






Systematic Review

Surveillance of Healthcare-Associated Infections in the WHO African Region: Systematic Review of Literature from 2011 to 2024

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Abstract

Background: Evidence on HAIs in Africa is fairly common. **Objectives:** The main objective was to identify the surveillance tools used for healthcare-associated infections (HAIs) in countries in the WHO African Region. Secondary objectives focused on the organization of surveillance, the pathogens involved, and the frequency of multidrug-resistant species. **Inclusion and exclusion criteria:** Observational or interventional studies on healthcare-associated infections in humans, published between January 2011 and December 2024, in French or English, were included. However, the following publications were not included: animal studies, healthcare-associated infections not related to healthcare, literature reviews, studies outside the period or geographical area, and studies in languages other than French or English. **Sources of information and search date:** The databases consulted were PubMed, Web of Science, EMBASE, Cochrane, African Index Medicus, Google Scholar, and AJOL. The search was conducted between January and March 2025. **Risk of bias assessment:** The risk of bias was assessed using a specific grid (eleven criteria), scored from one (low) to three (high). The studies were classified into three levels of methodological quality. The results of the bias assessment showed that the publications were excellent (strong and moderate) with a cumulative rate of 99.9%. **Methods of synthesizing results:** Data were extracted using a standardized grid and synthesized narratively. No meta-analysis was performed. **Number of studies and characteristics:** 95 studies were included, mostly cross-sectional studies (82.1%), cohorts (10.4%), and a few case reports. Most were from West Africa (60.0%), particularly Nigeria (16.8%) and South Africa (14.7%). **Main results:** • Most common pathogens: *Staphylococcus aureus* (53.7%), *Escherichia coli* (43.2%), *Klebsiella pneumoniae* (32.6%). • Resistance profile: ESBL (27.4%), MRSA (21.1%), multidrug resistance (13.7%). • Sources of HAIs: mainly exogenous (83.2%). • Laboratory methods: phenotypic (70.5%), genotypic or genomic rare (3.1%). • Scope of studies: local (96.8%), national (3.2%). **Limitations of evidence:** Risk of bias due to underreporting of HAIs, methodological heterogeneity, predominance of cross-sectional studies, low use of molecular methods, lack of modeling, and uneven geographical coverage. **Overall interpretation and implications:** surveillance of HAIs in Africa remains fragmented and poorly



Academic Editor: Masafumi Seki

Received: 12 September 2025

Revised: 11 November 2025

Accepted: 14 November 2025

Published: 18 December 2025

Citation: Gahimbare, L.; Guessennd, N.K.; Muvunyi, C.M.; Fuller, W.; Coulibaly, S.O.; Cihambanya, L.; Kariyo, P.C.; Perovic, O.; Mwamelo, A.J.; Maxime, D.K.; et al. Surveillance of Healthcare-Associated Infections in the WHO African Region: Systematic Review of Literature from 2011 to 2024. *Antibiotics* **2025**, *14*, 1287. <https://doi.org/10.3390/antibiotics14121287>

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standardized. There is a need to strengthen national systems, integrate molecular methods, train professionals, and promote interventional research. The WHO GLASS program can serve as a framework for harmonizing surveillance.

Keywords: healthcare-associated infection; hospital-acquired infection; nosocomial infection; infection control surveillance

1. Introduction

Nosocomial infections (NI) or healthcare-associated infections have become a major global safety concern for both patients and healthcare professionals [1–5]. This term encompasses infections that occur during or after diagnostic, therapeutic, palliative, preventive, or educational care, provided they were neither present nor incubating at the beginning of the care [6].

Nosocomial infections (NI) are infections contracted in hospital. The term ‘healthcare-associated infections’ (HAIs) [7] is now preferred because it includes not only infections contracted in hospital, but also in other healthcare settings (nursing practices, convalescent homes) and even during home care. For ease of reading, the term HAI will be used throughout this article.

HAIs disproportionately affect low- and middle-income countries, where infrastructures are often underdeveloped [8]. Despite global efforts, such as the World Health Organization (WHO) guidelines for infection prevention in hospital practices, implementation remains uneven, particularly in resource-limited settings [8]. Although the public health problem of HAIs in developing countries has been known for decades, these HAIs affect at least 7% of patients in developed countries and 10% in developing countries [9]. Middle-income countries had a national HAIs surveillance system in 12.5% of cases. Most systems operated on a voluntary basis by health services. These systems monitored the incidence of HAIs using the definitions of the Center for Disease Control and Prevention [10]. This paradigm shift has been largely driven by evidence from seminal studies such as the Study on the Efficacy of Nosocomial Infection Control, or SENIC Project, which demonstrated the effectiveness of surveillance and control programs [11].

However, these infections are insufficiently researched, diagnosed, and reported in intertropical Africa [12]. They are a cause for concern due to their high morbidity and mortality rates, and especially due to the emergence of multi-resistant bacteria (MDR).

In intertropical Africa, the risk of HAIs may seem marginal compared to major public health problems such as malnutrition, childhood infections, malaria, AIDS, and violence-related diseases. HAIs are underreported.

The link between HAIs and multidrug-resistant bacteria is an integral part of the HAI problem that can no longer be ignored in Africa [13,14].

Surveillance systems exist in developed countries and provide regular reports on national trends in endemic HAIs [15–20]. African countries have established national HAIs surveillance systems, as highlighted in the WHO module on patient safety [21].

Assessing the significance of HAIs in sub-Saharan Africa requires first examining published data. Over two decades, only a few dozen publications have addressed HAIs in patients in intertropical Africa. Most of these are retrospective studies [4].

Previous literature reviews of HAIs in developing countries [22] covered the period between 1995 and 2008, while another, which focused on the World Health Organization (WHO) African Region, covered the period between 1995 and 2009 [23].

These reviews highlighted the need to strengthen frequent surveillance of HAIs. In addition, HAI databases from different healthcare facilities across Africa differ in their

methodological approach. Several literature reviews have provided updates on the occurrence of HAIs in Africa from 2010 to 2017, as well as on the contribution of emerging antimicrobial resistance to healthcare delivery in Africa [24].

From 2009 to 2018, a review addressed healthcare-associated bacterial infections in Africa [25]. A systematic review of recent articles published between 2013 and 2016 on antimicrobial resistance in Africa was conducted [26].

An assessment of HAI surveillance in certain African countries was conducted. In Ethiopia, a literature review was conducted by Shiferaw et al. on articles published between 2000 and 2019 on surgical site infections and associated factors [27].

In South Africa, a 10-year literature review was conducted from 2005 to 2014 on bloodstream infections with ESBL-producing and non-ESBL-producing *Escherichia coli* in children in a tertiary referral hospital [28] and then from 2009 to 2013 on exposure to infectious diseases and epidemics in a South African neonatal unit and examination of the epidemiology of neonatal epidemics in Africa [29].

These previous studies reported a wide range of surveillance methods for HAIs-related pathogens. Given the significant differences between the surveillance methods reported in the literature, there is an urgent need for a systematic analysis to synthesize existing knowledge and identify gaps in our understanding.

The main research question addressed is ‘What surveillance tools are used for HAIs in countries in the WHO African Region?’ The secondary questions are ‘What are the types and sources of HAIs?’; ‘How is HAI surveillance organized in the WHO African Region? Which pathogens contribute to these infections and how common are multidrug-resistant (MDR) species in HAIs?’

In this context, the present study aims to describe the organizational methods of surveillance, the pathogens involved, and the frequency of multidrug-resistant HAI species in the WHO African region.

2. Materials and Methods

A systematic literature review assessed infection surveillance tools associated with healthcare in WHO African countries. The literature search for this systematic review was conducted in accordance with the PRISMA (Preferred Reporting Items for Systematic Reviews and Meta-Analyses) [30]. All steps of the systematic review were carried out using the free web tool RAYYAN.

2.1. PICO Elements (Population (Or Patient/Problem), Intervention, Comparison, and Outcome (Result)) [31,32]

- Population (P): the target population consists of patients receiving care in any type of health facility (university hospitals, district health centers, clinics, etc.) located in member countries of the WHO African Region.
- Intervention (I): involved the implementation of an active and continuous surveillance program for HAIs or hospital-acquired infections using epidemiological methods.
- Comparison (C): the local or national scope of the HAI surveillance methods implemented and the existence of a structured or unstructured surveillance and prevention program.
- Outcome (O): Reduction in healthcare-associated infections, prevention of antimicrobial resistance, improvement of patient safety.

PICO question: What are the HAI surveillance methods and their scope in WHO African countries compared to limited or no implementation of surveillance to reduce healthcare-associated infections, prevent antimicrobial resistance, and improve safety outcomes?

2.2. Search Strategy

For this systematic review, we undertook a literature search and review process according to a protocol designed prior to data collection. The search was conducted between January and March 2025.

To assess methods of HAI surveillance in WHO Afro countries, we extracted studies from *PubMed*, *Web of Science*, *Science Daily*, the Cochrane Database of Systematic Reviews, *African Journals Online Library*, and *free-text web* searches using *Google Scholar*. Reviewers, KKF, DKM, and GKN independently searched these databases to identify articles published from January 2011 to December 2024. Our search terms included a combination of keywords and MeSH terms (*Medical Subject Headings*) such as “nosocomial infection”, “hospital-acquired infection”, “healthcare-associated infection”, “surveillance”, “Africa”, “individual country name”. Boolean operators (AND/OR) were used to refine and combine terms.

Additionally, we reviewed the references of all included studies to identify further relevant research, ensuring a comprehensive synthesis of available evidence on HAI surveillance methods in WHO Afro countries.

A separate search was conducted in the WHO regional medical database for Africa, African Index Medicus, using a shorter list of essential keywords and time restrictions.

2.3. Selection Process

The selection process consisted of a two-stage selection process to compile a list of documents relevant to our research question. Reviewers from the team conducted independent blind checks to ensure objectivity.

First, inclusion and exclusion criteria were defined for the review of titles and abstracts of the identified documents (results of the search in bibliographic databases), which allowed an initial filtering of the literature. In accordance with the PICO principles, the inclusion criteria were population, humans, exposure, healthcare-associated or hospital-acquired infections and the outcome, the HAI surveillance method.

Exclusion criteria were applied articles concerning literature reviews on HAI; articles outside the period and area of study; articles published in languages other than French or English.

If the necessary information in a file was not clearly indicated in the title or abstract, it was selected for the next selection stage.

In the second stage, a full-text selection was carried out for reports (full-text documents) that passed the first filter using additional criteria, full-text accessibility with English or French as the language and reporting an HAIs, accompanied by a description of the methodology used to obtain the data.

In the event of discrepancies between the results obtained by the two reviewers during either screening phase, discussions took place to reach a consensus on the inclusion or exclusion of problematic documents. Following these checks and harmonization, a final list of documents to be extracted was drawn up. The reference lists of the included documents was also reviewed to identify and add relevant records that may have been missed during the initial search. If multiple studies were identified in a single document, they were treated individually during the data extraction phase.

2.4. Data Extraction

A data extraction framework was established to facilitate a systematic and efficient analysis of the studies by extracting relevant information. This framework was applied to the selected studies. The categories and labels, designed to be as comprehensive as possible, are presented in Table 1.

Table 1. Data extraction categories, labels, and information to be collected.

Category	Label	Information to Be Collected
General information	Item ID	Awarded by team
	Authors	Authors' names
	Year of publication	Year of publication of the study
	Title	Article Title
	DOI/PMID/Link	Study ID
Study characteristics	Type of study	Case–control, Descriptive cohort, Before–after study; Case report; Case series Cross–sectional study
	Objectives of the study	Describe the objective of the study
	WHO Region	Region of the world defined by the WHO
	Country	Countries where the study was conducted
	Study period	Duration of the study
Population details	Population or subject studied	General population, children, elderly, etc.
	Sources of infection	Acquired in hospital
	Population information	Gender, age, health status, inpatient/outpatient service, etc.
Exhibition details	Mode of transmission	Individual cases of illness, epidemics, etc.
	HAIs source	Patients, staff; equipment
	Type of samples	Pus swab, urine, blood, catheters, sputum, nasopharyngeal aspirate, expectorations, and hospital consumables, environmental swab
	Number of samples	Number
Laboratory methods	Pathogens identified	Genera and species
	Type of pathogens	Bacteria, fungi, viruses
	Laboratory identification method	Genomic, genotypic, unspecified, phenotypic
	Antibiogram	Antibiotics tested
	AST Guidelines	CLSI; EUCAST–CASFM
	Resistance profile	ESBL, CRAB; CRE; MRSA
Main findings and results	Main results	Key findings of the article
	Main conclusions	Conclusions drawn by the authors
	Additional notes	Any other relevant information

2.5. Assessment of the Quality or Risk of Bias of Articles

We developed a tool with a series of criteria (Table 2) to identify potential sources of bias, which allowed us to assess the rigor of each study. This tool is an adaptation of the Joanna Briggs Institute (JBI) Critical Appraisal Checklist [30]. The criteria, presented in Table 2, allowed the reviewers to rate the included articles on a scale from low (1 point) to moderate (2 points) or high (3 points). Higher scores indicate that the study had minimal bias, used a sound methodology, and carefully considered potential limitations.

Table 2. Criteria used to assess the risk of bias in the selected articles.

