

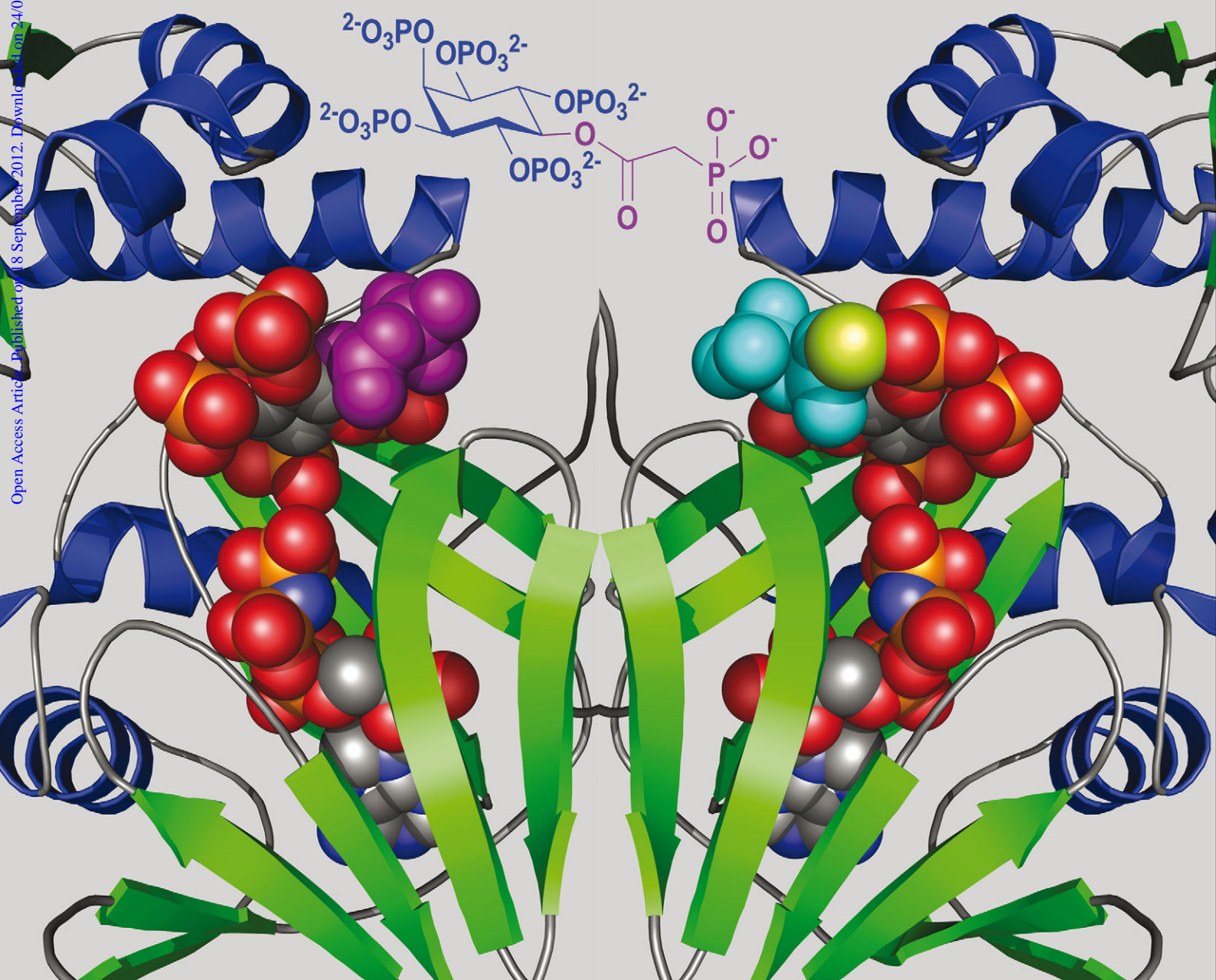
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COMMUNICATION

First synthetic analogues of diphosphoinositol polyphosphates: interaction with PP-InsP₅ kinase†Andrew M. Riley,^a Huanchen Wang,^b Jeremy D. Weaver,^b Stephen B. Shears^b and Barry V. L. Potter^{*a}

