

Synthesis of *meta*-substituted arene bioisosteres from [3.1.1]propellane

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Summary Paragraph: Small-ring cage hydrocarbons are common bioisosteres for *para*-substituted benzene rings in drug design¹. The popularity of these structures derives from the superior pharmacokinetic properties they exhibit compared to the parent aromatics, including improved solubility and reduced susceptibility to metabolism^{2,3}. A prime example is the bicyclo[1.1.1]pentane motif, which is mainly synthesised by ring-opening of the inter-bridgehead bond of the strained hydrocarbon [1.1.1]propellane with radicals or anions⁴. In contrast, scaffolds mimicking *meta*-substituted arenes are lacking due to the challenge of synthesising saturated isosteres that accurately reproduce substituent vectors⁵. Here we show that bicyclo[3.1.1]heptanes (BCHepts), hydrocarbons whose bridgehead substituents map precisely onto the geometry of *meta*-substituted benzenes, can be conveniently accessed from [3.1.1]propellane. We found that [3.1.1]propellane can be synthesized on multigram scale, and readily undergoes a range of radical-based transformations to generate medicinally-relevant carbon- and heteroatom-substituted BCHepts, including pharmaceutical

analogues. Comparison of ADME properties of these analogues revealed enhanced metabolic stability relative to their parent arene-containing drugs, validating the potential of this *meta*-arene analogue as an sp^3 -rich motif in drug design. Collectively, our results show that BCHePs can be prepared on useful scales using a variety of methods, offering a novel surrogate for *meta*-substituted benzene rings for implementation in drug discovery programmes.

Main Text: Strategies for the structural modification of lead molecules that improve physicochemical and pharmacokinetic properties such as metabolic stability are increasingly sought in drug development⁶. One example is the replacement of aromatic rings with non-classical bioisosteres such as small-ring cage hydrocarbons^{1,3,5,7}. Such structures display a higher fraction of saturated carbon atoms compared to the parent arenes (F_{sp^3} , corresponding to greater three-dimensionality), a property linked to greater clinical success rates⁸. Amongst these motifs, the replacement of planar *para*-substituted arenes with bicyclo[1.1.1]pentanes (BCPs, Fig. 1a), which have similar dimensions and identical substituent vectors to the parent aromatic, has emerged as a popular strategy^{2,9,10}. For instance, substitution of the fluorinated arene in the Alzheimer's treatment avagacestat with a BCP resulted in an analogue that maintained the bioactivity of the parent compound, but displayed an improved pharmacokinetic profile². More generally, cage hydrocarbons expand the vector space around a molecular core, offering new opportunities in drug design.

Meta-substituted arenes are also commonplace in pharmaceuticals and agrochemicals¹¹. However, in stark contrast to the numerous sp^3 -rich bioisosteres for *ortho*- and *para*-substituted arenes^{2,9,10,12}, a geometrically-accurate bioisostere for *meta*-arenes is yet to be discovered. Recent reports on the use of (hetero)bicyclo[2.1.1]hexanes¹³⁻¹⁷ and bridge-substituted BCPs¹⁸⁻²⁰ have contributed to this arena (Fig. 1b). However, those motifs fail to recreate the bond vectors displayed in the *meta*-substituted aromatic, and a precise and accessible mimic remains absent from the arsenal of the medicinal chemist.

50 Here we report a solution to this challenge in the form of the saturated carbocycle
51 bicyclo[3.1.1]heptane (BCHeP, Fig. 1c), the bridgehead substituent vectors of which precisely
52 replicate those of the parent *meta*-arene ($\sim 119^\circ$ and $\sim 120^\circ$ respectively). While BCHePs have been
53 prepared by ring expansion of BCPs²¹ and by cyclization of cyclohexane dicarboxylates²², these
54 approaches can be limited in substituent scope or involve lengthy synthetic sequences. We show that
55 BCHePs can instead be conveniently and directly accessed from [3.1.1]propellane (**1**), a homologue
56 of [1.1.1]propellane (**2**) which is widely used as the near-ubiquitous source of BCPs⁴. We found **1** to
57 be a versatile precursor that undergoes a variety of radical-based transformations to access a wide
58 range of functionalized BCHePs, including drug analogues. Profiling of the ADME properties of these
59 analogues reveals that, like BCPs, BCHePs significantly improve physicochemical properties
60 compared to their arene parents. As such, we anticipate that this novel scaffold should offer a readily
61 accessible bioisostere for broad implementation in drug discovery programmes.