Criteria	Source of Bias	Applied Scale
Study using data from one's own work or another source	Data source	1—External; 2—Mixed; 3—Internal
Data used clearly indicated (with source if necessary)	Data availability	1—No; 2—Unclear; 3—Yes
Estimate or measure the size of the exposed population	Size of exposed population	1—Unclear; 2—Estimated; 3—Measured
Estimation or measurement of the size of the sick population	Size of the sick population	1—Unclear; 2—Estimated; 3—Measured
Characteristics of individuals clearly presented	Population health status	1—Not described; 2—Described
Assessment of population size with critical evaluation by the author	Population size	1—No; 2—Yes
Assessment of the author's intervention method	Type of study	1—Not described; 2—Described
Appreciate the different sources of HAIs	HAIs Sources	1—Not described; 2—Described
Clearly identified HAIs source	HAIs Sources	1—No; 2—Unclear; 3—Yes
Critical evaluation by the authors of the study results and comparison with other studies	Discussion of results	1—No critical evaluation by the authors; 2—Critical evaluation by the authors
Critical assessment by the authors of factors that may influence their results	Discussion of factors influencing the results	1—No critical evaluation by the authors; 2—Critical evaluation by the authors

Our tool to assess the overall risk of bias in the included studies considered scales, checklists, and individual components. This tool had scales in which various quality components are rated and combined to give a summary score [32].

The methodological quality assessment scores allowed the articles to be categorized into 3 groups (Table 3).

Table 3. Different score groups of methodological quality assessment.

Band	Methodological Quality	Score (%)
I	Forte	(28–33) 75–100
II	Moderate	(17–27) 50–75
III	Weak	(<16) <50

This tool assesses the methodological rigor and reliability of studies in several areas, including the clarity of inclusion criteria, the appropriateness of the study design, the validity of the outcome measure, and the adequacy of the data analysis. Two reviewers independently conducted the quality assessment, and any disagreements were resolved after consultation with a third reviewer.

To assess the different sources of HAIs, a list of descriptions was selected and detailed in Table 4.

Table 4. Description of HAIs sources.

Source Category	Origin of the Microorganism	Specific Sources	Mode of Transmission Predominant
1. Endogenous Source (Self-contamination)	The patient himself	Patient flora (digestive, cutaneous, respiratory) which enters through an artificial or natural entry point.	Migration/Direct inoculation (during an invasive or surgical procedure): the patient's germs are introduced into a sterile site (e.g., catheter, surgical site).
2. Exogenous Source (Cross-Contamination)	External to the patient		
	Humans: Other Patients	Infected patients or asymptomatic carriers (especially MDR).	Direct or indirect contact (through the hands of staff or shared equipment).
	Humans: Healthcare Staff	Asymptomatic carrier staff members. Lack of hygiene (hands, clothing).	Handling (hands not disinfected between patients) or airborne (in case of unmasked respiratory infection).
	Environmental/Material: Equipment and Devices	Invasive devices (catheters, probes, ventilators, etc.). Improperly sterilized surgical instruments. Shared surfaces (bed, table, handles) and objects.	Direct inoculation (via the device) or indirect contact (via surfaces/materials).
	Environmental/Material: Inanimate Environment	Water (taps, showers, air conditioning systems). Air (works, ventilation). Food (very rare).	Airborne (inhalation) or waterborne (ingestion, nebulization).

The validated protocol was submitted to the Prospero platform and was registered under number CRD420251032268.

Statistical analysis was performed with Microsoft Excel for all frequency description calculations and graphs. The map was created using QGIS Desktop 3.30.1.

3. Results

3.1. Document Research, Selection Process, and Characteristics of the Selected Studies

3.1.1. Document Research and Selection Process

The database search yielded 10,362 unique articles (Figure 1). After screening titles and abstracts, 95 articles met the inclusion criteria and were therefore eligible for full-text review. Five [5] additional articles were identified through reference review.

Each included study underwent a risk of bias assessment, which identified potential bias that could affect the reliability of the results. No study had a risk of bias sufficiently high to warrant exclusion from this analysis.

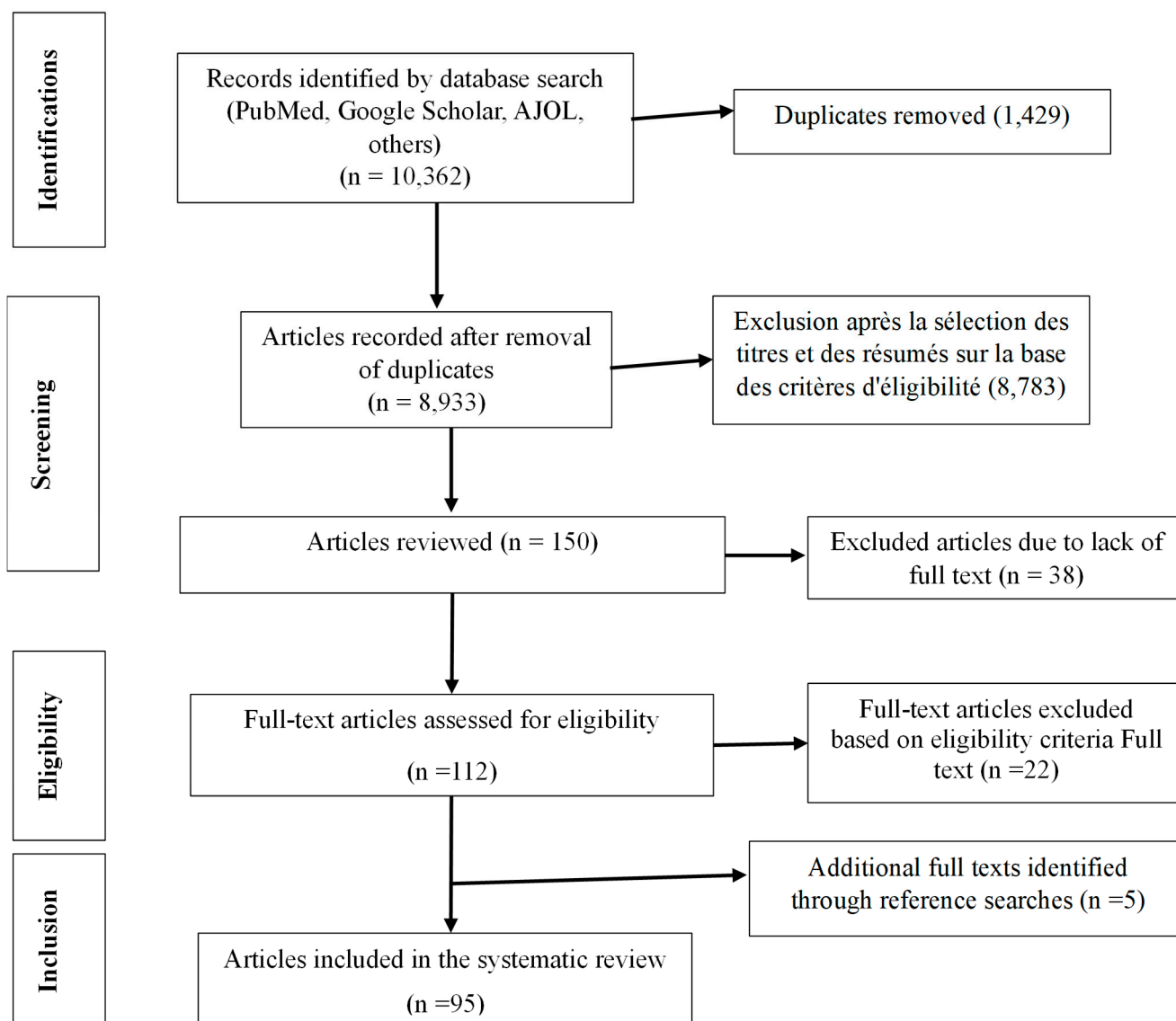


Figure 1. Summary of the selection of studies included in the review according to PRISMA guidelines.

3.1.2. Characteristics of the Selected Studies

In Table 5, this review included 95 observational studies, including 78 cross-sectional studies (82.1%), 10 cohort studies (10.4%), and a single quasi-experimental interventional study of the before-after design. The 78 included detailed descriptions of multiple cases of healthcare ward patients following exposure to a single source of hazard, including contamination by bacterial, viral, or myco-parasitic pathogens. The three case reports and case series documented sporadic incidents where individuals with unique characteristics became ill after being admitted to a healthcare ward. The quasi-experimental study was an evaluation of the effectiveness of a nosocomial infection control program in a Senegalese neonatal unit to reduce nosocomial bacteremia and bacterial resistance conducted by C. Landre-Peigne in 2011 [32].

Table 5. Typology of selected studies.

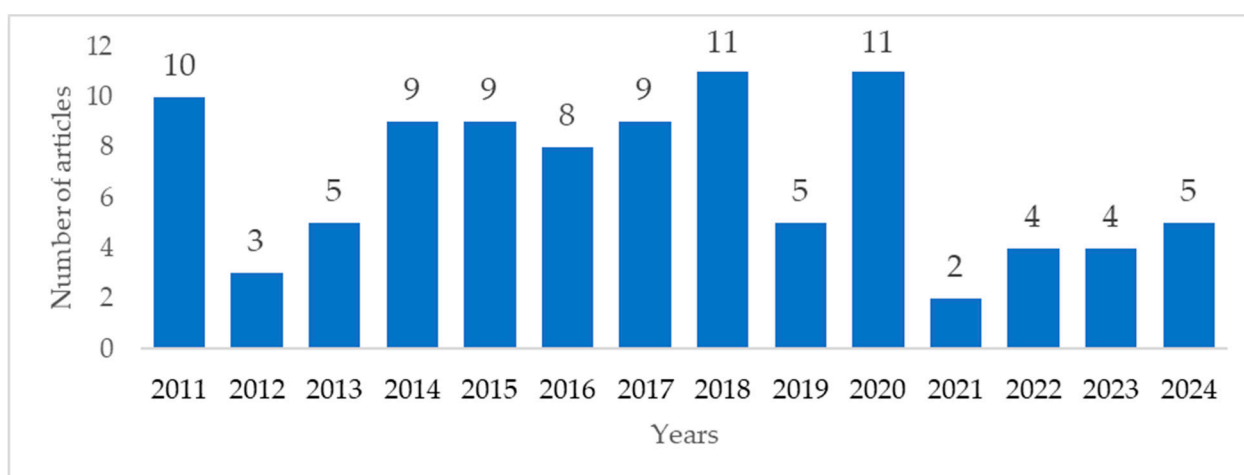
Type of Study	Bibliographic References	n (%)
Case-control	[33,34]	2 (2.1)
Cohort	[35,36]	10 (10.4)
Before-and-after study	[37]	1 (1.1)
Case report	[38–40]	3 (3.2)
Case series	[41]	1 (1.1)
Transversal	[42–119]	78 (82.1)
Total		95

The use of cross-sectional studies to monitor HAIs in our countries began a long time ago and remains a predominant method, often based on data from documented epidemics of different pathogens. The oldest evaluative interventional study of the “before and after” and “here and elsewhere” type was conducted in 2011 and has not been conducted since 2011 or in any other country, while infection prevention and control programs are underway in all African countries. Case reports and series have been used over time to determine HAIs in real-life conditions.

Among the publications studied, five were produced during an epidemic caused by bacteria [35,77,120,121] and one study was due to a virus [66]. These epidemics were described in two cross-sectional studies, two cohort studies, and one case-control study [34]. The origin of the epidemics was discussed and linked mainly to hygiene, either through interpatient transmission (overcrowding [120,122]) or inappropriate practices by staff. These epidemics mainly involved highly resistant bacteria such as NDM-1-producing bacteria [34] and ESBL [120]. All of these epidemics were brought under control. However, there were no mathematical modeling studies on outbreaks of diseases related to healthcare-associated infections, combined with mathematical tools to compensate for the lack of field data [35,66,77,120,121].

Figure 2 shows the temporal distribution of the different types of studies, from 2011 to 2024. The evolution of the number of publications related to HAIs was sawtooth. The lowest number of articles was in 2021 (2 articles, 2.1%) and the highest number of publications was in 2018 and 2020 (11 articles, 11.6%).

Table 6 presents the number of studies identified on HAIs and the spread of antimicrobial resistance in a nosocomial environment according to the year of publication.

**Figure 2.** Number of publications on HAIs over time (2011–2024).

The geographical distribution of data sources (Table 6) was another relevant parameter, as the distribution of HAI prevalence is not uniform across Africa. Thus, the findings of these studies are not always applicable to HAIs present in other regions or countries. In addition, HAI control methods vary across regions, influencing both the hazard and the level of risk. The West Africa, Southern Africa, and East Africa regions accounted for 57 (60.0%), 16 (16.8%), and 9 (9.5%) of the studies, respectively.

Table 6. Temporal and Afro-regional distribution by study types of HAIs.

	Case-Control	Descriptive Cohort	Before-and-After Study	Cross-Sectional Study	Case Report	Case Series	Total n (%)
Years							
2011–2015	2	2	1	29	2	0	36 (37.9)
2016–2020	0	3	0	39	1	1	44 (46.3)
2021–2024	0	5	0	10	0	0	15 (15.8)
African regions							
West Africa	0	7	1	45	3	1	57 (60.0)
Central Africa	0	1	0	7	0	0	8 (8.4)
East Africa	0	0	0	9	0	0	9 (9.5)
South Africa	2	2	0	12	0	0	16 (16.8)
North Africa	0	0	0	5	0	0	5 (5.3)
Total	2	10	1	78	3	1	95

The characteristics of the selected studies are summarized in Table 6. This highlights the frequencies of case reports, case series, before-and-after studies, and cross-sectional studies, as well as the regions of origin of these studies, over the following three publication periods: 2011–2015; 2016–2020; 2021–2024.