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We synthesised analogues of diphosphoinositol polyphosphates (PP-InsPs) in which the diphosphate is replaced by an α -phosphonoacetic acid (PA) ester. Structural analysis revealed that 5-PA-InsP₅ mimics 5-PP-InsP₅ binding to the kinase domain of PPIP5K2; both molecules were phosphorylated by the enzyme. PA-InsPs are promising candidates for further studies into the biology of PP-InsPs.

myo-Inositol polyphosphates (InsPs) regulate many cellular processes.¹ The chemical synthesis of InsPs, plus various analogues and probes, has been vital to the functional characterisation of enzymes and receptors involved in InsP metabolism and signalling.² In the early 1990s, InsPs containing diphosphate groups were discovered.³ These diphosphoinositol polyphosphates (PP-InsPs, Fig. 1a) have been proposed to regulate a range of cellular functions including vesicular trafficking, apoptosis, DNA repair, telomere maintenance, insulin signalling and stress responses.⁴ However, little is known about the molecular mechanisms that underlie these effects. There is evidence that PP-InsPs can act through non-covalent interactions with receptor proteins.⁵ A more provocative mechanistic proposal involves non-enzymatic transfer of the β -phosphoryl of the diphosphate (PP) group in PP-InsPs to a phosphorylated serine residue in target proteins, to give a diphosphorylated protein.⁶

Several chemical syntheses of PP-InsPs have been published,⁷ although no analogues or probes based on PP-InsPs have been described. We set out to synthesise analogues that cannot transfer phosphoryl groups, yet can mimic non-covalent interactions of PP-InsPs with target proteins. Such stabilised analogues could potentially be used for the mechanistic dissection of PP-InsP signalling. They would also be open to synthetic modifications such as tagging with fluorophores or photoreactive groups and the synthesis of biotinylated and resin-immobilised versions.

Diphosphate mimics such as methylenebisphosphonates and imidodiphosphates, often used in non-hydrolysable analogues of nucleotide diphosphates,⁸ are not easily introduced onto the secondary hydroxyl groups of *myo*-inositol. We therefore chose to explore α -phosphonoacetic acid (phosphonoacetate, PA) esters⁹ as mimics of the PP motif in PP-InsPs. The PA ester can be introduced by straightforward *O*-acylation, without disturbing the *myo*-inositol stereochemistry. Due to the higher activation energy for cleavage of a C–P bond *versus* an O–P bond,¹⁰ it is not plausible that PA-InsPs could act as non-enzymatic phosphoryl donors. We identified 5-PA-InsP₄ (**1**) and 5-PA-InsP₅ (**2**) (Fig. 1b) as initial targets. Noting previous evidence that a 2-*O*-benzyl group enhances the anti-tumour and pro-apoptotic properties of InsP₅,¹¹ we also synthesised 2-*O*-benzyl-5-PA-InsP₄ (**3**).

The syntheses of **1**, **2** and **3** (Scheme 1) began with diol **4**, directly available from *myo*-inositol in multi-gram quantities.¹² For the synthesis of **1**, alkylation of **4** was used to introduce a benzyl ether selectively at the axial O-2 atom, giving **5**. The origin of

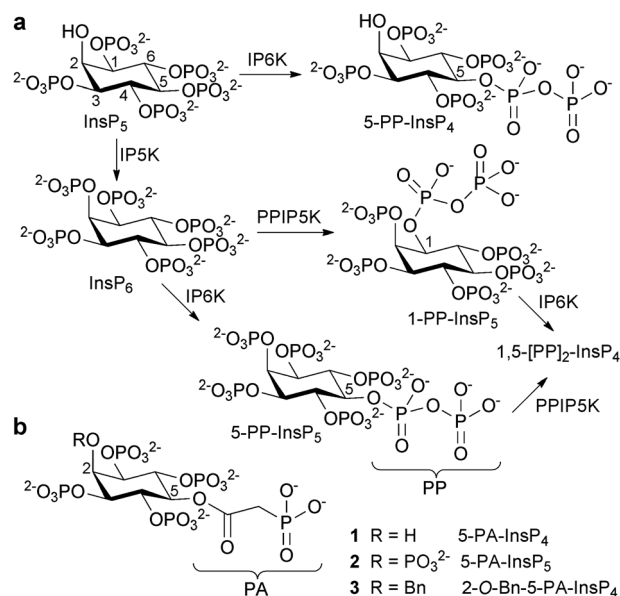
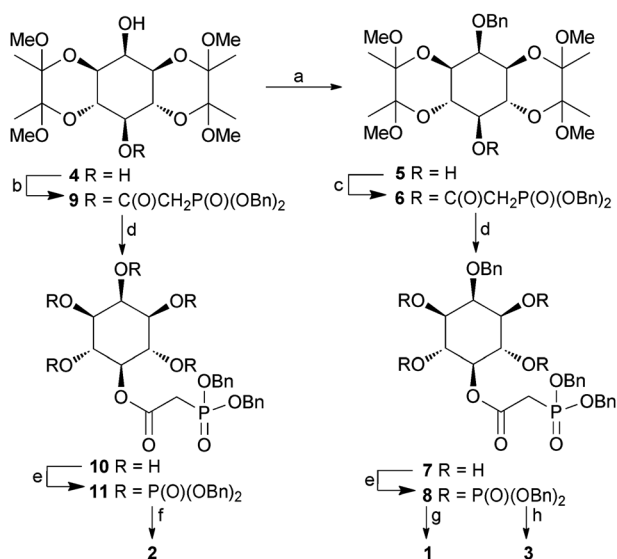