62 Higher [n.1.1]propellanes such as **1** have to date been of predominantly theoretical interest^{23,24}; these
63 elusive molecules have likely been overlooked in synthetic and medicinal chemistry due to the
64 challenge of their synthesis and reported instability^{25,26}. We devised a strategy to synthesise **1** on
65 multigram scale (Fig. 1d), which began with Kulinkovich cyclopropanation of commercially
66 available γ -chloroester **3**²⁷. Mesylation of the resulting alcohol **4**, followed by TiCl₄-mediated
67 cyclopropyl–allyl chlorinative rearrangement and dibromocyclopropanation, afforded cyclopropane
68 **5** in 58% yield over four steps on >30 mmol scale with only one chromatographic purification.
69 Reaction of **5** with two equivalents of phenyllithium generated **1** in yields of 43–61% after distillation;
70 the resulting solution of **1** (0.25–0.50 M in dibutyl ether) can be stored at -20°C under an inert
71 atmosphere for several months with negligible decomposition.

72 Aside from solvolysis²⁵, previous reports on the chemistry of **1** detail only one productive reaction
73 – the addition of thiophenol to generate BCHeP phenyl thioether²⁶. As a prelude to exploring the
74 wider reactivity of **1**, we first compared the calculated reaction barriers for the addition of a
75 prototypical radical (CH_3^\bullet) and nucleophile (NH_2^-) with those for **2** (Fig. 2a)²⁸. These calculations

predict that **1** should be similarly susceptible to the addition of radicals to the inter-bridgehead C–C bond ($\Delta\Delta G^\ddagger = -1.1$ kcal mol⁻¹ between **1** and **2**), but that the reaction of **1** with anions is less favourable ($\Delta\Delta G^\ddagger = +5.4$ kcal mol⁻¹). This in part may relate to the greater increase of charge density inside the propellane cage in anionic additions, which can be better accommodated by **2** due to the presence of a third three-membered ring that enables enhanced charge delocalization^{28,29}. Our calculations therefore suggest that **1** should be amenable to the same panoply of radical functionalization chemistry established in the [1.1.1]propellane/BCP arena.

This theoretical analysis correlated well with experimental findings. We first explored atom transfer radical addition (ATRA) reactions, which are a powerful method to access disubstituted BCPs from **1**. Both Et₃B-initiated³⁰ and Ir(ppy)₃-catalyzed³¹ addition of a variety of C–I bonds to **1** proceeded efficiently to afford diverse BCHeP scaffolds (Fig. 2b). The photoredox-catalyzed variant (Ir(ppy)₃) proved more general and higher yielding, affording iodo-BCHePs from α -iodocarbonyls (**6a–6d**) benzyl iodides (**6e–6f**), alkyl iodides (**6g–6k**), α -amino acids (**6l**), and heteroaryl iodides (**6m**). In contrast to ATRAs with [1.1.1]propellane³⁰, Et₃B initiation was suitable mainly for electrophilic radicals such as α -iodocarbonyls (**6a**, **6b**) and azetidines (**6h**). Notably, the addition of iodotrifluoromethane to **1** proceeded in the absence of an external initiator to afford **6n**, which could be a valuable building block for the synthesis of bioisosteres of *meta*-CF₃-substituted arenes. Addition to alkyl bromides such as bromomalonate (**6o**, 57%) and bromotrichloromethane (**6p**, 68%) also proved feasible, the latter proceeding without an initiator. The chemistry could further be applied to the late-stage bicycloheptylation of various drug analogues, affording BCHeP derivatives of corticosterone (**6q**), nicotinic acid (**6r**), brequinar (**6s**) and indomethacin (**6t**) which were obtained from the corresponding alkyl iodides. Notably, in contrast to equivalent ATRA reactions with [1.1.1]propellane, no ‘staffane’ byproducts arising from [3.1.1]propellane oligomerization were observed.