In all countries, three recorded the highest number of publications on healthcare-associated infections (HAIs). The first country was Nigeria with 16 articles, followed by South Africa (14 publications), and Ghana with 11 publications. Other countries published very few articles over the ten years covered by this study. However, even the leading countries have an average of only 1 to 1.8 publications per year (Figure 3).

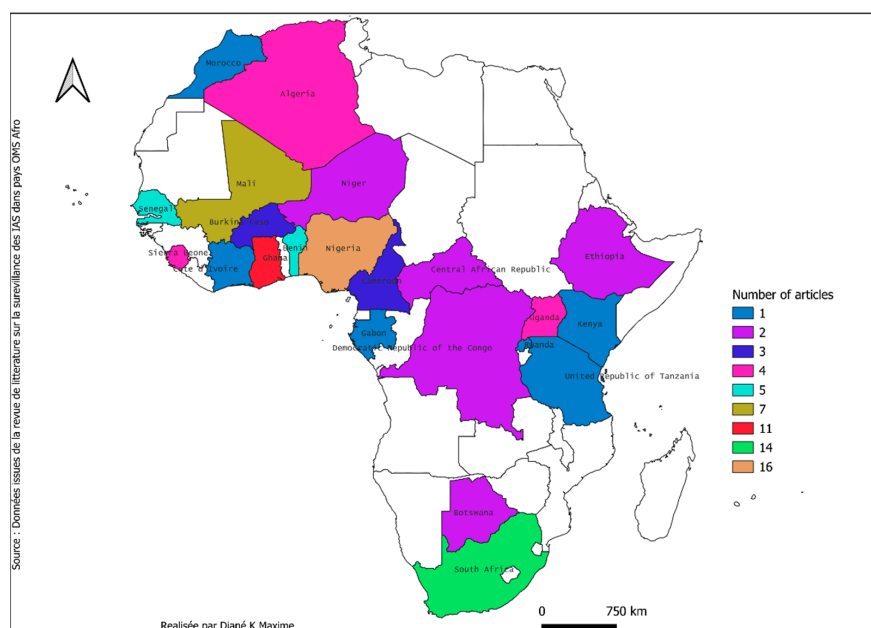


Figure 3. Number of publications on HAIs per countries over time (2011–2024).

When analyzing the sources of HAIs, Table 7 shows that the main sources of contamination were exogenous. The microorganism comes from a source external to the patient in 79 articles (83.2%). Some infections are both endogenous (the index patient was infected) and exogenous (transmission to other patients and caregivers) in 8.4% of cases.

Table 7. Distribution of types of HAIs according to source.

Type of HAIs Sources	Bibliographic References	n (%)
Endogenous	[44,47,50,57,71,79,90,113]	8 (8.4)
Exogenous	[33–43,45,46,48,49,52,53,55,56,58–65,67–70,72–78,80,83,84,86–89,91–98,100–112,116–118,120–127]	79 (83.2)
Exogenous and Endogenous	[54,66,81,82,85,99,114,115]	8 (8.4)
Total		95

Figure 4 presents a summary analysis of the origins of HAIs in our literature review. With nearly half of the references (43.2%), HAIs remain the central concern in the literature, reflecting their high incidence and the complexity of their prevention in the intraoperative setting. The burden of invasive infections represented by bacteremia/bloodstream infections (35.8%) and respiratory infections (27.4%) is significant, often associated with morbidity.

Table 8 summarizes the results of the risk of bias assessment of the selected articles. The articles had excellent methodological rigor (85.2%) with an overall low risk of bias.

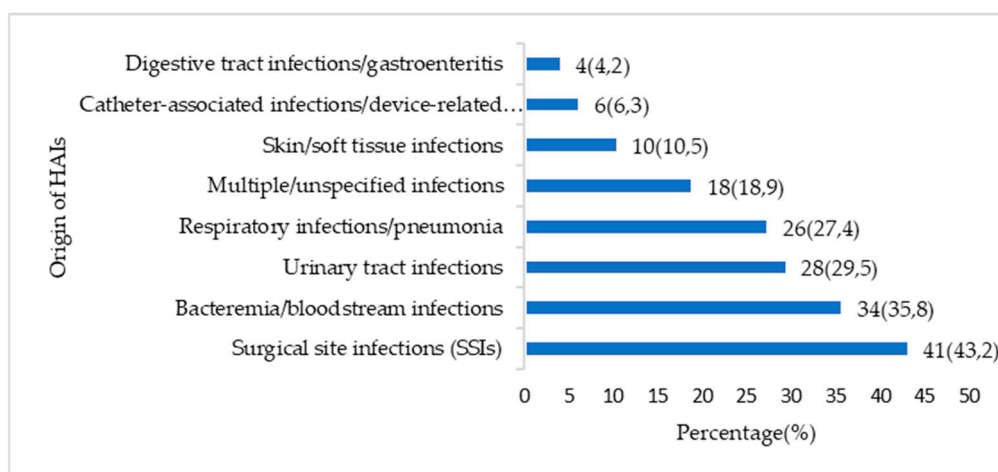


Figure 4. Distribution of healthcare-associated infections by origin.

Table 8. Risk of bias assessments for each included study.

Methodological Quality	Bibliographic References	n (%)
High (>75%)	[33–37,40–43,45–55,57–61,63–67,69,71,75–82,86,88–104,106–109,111–118,120–127]	81 (85.2)
Moderate (50–75%)	[38,39,44,56,62,68,70,73,74,83,87,105,110]	13 (13.7)
Acceptable (<50%)	[72]	1 (1.1)

3.2. Organization of HAIs Surveillance

The analysis of the scope of the studies shows that 96.8% of surveillance interventions have a local scope (92 studies), while 3 studies have a national scope (3.2%).

Only three countries (Ghana, South Africa, and Benin) reported studies based on national data. These studies used cross-sectional methods [80,92,101]. The vast majority

of studies were local in scope, employing various methods. Among these methods, cross-sectional studies were widely reported in 78 articles, representing 82.1% (Table 9).

Table 9. Typical distribution and scope of HAIS studies by country.

Type of Study	Scope	Country	Bibliographic Reference	n (%)
Case-control n = 2 (2.1)	Local n = 2 (2.1)	South Africa	[34,35]	2 (2.1)
		South Africa	[120,122]	2 (2.1)
Descriptive cohort n = 10 (10.5)	Local n = 10 (10.5)	Cameroon	[126]	1 (1.1)
		Ivory Coast	[125]	1 (1.1)
		Ghana	[123]	1 (1.1)
		Mali	[121]	1 (1.1)
		Niger	[124]	1 (1.1)
		Sierra Leone	[36,37,127]	3 (3.2)
		Before-after study n = 1 (1.1)	Local n = 1 (1.1)	Senegal
Cross-sectional study n = 78 (82.1)	Local n = 75 (78.9)	South Africa	[26,51,58,64,94,97, 99,100,102]	9 (9.5)
		Algeria	[48,60,72,113]	4 (4.2)
		Benin	[45,75,82,90]	4 (4.2)
		Botswana	[117]	1 (1.1)
		Botswana and South Africa	[96]	1 (1.1)
		Burkina Faso	[52,65,76]	3 (3.2)
		Cameroon	[54,61]	2 (2.1)
		Ethiopia	[62,117]	2 (2.1)
		Gabon	[67]	1 (1.1)
		Gambia	[74,77]	2 (2.1)
		Ghana	[47,59,70,78,83– 85,87,104]	9 (9.5)
		Kenya	[57]	1 (1.1)
		Mali	[86,89,110,112,115]	5 (5.3)
		Morocco	[115]	1 (1.1)
		Niger	[71]	1 (1.1)
		Nigeria	[42,44,46,50,53,55, 68,69,73,79,81,87,95, 98,106,109]	16 (16.8)
		Uganda	[63,105,107,111]	4 (4.2)
Central African Republic	[66,109]	2 (2.1)		

Table 9. Cont.

Type of Study	Scope	Country	Bibliographic Reference	n (%)
Cross-sectional study n = 78 (82.1)	Local n = 75 (78.9)	Democratic Republic of Congo (DRC)	[93,103]	2 (2.1)
		Rwanda	[49]	1 (1.1)
		Senegal	[88,91]	2 (2.1)
		Sierra Leone	[43]	1 (1.1)
	Tanzania	[56]	1 (1.1)	
	National n = 3 (3.2)	South Africa	[101]	1 (1.1)
		Benin	[92]	1 (1.1)
Ghana		[80]	1 (1.1)	
Case report n = 3 (3.2)	Local n = 3 (3.2)	Mali	[38]	1 (1.1)
		Senegal	[39,40]	2 (2.1)
Case series n = 1 (1.1)	Local n = 1 (1.1)	Gambia	[41]	1 (1.1)
Total				95 (100.0)

A certain number of studies (94.7%, 90 studies) specified the laboratory investigation technique for HAIs samples. Among the publications that did so, only two studies (2.1%) explored HAIs samples using a genotypic technique and two studies with genomics (2.1%). Regarding the methods, 67 studies used purely phenotypic methods (70.5%) compared to 16 studies that used both phenotypic and genotypic methods (16.8%). National studies conducted in three African countries used phenotyping as the method for investigating HAIs samples in the laboratory (Table 10).

Table 10. Distribution of study scopes of HAIs investigation methods in the HAI laboratory.

Scope n (%)	Investigation Methods n (%)	Country	Bibliographic Reference	n (%)
Local n = 92 (96.8)	Clinical n = 5 (5.3)	South Africa	[58]	1 (1.1)
		Ghana	[78]	1 (1.1)
		Uganda	[63,105]	2 (2.1)
		Sierra Leone	[37]	1 (1.1)
		Gambia	[74]	1 (1.1)
	Genomics n = 2	Ghana	[87]	1 (1.1)
		Nigeria	[98]	1 (1.1)
	Genotypic n = 2	Central African Republic	[66]	1 (1.1)
		Phenotypic n = 67 (70.5)	South Africa	[26,35,51,64,94,97,99,102,122]
	Algeria		[48,72,113]	3 (3.2)

Table 10. Cont.

Scope n (%)	Investigation Methods n (%)	Country	Bibliographic Reference	n (%)	
Local n = 92 (96.8)	Phenotypic n = 67 (70.5)	Benin	[45,90]	2 (2.1)	
		Botswana	[117]	1 (1.1)	
		Botswana and South Africa	[96]	1 (1.1)	
		Burkina Faso	[52,65,76]	3 (3.2)	
		Cameroon	[54,61,126]	3 (3.2)	
		Ivory Coast	[125]	1 (1.1)	
		Ethiopia	[62,117]	2 (2.1)	
		Gabon	[67]	1 (1.1)	
		Gambia	[41,77]	2 (2.1)	
		Ghana	[84,104]	2 (2.1)	
		Kenya	[57]	1 (1.1)	
		Mali	[38,86,89,110,112,116, 121]	7 (7.4)	
		Morocco	[115]	1 (1.1)	
		Niger	[71,124]	2 (2.1)	
		Nigeria	[42,44,50,53,55,68,69,79, 81,86,95,106,109]	13 (13.7)	
		Uganda	[107,112]	2 (2.1)	
		Central African Republic	[108]	1 (1.1)	
		Democratic Republic of Congo (DRC)	[93,103]	2(2.1)	
		Rwanda	[49]	1 (1.1)	
		Senegal	[33,88,91]	3 (3.2)	
		Sierra Leone	[36,43,127]	3 (3.2)	
		Tanzania	[56]	1 (1.1)	
		South Africa	[34,100,120]	3 (3.2)	
		Phenotypic + genotypic n = 16 (16.8)	Algeria	[60]	1 (1.1)
			Benin	[75,82]	2 (2.1)
			Ghana	[47,59,70,83,85,123]	6 (6.3)
Nigeria	[46,73]		2 (2.1)		
Senegal	[39,40]		2 (2.1)		
South Africa	[101]		1 (1.1)		
National n = 3 (3.2)	Phenotypic n = 3 (3.2)	Benin	[92]	1 (1.1)	
		Ghana	[80]	1 (1.1)	
		Total		95	

3.3. Reported Pathogens and Resistance Profile

3.3.1. Reported Pathogens

Table 11 shows that in the 95 publications the vast majority of reported HAIs concerned the 48 bacterial species (50.5%). Only 4 species of *Candida fungi* (4.2%) were reported, and 4 species of viruses (4.2%) were reported.

The top five most monitored bacteria in the articles were *Staphylococcus aureus* in 51 articles (53.7%), *Escherichia coli* in 41 articles (43.2%), *Pseudomonas aeruginosa* in 35 publications (36.8%), *Klebsiella pneumoniae* was concerned by 31 articles (32.6%), and finally *Acinetobacter baumannii* cited in 21 articles (22.1%).

Table 11. Distribution of pathogens involved in HAIs studies.

