



Current perspectives on invasive nontyphoidal *Salmonella* disease

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Purpose of review

We searched PubMed for scientific literature published in the past 2 years for relevant information regarding the burden of invasive nontyphoidal *Salmonella* disease and host factors associated with nontyphoidal *Salmonella* infection and discuss current knowledge on vaccine development. The following search terms were used: *Salmonella*, non typhoidal/nontyphoidal, NTS, disease, bloodstream infection, invasive, sepsis/septicaemia/septicemia, bacteraemia/bacteremia, gastroenteritis, incidence, prevalence, morbidity, mortality, case fatality, host/risk factor, vaccination, and prevention/control.

Recent findings

Estimates of the global invasive nontyphoidal *Salmonella* disease burden have been recently updated; additional data from Africa, Asia, and Latin America are now available. New data bridge various knowledge gaps, particularly with respect to host risk factors and the geographical distribution of iNTS serovars. It has also been observed that *Salmonella Typhimurium* sequence type 313 is emergent in several African countries. Available data suggest that genetic variation in the sequence type 313 strain has led to increased pathogenicity and human host adaptation. A bivalent efficacious vaccine, targeting *Salmonella* serovars *Typhimurium* and *Enteritidis*, would significantly lower the disease burden in high-risk populations.

Summary

The mobilization of surveillance networks, especially in Asia and Latin America, may provide missing data regarding the invasive nontyphoidal *Salmonella* disease burden and their corresponding antimicrobial susceptibility profiles. Efforts and resources should be directed toward invasive nontyphoidal *Salmonella* disease vaccine development.

Keywords

epidemiology, invasive nontyphoidal *Salmonella* disease, invasive, nontyphoidal *Salmonella*, vaccination

INTRODUCTION

Salmonella enterica serovar *Typhi* (*S. Typhi*) and the various pathovars of *S. Paratyphi* are commonly referred to as typhoidal *Salmonella* serovars. These agents are restricted to human hosts. *Salmonella* serovars that fall outside of this group are typically referred to as the nontyphoidal *Salmonella* (NTS) serovars and are considered to have the potential to interact with human and nonhuman hosts [1[¶]]. Poor access to improved water supplies and adequate sanitation facilities, combined with growing urbanization, favor the transmission of NTS serovars through food or water sources and contact with animals [2[¶]]. In addition to animal reservoirs, humans may be a growing substantial secondary pathogen reservoir [3]. Typical NTS disease in immunocompetent hosts manifests as a mild, self-limiting gastroenteritis. In contrast, invasive nontyphoidal *Salmonella* (iNTS) disease commonly presents as a febrile bacteremia, which can be fatal if left

untreated. Invasive NTS disease is associated with the extremes of age, malnutrition, clinical malaria, and HIV infections, especially in Africa [4[¶], 5[¶]–7[¶]]. In this study, we review available literature published

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KEY POINTS

- Global estimates suggest 3.4 million iNTS illnesses and 618 316 iNTS disease-related deaths per year.
- The most common iNTS serovars are *S. Enteritidis*, *S. Dublin*, *S. Typhimurium*; of particular concern is the *S. Typhimurium* ST313 variant.
- Bivalent *S. Enteritidis* and *S. Typhimurium* vaccines may decrease the global disease burden dramatically.

from 2015 to present on the global burden of iNTS disease, host risk factors, and the implications of these data for vaccination.

GLOBAL DISEASE BURDEN

A recent review on the global distribution of iNTS has indicated a low number of reported cases in South and Southeast Asia [8[■]]. The overwhelming majority of the estimated 3.4 million annual iNTS infections and 618 316 iNTS-related deaths occur in Africa [4[■],8[■],9]. *Salmonella enterica* serovars *Enteritidis*, *Dublin*, and *Typhimurium* are the most common serovars associated with iNTS disease [4[■],5[■]]. Of specific concern is *S. Typhimurium* sequence type 313 (ST313), which is more frequently associated with bacteremia than gastrointestinal infections [10,11], and is commonly multidrug resistant (MDR) [2[■],12]. Additional data suggest that *S. Typhimurium* ST313 may have a greater propensity for transmission between humans, and an animal reservoir has not, as yet, been well defined. Keddy *et al.* [13] recently found a significant association of an MDR phenotype in ST313 [odds ratio (OR) 6.6; 95% confidence interval (95%CI) 2.5–17.2] in comparison to *S. Typhimurium* sequence type 19 (ST19). ST19 arises mostly in Europe and Northern America, whereas ST313 isolates are more commonly found in Africa [14[■]].