Fig. 1 (a) Origin of diphosphoinositol polyphosphates. IP5K, inositol pentakisphosphate 2-kinase; IP6K, inositol hexakisphosphate 5-kinase; PPIP5K, diphosphoinositol pentakisphosphate kinase. (b) Structures of phosphonoacetic acid ester (PA) analogues **1**, **2** and **3**. Bn = benzyl.

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† Electronic supplementary information (ESI) available: Data deposition: the atomic coordinates and structure factors have been deposited in the Protein Data Bank, www.pdb.org (PDB ID code 4GB4). See DOI: 10.1039/c2cc36044f



Scheme 1 Synthesis of PA analogues **1**, **2** and **3**. Reagents and conditions: a. BnBr, NaH, DMF, 55%; b. (BnO)₂P(O)CH₂COOH, DCC, CH₂Cl₂, 73%; c. (BnO)₂P(O)CH₂COOH, EDAC, DMAP, CH₂Cl₂, 95%; d. TFA, H₂O, 5 min, 50–62%; e. (i) (BnO)₂PNPr₂, 5-phenyl-1H-tetrazole, CH₂Cl₂; (ii) mCPBA, CH₂Cl₂, 84–95%; f. H₂, Pd(OH)₂/C, MeOH, H₂O, 92%; g. H₂, Pd(OH)₂/C, MeOH, H₂O, 50 psi, 79%; h. H₂, Pd(OH)₂/C, MeOH, aqueous triethylammonium bicarbonate, 79%. Bn = benzyl.

the regioselectivity observed in this reaction is not clear. The free 5-OH group of **5** was then acylated using dibenzylphosphonoacetic acid and EDAC giving **6**. Careful removal of the butane diacetate (BDA) groups using aqueous trifluoroacetic acid gave tetraol **7**. Phosphitylation of **7** using bis(benzylphosphoryl)diisopropylaminophosphine, followed by oxidation gave **8**, and global deprotection by hydrogenolysis gave 5-PA-InsP₄ (**1**). For the synthesis of 5-PA-InsP₅ (**2**), regioselective carbodiimide-mediated acylation of the equatorial 5-hydroxyl group of diol **4** with dibenzylphosphonoacetic acid gave alcohol **9**. Removal of the BDA groups to give pentaol **10**, followed by phosphitylation/oxidation to give **11** and hydrogenolytic deprotection as before gave 5-PA-InsP₅ (**2**).

2-O-Bn-5-PA-InsP₄ (**3**) was prepared by selective deprotection of **8**. We have previously noted that triethylammonium ions suppress hydrogenolytic cleavage of 2-O-benzyl ethers in *myo*-inositol derivatives. For **8**, we found that hydrogenolysis in the presence of aqueous triethylammonium bicarbonate cleaved the benzylphosphate esters within 30 minutes, while the 2-O-benzyl ether was unaffected, giving **3** as the triethylammonium salt.