Name of the Pathogen	Bibliographic References	n (%)
Bacteria n = 48		(50.5)
<i>Acinetobacter baumannii</i>	[26,33,35,43,45,48,51,56,83,86,90–93,96,100,106,114,116,123,124]	21 (22.1)
<i>Acinetobacter</i> spp.	[67,70,79,85,90,91,112,116,126]	9 (9.5)
<i>Alcaligenes</i>	[69]	1 (1.1)
<i>Bacillus</i> spp.	[104]	1 (1.1)
Gram–negative bacteria	[53,88,110,114,121,127]	6 (6.3)
Gram–positive bacteria	[53,88,127]	3 (3.2)
<i>Bacteroides</i> spp.	[69,79]	2 (2.1)
<i>Bacteroides fragile</i>	[42]	1 (1.1)
<i>Citrobacter</i> spp.	[62,85,92,106,126]	5 (5.3)
<i>Citrobacter friends</i>	[93]	1 (1.1)
<i>Clostridium</i> spp.	[69]	1 (1.1)
Coagulase–negative <i>Staphylococcus</i>	[91,92,117]	3 (3.2)
<i>Corynebacterium aurimucosum</i>	[39]	1 (1.1)
<i>Enterobacter</i> spp.	[61,62,67,85,106,116,126]	7 (7.4)
<i>Enterobacter cloacae</i>	[33,34,45,51,54,61,86,116,125]	9 (9.5)
<i>Enterobacter faecalis</i>	[86]	1 (1.1)
<i>Enterococcus faecalis</i>	[43,56,93,94,124]	4 (4.2)
<i>Enterococcus faecium van A</i>	[60]	1 (1.1)
<i>Enterococcus</i> spp.	[36,67,68,92] [36,42,44–49,52,54–	4 (4.2)
<i>Escherichia coli</i>	57,59,61,65,67,68,71,76,80,82,84–86,89,90,92, 93,95,101,103,104,106,111,112,116,122– 124,126]	41 (43.2)
<i>Hafnia alvei</i>	[46,106]	2 (2.1)
<i>Klebsiella oxytoca</i>	[36,46]	2 (2.1)
<i>Klebsiella pneumoniae</i>	[33,34,36,42,43,45,48,49,51,54,56,57,61,65,67, 68,71,72,84,86,89,90,95–97,99,106,112,116,123– 125]	31 (32.6)
<i>Klebsiella</i> spp.	[53,58,62,63,81,85,86,88,91,97,108,111,113,114, 127]	15 (15.8)

Table 11. Cont.

Name of the Pathogen	Bibliographic References	n (%)
<i>Listeria monocytogenes</i>	[96]	1 (1.1)
<i>Morganella</i>	[84]	1 (1.1)
<i>Peptococcus</i> spp.	[42]	1 (1.1)
<i>Proteus mirabilis</i>	[36,42,46,54,56,61,86,106,113]	9 (9.5)
<i>Proteus</i> spp.	[55,62,68,79,87,95,106]	7 (7.4)
<i>Proteus vulgaris</i>	[126]	1 (1.1)
<i>Providencia</i> spp.	[62,69]	2 (2.1)
<i>Providencia stuartii</i>	[56]	1 (1.1)
<i>Pseudomonas aeruginosa</i>	[36,42–46,48,51,54,56,57,61,62,68,71,76,79,80,84–86,89–93,95,102–104,106,112,113,115,116,123]	35 (36.8)
<i>Pseudomonas</i> spp.	[102,111,126]	3 (3.2)
<i>Salmonella arizonae</i>	[126]	1 (1.1)
<i>Salmonella enterica</i>	[40,120]	2 (2.1)
Non-typhoidal <i>Salmonella</i>	[124]	1 (1.1)
<i>Salmonella</i> spp.	[92,103]	2 (2.1)
<i>Salmonella typhi</i>	[103]	1 (1.1)
<i>Serratia liquefaciens</i>	[77]	1 (1.1)
<i>Serratia marcescens</i>	[34,61]	2 (2.1)
<i>Shigella</i> spp.	[103]	1 (1.1)
<i>Staphylococcus aureus</i>	[33,36,42–45,48,49,51,52,54–57,59,61,62,64,67–69,71–76,80,86,87,89–93,95–97,99,101–104,106,111,112,115,117,123]	51 (53.7)
<i>Staphylococcus haemolyticus</i>	[59]	1 (1.1)
<i>Staphylococcus</i> spp.	[52,56]	2 (2.1)
Group B <i>Streptococcus</i>	[96,101]	2 (2.1)
<i>Streptococcus</i> spp.	[69,91]	2 (2.1)
<i>Streptococcus viridans</i>	[69]	1 (1.1)
Fungi/mycoses n = 4		(4.2)
<i>Candida albicans</i>	[26,51,56,86,92,101,113,115,122]	9 (9.5)
<i>Candida krusei</i>	[122]	1 (1.1)
<i>Candida parapsilosis</i>	[122]	1 (1.1)
<i>Candida</i> spp.	[42,44,52,53,81,92,97,99,113,116]	11 (11.6)
Virus n = 4	[4]	4 (4.2)
Virus (HVA, HVB, HVC)	[72]	1 (1.1)
Virus (RSV, Adenovirus)	[97,99]	2 (2.1)
<i>Lassa fever virus</i>	[98]	1 (1.1)
<i>Monkeypox virus</i> (Mpox)	[66]	1 (1.1)
UNSPECIFIED n = 9	[9]	(9.5)
Not specified	[37,41,50,58,63,78,94,105,107]	9 (9.5)

NB: Individual studies have reported several agents.

Although HAIs are often associated with antibiotic-resistant bacteria, eight articles (8.4%) did not specify a particular pathogen.

3.3.2. Resistance Profile

Table 12 shows that the ESBL bacterial phenotype was the most frequently reported profile (26 references, 27.4%) and the MRSA profile came second (20 references, 21.1%). Multidrug resistance (MDR) was also well documented with thirteen references, 13.7%. Resistance to carbapenems and methicillin is well represented. Many specific molecular profiles were identified but with fewer references. An important category concerns unspecified or unstudied resistance (twenty-five references).

Table 12. Characteristics of phenotypic resistance profiles of bacteria in the articles studied.

Phenotypic Profile	Bibliographic References	Number n = 95 (%)
ESBL	[26,43,45,48,56,62,65,67,76,80,85,86,89–91,96,98,100,102,107,120,121,123–125,127]	26 (27.4)
MRSA/Methicillin-resistant	[33,48,53,56,62,64,67,73–75,88,90,92,95,97,99,101,111,113,123]	20 (21.1)
MDR/Multi-resistance	[53,54,56,60–62,70,84,100,111,117,120,126]	13 (13.7)
CRE/Carbapenemases	[35,36,45,80,85,96,100,123,124,127]	10 (10.5)
C3GR	[33,38,80,85,95,123]	6 (6.3)
Tetracycline R	[44,59,68,84,86,87]	6 (6.3)
Beta-lactamase	[44,68,89,125]	4 (4.2)
Ceftazidime R	[88,92,115]	3 (3.2)
Fluconazole R	[97,122]	2 (2.1)
Cephalosporinase	[110]	1 (1.1)
Amphotericin B (sensitive)	[122]	1 (1.1)
CRAB	[26,48,100]	3 (3.2)
VRE	[92,113]	2 (2.1)
Partial resistance/not specified	[34,35,39,41,49,50,55,58,61,63,69,72,77–79,87,94,98,102–105,107,109]	25 (26.3)

ESBL: Extended-spectrum beta-lactamase-producing enterobacteria; MRSA: Methicillin-Resistant *Staphylococcus aureus*; MDR: Multidrug-Resistant Bacteria; CRE: Carbapenem-Resistant Enterobacteriaceae; C3GR: Resistance to third-generation cephalosporins; CRAB: Carbapenem-Resistant *Acinetobacter baumannii*; R: Resistance; VRE: vancomycin-resistant enterococcus.

Table 13 shows very high rates of resistance of the usual isolates in the studies. *Escherichia coli* showed a rate of 97.6% against ampicillin. For *Acinetobacter baumannii*, the rate of MDR varied from 62.1 to 90%, and for carbapenems, it was from 47.6 to 75.3%. All the listed germs are MDR.

Table 13. Resistance rates and phenotypic profiles of bacteria to antibiotics in articles.

Name of the Pathogen	Resistance Rate (%)	Common Resistance Phenotypes	Bibliographic References
<i>Escherichia coli</i>	ESBL (56–79.2) Ampicillin resistance (97.6) Amoxicillin resistance (95.2)	ESBL, resistance to 3rd generation cephalosporins, multi-resistance	[36,65,82,84,111]
<i>Klebsiella pneumoniae</i>	ESBL (59.2–80.3) Carbapenem resistance (1.3–5.24)	ESBL, CRE, resistance to beta-lactams, cephalosporins	[26,38,72,85,96,124]
<i>Staphylococcus aureus</i>	MRSA (15–100) Penicillin resistance (88.6–100)	MRSA, oxacillin resistance, multi-drug resistance to beta-lactams	[26,48,62,64,75,87]
<i>Pseudomonas aeruginosa</i>	Ciprofloxacin resistance (50–68.2) Resistance to meropenem (31)	Resistance to fluoroquinolones, cephalosporins, carbapenems	[35,48,76,91,123]
<i>Acinetobacter baumannii</i>	Multidrug resistance (MDR) (62.1–90) Carbapenem resistance (47.6–75.3)	CRAB, resistance to cephalosporins, carbapenems, colistin (rare)	[26,35,48,70,83,100]
<i>Enterobacter</i> spp.	ESBL (58.3) Resistance to C3G frequent Vancomycin resistance (67.5)	ESBL, cephalosporin resistance, variable sensitivity to carbapenems VRE, resistance to glycopeptides, aminopenicillins	[34,72,85,125]
<i>Enterococcus</i> spp.	Ampicillin resistance frequent Fluconazole resistance: variable		[60,67,92]
<i>Candida</i> spp.	<i>C. krusei</i> : intrinsic resistance	Resistance to azoles, sensitivity to amphotericin B	[81,97,122]
<i>Salmonella</i> spp.	ESBL: detected Multi-resistance ≥ 6 antibiotics	Resistance to beta-lactams, aminoglycosides, cotrimoxazole	[40,120]
<i>Proteus</i> spp.	Ampicillin resistance (100) Multi-resistance frequent	Resistance to beta-lactams, aminopenicillins, quinolones	

ESBL: Extended-spectrum beta-lactamase-producing enterobacteria; MRSA: Methicillin-Resistant *Staphylococcus aureus*; MDR: Multidrug-Resistant Bacteria; CRE: Carbapenem-Resistant Enterobacteriaceae; C3GR: Resistance to third-generation cephalosporins; CRAB: Carbapenem-Resistant *Acinetobacter baumannii*; VRE: vancomycin-resistant *enterococcus*.

Apart from resistance phenotypes, only 11 articles out of 95 reported cases of resistance genotypes. The resistance genotypes most reported in this study concerned beta-lactams, fluoroquinolones, and aminoglycosides, respectively. These genotypes were represented by the CTX-M-15 gene in 2 articles [47,108], qnr (*qnrB*, *qnrS*) and AAC(6′)-Ib-cr in 1 article [108], rendering several families of antibiotics ineffective. Other resistance genes were also reported, including the *mecA* gene (MRSA, methicillin resistance) in one article [74] and VanA in 1 article [60].

4. Discussion

4.1. Summary of the Main Results

This systematic review aimed to identify healthcare-associated infection (HAI) surveillance tools in the WHO African Region. A total of 95 studies were included, predominantly cross-sectional (82.1%) and cohort (10.5%) studies. Most studies were from West Africa (60%), with Nigeria, South Africa, and Ghana as the main contributors. The most frequently reported pathogens were *Staphylococcus aureus* (53.7%), *Escherichia coli* (43.2%), and *Pseudomonas aeruginosa* (36.8%). The main resistance patterns were ESBL (27.4%) and MRSA (21.1%). The majority of studies (96.8%) were local in scope and used phenotypic methods (70.5%), with only a few studies incorporating genotypic or genomic approaches. This significant variability in the types of studies and methodologies employed included differences in the populations exposed, the pathogens incriminated, and the laboratory investigation techniques.

Our results are broadly consistent with those of other recent systematic reviews of HAIs in Africa, but some methodological and contextual differences are worth highlighting.

In a review Irek et al. also identified a predominance of cross-sectional studies and underreporting of HAIs in sub-Saharan Africa. However, she highlighted a lack of data on fungal and viral infections, which our study confirms (only 4.2% for each category) [22].

In their study, Talaat et al. highlighted the emergence of multidrug-resistant bacteria (MDR) in HAIs in Africa, with an increasing prevalence of ESBL and carbapenem-resistant Enterobacteriaceae. Our results are consistent, with 27.4% of studies reporting ESBL and 10.5% CRE [119].

The Global Report on Infection Prevention and Control 2024 [122] assessed HAI surveillance systems in referral hospitals in Africa and found that less than 30% of countries had national surveillance systems. Our findings go even further: only three studies (3.2%) were national in scope, highlighting the critical lack of harmonized data at the national level.

During our literature search, we identified several studies reporting HAIs related to case series and surgical site infections. These studies provide valuable data on contamination levels in different matrices [38,41].

The most frequently studied pathogens represented in the literature were bacteria. The most isolated bacterial species were *Escherichia coli*, *Staphylococcus aureus*, and *Klebsiella pneumoniae*.

HAI surveillance in the WHO African Region represents a major public health challenge, as demonstrated by this systematic review covering the period from 2011 to 2024. The results highlight the methodological, epidemiological, and structural challenges faced by African countries in HAI surveillance, as well as the progress made and persistent gaps, as well as opportunities to improve HAI surveillance and prevention in the African region.

4.2. Current Status of HAI Surveillance in Africa

The results of this review showed marked heterogeneity in HAI surveillance systems across the African region. Although a few countries, such as Nigeria, South Africa, and Ghana, have published a relatively high number of HAI studies, the majority of African countries remain underrepresented in the scientific literature. This disparity likely reflects differences in the infrastructural capacity, financial resources, and technical expertise available to conduct robust HAIs studies.