A review published in 2017 assessed the occurrence of iNTS disease across Africa [5[■]]. The authors described disease incidence estimates that ranged from 1.4 per 100 000 population/year (all ages, South Africa, 2003–2004) to 2520 per 100 000 population/year (children <5 years old, Ghana, 2007–2009) [5[■]], with highest incidences in those infected with HIV, in patients with sickle cell disease, in young children, and in those residing in rural settings [5[■]]. The prevalence of NTS-related community-acquired bacteremia ranged from 8% in Nigeria and South Africa to 45% in the Central African Republic [5[■]], with an overall case fatality rate of 20.6% (548 deaths/2656 iNTS disease cases) [5[■]].

The emergence of iNTS organisms in Africa exhibiting resistance to various commonly used

antimicrobials, including chloramphenicol, ampicillin, and co-trimoxazole, has been reported [2[■],4[■],15,16]. These ‘baseline’ antimicrobial resistance profiles have been followed by the advent of resistance against third-generation cephalosporins; iNTS isolates with resistance to ceftriaxone has now been reported in the Democratic Republic of the Congo (DRC) [17,18], Kenya [19,20], Malawi [15,21] and South Africa [2[■]].

The Typhoid Fever Surveillance in Africa Program (TSAP), a population-based surveillance, conducted at 13 sentinel sites in 10 countries (Burkina Faso, Ethiopia, Ghana, Guinea-Bissau, Kenya, Madagascar, Senegal, South Africa, Sudan, and Tanzania) during 2010–2014, revealed an overall iNTS disease prevalence of 17% (94/568) among those with bacteremia [22]. The serovars *S. Typhimurium* (40%, 38 out of 94 NTS positive cases), *S. Enteritidis* (12%, 11 out of 94 NTS positive cases) and *S. Dublin* (11%, 10/94 NTS positive cases) were the most prevalent [22–24], which is largely concordant with findings reported by Crump and Heyderman [4[■]] and Uche *et al.* [5[■]]. In TSAP, the adjusted incidences of iNTS disease were highest among children aged more than 1 year, ranging from 291 (95%CI 176–482) per 100 000 person-years-of-observations (PYO) (Guinea-Bissau) to 1733 (95%CI 1373–2188) per 100 000 PYO (Ghana) [22]. The iNTS disease incidences among children aged 2 to 4 years ranged from 49 (95%CI 7–348) per 100 000 PYO in Kenya to 1908 (95%CI 1469–2479) per 100 000 PYO in Ghana [22]. In addition, several independent reports on blood-culture-based surveillance data have shown that iNTS disease is present in other locations in Africa such as the DRC [18], the Gambia [25], and Ghana [26,27]. Data from different sites in Kenya found an incidence of 4134 per 100 000 person-years [20] and 174 per 100 000 person-years [28] in infants. Children under 5 years of age had an overall incidence of 36.6 per 100 000 person-years [28,29]; incidences among children less than 5 years of age differed considerably by setting (rural setting: 3914 per 100 000 person-years, urban setting: 997.9 per 100 000 person-years) [30]. The presence of iNTS disease has also been reported from Mali [31], Mozambique where two studies were conducted [predominantly ST313 isolates [32]; infant incidence: 217.7 per 100 000 child-years [33[■]], and South Africa [34[■]].

In contrast to Africa, the epidemiology of iNTS disease and corresponding antimicrobial susceptibility patterns are poorly described in Asia and South America, suggesting either a lower disease burden or a lack of epidemiological reporting. A multicenter, hospital-based study investigating community-acquired bacteremia in Indonesia, Thailand, and

Vietnam from 2013 to 2015 identified an overall NTS-associated bacterial positivity rates of 27.5% (11/40 bacteremia cases) in children and 11.7% (7/60 bacteremia cases) in adults [35]. Limited iNTS prevalence (20/12 940 bacteremia patients) and a 25% case fatality were reported among bacteremic patients hospitalized from 2009 to 2013 in Bangladesh [36]. A longitudinal study of community-acquired bacteremia in hospitalized children conducted in Malaysia from 2001 to 2011 found an iNTS prevalence of 16.2% (36/222), with most NTS isolated from infants below 1 year of age [37]. A surveillance study from Colombia investigated a sample of 4010 *S. Enterica* isolates collected from blood and feces samples and found that 32.5% were *S. Typhimurium*, 28.2% were *S. Enteritidis*, and 2.9% were *S. Dublin* cases over a 6-year period [38]. These numbers are considerably lower than those reported from Asia and sub-Saharan Africa. Notably, *S. Typhimurium* ST313 variants have been isolated from humans and poultry in Brazil [39]. On the basis of the investigations of Almeida *et al.* [39], the organisms identified appear genetically distinct from the ST313 variants isolated in sub-Saharan Africa.