To determine whether **1**, **2** and **3** could act as mimics of 5-PP-InsP₅, we examined whether they were recognized by the highly specific catalytic domain of human PPIP5K2 (PPIP5K2^{KD}).¹³ To increase the throughput of these experiments, we took advantage of the “reversibility” of the kinases that synthesise PP-InsPs.¹⁴ We used luciferin/luciferase to assay the ATP that was formed when PPIP5K2^{KD} was incubated with ADP and 1,5-[PP]₂-InsP₄. Analogues **1**, **2** and **3** inhibited ATP formation (Fig. 2) indicating that they were accommodated within the enzyme’s active site; 5-PA-InsP₅ (**2**) showed the strongest inhibition, while 5-PA-InsP₄ (**1**) was a ten-fold weaker inhibitor. This is consistent with the finding that 5-PP-InsP₄ is a poor

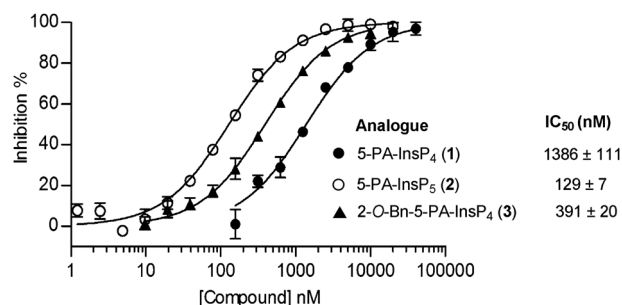


Fig. 2 Inhibition of human PPIP5K2 by analogues **1**, **2** and **3** in the presence of 100 nM 1,5-[PP]₂-InsP₄.

substrate for PPIP5K2.¹³ Unexpectedly, the 2-O-benzyl analogue **3** was only 3-fold less potent than 5-PA-InsP₅ (**2**), suggesting that an axial 2-O-benzyl group can partly compensate for the loss of the axial 2-phosphate. This may have implications for inhibitor design.

We obtained an X-ray structure of the most potent inhibitor, 5-PA-InsP₅ (**2**) in complex with PPIP5K2^{KD} (Fig. 3). Superposition of this structure with that of PPIP5K2^{KD} in complex with 5-PP-InsP₅ (PDB accession code 3T9D), yielded RMSDs (root mean square deviations) of 0.13 Å for the C α atoms of the residues in the protein (Fig. 3a). A simulated-annealing omit map shows that 5-PA-InsP₅ binds at the same site as 5-PP-InsP₅ with a similar orientation (Fig. 3a) although there were some conformational changes of the phosphate groups on the side of inositol carbon positions C-4 to C-6. This reflects the substitution of a PA group for the diphosphate group at C-5; as a result of accommodating 5-PA-InsP₅ in the active site rather than 5-PP-InsP₅ a magnesium atom is lost and the side chains of residues Lys214, Arg262, Arg273 and Arg281 slightly adjust their orientations (Fig. 3a and b). On the other hand, the position of the 1-phosphate of 5-PA-InsP₅ relative to the γ -phosphorus atom of AMPPNP and the key catalytic residue Lys248 is very similar to the 1-phosphate of 5PP-InsP₅ in complex with PPIP5K2 [ref. 13 and Fig. 3]. Indeed, by using a gel electrophoresis method,¹⁵ phosphorylation of 5-PA-InsP₅ could be detected (Fig. 3d) although, due to limitations of sensitivity, an accurate comparison of reaction rates with natural substrate, InsP₆ (Fig. 1a), was not possible.

In conclusion, PA esters **1**, **2** and **3** may be regarded as first-generation synthetic mimics of PP-InsPs. Proof-of-principle that PA substitution need not prevent target recognition was obtained from studies using PPIP5K2. By using other protecting group arrangements, the synthetic methods can be extended to give other PA-InsPs including [PA]_n-InsPs with multiple PA esters, e.g. as mimics of [PP]₂-InsP₄ isomers. By scaling up the enzymatic phosphorylation of 5-PA-InsP₅ (Fig. 3d), it should be possible to prepare sufficient quantities of 1-PP-5-PA-InsP₄ to dissect the separate roles of the two diphosphate groups in 1,5-[PP]₂-InsP₄. Stabilised PP-InsP mimics such as PA-InsPs could help to identify effects of PP-InsPs that do not involve protein dephosphorylation. They also have the potential for further synthetic elaboration, to provide new probes for the study of InsP and PP-InsP signalling.