100 Bridgehead amine substituents are highly attractive as potential *meta*-substituted aniline bioisosteres.

101 We found that the three-component metallaphotoredox catalyzed coupling of iodonium
102 dicarboxylates, [1.1.1]propellane and *N*-heteroarenes described by the Macmillan group³² translated
103 smoothly to [3.1.1]propellane (Fig. 2c), affording azole- and sulfonamide-substituted BCHePs **7a–7g**
104 in good yields, including pharmaceutical derivatives (gemfibrozil, **7g**). The synthesis of *N*-substituted
105 iodo-BCHePs was achieved using other methods, such as pyrazole BCHeP **7h** by reaction of **1** with
106 pyrazole / I₂³³, and allyl sulfonamide BCHeP **7i** from radical fragmentation of an iodomethyl
107 aziridine³⁴. As well as *C*- and *N*-centered radicals, other heteroatoms proved excellent substrates for
108 reactions with **1** (Fig. 2d): Thioether **8a** and selenoether **8b** were formed in quantitative yields at room
109 temperature, sulfonylthioate addition (**8c**, **8d**) proceeded efficiently under heating^{35,36}, and reaction
110 with a disulfide could be achieved under UV irradiation (**8e**)³⁷. The successful bicycloheptylation of
111 protected cysteine (**8f**) in diethyl ether highlights the potential for applications in peptide
112 modification³⁸; unexpectedly, reaction of a similar cysteine residue in a tripeptide ((L,L,D) δ-(α-
113 amino adipoyl)-Cys-Val, ACV) in aqueous buffer afforded the rearranged adducts **9** and **10** (Fig. 2e),
114 which may arise from a cationic reaction pathway. While the reason for this reactivity difference is
115 unknown, it is clear that selective *S*-alkylation of cysteine is possible under physiologically-relevant
116 conditions.

117 Iodinated BCHePs offer opportunities for C–I functionalization towards medically-relevant
118 difunctionalized scaffolds. Investigation of iron-catalyzed Kumada cross-coupling³⁹ revealed
119 efficient reaction of iodo-BCHePs with both aryl and heteroaryl Grignard reagents to afford
120 (hetero)aryl BCHePs in excellent yields (**11a–11f**, Fig. 3a). BCHeP functionalization was also
121 possible by lithiation of the iodide; reaction of the resulting bridgehead carbanion with CO₂ or *i*-
122 PrOBpin gave carboxylic acid **11g** and hydroxy-BCHeP **11h** (after *in situ* oxidation) respectively, the
123 latter of which corresponds to a *meta*-phenol bioisostere.

124 X-Ray structural determination of several crystalline BCHePs enabled us to study the geometry of
125 the scaffold in more detail (Fig. 3b). Two substituent vector angles were considered: the exit vector

126 angle α ($\sim 120^\circ$ for *m*-arenes), and the out-of-plane vector angle ϕ (the dihedral angle along the
127 BCHeP inter-bridgehead axis, $\sim 0^\circ$ for *m*-arenes). Comparison of the BCHeP solid state structures
128 with computed structures of the BCHeP and the equivalent *meta*-arene showed excellent agreement
129 for both angles ($\Delta\alpha = 0\text{--}7^\circ$, $\Delta\phi = 3\text{--}11^\circ$), validating our hypothesis that the replacement of *meta*-
130 substituted arenes with a BCHeP conserves the critical substituent geometry.