Most of the included studies (82.1%) were cross-sectional in nature, suggesting a predominance of one-off surveys rather than continuous surveillance systems. Only a few longitudinal or cohort studies were identified, limiting the ability to assess temporal trends in HAIs. Furthermore, only one interventional “before–after” study was identified,

highlighting the lack of impact evaluations of HAI prevention programs in the region, where they exist.

The scope of the studies was predominantly local (96.8%), with very little national or regional data. This focus on specific contexts, while valuable for understanding local dynamics, makes it difficult to generalize the results and implement coordinated strategies on a larger scale.

4.3. Heterogeneity of Monitoring Methods

This review identified methodological diversity in HAI studies, with a predominance of cross-sectional studies (82.1%), followed by cohort studies (10.5%), and case reports (3.2%). This heterogeneity reflects the logistical and financial constraints of African health systems, where one-off cross-sectional studies are often preferred due to their feasibility and low cost. However, these studies offer a limited view over time and do not allow monitoring the evolution of HAIs or assessing the real impact of interventions in the long term. Only one quasi-experimental “before-after” study was identified, conducted in Senegal in 2011, highlighting the glaring lack of intervention evaluations in the region [32].

Geographic disparities are also notable, with studies concentrated in West Africa (60.0%) and Southern Africa (16.8%), while Central and Northern Africa are underrepresented. Nigeria, South Africa, and Ghana dominate publications, perhaps reflecting more developed research capacities in these countries. These disparities highlight the need to strengthen surveillance capacities in less-covered regions to obtain a more balanced view of the HAI situation at the continental level.

4.4. Pathogens Involved in HAIs and Predominance of Multidrug-Resistant Pathogens

The most frequently reported pathogens in HAIs include *Staphylococcus aureus* (51 studies), *Escherichia coli* (41 studies), and *Klebsiella pneumoniae* (31 studies). These bacteria, often associated with antibiotic resistance profiles (ESBL, MRSA, CRE), pose a major challenge for the management of infections in hospital settings. The presence of *Acinetobacter baumannii* and *Pseudomonas aeruginosa*, also documented, worsens this picture due to their resistance to last-resort antibiotics [23].

Monkeypox virus (one study) and Lassa fever virus in the context of HAIs are a reminder that threats are not limited to bacteria. The COVID-19 pandemic has highlighted the risks of HAIs transmission of viruses, particularly in healthcare settings with limited resources [20,124]. These findings highlight the importance of integrated surveillance, covering bacteria, viruses, and other pathogens.

The most frequently cited sources of HAIs were invasive medical devices and the hospital environment. These findings highlight the need to strengthen hygiene practices, equipment sterilization, and staff training to reduce the risk of HAIs.

4.5. Laboratory Methods and Technical Limitations

Of the studies analyzed, 93.4% specified the laboratory methods used to identify pathogens. However, the majority (78 studies) used phenotypic techniques, while only 3 studies used genotypic methods, and 2 used genomic approaches. This predominance of phenotypic methods, although more accessible and less expensive, limits the ability to accurately characterize resistance mechanisms and detect outbreaks linked to specific species.

Genotypic and genomic methods, although rarer, offer significant advantages for HAI surveillance, including better resolution for species typing and detection of resistance genes. Their wider adoption in Africa could improve the quality of surveillance data and facilitate international comparisons.

Limitations of the evidence presented in the analysis: The identification of few eligible studies or studies involving small numbers of participants, leading to imprecise estimates; the risk of bias in studies or missing results; or identifying studies that only partially or indirectly answer the review question, raising doubts about their relevance and applicability to particular patients, settings, or other target audiences.

Limitations of the review processes used: The decision to restrict inclusion to studies in English and French only, to search only a limited number of databases, to entrust file review or data collection to a single reviewer, or not to contact study authors to clarify ambiguous information. We were probably unable to access all potentially eligible study reports or conduct some of the planned analyses due to insufficient data.

Despite the efforts described in this review, several limitations persist in the current monitoring system:

Limited scope of studies: The majority of studies (96.2%) are local in scope, focusing on individual hospitals or specific regions. Only three studies are national in scope, and only one is regional. This fragmentation limits the generalizability of results and the implementation of coordinated policies at the continental level.

Laboratory methods: Although 93.4% of studies specify investigation techniques, only 3% use genotypic or genomic methods, which are essential for understanding epidemic dynamics and antibiotic resistance. The majority of studies rely on phenotypic methods, which are less precise for identifying resistance mechanisms [21].

Underreporting and HAIs: As several authors point out, HAIs are often underreported in sub-Saharan Africa due to the lack of standardized surveillance systems, low awareness among health professionals, and diagnostic constraints [7]. This underreporting distorts the true estimate of the HAI burden and hinders prevention efforts.

Lack of modeling: No mathematical modeling studies were identified, although these tools could compensate for the lack of field data and anticipate epidemiological trends [22].

4.6. Recommendation to Public Health Policies and Professionals

The results of this review highlight the urgency of strengthening HAI surveillance systems in Africa. The following recommendations could guide policymakers and health professionals.

Develop national surveillance systems: African countries should invest in continuous and standardized surveillance systems, aligned with WHO guidelines, to generate comparable and actionable data.

Promote the use of molecular methods: The integration of genotypic and genomic techniques in reference laboratories would enable more accurate monitoring of pathogens and AMR.

Strengthening local capacity: Targeted training for health personnel and laboratory technicians is needed to improve data quality and the implementation of prevention protocols.

Improve hygiene and infection control practices: Interventions such as hand hygiene promotion, equipment sterilization, and medical waste management could significantly reduce the incidence of HAIs.

Encourage intervention research: More “before-and-after” or randomized studies are needed to evaluate the effectiveness of HAI prevention programs.

4.7. Recommendations for GLASS

Following this literature review on HAIs in the WHO Afro region, it is recommended that a laboratory surveillance program for AMR-related HAIs be strengthened in countries of the WHO Afro region. For a national laboratory surveillance program, government com-

mitment and support are non-negotiable. GLASS should also encourage the establishment of supranational coordination centers that will coordinate national reference centers, which in turn will systematically collect, analyze, and share AMR-related HAI data nationally and internationally. These regional reference laboratories will provide technical support, including training, capacity building, and policy advice.

5. Conclusions

This systematic review provides an overview of the challenges and opportunities related to HAI surveillance in Africa. Although progress has been made in publishing HAI surveillance data, particularly in leading countries such as Nigeria and South Africa, significant gaps persist in terms of geographical coverage, surveillance methods, and access to advanced technologies. A coordinated approach, involving governments, health institutions, and international partners, is essential to improve HAI surveillance and prevention in the region. By addressing these gaps, Africa can better protect the health of patients and healthcare workers, while contributing to the global fight against antimicrobial resistance.

The capacity of countries in the WHO Afro region varies greatly in the area of combating AMR and consequently AMR related to HAIs. However, this capacity has been significantly hampered by the availability of capacity building and, more recently, by the emergence of the COVID-19 pandemic, which has also been added to the long list of healthcare-associated infections.

Thus, the true burden of HAIs in this systematic review is underreported and is perhaps greater in countries with weaker health infrastructure. However, this trend is evolving with the growing commitment in Africa to address the global threat of AMR. National technical capacities with the support of partners are now evolving not only to develop national AMR action plans but also to institute national AMR surveillance. The WHO Global Antimicrobial Surveillance System (GLASS) provides a tool to standardize data collection, sharing, and analysis across participating institutions and countries globally to monitor trends and implement controls.

Data on the prevalence of antimicrobial resistance and HAIs in developing countries in African WHO Afro are scarce and unsystematic; thus, authors suggest that intensive survey and surveillance are warranted.

Author Contributions: Study conception and design: L.G., N.K.G., C.M.M. and D.K.M. Data acquisition: D.K.M. and K.K.F. Analysis and interpretation of data: L.G. and N.K.G. Writing the manuscript: D.K.M., K.K.F., N.K.G. and L.G. Revising the manuscript critically for important intellectual content: O.P., Y.A.A., V.G., S.O.C., W.F., L.C., P.C.K., B.N. and A.J.M. Approval of the version of the manuscript to be published: L.G. and N.K.G. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflicts of interest.

Registration: The protocol was registered in PROSPERO under number CRD420251032268.

Abbreviations

AST	Antimicrobial Susceptibility Test
C3GR	Resistance to third-generation cephalosporins
CASFM	Antibiogram Committee of the French Society for Microbiology
CLSI	Clinical and Laboratory Standards Institute
CRAB	Carbapenem-Resistant <i>Acinetobacter baumannii</i>
CRE	Carbapenem-Resistant Enterobacteriaceae
DRC	Democratic Republic of Congo
ESBL	Extended-spectrum beta-lactamase-producing enterobacteria
EUCAST	Extended Spectrum Beta-lactamases
GLASS	Global Surveillance System for this Resistance
HAI	Healthcare-Associated Infections
HINARI	Health InterNetwork Access to Research Initiative
MDR	Multidrug-Resistant Bacteria
Mpox	<i>Monkeypox</i>
MR	Multidrug-Resistant
MRSA	Methicillin-Resistant <i>Staphylococcus aureus</i>
NDM	New Delhi metallo-beta-lactamase
NI	Nosocomial Infections
PICO	Population (or Patient/Problem), Intervention, Comparison, and Outcome (Result)
PRISMA	Preferred Reporting Items for Systematic Reviews and Meta-Analyses
R	Resistance
SARS-CoV-2	Severe Acute Respiratory Syndrome CoronaVirus 2 (COVID-19)
VRE	vancomycin-resistant <i>enterococcus</i>
WHO	World Health Organization

References

- World Health Organization. *Prevention of Hospital-Acquired Infections: A Practical Guide*; World Health Organization: Geneva, Switzerland, 2002.
- Burke, J.P. Infection control—A problem for patient safety. *N. Engl. J. Med.* **2003**, *348*, 651–656. [CrossRef]
- Bates, D.W.; Larizgoitia, I.; Prasopa-Plaizier, N.; Jha, A.K. Global priorities for patient safety research. *BMJ* **2009**, *338*, b1775. [CrossRef] [PubMed]
- Simon, F.; Kraemer, P.; De Pina, J.J.; Demortiere, E.; Rapp, C. Le risque nosocomial en Afrique intertropicale—Partie 2: Les infections des patients. *Med. Trop.* **2007**, *67*, 197–203.
- Rebaudet, S.; Kraemer, P.; Savini, H.; De Pina, J.J.; Rapp, C.; Demortiere, E. Le risque nosocomial en Afrique intertropicale Partie 3: Les infections des soignants. *Méd. Trop.* **2007**, *67*, 291.
- HAS. Évaluation de la Prévention des Infections Associées Aux Soins Selon le Référentiel de Certification. 2022. Available online: https://www.has-sante.fr/upload/docs/application/pdf/2020--11/fiche_pedagogique_prevention_infection_soins_certification.pdf (accessed on 26 October 2025).
- Horan, T.C.; Andrus, M.; Dudeck, M.A. CDC/NHSN surveillance definition of health care-associated infection and criteria for specific types of infections in the acute care setting. *Am. J. Infect. Control* **2008**, *36*, 309–332. [CrossRef]
- Hinson, C.; Kilpatrick, C.; Rasa, K.; Ren, J.; Nthumba, P.; Sawyer, R.; Ameh, E. Global surgery is stronger when infection prevention and control is incorporated: A commentary and review of the surgical infection landscape. *BMC Surg.* **2024**, *24*, 397. [CrossRef]
- Storr, J.; Twyman, A.; Zingg, W.; Damani, N.; Kilpatrick, C.; Jacqui, R.; Lesley, P.; Matthias, E.; Lindsay, G.; Kelley, E.; et al. Core components for effective infection prevention and control programmes: New WHO evidence-based recommendations. *Antimicrob. Resist. Infect. Control* **2017**, *6*, 6. [CrossRef]
- Takaya, S.; Hayakawa, K.; Matsunaga, N.; Moriyama, Y.; Katanami, Y.; Tajima, T.; Tanaka, C.; Kimura, Y.; Saito, S.; Kusama, Y.; et al. Surveillance systems for healthcare-associated infection in high and upper-middle income countries: A scoping review. *J. Infect. Chemother.* **2020**, *26*, 429–437. [CrossRef]
- Haley, R.W.; Quade, D.; Freeman, H.E.; Bennett, J.V.; The Cdc Senic Planning Committee. Study on the efficacy of nosocomial infection control (SENIC Project): Summary of study design. *Am. J. Epidemiol.* **1980**, *111*, 472–485. [CrossRef]