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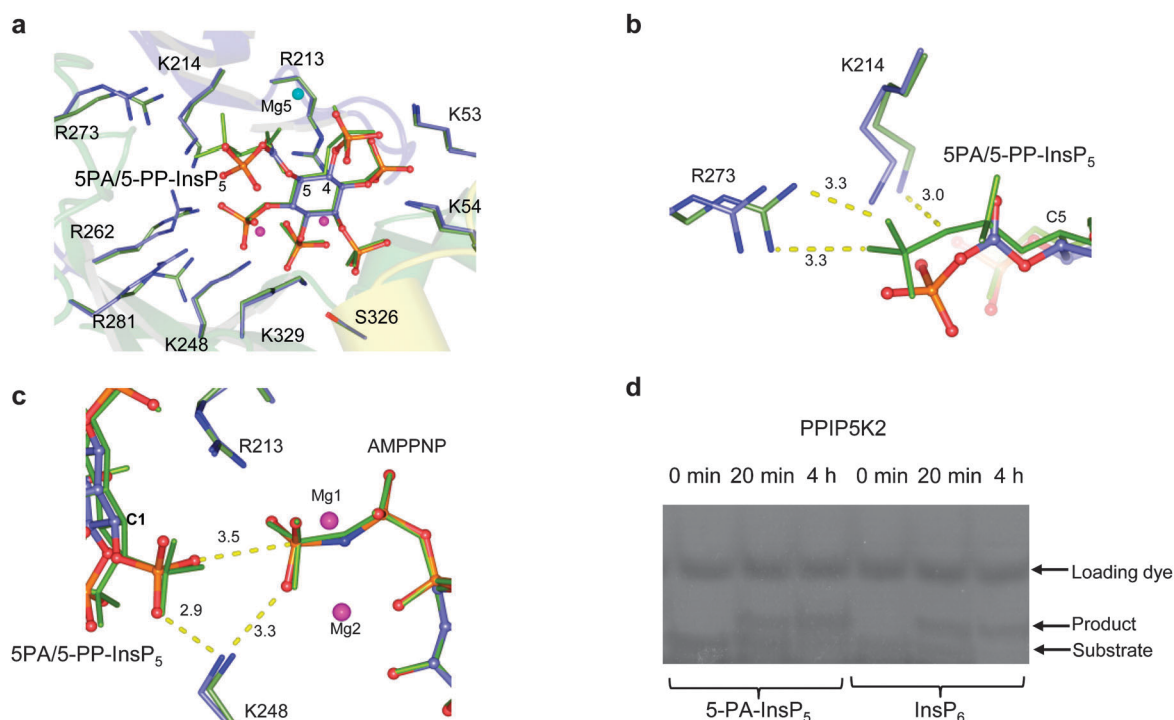


Fig. 3 Binding of 5-PA-InsP₅ (**2**) to human PPIP5K2^{KD} (a–c) and phosphorylation of **2** by PPIP5K2^{KD} (d). (a) 5-PA-InsP₅ (**2**, ball-and-stick model) and its interacting residues (stick model) for hPPIP5K2 are shown in light blue for carbon, blue for nitrogen, orange and red for phosphate groups. Residues that make contacts with 5-PP-InsP₅ (green stick) are also shown in green carbon. Magnesium atoms are shown as spheres. C-4 and C-5 in the inositol ring are labelled. (b) Residues that interact with PA or PP are shown. Bond distances (dashed lines) are denoted in Å. (c) Key catalytic residues are shown in stick model. Inositol phosphates and nucleotide cofactors are shown in ball-and-stick model. Colours are as (a). Also shown are catalytic magnesium atoms (magenta spheres). C-1 of the inositol ring is labelled. Bond distances (dashed lines) are denoted in Å. (d) Phosphorylation of 5-PA-InsP₅ by PPIP5K2^{KD}.

Team (SER-CAT) 22-beamline at the Advanced Photon Source, Argonne National Laboratory. Supporting institutions may be found at www.ser-cat.org/members.html. Use of the Advanced Photon Source was supported by the U. S. Department of Energy, Office of Science, Office of Basic Energy Sciences, under Contract No. W-31-109-Eng-38.

