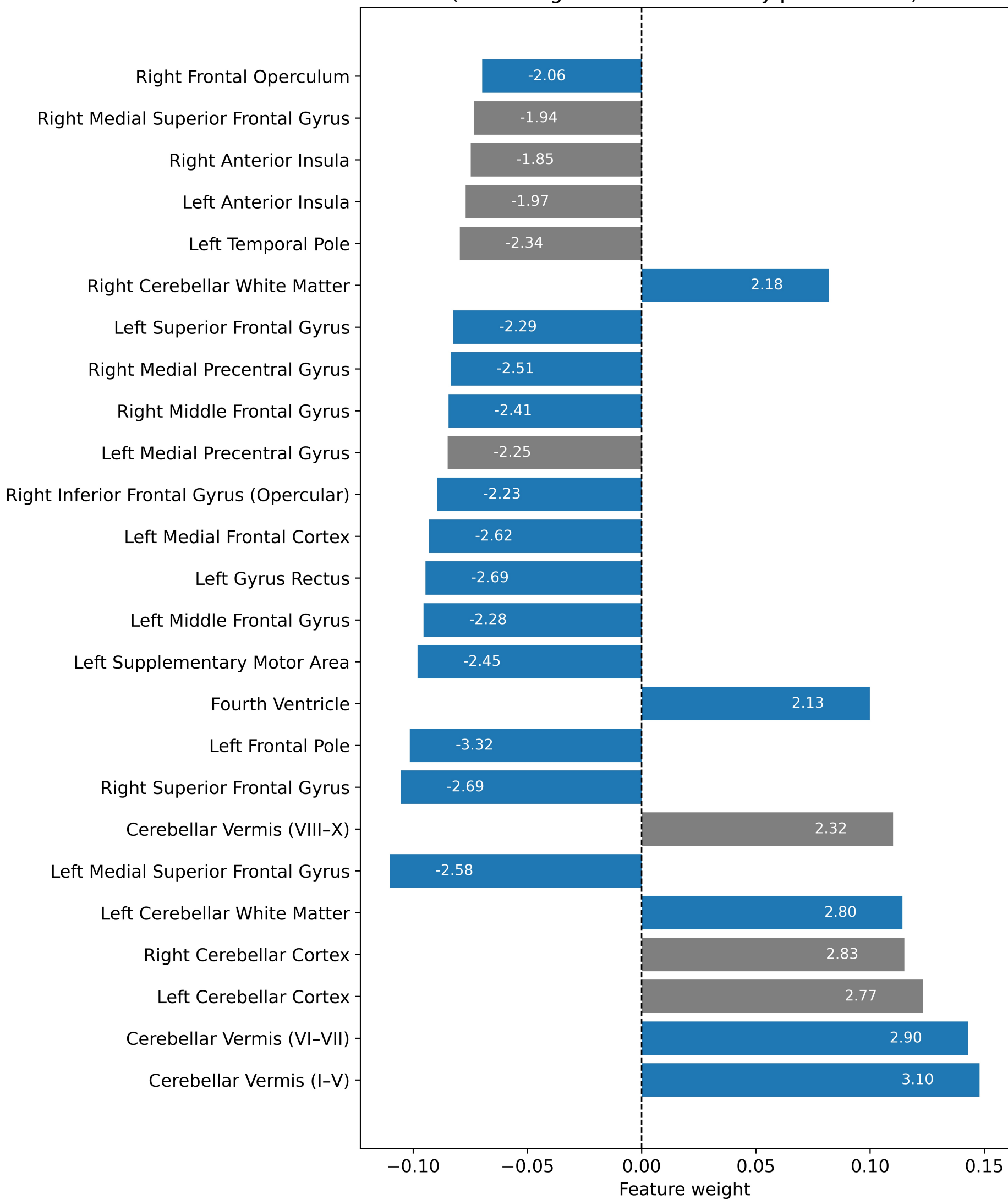
**Figure 3.** Top neuroanatomical predictors of ketamine treatment response (**Panel A**) and non-response (**Panel B**), derived from a linear SVC model trained on regional gray matter volumes using the Neuromorphometrics brain atlas. Images were generated using atlas-based overlays. In Panel A, greater volume in the highlighted regions was associated with increased likelihood of treatment response. In Panel B, greater volume was associated with increased likelihood of non-response.