HOST-ASSOCIATED FACTORS

Common factors contributing to iNTS disease include extremes in age, the occurrence of immunosuppressive conditions, and other underlying comorbidities (e.g., diabetes, cancer, and cardiovascular diseases) [40]. In addition, climatic conditions such as increased rainfall or drought that can result in food scarcity, leading to malnutrition and increased transmission of malaria parasites are factors that may favor the transmission of NTS organisms [7[■]]. Particularly in Africa, the association of iNTS disease with malnutrition (OR 1.44–2.42) and sickle cell disease (OR 35.6) has been described predominantly in children, whereas *Plasmodium falciparum* malaria (OR 1.5–4.1), anemia, and HIV infection (OR 3.2–48.2) are risk factors that are not generally associated with age [2[■],4[■],5[■],7[■],24,29,41[■]]. Adjusted odds ratios of 4.0 and 5.0 were calculated for the association of iNTS disease with moderate and severe anemia, respectively [33[■]]. Keddy *et al.* [34[■]] found a significant association between an increased usage of antiretroviral therapy and a decrease in incident iNTS disease infections ($P < 0.001$) in a South African province. Similar observations were made by Lan *et al.* [42] in Vietnam. ST313, the most common *S. Typhimurium* variant associated with iNTS disease, was initially identified in HIV-infected patients [2[■]]. In comparison to other *S. Typhimurium* types (e.g., ST19), the genomically degraded ST313 may cause systemic infections and induce a lower inflammatory reaction in the

intestine, exerted by evasion mechanisms from the immune response [43,44]. The genomic degradation includes the downregulation of gene expression involved in active cell invasion through effector proteins [43,44]. The reduced activation of macrophages is assumed to be caused by lower flagellin expression [43,44]. The survival time and replication rate were found to be more efficient in the investigated ST313 isolates compared with ST19 [43,44]. Therefore, the ST313 phenotype appears to become closer to that of typhoidal *Salmonella*, suggesting analogical adaptation toward a more invasive phenotype in humans [1[■]].

In addition, an MDR phenotype may allow for rapid ST313 dissemination throughout susceptible populations [2[■]]. Advanced HIV disease leads to a reduced immune response in the gastrointestinal mucosa and poses a higher invasion risk of iNTS [7[■]]. Changes in the gastrointestinal microbiota, induced by the intake of acid blockers, gastric surgery, and antimicrobial pretreatment, are also suggested to favor iNTS disease [7[■],45,46]. Martz *et al.* [46] found stabilizing effects on the gastrointestinal microbiome associated with the ingestion of probiotics in mice, which may improve the functionality of the intestinal barrier.

VACCINE DEVELOPMENT

Effective vaccines preventing iNTS disease are likely to differ inherently from those protecting against *S. Typhi* infections. Studies from Africa have shown that naturally acquired antibodies against NTS correspond with a reduced risk of iNTS disease [47,48]. Several vaccine candidates targeting *S. Typhimurium* and *S. Dublin* are currently under development, some of which may provide protection against both serovars. The current status of iNTS vaccine considerations has been described in a recent review [49[■]]: several potential iNTS vaccines are under development, including live-attenuated, subunit-based, and recombinant antigen-based substances. Both humoral and cellular immunities are likely required to achieve full protection against iNTS disease. Live-attenuated vaccines provide both types of immune response; however, they may pose a risk for immunocompromised individuals [41[■]]. Inactivated iNTS vaccines may induce humoral immunity only and suppress NTS during the acute phase of infection, but are likely not to achieve systematic clearance in infected individuals [41[■]]. The lack of disease burden data from Asia and South America, coupled with the enormous number of NTS serovars, and the role of alternate prevention measures (i.e., access to improved water and sanitation and food safety) have contributed to a delay and a lack of investment in the development of iNTS disease vaccines.