Notes and references

- A. J. Hatch and J. D. York, *Cell*, 2010, **143**, 1030–1030.e1.
- M. D. Best, H. Zhang and G. D. Prestwich, *Nat. Prod. Rep.*, 2010, **27**, 1403–1430.
- G. N. Europe-Finner, B. Gammon and P. C. Newell, *Biochem. Biophys. Res. Commun.*, 1991, **81**, 191–196; F. S. Menniti, R. N. Miller, J. W. Putney Jr. and S. B. Shears, *J. Biol. Chem.*, 1993, **268**, 3850–3856; L. Stephens, T. Radenberg, U. Thiel, G. Vogel, K. H. Khoo, A. Dell, T. R. Jackson, P. T. Hawkins and G. W. Mayr, *J. Biol. Chem.*, 1993, **268**, 4009–4015.
- For access to the literature, see; A. Saiardi, in: *Phosphoinositides II: The Diverse Biological Functions: Subcellular Biochemistry 59*, ed. T. Balla, M. Wymann and J. D. York, Springer, Dordrecht, Heidelberg, London, New York, 2012, ch. 14, pp. 413–443.
- Y. S. Lee, K. Huang, F. A. Quirocho and E. K. O'Shea, *Nat. Chem. Biol.*, 2008, **4**, 25–32.
- R. Bhandari, A. Saiardi, Y. Ahmadibeni, A. M. Snowman, A. C. Resnick, T. Z. Kristiansen, H. Molina, A. Pandey, J. K. Werner Jr, K. R. Juluri, Y. Xu, G. D. Prestwich, K. Parang and S. H. Snyder, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 15305–15310.
- J. R. Falck, K. K. Reddy, J. Ye, M. Saady, C. Mioskowski, S. B. Shears, Z. Tan and S. Safrany, *J. Am. Chem. Soc.*, 1995, **117**, 12172–12175; K. M. Reddy, K. K. Reddy and J. R. Falck, *Tetrahedron Lett.*, 1997, **38**, 4951–4952; C. Albert, S. T. Safrany, M. E. Bembenek, K. M. Reddy, K. K. Reddy, J. R. Falck, M. Bröcker, S. B. Shears and G. W. Mayr, *Biochem. J.*, 1997, **327**, 553–560; H. Zhang, J. Thompson and G. D. Prestwich, *Org. Lett.*, 2009, **11**, 1551–1554.
- T. S. Elliott, A. Slowey, Y. Ye and S. J. Conway, *Med. Chem. Commun.*, 2012, **3**, 735–751.
- R. W. Lambert, J. A. Martin, G. J. Thomas, I. B. Duncan, M. J. Hall and E. P. Heimer, *J. Med. Chem.*, 1989, **32**, 367–374; S. Vincent, S. Grenier, A. Valleix, C. Salesse, L. Lebeau and C. Mioskowski, *J. Org. Chem.*, 1998, **63**, 7244–7257.
- J. P. Quinn, A. N. Kulakova, N. A. Cooley and J. W. McGrath, *Environ. Microbiol.*, 2007, **9**, 2392–2400.
- M. Falasca, D. Chiozzotto, H. Y. Godage, M. Mazzeletti, A. M. Riley, S. Previdi, B. V. L. Potter, M. Brogginini and T. Maffucci, *Br. J. Cancer*, 2010, **102**, 104–114.
- J. L. Montchamp, F. Tian, M. E. Hart and J. W. Frost, *J. Org. Chem.*, 1996, **61**, 3897–3899; A. M. Riley, D. J. Jenkins and B. V. L. Potter, *Carbohydr. Res.*, 1998, **314**, 277–281; D. K. Baeschlin, A. R. Chaperon, L. G. Green, M. G. Hahn, S. J. Ince and S. V. Ley, *Chem.-Eur. J.*, 2000, **6**, 172–186; For an optimised method, see ESI†.
- H. Wang, J. R. Falck, T. M. Hall and S. B. Shears, *Nat. Chem. Biol.*, 2012, **8**, 111–116.
- S. M. Voglmaier, M. E. Bembenek, A. I. Kaplin, G. Dorman, J. D. Olszewski, G. D. Prestwich and S. H. Snyder, *Proc. Natl. Acad. Sci. U. S. A.*, 1996, **93**, 4305–4310.
- O. Losito, Z. Sziygyarto, A. C. Resnick and A. Saiardi, *PLoS One*, 2009, **4**, e5580.