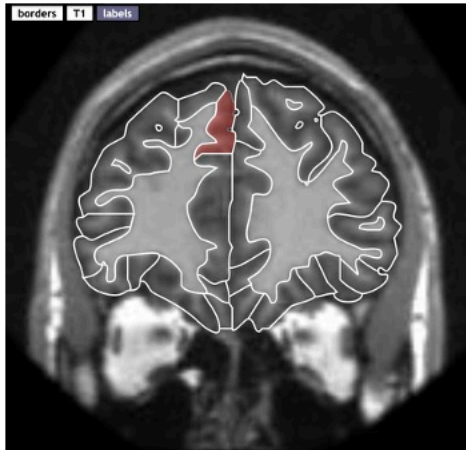
**Figure 4. Normalized gene expression of ketamine-relevant receptors across top predictive brain regions, mapped using the Allen Human Brain Atlas (AHBA).** This heatmap shows normalized gene expression values for seven receptor subunit proteins implicated in ketamine's mechanism of action (columns) across six brain regions (rows) identified as the top predictors of treatment response from the ML model. Gene expression values were extracted from the AHBA using the abagen toolbox, which aggregates postmortem microarray data from six adult donors and maps it to the MNI space using the Neuromorphometrics atlas. The brain regions are ordered by ML-derived response relevance: The bottom three rows correspond to frontal regions (Left FRP frontal pole, Left MSFG superior frontal gyrus medial segment, Right SFG superior frontal gyrus), where greater gray matter volume predicted treatment response. The top three rows correspond to cerebellar regions (Left Cerebellum Exterior, Vermal Lobules I–V and VI–VII), where greater volume predicted non-response. Frontal regions showed consistently higher expression of NMDA receptor subunits (GRIN1, GRIN2A, GRIN2B), AMPA receptors (GRIA1, GRIA2), and GABA receptor subunits (GABRA1, GABRB2), relative to cerebellar predictors. Color intensity represents normalized AHBA expression values per gene-region pair, with numeric annotations indicating exact expression levels. This analysis was intended to provide biological contextualization of predictive regions using genes implicated in ketamine

pharmacology, rather than to perform a genome-wide enrichment analysis of receptor gene expression.

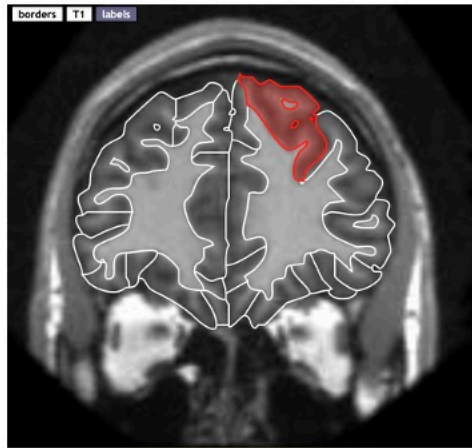
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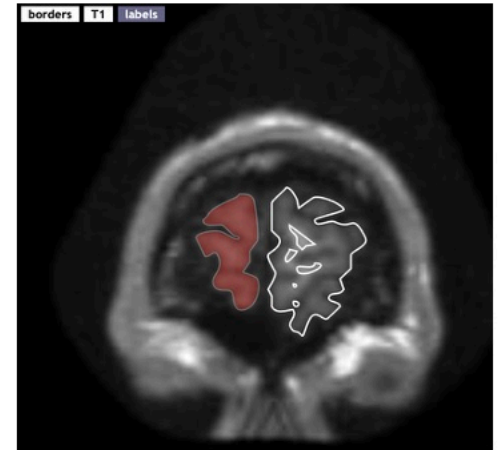
Top 25 Features: SVM Weight + CV Ratio  
(blue = sign-based consistency pFDR < 0.05)

**A**

1- Left medial superior frontal gyrus



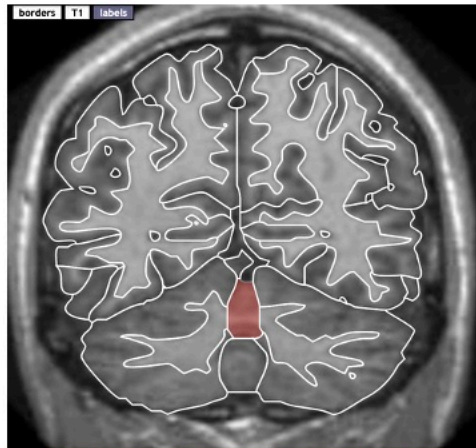
2- Right superior frontal gyrus



3- Left frontal pole

**B**

1- Cerebellar Vermal Lobules I-V



2- Cerebellar Vermal Lobules VI-VII



3- Cerebellum Exterior (L)

## Receptor Gene Expression in Predictive Brain Regions