Previously, when animals were considered to be the only reservoir of NTS organisms, the implementation of hygiene and safety measures along a regulated and appropriate food chain was thought to be sufficient for the reduction of iNTS transmission. However, with the speculation that humans may be a growing alternative reservoir for ST313 [50], the development and deployment of iNTS disease vaccines appear to be a more viable solution. However, iNTS disease vaccines would not only require considerable funding to progress existing vaccine candidates, but also will require parallel vaccine deployment strategies to identify appropriate target age groups, schedules, formulations, and potential vaccine adjuvants.

An iNTS disease vaccine would need to be administered in infants to ensure protection before the peak occurrence of disease. This strategy, however, poses the challenge of combining iNTS and *S. typhi* vaccines, as the peak disease incidence for typhoid (5–8 years of age) is later than that for iNTS disease [51], except in highly endemic areas. A potential byproduct after the widespread use of a bivalent iNTS vaccine conferring protection against *S. Typhimurium* and *S. Enteritidis* would be serovar replacement by other *Salmonella* variants, such as *S. Dublin*. Such a serotype replacement was observed following the MenAfriVac campaign, when large populations in the African meningitis belt were vaccinated against *Neisseria meningitidis* serotype A and other serotypes subsequently emerged [52–54]. Another consideration would be a combined iNTS disease/malaria vaccine; this approach may be particularly prudent, given that malaria is associated with the severity of iNTS disease. Such a vaccine would then be tailor-made for sub-Saharan Africa, but may be less applicable for low and nonendemic malaria regions (i.e., Brazil) [39]. This would potentially suggest the need to develop an independent, nonmalaria-combined vaccine that is applicable to all iNTS endemic regions.

CONCLUSION

iNTS is a major public health issue in sub-Saharan Africa. ST313 appears to be better adapted to humans than other *S. Typhimurium*, is associated with an increased disease severity, and has acquired an MDR phenotype. Observations of the Brazilian ST313 lead to some insights on this serotype that are also relevant for Africa. First, ST313 has the ability to arise in new locations independently and does not appear to be confined to sub-Saharan Africa. The fact that some Brazilian ST313 isolates exhibit different antimicrobial susceptibility profiles in comparison to African variants suggests that iNTS disease has the potential to evolve *de novo* outside of Africa, which

may result in new and unlinked epidemics. Improvement in water and sanitation, a reduction in malaria incidences and malnutrition and improved management of HIV infections should additionally prevent iNTS disease from becoming an even bigger global health threat. However, the rapid emergence of ST313, the possible de-novo occurrence and spread of the future MDR and pathogenic variants place iNTS disease increasingly on the vaccine development agenda. Now is also a prime time to invest in enhanced iNTS disease surveillance. This enhanced surveillance is particularly important in Asia and Latin America, and is required to assess the actual extent of disease in these locations. Typhoid fever should be used as an example, when, despite the availability of vaccines, a lack of appropriate disease burden data stalled the global commitment, resulting in limited vaccine uptake and dampened efforts to develop conjugated vaccines. The persistence of typhoid fever culminated in the evolution of a highly antimicrobial resistant *S. Typhi* genotype (H58), which is spreading globally [55].

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Conflicts of interest

The sponsors had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

The findings and conclusions contained within are our own and do not necessarily reflect positions or policies of the IVI.

The authors declare no conflicts of interest.

REFERENCES AND RECOMMENDED READING

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
- of outstanding interest

1. Singletary LA, Karlinsey JE, Libby SJ, *et al.* Loss of multicellular behavior in epidemic African nontyphoidal *Salmonella enterica* serovar typhimurium ST313 Strain D23580. *MBio* 2016; 7:02265–02315.

This article provides an update on the adapted selection advantages of the ST313 genome.