131 Kumada cross-coupling was deployed to synthesise two BCHeP drug analogues (Fig. 4a). The BCHeP
132 analogue of the anticancer agent sonidegib was accessed from coupling product **11e** by pivaloate ester
133 hydrolysis, oxidation of the resulting primary alcohol to the carboxylic acid **13**, and amide formation
134 with aminopyridine **14** (53% yield over three steps, 19% from **1**). BCHeP-URB597, the parent *meta*-
135 arene of which was developed as a fatty acid amide hydrolase inhibitor, was synthesized from **6j** by
136 a similar cross-coupling / hydrolysis / oxidation sequence, followed by amide formation,
137 debenzoylation, and carbamoylation with cyclohexyl isocyanate (16% over 5 steps). Computational
138 conformer sampling once again revealed a similar global topology between BCHeP-sonidegib and
139 the parent drug for the vector angles α and ϕ (Fig. 4b). Here, an additional parameter was considered:
140 the rotational orientation of the planes between the two substituent groups as defined by the dihedral
141 angle ψ ($\Delta\psi = 13^\circ$). The BCHeP displayed a shallow potential energy profile for rotation around the
142 BCHeP–substituent C–C bond ($\Delta G = 1.5 \text{ kcal mol}^{-1}$), reflecting a low conformational preference of
143 the substituents adjacent to the quaternary carbons of the BCHeP, whereas for the parent arene more
144 defined minima exist ($\Delta G = 12 \text{ kcal mol}^{-1}$, see the Supplementary Information for details). This may
145 suggest that BCHePs offer significant flexibility in substituent conformation, which could be a
146 valuable property for drug design by facilitating a more adaptable association with protein targets.

147 Synthesis of these drug analogues raises the question of how the physicochemical and
148 pharmacological properties of the BCHeP compare with the parent arene (Fig. 4c). The cLogP,
149 Topological Polar Surface Area (TPSA) and solubility of each drug / analogue pair are remarkably
150 similar, demonstrating that BCHePs can be readily deployed in drug design as true *meta*-arene

151 bioisosteres. In keeping with their well-established BCP cousins, BCHePs showed reduced clearance
152 rates in mouse and human liver microsomes compared to their arene equivalents, while membrane
153 permeability (Caco-2) was improved. The BCHeP analogues were also tested for CYP inhibition and
154 also generally showed an improvement compared to their corresponding arenes (Fig 4c). URB597
155 inhibits CYP1A2 and CYP2C9 with IC₅₀'s below 10 μ M, but BCHeP-URB597 is 7- and 3-fold
156 weaker against these two polymorphic enzymes. Collectively, these data underline the potential
157 power of the BCHeP scaffold as a beneficial motif for improving the pharmacokinetic and
158 physicochemical properties of drug candidates.

159

160 Bridgehead-disubstituted bicyclo[3.1.1]heptanes (BCHePs) accurately replicate the geometry of
161 *meta*-substituted arenes, and are readily accessed by radical ring opening of [3.1.1]propellane. This
162 unusual member of the propellane family can be prepared on multigram scale, but to date has barely
163 been explored as a BCHeP precursor. A wide variety of radical-based functionalizations demonstrate
164 the potential of this entry to novel bioisosteres; application to the synthesis of pharmaceutical
165 analogues provides a first demonstration of BCHeP installation for use in sp³-rich drug design.

166

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Fig. 1. Comparison of *para*- and *meta*-substituted arene bioisosteres, and synthesis of [3.1.1]propellane. **a**, Bicyclo[1.1.1]pentanes (BCPs) derived from [1.1.1]propellane (**2**) are bioisosteres for *para*-substituted benzene rings. **b**, Previous mimics of *meta*-substituted arenes, which do not accurately reproduce the geometry of the aromatic. **c**, Bicyclo[3.1.1]heptanes (BCHepts) derived from [3.1.1]propellane (**1**) exactly mimic the geometry of *meta*-substituted arenes. **d**, A multigram scale synthesis of [3.1.1]propellane.

Fig. 2. Theoretical analysis of [1.1.1] and [3.1.1]propellane reactivity and synthesis of BCHepts from [3.1.1]propellane. **a**, Reactivity profile of **1** calculated at SMD(THF)-DLPNO-CCSD(T)/ma-def2-QZVPP//SMD(THF)-B2PLYP-D3BJ/def2-TZVP (ma-def2-TZVP on N) level of theory. **b**, Carbon/halogen-substituted BCHepts prepared from organohalides using a) Ir(ppy)₃ (2.5 mol%), blue LED irradiation, or b) Et₃B (10 mol%) as initiator, or c) without an initiator. **c**, Nitrogen-substituted BCHepts prepared using d) dual photoredox / Cu-catalyzed coupling of iodonium dicarboxylates and N-heteroarenes, or e) pyrazole / I₂, or f) α -iodoaziridine, Ir(ppy)₃ (2.5 mol%), blue LEDs. **d**, **g**, Chalcogen-substituted BCHepts prepared by direct reaction with the chalcogen-X precursor. **e**, Cysteine-selective conjugation studies using the (L,L,D) δ -(α -aminoadipolyl)-Cys-Val (ACV) tripeptide in aqueous phosphate buffer (50 mM, pH 8.0). Reactions run on 0.1–0.2 mmol scale unless shown otherwise. See the Supplementary Information for details.