12. Simon, F.; Demortiere, E.; Chadli, M.; Kraemer, P.; De Pina, J.J. Le risque nosocomial en Afrique intertropicale. Partie 1: Le contexte. *Méd. Trop.* **2006**, *66*, 91–96.
13. Okeke, I.N.; Laxminarayan, R.; Bhutta, Z.A.; Duse, A.G.; Jenkins, P.; O'Brien, T.F.; Pablos-Mendez, A.; Klugman, K.P. Antimicrobial resistance in developing countries. Part I: Recent trends and current status. *Lancet Infect. Dis.* **2005**, *5*, 481–493. [[CrossRef](#)]
14. Dellamonica, P. Antibiorésistance et maladies transmissibles [zone Afrique]. *Méd. Trop.* **1998**, *58*, 73–77.
15. Wen, R.; Li, X.; Liu, T.; Lin, G. Effect of a real-time automatic nosocomial infection surveillance system on hospital-acquired infection prevention and control. *BMC Infect. Dis.* **2022**, *22*, 857. [[CrossRef](#)] [[PubMed](#)]
16. Haley, R.W.; Culver, D.H.; White, J.W.; Morgan, W.M.; Emori, T.G.; van Munn, P.; Hooton, T.M. The efficacy of infection surveillance and control programs in preventing nosocomial infections in US hospitals. *Am. J. Epidemiol.* **1985**, *121*, 182–205. [[CrossRef](#)]
17. Pittet, D.; Allegranzi, B.; Sax, H.; Bertinato, L.; Concia, E.; Cookson, B.; Fabry, J.; Richet, H.; Philip, P.; Spencer, R.C. Considerations for a WHO European strategy on health-care-associated infection, surveillance, and control. *Lancet Infect. Dis.* **2005**, *5*, 242–250. [[CrossRef](#)] [[PubMed](#)]
18. Jafari, Y.; Yin, M.; Lim, C.; Pople, D.; Evans, S.; Stimson, J.; Pham, T.M.; Read, J.M.; Robotham, J.V.; Cooper, B.S.; et al. Effectiveness of infection prevention and control interventions, excluding personal protective equipment, to prevent nosocomial transmission of SARS-CoV-2: A systematic review and call for action. *Infect. Prev. Pr.* **2022**, *4*, 100192. [[CrossRef](#)]
19. Kamath, K.; Kumar, H.S.; Shabaraya, A.R. A comprehensive review on prevention and management of hospital-acquired infections: Current strategies and best practices. *Int. J. Biol. Pharm. Sci. Arch.* **2023**, *6*, 149–155. [[CrossRef](#)]
20. Qattan, S.Y.M.; Al Zaydan, S.M.S.; Almalki, A.A.S.; Almari, B.M.S.; Jassas, M.M.A. Nosocomial Infections: Prevention, Control and Surveillance. *IJSR* **2023**, *2*, 534–545. [[CrossRef](#)]
21. WHO. *WHO Guidelines on Hand Hygiene in Health Care: First Global Patient Safety Challenge Clean Care is Safer Care*; World Health Organization: Geneva, Switzerland, 2009; WHO Guidelines Approved by the Guidelines Review Committee. Available online: <http://www.ncbi.nlm.nih.gov/books/NBK144013/> (accessed on 6 November 2021).
22. Allegranzi, B.; Bagheri Nejad, S.; Combescure, C.; Graafmans, W.; Attar, H.; Donaldson, L.; Pittet, D. Burden of endemic health-care-associated infection in developing countries: Systematic review and meta-analysis. *Lancet* **2011**, *377*, 228–241. [[CrossRef](#)]
23. Nejad, S.B.; Allegranzi, B.; Syed, S.B.; Ellis, B.; Pittet, D. Health-care-associated infection in Africa: A systematic review. *Bull. World Health Organ.* **2011**, *89*, 757–765. [[CrossRef](#)]
24. Irek, E.O.; Amupitan, A.A.; Obadare, T.O.; Aboderin, A.O. A systematic review of healthcare-associated infections in Africa: An antimicrobial resistance perspective. *Afr. J. Lab. Med.* **2018**, *7*, 796. [[CrossRef](#)]
25. Fraser, J.L.; Mwatondo, A.; Alimi, Y.H.; Varma, J.K.; Vilas, V.J.D.R. Healthcare-associated outbreaks of bacterial infections in Africa, 2009–2018: A review. *Int. J. Infect. Dis.* **2020**, *103*, 469–477. [[CrossRef](#)]
26. Tadesse, B.T.; Ashley, E.A.; Ongarello, S.; Havumaki, J.; Wijegoonewardena, M.; González, I.J.; Dittrich, S. Antimicrobial resistance in Africa: A systematic review. *BMC Infect. Dis.* **2017**, *17*, 616. [[CrossRef](#)]
27. Shiferaw, W.S.; Aynalem, Y.A.; Akalu, T.Y.; Petrucka, P.M. Surgical site infection and its associated factors in Ethiopia: A systematic review and meta-analysis. *BMC Surg.* **2020**, *20*, 107. [[CrossRef](#)] [[PubMed](#)]
28. Malande, O.O.; Nuttall, J.; Pillay, V.; Bamford, C.; Eley, B. A ten-year review of ESBL and non-ESBL *Escherichia coli* bloodstream infections among children at a tertiary referral hospital in South Africa. *PLoS ONE* **2019**, *14*, e0222675. [[CrossRef](#)] [[PubMed](#)]
29. Dramowski, A.; Madide, A.; Bekker, A. Neonatal nosocomial bloodstream infections at a referral hospital in a middle-income country: Burden, pathogens, antimicrobial resistance and mortality. *Paediatr. Int. Child Health* **2015**, *35*, 265–272. [[CrossRef](#)] [[PubMed](#)]
30. Page, M.J.; McKenzie, J.E.; Bossuyt, P.M.; Boutron, I.; Hoffmann, T.C.; Mulrow, C.D.; Shamseer, L.; Tetzlaff, J.M.; Akl, E.A.; Brennan, S.E.; et al. The PRISMA 2020 statement: An updated guideline for reporting systematic reviews. *BMJ* **2021**, *372*, n71. [[CrossRef](#)]
31. Huang, X.; Lin, J.; Demner-Fushman, D. Evaluation of PICO as a Knowledge Representation for Clinical Questions. *AMIA Annu. Symp. Proc.* **2006**, *2006*, 359–363.
32. Nishikawa-Pacher, A. Research Questions with PICO: A Universal Mnemonic. *Publications* **2022**, *10*, 21. [[CrossRef](#)]
33. de Jager, P.; Chirwa, T.; Naidoo, S.; Perovic, O.; Thomas, J. Nosocomial Outbreak of New Delhi Metallo- β -Lactamase-1-Producing Gram-Negative Bacteria in South Africa: A Case-Control Study. *PLoS ONE* **2015**, *10*, e0123337. [[CrossRef](#)]
34. Reddy, D.; Morrow, B.M.; Argent, A.C. Acinetobacter baumannii infections in a South African paediatric intensive care unit. *J. Trop. Pediatr.* **2015**, *61*, 182–187. [[CrossRef](#)]
35. Bediako-Bowan, A.A.A.; Kurtzhals, J.A.L.; Mølbak, K.; Labi, A.K.; Owusu, E.; Newman, M.J. High rates of multi-drug resistant gram-negative organisms associated with surgical site infections in a teaching hospital in Ghana. *BMC Infect. Dis.* **2020**, *20*, 890. [[CrossRef](#)] [[PubMed](#)]

36. Carshon–Marsh, R.; Squire, J.S.; Kamara, K.N.; Sargsyan, A.; Delamou, A.; Camara, B.S.; Manzi, M.; Guth, J.A.; Khogali, M.A.; Reid, A. Incidence of surgical site infection and use of antibiotics among patients who underwent caesarean section and herniorrhaphy at a regional referral hospital, Sierra Leone. *Int. J. Environ. Res. Public Health* **2022**, *19*, 4048. [[CrossRef](#)] [[PubMed](#)]
37. Landre–Peigne, C.; Ka, A.S.; Peigne, V.; Bougere, J.; Seye, M.N.; Imbert, P. Efficacy of an infection control programme in reducing nosocomial bloodstream infections in a Senegalese neonatal unit. *J. Hosp. Infect.* **2011**, *79*, 161–165. [[CrossRef](#)] [[PubMed](#)]
38. Meli, H.; Cissoko, Y.; Konaté, I.; Soumaré, M.; Fofana, A.; Dembélé, J.P.; Kaboré, M.; Cissé, M.A.; Zaré, A.; Dao, S. Co–infection tuberculose–VIH compliquée d’une sur infection nosocomiale à *Klebsiella pneumoniae*: À propos de 4 observations dans un Service de Maladies Infectieuses au Mali. *Pan Afr. Med. J.* **2020**, *37*, 141. [[CrossRef](#)] [[PubMed](#)]
39. Lo, S.; Thiam, I.; Fall, B.; Ba–Diallo, A.; Diallo, O.F.; Diagne, R.; Dia, M.L.; Ka, R.; Sarr, A.M.; Sow, A.I. Urinary tract infection with *Corynebacterium aurimucosum* after urethroplasty stricture of the urethra: A case report. *J. Med. Case Rep.* **2015**, *9*, 156.
40. Moissenet, D.; Weill, F.X.; Arlet, G.; Harrois, D.; Girardet, J.P.; Vu–Thien, H. *Salmonella enterica* serotype Gambia with CTX–M–3 and armA resistance markers: Nosocomial infections with a fatal outcome. *J. Clin. Microbiol.* **2011**, *49*, 1676–1678.
41. Aulakh, A.; Idoko, P.; Anderson, S.T.; Graham, W. Caesarean section wound infections and antibiotic use: A retrospective case–series in a tertiary referral hospital in The Gambia. *Trop. Doct* **2018**, *48*, 192–199. [[CrossRef](#)]
42. Nwankwo, E.O.; Mofolorunsho, C.K.; Akande, A.O. Aetiological agents of surgical site infection in a specialist hospital in Kano, north–western Nigeria. *Tanzan. J. Health Res.* **2014**, *16*, 289–295.
43. Lakoh, S.; Li, L.; Sevalie, S.; Guo, X.; Adekanmbi, O.; Yang, G.; Oladimeji, A.; Le, Y.; Joshua, M.C.; Shuchao, W.; et al. Antibiotic resistance in patients with clinical features of healthcare–associated infections in an urban tertiary hospital in Sierra Leone: A cross–sectional study. *Antimicrob. Resist. Infect. Control* **2020**, *9*, 38. [[CrossRef](#)]
44. Akinkunmi, E.O.; Adesunkanmi, A.R.; Lamikanra, A. Pattern of pathogens from surgical wound infections in a Nigerian hospital and their antimicrobial susceptibility profiles. *Afr. Health Sci.* **2014**, *14*, 802–809. [[CrossRef](#)]
45. Yehouenou, C.L.; Kpangon, A.A.; Affolabi, D.; Rodriguez–Villalobos, H.; Van Bambeke, F.; Dalleur, O.; Simon, A. Antimicrobial resistance in hospitalized surgical patients: A silently emerging public health concern in Benin. *Ann. Clin. Microbiol. Antimicrob.* **2020**, *19*, 54. [[CrossRef](#)] [[PubMed](#)]
46. Olowo–okere, A.; Ibrahim, Y.K.E.; Olayinka, B.O. Molecular characterisation of extended–spectrum β –lactamase–producing Gram–negative bacterial isolates from surgical wounds of patients at a hospital in North Central Nigeria. *J. Glob. Antimicrob. Resist.* **2018**, *14*, 85–89. [[CrossRef](#)] [[PubMed](#)]
47. Labi, A.K.; Nielsen, K.L.; Marvig, R.L.; Bjerrum, S.; Enweronu–Laryea, C.; Bennedbæk, M.; Newman, M.J.; Ayibor, P.K.; Andersen, L.P.; Kurtzhals, J.A. Oxacillinase–181 Carbapenemase–Producing *Klebsiella pneumoniae* in Neonatal Intensive Care Unit, Ghana, 2017–2019. *Emerg. Infect. Dis.* **2020**, *26*, 2235–2238. [[CrossRef](#)] [[PubMed](#)]
48. Guetarni, N.; Zouagui, S.; Besbes, F.; Derkaoui, A.; Hanba, M.; Ahmed Fouatih, Z. Infections Nosocomiales (IN): Enquête de prévalence et d’identification des facteurs de risque dans un centre hospitalier universitaire de la région ouest d’Algérie, 2016. *Rev. Méd. L Hmruo* **2017**, *4*, 584–590.
49. Bayingana, C.; Sendegeya, A.; Habarugira, F.; Mukumpunga, C.; Lyumugabe, F.; Ndoli, J. Hospital acquired infections in pediatrics unit at Butare University Teaching Hospital (CHUB). *Rwanda J. Med. Health Sci.* **2019**, *2*, 272–276. [[CrossRef](#)]
50. Kolawole, O.M.; Idakwo, A.I.; Ige, O.; Anibijuwon, I.O. Prevalence of Hospital Acquired Infections in The Intensive Care Unit of University of Ilorin Teaching Hospital, Ilorin Nigeria. *Sudan J. Med. Sci.* **2015**, *10*, 59–66.
51. Nair, A.; Steinberg, W.J.; Habib, T.; Saeed, H.; Raubenheimer, J.E. Prevalence of healthcare–associated infection at a tertiary hospital in the Northern Cape Province, South Africa. *S. Afr. Fam. Pract.* **2018**, *60*, 162–167. [[CrossRef](#)]
52. Sanou, I.; Kabore, A.; Tapsoba, E.; Bicaba, I.; Ba, A.; Zango, B. Nosocomial Urinary Infections at the Urogoly Unit of the National University Hospital (Yalgado Ouedraogo), Ouagadougou: Feb.–Sept 2012. *Afr. J. Clin. Exp. Microbiol.* **2015**, *16*, 1–6. [[CrossRef](#)]
53. Iwuafor, A.A.; Ogunsola, F.T.; Oladele, R.O.; Oduyebo, O.O.; Desalu, I.; Egwuatu, C.C.; Nnachi, A.U.; Akujobi, C.N.; Ita, I.O.; Ogban, G.I. Incidence, Clinical Outcome and Risk Factors of Intensive Care Unit Infections in the Lagos University Teaching Hospital (LUTH), Lagos, Nigeria. *PLoS ONE* **2016**, *11*, e0165242. [[CrossRef](#)]
54. Kihla, A.J.F.T.; Ngunde, P.J.; Mbianda, S.E.; Nkwelang, G.; Ndip, R.N. Risk factors for wound infection in health care facilities in Buea, Cameroon: Aerobic bacterial pathogens and antibiogram of isolates. *Pan Afr. Med. J.* **2014**, *18*, 6. [[CrossRef](#)]
55. Madu, K.A.; Enweani, U.N.; Katchy, A.U.; Madu, A.J.; Aguwa, E.N. Implant Associated Surgical Site Infection in Orthopaedics: A Regional Hospital Experience. *Niger. J. Med.* **2011**, *20*, 435–440.
56. Manyahi, J.; Matee, M.I.; Majigo, M.; Moyo, S.; Mshana, S.E.; Lyamuya, E.F. Predominance of multi–drug resistant bacterial pathogens causing surgical site infections in Muhimbili National Hospital, Tanzania. *BMC Res. Notes* **2014**, *7*, 500. [[CrossRef](#)]
57. Victor, D.; Revathi, G.; Sam, K.; Abdi, H.; Asad, R.; Andrew, K. Pattern of pathogens and their sensitivity isolated from surgical site infections at the Aga Khan University Hospital, Nairobi, Kenya. *Ethiop. J. Health Sci.* **2013**, *23*, 141–149.
58. Khan, N.B.; Charles, C.R.; Naidoo, N.; Nokubonga, A.; Mkhwanazi, N.A.; Moustache, H.M.T.E. Infection prevention and control measures in audiology practice within public healthcare facilities in KwaZulu–Natal province, South Africa. *S. Afr. J. Commun. Disord.* **2019**, *66*, e1–e14. [[CrossRef](#)] [[PubMed](#)]