2. Mahon BE, Fields PI. Invasive infections with nontyphoidal *Salmonella* in Sub-Saharan Africa. *Microbiol Spectr* 2016; 4:0015–2016.
- Authors deepen the evidence on risk factors, including antimicrobial resistance patterns, and discuss vaccine candidates.
3. Wikswo ME, Kambhampati A, Shioda K, *et al*. Outbreaks of acute gastroenteritis transmitted by person-to-person contact, environmental contamination, and unknown modes of transmission—United States. *MMWR Surveill Summ* 2015; 64:1–16.
4. Crump JA, Heyderman RS. A perspective on invasive *Salmonella* disease in Africa. *Clin Infect Dis* 2015; 1:S235–S240.
- Authors highlight the disease burden, antimicrobial resistance patterns, and transmission modes of NTS.
5. Uche IV, MacLennan CA, Saul A. A systematic review of the incidence, risk factors and case fatality rates of invasive nontyphoidal *Salmonella* (iNTS) disease in Africa (1966 to 2014). *PLoS Negl Trop Dis* 2017; 11:e0005118.
- A comprehensive overview on the iNTS disease incidence as well as risk factors and mortality across African countries.
6. Okoro CK, Barquist L, Connor TR, *et al*. Signatures of adaptation in human invasive *Salmonella typhimurium* ST313 populations from sub-Saharan Africa. *PLoS Negl Trop Dis* 2015; 9:e0003970.
- The authors provide comprehensive phenotypic as well as genotypic data on ST313.
7. Crump JA, Sjolund-Karlsson M, Gordon MA, Parry CM. Epidemiology, clinical presentation, laboratory diagnosis, antimicrobial resistance, and antimicrobial management of invasive *Salmonella* infections. *Clin Microbiol Rev* 2015; 28:901–937.
- Authors presented a broad and in-depth overview on the clinical picture as well as host-related risk and protective effects on iNTS disease.
8. Gilchrist JJ, MacLennan CA, Hill AV. Genetic susceptibility to invasive *Salmonella* disease. *Nat Rev Immunol* 2015; 15:452–563.
- The publication provides a comprehensive update on the global iNTS disease distribution
9. Ao TT, Feasey NA, Gordon MA, *et al*. Global burden of invasive nontyphoidal *Salmonella* disease, 2010(1). *Emerg Infect Dis* 2015; 21.
10. Graham SM, English M. Nontyphoidal *Salmonellae*: a management challenge for children with community-acquired invasive disease in tropical African countries. *Lancet* 2009; 373:267–269.
11. Hawkey PM, Livermore DM. Carbapenem antibiotics for serious infections. *BMJ* 2012; 344:e3236.
12. Kingsley RA, Msefula CL, Thomson NR, *et al*. Epidemic multiple drug resistant *Salmonella typhimurium* causing invasive disease in sub-Saharan Africa have a distinct genotype. *Genome Res* 2009; 19:2279–2287.
13. Keddy KH, Sooka A, Musekiwa A, *et al*. Clinical and microbiological features of *Salmonella* meningitis in a South African Population, 2003–2013. *Clin Infect Dis* 2015; 1:S272–S282.
14. Lokken KL, Walker GT, Tsolis RM. Disseminated infections with antibiotic-resistant nontyphoidal *Salmonella* strains: contributions of host and pathogen factors. *Pathog Dis* 2016; 74; ftw103.
- This article describes the differences and similarities of the ST313 and ST19 phenotypes in detail.
15. Gordon MA, Graham SM, Walsh AL, *et al*. Epidemics of invasive *Salmonella enterica* serovar enteritidis and *S. enterica* serovar typhimurium infection associated with multidrug resistance among adults and children in Malawi. *Clin Infect Dis* 2008; 46:963–969.
16. Kariuki S, Revathi G, Kariuki N, *et al*. Characterisation of community acquired nontyphoidal *Salmonella* from bacteraemia and diarrhoeal infections in children admitted to hospital in Nairobi, Kenya. *BMC Microbiol* 2006; 6:101.
17. Lunguya O, Lejon V, Phoba MF, *et al*. Antimicrobial resistance in invasive nontyphoid *Salmonella* from the Democratic Republic of the Congo: emergence of decreased fluoroquinolone susceptibility and extended-spectrum beta lactamases. *PLoS Negl Trop Dis* 2013; 7:14.
18. Kalonji LM, Post A, Phoba MF, *et al*. Invasive *Salmonella* infections at multiple surveillance sites in the Democratic Republic of the Congo, 2011–2014. *Clin Infect Dis* 2015; 1:S346–S353.
19. Kariuki S, Gordon MA, Feasey N, Parry CM. Antimicrobial resistance and management of invasive *Salmonella* disease. *Vaccine* 2015; 19:23.
20. Onoko M, Kariuki S, Muturi-Kioi V, *et al*. Emergence of community-acquired, multidrug-resistant invasive nontyphoidal *Salmonella* disease in rural Western Kenya, 2009–2013. *Clin Infect Dis* 2015; 1:S310–S316.
21. Msefula CL, Kingsley RA, Gordon MA, *et al*. Genotypic homogeneity of multidrug resistant *S. Typhimurium* infecting distinct adult and childhood susceptibility groups in Blantyre, Malawi. *PLoS One* 2012; 7:27.
22. Marks F, von Kalkreuth V, Aaby P, *et al*. Incidence of invasive *Salmonella* disease in sub-Saharan Africa: a multicentre population-based surveillance study. *Lancet Glob Health* 2017; 5:e310–e323.
23. Al-Emran HM, Krumkamp R, Dekker DM, *et al*. Validation and identification of invasive *Salmonella* serotypes in Sub-Saharan Africa by multiplex polymerase chain reaction. *Clin Infect Dis* 2016; 15:S80–S82.
24. Park SE, Pak GD, Aaby P, *et al*. The relationship between invasive nontyphoidal *Salmonella* disease, other bacterial bloodstream infections, and malaria in Sub-Saharan Africa. *Clin Infect Dis* 2016; 15:S23–S31.
25. Kwambana-Adams B, Darboe S, Nabwera H, *et al*. *Salmonella* infections in the Gambia, 2005–2015. *Clin Infect Dis* 2015; 1:S354–S362.