275 **Fig. 3. BCHeP functionalization and topological analysis of crystalline derivatives. a,**
276 Functionalization of iodo-BHePs through a) iron-catalyzed Kumada coupling, or b) lithiation /
277 electrophilic quench. **b,** Comparison of angles between substituent vectors from single crystal X-ray
278 structures ('X-ray'), and computed structures for BCHePs and the parent arenes ('BCHeP_{calc}' and '*m*-
279 Ar_{calc}', CPCM(THF)-B2PLYP-D3BJ/def2-TZVP level of theory). See the Supplementary
280 Information for details.

281

282 **Fig. 4. Synthesis of BCHeP pharmaceutical analogues and comparison of pharmacokinetic**
283 **profile and metabolic stability. a,** Synthesis of BCH analogues of sonidegib and URB597. **b,**
284 Computational investigation of the topology of BCHeP-sonidegib. **c,** Physicochemical and metabolic
285 profile of BCHeP-sonidegib and BCHeP-URB597 along with their parent compounds. See the
286 Supplementary Information for details.

287

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296 **Author contribution**

297 E.A.A., N.F., J.N. and A.J.S. conceived the project. Experimental work was carried out by N.F., J.N.,
298 B.R.S., P.R., T.Z.G. and T.G.. H.D.P. collected the crystallographic data. N.F. and A.J.S. carried out

the computational analysis. The project was supervised by E.A.A., F.D., P.E.B. and C.J.S.. E.A.A., N.F., J.N. and F.D. wrote the initial manuscript which was reviewed and edited by E.A.A., N.F., J.N., R.C.S., P.E.B. and F.D..

Competing interest declaration

The authors declare no competing interests.

Additional information

Supplementary Information is available for this paper. Crystallographic data for **6s**, **7f**, **7h**, **8d** and **12** are deposited with the CCDC (2175923–2175926 and 2176171). The minimum datasets (physicochemical and pharmacokinetic property analysis in Fig. 4c) have been deposited in the Oxford Research Archive (<https://ora.ox.ac.uk>). Correspondence and requests for materials should be addressed to Edward A. Anderson. Reprints and permissions information is available at www.nature.com/reprints.

Online-only Methods

Synthesis of [3.1.1]propellane

1-(3-Chloropropyl)cyclopropan-1-ol, S1.

A solution of ethyl 4-chlorobutanoate (5.60 mL, 40.0 mmol, 1.0 equiv.), and Ti(Oi-Pr)₄ (1.20 mL, 4.0 mmol, 0.10 equiv.) in anhydrous diethyl ether (60 mL) was cooled to 0 °C and a solution of EtMgBr (33.3 mL, 3.0 M in Et₂O, 100 mmol, 2.5 equiv.) was added dropwise over 90 minutes. The mixture was stirred for further 30 minutes at 0 °C, then the mixture was *slowly* quenched by dropwise addition of 10% aqueous H₂SO₄ (50 mL). The organic layer was washed sequentially with H₂O (70 mL), NaHCO₃ (sat., aq., 70 mL), brine (70 mL), then dried (Na₂SO₄), filtered and

323 concentrated *in vacuo* to afford **S1** (4.73 g, 35.1 mmol, 88%) as a colourless oil which was used
324 without further purification.