59. Afeke, I.; Amegan–Aho, K.H.; Adu–Amankwaah, J.; Orish, V.N.; Mensah, G.L.; Mbroh, H.K.; Jamfaru, I.; Hamid, A.W.M.; Mac Ankrah, L.; Korbuvi, J. Antimicrobial profile of coagulase–negative staphylococcus isolates from categories of individuals at a neonatal intensive care unit of a tertiary hospital, Ghana. *Pan Afr. Med. J.* **2023**, *44*, 92. [[CrossRef](#)] [[PubMed](#)]
60. Benammar, S.; Pantel, A.; Aujoulat, F.; Benmehidi, M.; Courcol, R.; Lavigne, J.P.; Romano–Bertrand, S.; Marchandin, H. First molecular characterization of related cases of healthcare–associated infections involving multidrug–resistant *Enterococcus faecium* vanA in Algeria. *Infect. Drug Resist.* **2018**, *11*, 1483. [[CrossRef](#)]
61. Palle, J.N.; Bassah, N.; Kamga, H.L.F.; Nkwelang, G.; Akoachere, J.F.; Mbianda, E.; Nwarie, U.G.; Njunda, A.L.; Assob, N.J.C.; Ekane, G.H. Current antibiotic susceptibility profile of the bacteria associated with surgical wound infections in the buea health district in Cameroon. *Afr. J. Clin. Exp. Microbiol.* **2014**, *15*, 27–34. [[CrossRef](#)]
62. Godebo, G.; Kibru, G.; Tassew, H. Multidrug–resistant bacterial isolates in infected wounds at Jimma University Specialized Hospital, Ethiopia. *Ann. Clin. Microbiol. Antimicrob.* **2013**, *12*, 17. [[CrossRef](#)]
63. Ogwang, M.; Paramatti, D.; Molteni, T.; Ochola, E.; Okello, T.R.; Salgado, J.O.; Kayanja, A.; Greco, C.; Kizza, D.; Gondoni, E. Prevalence of hospital–associated infections can be decreased effectively in developing countries. *J. Hosp. Infect.* **2013**, *84*, 138–142. [[CrossRef](#)]
64. Heysell, S.K.; Sheno, S.V.; Catterick, K.; Thomas, T.A.; Friedland, G. Prevalence of methicillin–resistant *Staphylococcus aureus* nasal carriage among hospitalised patients with tuberculosis in rural Kwazulu–Natal. *S. Afr. Med. J.* **2011**, *101*, 332–334. [[CrossRef](#)] [[PubMed](#)]
65. Hien, H.; Drabo, K.M.; Ouédraogo, L.; Konfé, S.; Zeba, S.; Sangaré, L.; Compaoré, S.C.; Ouédraogo, J.B.; Ouendo, E.M.; Makoutodé, M.; et al. Healthcare–associated infection in Burkina Faso: An assessment in a district hospital. *J. Public Health* **2012**, *3*, e29. [[CrossRef](#)] [[PubMed](#)]
66. Nakoune, E.; Lampaert, E.; Ndjapou, S.G.; Janssens, C.; Zuniga, I.; Van Herp, M.; Fongbia, J.P.; Koyazegbe, T.D.; Selekon, B.; Komoyo, G.F.; et al. A Nosocomial Outbreak of Human Monkeypox in the Central African Republic. *Open Forum Infect. Dis.* **2017**, *4*, ofx168. [[CrossRef](#)] [[PubMed](#)]
67. Scherbaum, M.; Kösters, K.; Mürbeth, R.E.; Ngoa, U.A.; Kremsner, P.G.; Lell, B.; Alabi, A. Incidence, pathogens and resistance patterns of nosocomial infections at a rural hospital in Gabon. *BMC Infect. Dis.* **2014**, *14*, 124. [[CrossRef](#)]
68. Esebelahie, N.O.; Newton–Esebelahie, F.O.; Omoregie, R. Aerobic bacterial isolates from infected wounds. *Afr. J. Clin. Exp. Microbiol.* **2013**, *14*, 155–159. [[CrossRef](#)]
69. Osaiyuwu, O. Occurrence of bacteraemia following oral and maxillofacial surgical procedures in Port Harcourt, Nigeria. *Afr. Health Sci.* **2021**, *21*, 1692–1700. [[CrossRef](#)]
70. Olu–Taiwo, M.A.; Opintan, J.A.; Codjoe, F.S.; Obeng Forson, A. Metallo–Beta–Lactamase–Producing *Acinetobacter* spp. from Clinical Isolates at a Tertiary Care Hospital in Ghana. *Biomed. Res. Int.* **2020**, *2020*, 3852419. [[CrossRef](#)]
71. Abdoulaye, O.; Amadou, M.L.H.; Amadou, O.; Adakal, O.; Larwanou, H.M.; Boubou, L.; Oumarou, D.; Abdoulaye, M.; Mamadou, S. Aspects épidémiologiques et bactériologiques des infections du site opératoire (ISO) dans les services de chirurgie à l’Hôpital National de Niamey (HNN). *Pan Afr. Med. J.* **2018**, *31*, 33. [[CrossRef](#)]
72. Roubi, I.; Ghemri, S. Enquête sur la Prévalence des Infections Associées Aux Soins (IAS) Dans Les Établissements de Santé. Université Mohamed Khider de Biskra. 2024. Available online: http://archives.univ--biskra.dz/bitstream/123456789/30034/1/Im%C3%A9ne_ROUBI_Selma_GHEMRI.pdf (accessed on 16 May 2025).
73. Joshua, I.A.; Giwa, F.J.; Kwaga, J.K.P.; Kabir, J.; Owolodun, O.A.; Umaru, G.A.; Habib, A.G. Molecular Characterisation of *Staphylococcus aureus* isolated from Patients in Healthcare facilities in Zaria Metropolis, Kaduna State, Nigeria. *J. Epidemiol. Soc. Niger.* **2021**, *4*, 87–101.
74. Bojang, A.; Chung, M.; Camara, B.; Jagne, I.; Guérillot, R.; Ndure, E.; Howden, B.P.; Roca, A.; Ghedin, E. Genomic approach to determine sources of neonatal *Staphylococcus aureus* infection from carriage in the Gambia. *BMC Infect. Dis.* **2024**, *24*, 941. [[CrossRef](#)]
75. Ahoyo, T.A.; Martin–Odoom, A.; Bankolé, H.S.; Baba–Moussa, L.; Zonon, N.; Loko, F.; Prevost, G.; Sanni, A.; Dramane, K. Epidemiology and prevention of nosocomial pneumonia associated with Panton–Valentine Leukocidin (PVL) producing *Staphylococcus aureus* in Departmental Hospital Centre of Zou Collines in Benin. *Ghana Med. J.* **2012**, *46*, 234.
76. Ouedraogo, A.S.; Some, D.A.; Dakouré, P.W.H.; Sanon, B.G.; Poda, G.E.A.; Kambou, T. Profil bactériologique des infections du site opératoire au centre hospitalier universitaire Souro Sanou de Bobo Dioulasso. *Med. Trop.* **2011**, *71*, 49–52.
77. Ikumapayi, U.N.; Kanteh, A.; Manneh, J.; Lamin, M.; Mackenzie, G.A. An outbreak of *Serratia liquefaciens* at a rural health center in The Gambia. *J. Infect. Dev. Ctries.* **2016**, *10*, 791–798. [[CrossRef](#)]
78. Tabiri, S.; Yenli, E.; Kyere, M.; Anyomih, T.T.K. Surgical Site Infections in Emergency Abdominal Surgery at Tamale Teaching Hospital, Ghana. *World J. Surg.* **2018**, *42*, 916–922. [[CrossRef](#)]
79. Mafikoya, B.O.; Niemogha, M.T.; Ogunsola, F.T.; Atoyebi, O.A. Predictors of surgical site infections of the abdomen in Lagos, Nigeria. *Niger. Q. J. Hosp. Med.* **2021**, *21*, 124–128.