26. Eibach D, Al-Emran HM, Dekker DM, *et al*. The emergence of reduced ciprofloxacin susceptibility in *Salmonella enterica* causing bloodstream infections in rural Ghana. *Clin Infect Dis* 2016; 15:S32–S36.
27. Andoh LA, Ahmed S, Olsen JE, *et al*. Prevalence and characterization of *Salmonella* among humans in Ghana. *Trop Med Health* 2017; 45:3.
28. Muthumbi E, Morpeth SC, Ooko M, *et al*. Invasive salmonellosis in Kilifi, Kenya. *Clin Infect Dis* 2015; 1:S290–S301.
29. Kariuki S, Onsare RS. Epidemiology and genomics of invasive nontyphoidal *Salmonella* infections in Kenya. *Clin Infect Dis* 2015; 1:S317–S324.
30. Verani JR, Toroitich S, Auko J, *et al*. Burden of invasive nontyphoidal *Salmonella* disease in a rural and urban site in Kenya, 2009–2014. *Clin Infect Dis* 2015; 61(Suppl 4):S302–S309.
31. Tapia MD, Tennant SM, Bornstein K, *et al*. Invasive nontyphoidal *Salmonella* infections among children in Mali, 2002–2014: microbiological and epidemiologic features guide vaccine development. *Clin Infect Dis* 2015; 1:S332–S338.
32. Moon TD, Johnson M, Foster MA, *et al*. Identification of invasive *Salmonella enterica* serovar typhimurium ST313 in ambulatory HIV-infected adults in Mozambique. *J Glob Infect Dis* 2015; 7:139–142.
33. Mandomando I, Bassat Q, Sigauque B, *et al*. Invasive *Salmonella* infections among children from rural Mozambique, 2001–2014. *Clin Infect Dis* 2015; 1:S339–S345.
- This study provides evidence on causal relationships between iNTS diseased and their host factors.
34. Keddy KH, Takuva S, Musekiwa A, *et al*. An association between decreasing incidence of invasive nontyphoidal salmonellosis and increased use of antiretroviral therapy, Gauteng Province, South Africa, 2003–2013. *PLoS One* 2017; 12:e0173091.
- Most recent findings which suggest that antiretroviral therapy may be an important factor on the occurrence of incident NTS infections.
35. SAIDCR Network. Causes and outcomes of sepsis in southeast Asia: a multinational multicentre cross-sectional study. *Lancet Glob Health* 2017; 5:e157–e167.
36. Shahunja KM, Leung DT, Ahmed T, *et al*. Factors associated with nontyphoidal *Salmonella* bacteremia versus typhoidal *Salmonella* bacteremia in patients presenting for care in an urban diarrheal disease hospital in Bangladesh. *PLoS Negl Trop Dis* 2015; 9:e0004066.
37. Nor Azizah A, Fadzilah MN, Mariam M, *et al*. Community-acquired bacteremia in paediatrics: epidemiology, aetiology and patterns of antimicrobial resistance in a tertiary care centre, Malaysia. *Med J Malaysia* 2016; 71:117–121.
38. Rodriguez EC, Diaz-Guevara P, Moreno J, *et al*. Laboratory surveillance of *Salmonella enterica* from human clinical cases in Colombia 2005–2011. *Enferm Infecc Microbiol Clin* 2016; 30:30008–30018.
39. Almeida F, Seribelli AA, da Silva P, *et al*. Multilocus sequence typing of *Salmonella typhimurium* reveals the presence of the highly invasive ST313 in Brazil. *Infect Genet Evol* 2017; 51:41–44.
40. Turgeon P, Murray R, Nesbitt A. Hospitalizations associated with salmonellosis among seniors in Canada, 2000–2010. *Epidemiol Infect* 2017; 145: 1527–1534.
41. Mastroeni P, Rossi O. Immunology, epidemiology and mathematical modelling towards a better understanding of invasive nontyphoidal *Salmonella* disease and rational vaccination approaches. *Expert Rev Vaccines* 2016; 15:1545–1555.
- The authors address necessary considerations related to the vaccine development and introduction in different populations.
42. Phu Huong Lan N, Le Thi Phuong T, Nguyen Huu H, *et al*. Invasive nontyphoidal *Salmonella* infections in Asia: clinical observations, disease outcome and dominant serovars from an infectious disease hospital in Vietnam. *PLoS Negl Trop Dis* 2016; 10:e0004857.
43. Carden S, Okoro C, Dougan G, Monack D. Nontyphoidal *Salmonella typhimurium* ST313 isolates that cause bacteremia in humans stimulate less inflammasome activation than ST19 isolates associated with gastroenteritis. *Pathog Dis* 2015; 73:24.
44. Ramachandran G, Perkins DJ, Schmidlein PJ, *et al*. Invasive *Salmonella typhimurium* ST313 with naturally attenuated flagellin elicits reduced inflammation and replicates within macrophages. *PLoS Negl Trop Dis* 2015; 9:e3394.
45. Freeman R, Dabrera G, Lane C, *et al*. Association between use of proton pump inhibitors and nontyphoidal salmonellosis identified following investigation into an outbreak of *Salmonella mikawasima* in the UK, 2013. *Epidemiol Infect* 2016; 144:968–975.
46. Martz SL, McDonald JA, Sun J, *et al*. Administration of defined microbiota is protective in a murine *Salmonella* infection model. *Sci Rep* 2015; 5:16094.
47. Gordon MA. Invasive nontyphoidal *Salmonella* disease: epidemiology, pathogenesis and diagnosis. *Curr Opin Infect Dis* 2011; 24:484–489.
48. MacLennan CA, Gondwe EN, Msefula CL, *et al*. The neglected role of antibody in protection against bacteremia caused by nontyphoidal strains of *Salmonella* in African children. *J Clin Invest* 2008; 118:1553–1562.
49. Tennant SM, MacLennan CA, Simon R, *et al*. Nontyphoidal *Salmonella* disease: current status of vaccine research and development. *Vaccine* 2016; 34:2907–2910.
- This article discusses potential vaccine candidates and immune responses in different vaccine-receiving populations.