80. Labi, A.K.; Obeng–Nkrumah, N.; Owusu, E.; Bjerrum, S.; Bediako–Bowen, A.; Sunkwa–Mills, G.; Akufo, C.; Fenny, A.P.; Opintan, J.A.; Enweronu–Laryea, C. Multi–centre point–prevalence survey of hospital–acquired infections in Ghana. *J. Hosp. Infect.* **2019**, *101*, 60–68. [CrossRef] [PubMed]
81. Ezenwa, B.N.; Oladele, R.O.; Akintan, P.E.; Fajolu, I.B.; Oshun, P.O.; Oduyebo, O.O.; Ezeaka, V.C. Invasive candidiasis in a neonatal intensive care unit in Lagos, Nigeria. *Niger. Postgrad. Med. J.* **2017**, *24*, 150–154. [CrossRef] [PubMed]
82. Anago, E.; Ayi–Fanou, L.; Akpovi, C.D.; Hounkpe, W.B.; Agassounon–Djikpo Tchibozo, M.; Bankole, H.S.; Sanni, A. Antibiotic resistance and genotype of beta–lactamase producing *Escherichia coli* in nosocomial infections in Cotonou, Benin. *Ann. Clin. Microbiol. Antimicrob.* **2015**, *14*, 5. [CrossRef]
83. Agyepong, N.; Govinden, U.; Owusu–Ofori, A.; Allam, M.; Ismail, A.; Pedersen, T.; Sundsfjord, A.; Arnfinn, S.; Maresca, J.A. Whole–Genome Sequences of Two Multidrug–Resistant *Acinetobacter baumannii* Strains Isolated from Patients with Urinary Tract Infection in Ghana. *Microbiol. Resour. Announc.* **2019**, *8*, e00270–19. [CrossRef]
84. Duedu, K.O.; Offei, G.; Codjoe, F.S.; Donkor, E.S. Multidrug Resistant Enteric Bacterial Pathogens in a Psychiatric Hospital in Ghana: Implications for Control of Nosocomial Infections. *Int. J. Microbiol.* **2017**, *2017*, 9509087. [CrossRef]
85. Labi, A.K.; Bjerrum, S.; Enweronu–Laryea, C.C.; Ayibor, P.K.; Nielsen, K.L.; Marvig, R.L.; Newman, M.J.; Andersen, L.P.; Kurtzhals, J.A. High Carriage Rates of Multidrug–Resistant Gram–Negative Bacteria in Neonatal Intensive Care Units From Ghana. *Open Forum Infect. Dis.* **2020**, *7*, ofaa109. [CrossRef] [PubMed]
86. Coulibaly, Y.; Amadou, I.; Koné, O.; Coulibaly, O.M.; Diop, T.H.M.; Doumbia, A.; Kamaté, B.; Djiré, M.K.; Traoré, A.; Ouologuem, H. Infections associées aux soins en chirurgie pédiatrique au CHU Gabriel Touré, Bamako, Mali. *Mali. Méd.* **2020**, *35*, 15–19. [PubMed]
87. Aye, E.C.; Omoregie, R.; Ohiorenuan, I.I.; Onemu, S. Microbiology of Wound Infections and Its Associated Risk Factors Among Patients of a Tertiary Hospital in Benin City, Nigeria. 2011. Available online: https://www.sid.ir/EN/VEWSSID/J_pdf/129720110207.pdf (accessed on 18 May 2025).
88. Amissah, N.A.; Buultjens, A.H.; Ablordey, A.; van Dam, L.; Opoku–Ware, A.; Baines, S.L.; Bulach, D.; Tetteh, C.S.; Prah, I.; van der Werf, T.S. Methicillin Resistant *Staphylococcus aureus* Transmission in a Ghanaian Burn Unit: The Importance of Active Surveillance in Resource–Limited Settings. *Front. Microbiol.* **2017**, *8*, 1906. [CrossRef]
89. Ka, R. Aspects bactériologiques des infections liées aux cathéters veineux à l’hôpital Dalal Jamm de Guédiawaye (Sénégal). *Rev. Mali. Infect. Microbiol.* **2024**, *19*, 40–47. [CrossRef]
90. Diarra, A.; Keita, K.; Tounkara, I.; Traoré, A.; Koné, A.; Konaté, M.; Karembé, B.; Keita, M.A.; Traoré, I.; Togola, M. Infections du site opératoire en chirurgie générale du centre hospitalier universitaire bocar sidy sall de kati. *Mali. Méd.* **2020**, *35*, 20.
91. Afle, F.C.D.; Quenum, K.J.; Hessou, S.; Johnson, R.C. Etat des lieux des infections associées aux soins dans deux hôpitaux publics du sud Bénin (Afrique de l’ouest): Centre Hospitalier Universitaire de Zone d’Abomey–Calavi/Sô–Ava et Centre Hospitalier de Zone de Cotonou 5. *J. Appl. Biosci.* **2018**, *121*, 12192–12201. [CrossRef]
92. Dia, N.M.; Cissoko, Y.; Diouf, A.; Seydi, M. Results of a survey incidence of the cases of nosocomial infections with multidrug resistant bacteria in a hospital center in Dakar (Senegal). *Rev. Mali. D Infect. Microbiol.* **2015**, *5*, 8.
93. Ahoyo, T.A.; Bankolé, H.S.; Adéoti, F.M.; Gbohoun, A.A.; Assavèdo, S.; Amoussou–Guénou, M.; Kindé–Gazard, D.A.; Pittet, D. Prevalence of nosocomial infections and anti–infective therapy in Benin: Results of the first nationwide survey in 2012. *Antimicrob. Resist. Infect. Control* **2014**, *3*, 17. [CrossRef]
94. Lukuke, H.M.; Kasamba, E.; Mahuridi, A.; Ngatu, N.R.; Narufumi, S.; Mukengeshayi, A.N.; Malou, V.; Makoutode, M.; Kaj, F.M. L’incidence des infections nosocomiales urinaires et des sites opératoires dans la maternité de l’Hôpital Général de Référence de Katuba à Lubumbashi en République Démocratique du Congo. *Pan Afr. Med. J.* **2017**, *28*, 57. [CrossRef]
95. Lowman, W. Active surveillance of hospital–acquired infections in South Africa: Implementation, impact and challenges. *S. Afr. Med. J.* **2016**, *106*, 59. [CrossRef]
96. Iliyasu, G.; Dayyab, F.M.; Abubakar, S.; Inuwa, S.; Tambuwal, S.H.; Tiamiyu, A.B.; Habib, Z.G.; Gadanya, M.A.; Sheshe, A.A.; Mijinyawa, M.S. Laboratory–confirmed hospital–acquired infections: An analysis of a hospital’s surveillance data in Nigeria. *Heliyon* **2018**, *4*, e00720. [CrossRef] [PubMed]
97. Gezmu, A.M.; Bulabula, A.N.H.; Dramowski, A.; Bekker, A.; Aucamp, M.; Souda, S.; Nakstad, B. Laboratory–confirmed bloodstream infections in two large neonatal units in sub–Saharan Africa. *Int. J. Infect. Dis.* **2021**, *103*, 201–207. [CrossRef] [PubMed]
98. Dramowski, A.; Whitelaw, A.; Cotton, M.F. Burden, spectrum, and impact of healthcare–associated infection at a South African children’s hospital. *J. Hosp. Infect.* **2016**, *94*, 364–372. [CrossRef] [PubMed]
99. Dan–Nwafor, C.C.; Ipadeola, O.; Smout, E.; Ilori, E.; Adeyemo, A.; Umeokonkwo, C.; Nwidi, D.; Nwachukwu, W.; Ukponu, W.; Omabe, E.; et al. A cluster of nosocomial Lassa fever cases in a tertiary health facility in Nigeria: Description and lessons learned, 2018. *Int. J. Infect. Dis.* **2019**, *83*, 88–94. [CrossRef]
100. Dramowski, A.; Cotton, M.F.; Whitelaw, A. Surveillance of healthcare–associated infection in hospitalised South African children: Which method performs best? *S. Afr. Med. J.* **2016**, *107*, 56–63. [CrossRef]

101. Lowe, M.; Ehlers, M.M.; Ismail, F.; Peirano, G.; Becker, P.J.; Pitout, J.D.D.; Kock, M.M. Acinetobacter baumannii: Epidemiological and Beta-Lactamase Data From Two Tertiary Academic Hospitals in Tshwane, South Africa. *Front. Microbiol.* **2018**, *9*, 1280. [CrossRef]
102. Olivier, C.; Kunneke, H.; O'Connell, N.; Von Delft, E.; Wates, M.; Dramowski, A. Healthcare-associated infections in paediatric and neonatal wards: A point prevalence survey at four South African hospitals. *S. Afr. Med. J.* **2018**, *108*, 418–422. [CrossRef]
103. Spicer, K.B.; Green, J.; Dhada, B. Hospital-acquired infections in paediatric medical wards at a tertiary hospital in KwaZulu-Natal, South Africa. *Paediatr. Int. Child. Health* **2018**, *38*, 53–59. [CrossRef]
104. Kakupa, D.K.; Muenze, P.K.; Byl, B.; Wilmet, M.D. Study of the prevalence of nosocomial infections and associated factors in the two university hospitals of Lubumbashi, Democratic Republic of Congo. *Pan Afr. Med. J.* **2016**, *24*, 275.
105. Tagoe, D.N.A.; Baidoo, S.E.; Dadzie, I.; Tengey, D.; Agede, C. Potential sources of transmission of hospital acquired infections in the volta regional hospital in Ghana. *Ghana Med. J. Mars* **2011**, *45*, 22–26. [CrossRef]
106. Greco, D.; Magombe, I. Hospital acquired infections in a large north Ugandan hospital. *J. Prev. Med. Hyg.* **2011**, *52*, 55–58.
107. Makanjuola, O.B.; Fayemiwo, S.A.; Okesola, A.O.; Gbaja, A.; Ogunleye, V.A.; Kehinde, A.O.; Bakare, R.A. Pattern of multidrug resistant bacteria associated with intensive care unit infections in Ibadan, Nigeria. *Ann. Ib. Postgrad. Med.* **2018**, *16*, 162–169. [PubMed]
108. Okello, T.R.; Kansime, J.; Odora, J. Invasive procedures and Hospital Acquired Infection (HAI) in A large hospital in Northern Uganda. *East Cent. Afr. J. Surg.* **2014**, *19*, 77–84.
109. Rafai, C.; Frank, T.; Manirakiza, A.; Gaudeuille, A.; Mbecko, J.R.; Nghario, L.; Serdouma, E.; Tekpa, B.; Garin, B.; Breurec, S. Dissemination of IncF-type plasmids in multiresistant CTX-M-15-producing Enterobacteriaceae isolates from surgical-site infections in Bangui, Central African Republic. *BMC Microbiol.* **2015**, *15*, 15. [CrossRef] [PubMed]
110. Okolo, M.O.; Toma, B.O.; Onyedibe, K.I.; Emanghe, U.; Banwat, E.B.; Egah, D.Z. Bacterial Contamination In A Special Care Baby Unit Of A Tertiary Hospital In Jos, Nigeria. *Niger. J. Med.* **2016**, *25*, 259–263. [CrossRef]
111. Beye, S.A.; Maiga, A.; Cissoko, Y.; Guindo, I.; Dicko, O.A.; Maiga, M.; Abeghe, A.T.A.; Diakit , M.; Diallo, B.; Dao, S. Prevalence of nosocomial infections at the Centre Hospitalier Universitaire du Point, G. in Bamako, Mali. *Rev. Mali. D Infect. Microbiol.* **2024**, *19*, 45.
112. Seni, J.; Najjuka, C.F.; Kateete, D.P.; Makobore, P.; Joloba, M.L.; Kajumbula, H.; Kapesa, A.; Bwanga, F. Antimicrobial resistance in hospitalized surgical patients: A silently emerging public health concern in Uganda. *BMC Res. Notes* **2013**, *6*, 298. [CrossRef]
113. Demb l , G. Infection du Site Op ratoire dans le Service de Traumatologie   L'h pital de Sikasso. Ph.D. Thesis, USTTB, Bamako, Mali, 2020. Available online: <https://library.adhl.africa/handle/123456789/14015> (accessed on 13 October 2025).
114. Saidoun, A.A. Surveillance des Infections Nosocomiales en P diatrie au CHU B ni Messous d'Alger. Ph.D. Thesis, Facult  de M decine d'Alger, El Biar, Algeria, 2021. Available online: <https://theses.hal.science/tel-03374867/> (accessed on 13 October 2025).
115. Rim, L. Facteurs Pr dictifs des Infections Nosocomiales en Milieu de R animation 2016. Available online: <https://toubkal.imist.ma/handle/123456789/28014> (accessed on 16 May 2025).
116. Angoue, T.A.A. Pr valence des Infections Nosocomiales Dans 10 Services du CHU du Point G. Ph.D. Thesis, Th se de M decine USTTB, Bamako, Mali, 2020. Available online: <https://bibliosante.ml/bitstream/handle/123456789/4442/20M05.pdf?sequence=1> (accessed on 16 May 2025).
117. Mulu, W.; Kibru, G.; Beyene, G.; Damtie, M. Postoperative nosocomial infections and antimicrobial resistance pattern of bacteria isolates among patients admitted at Felege Hiwot Referral Hospital, Bahirdar, Ethiopia. *Ethiop. J. Health Sci.* **2012**, *22*, 7–18.
118. Mwita, J.C.; Souda, S.; Magafu, M.G.M.D.; Masseur, A.; Godman, B.; Mwandri, M. Prophylactic antibiotics to prevent surgical site infections in Botswana: Findings and implications. *Hosp. Pract.* **2018**, *46*, 97–102. [CrossRef]
119. Talaat, M.; Zayed, B.; Tolba, S.; Abdou, E.; Gomaa, M.; Itani, D.; Hutin, Y.; Hajjeh, R. Increasing antimicrobial resistance in world health organization eastern mediterranean region, 2017–2019. *Emerg. Infect. Dis.* **2022**, *28*, 717. [CrossRef]
120. Togo, A.; Samak , M.; Sarro, Y.S.; Youssouf, D.; Kanikomo, D.; Diop, M.; Yacaria, C.; Niani, M.; Mamadou, C.; Diarra, B. Healthcare-associated infection in surgery department: A big challenge for African surgical teams. *J. West Afr. Coll. Surg.* **2023**, *13*, 73–77. [CrossRef] [PubMed]
121. Andersen, C.T.; Langendorf, C.; Garba, S.; Sayinzonga-Makombe, N.; Mambula, C.; Mouniaman, I.; Hanson, K.E.; Grais, R.F.; Isanaka, S. Risk of community- and hospital-acquired bacteremia and profile of antibiotic resistance in children hospitalized with severe acute malnutrition in Niger. *Int. J. Infect. Dis.* **2022**, *119*, 163–171. [CrossRef] [PubMed]
122. Smith, A.M.; Mthanti, M.A.; Haumann, C.; Tyalisi, N.; Boon, G.P.G.; Sooka, A.; Keddy, K.H. Nosocomial Outbreak of Salmonella enterica Serovar Typhimurium Primarily Affecting a Pediatric Ward in South Africa in 2012. *J. Clin. Microbiol.* **2014**, *52*, 627–631. [CrossRef] [PubMed]
123. van Schalkwyk, E.; Iyaloo, S.; Naicker, S.D.; Maphanga, T.G.; Mpembe, R.S.; Zulu, T.G.; Mhlanga, M.; Mahlangu, S.; Maloba, M.B.; Ntlemo, G.; et al. Large Outbreaks of Fungal and Bacterial Bloodstream Infections in a Neonatal Unit, South Africa, 2012–2016. *Emerg. Infect. Dis.* **2018**, *24*, 1204–1212. [CrossRef]

124. Lasmé–Guillao, E.; Amon–Tanoh–Dick, F.; GBonon, V.; Akaffou, E.A.; Kabas, R.; Faye–Kette, H. Infections a Klebsiella pneumonia et Enterobacter cloacae en néonatalogie a Abidjan. *J. Pédiatrie Puéricult.* **2011**, *24*, 118–124. [[CrossRef](#)]
125. Nouetchognou, J.S.; Ateudjieu, J.; Jemea, B.; Mesumbe, E.N.; Mbanya, D. Surveillance of nosocomial infections in the Yaounde University Teaching Hospital, Cameroon. *BMC Res. Notes* **2016**, *9*, 505. [[CrossRef](#)]
126. Lakoh, S.; Yi, L.; Sevalie, S.; Guo, X.; Adekanmbi, O.; Smalle, I.O.; Williams, N.; Barrie, U.; Koroma, C.; Zhao, Y.; et al. Incidence and risk factors of surgical site infections and related antibiotic resistance in Freetown, Sierra Leone: A prospective cohort study. *Antimicrob. Resist. Infect. Control* **2022**, *11*, 39. [[CrossRef](#)]
127. Lakoh, S.; Yi, L.; Russell, J.B.W.; Zhang, J.; Sevalie, S.; Zhao, Y.; Kanu, J.S.; Liu, P.; Conteh, S.K.; Williams, C.E.E.; et al. High incidence of catheter–associated urinary tract infections and related antibiotic resistance in two hospitals of different geographic regions of Sierra Leone: A prospective cohort study. *BMC Res. Notes* **2023**, *16*, 301. [[CrossRef](#)]

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