50. Kariuki S, Revathi G, Kariuki N, *et al.* Invasive multidrug-resistant nontyphoidal *Salmonella* infections in Africa: zoonotic or anthroponotic transmission? *J Med Microbiol* 2006; 55(Pt 5):585–591.
51. von Kalckreuth V, Konings F, Aaby P, *et al.* The Typhoid Fever Surveillance in Africa Program (TSAP): clinical, diagnostic, and epidemiological methodologies. *Clin Infect Dis* 2016; 62(Suppl 1):S9–S16.
52. Mohammed I, Iliyasu G, Habib A G. Emergence and control of epidemic meningococcal meningitis in sub-Saharan Africa. *Pathog Glob Health* 2017; 111:1–6.
53. Collard JM, Issaka B, Zaneidou M, *et al.* Epidemiological changes in meningococcal meningitis in Niger from 2008 to 2011 and the impact of vaccination. *BMC Infect Dis* 2013; 13:1471–2334.
54. MacNeil JR, Medah I, Koussoube D, *et al.* *Neisseria meningitidis* serogroup W, Burkina Faso, 2012. *Emerg Infect Dis* 2014; 20:394–399.
55. Wong VK, Baker S, Connor TR, *et al.* An extended genotyping framework for *Salmonella enterica* serovar *typhi*, the cause of human typhoid. *Nat Commun* 2016; 7:12827.