325 ***1-(2-Chloroethyl)cyclopropyl methanesulfonate, 4.***

326 A solution of **S1** (4.73 g, 35.1 mmol, 1.00 equiv.) and triethylamine (7.05 mL, 50.7 mmol, 1.44
327 equiv.) in anhydrous CH₂Cl₂ (60 mL) was cooled to 0 °C and methanesulfonyl chloride (3.08 mL,
328 40.0 mmol, 1.14 equiv.) was added dropwise over 30 minutes. The mixture was stirred for a further
329 30 minutes at 0 °C, then quenched with water (30 mL). The layers were separated, and the organic
330 layer was washed sequentially with H₂O (60 mL), 10% H₂SO₄ (aq., 50 mL), NaHCO₃ (sat., aq., 50
331 mL) and brine (50 mL), then dried (Na₂SO₄), filtered and concentrated *in vacuo* to afford **4** (7.20 g,
332 33.9 mmol, 96%) as a pale-yellow oil which was used without further purification.

333 ***5-Chloro-2-(chloromethyl)pent-1-ene, S2.***

334 To solution of **4** (7.20 g, 33.8 mmol, 1.0 equiv.) in anhydrous CH₂Cl₂ (60 mL) was slowly added
335 TiCl₄ (5.72 mL, 52.4 mmol, 1.55 equiv.) at room temperature over 20 minutes. The mixture was
336 stirred at room temperature for 3 h, then slowly quenched with H₂O (60 mL) at 0 °C with vigorous
337 stirring. The layers were separated, and the organic layer was washed sequentially with H₂O (2 × 70
338 mL), NaHCO₃ (sat., aq., 70 mL), brine (70 mL), then dried (MgSO₄), filtered and concentrated *in*
339 *vacuo* (300 mbar, 30 °C) to afford **S2** (4.46 g, 29.1 mmol, 86%) as a clear pale-yellow liquid. *Note:*
340 *The product is volatile under reduced pressure (isolated with residual solvent) and was taken*
341 *forward without further purification. The state yield makes allowance for residual solvent.*

342 ***1,1-Dibromo-2-(chloromethyl)-2-(3-chloropropyl)cyclopropane, 5.***

343 To a vigorously stirred (1000 rpm) solution of **S2** (4.46 g, 29.1 mmol, 1.00 equiv.), CHBr₃ (20.4
344 mL, 233 mmol, 8.00 equiv.), dibenzo-18-crown-6 (524 mg, 1.47 mmol, 0.05 equiv.) and pinacol
345 (137 mg, 1.15 mmol, 0.04 equiv.) in CH₂Cl₂ (37 mL) was added 50% NaOH solution (22 mL)
346 dropwise over 20 minutes at 50 °C. The resulting mixture was stirred for 5 h at 50 °C, then cooled
347 to room temperature and diluted with *n*-pentane (100 mL) and distilled water (100 mL). The

348 resulting suspension was filtered through a pad of celite and washed with additional *n*-pentane (100
349 mL). Additional distilled water (100 mL) was added to the filtrate. The layers were separated and
350 the organic layer was washed with brine (150 mL), then dried (Na₂SO₄), filtered and concentrated
351 *in vacuo*. The crude product was purified by column chromatography (ø 4 cm, 40 g SiO₂, gradient
352 100% pentane to 95:5 pentane / EtOAc) to afford **5** (7.50 g, 23.1 mmol, 79%) as a colourless oil.

353 **[3.1.1]Propellane, 1.**

354 To a cooled (−78 °C) solution of **5** (9.74 g, 30.0 mmol, 1.0 equiv.) in anhydrous Et₂O (160 mL) was
355 slowly added phenyllithium (31.8 mL, 60.4 mmol, 2.01 equiv., 1.9 M in *n*-Bu₂O). The resulting
356 mixture was stirred at −78 °C for 15 minutes then warmed to room temperature and stirred for 7 h.
357 The mixture was then distilled using a rotary evaporator (25 °C water bath temperature) equipped
358 with a dry-ice cold finger condenser, with the receiving flask immersed in a dry ice / acetone bath.
359 The Et₂O fraction was removed by slowly decreasing the applied pressure to 150 mbar. This
360 fraction was then discarded. The remaining solution was distilled by slowly reducing the applied
361 pressure to <10 mbar to afford a solution of [3.1.1]propellane **1** in *n*-Bu₂O which was stored under
362 an inert atmosphere at −20 °C. The yield was determined by ¹H NMR spectroscopy using 1,2-
363 dichloroethane as an internal standard (see below). The concentration of the [3.1.1]propellane
364 solution ranged between 0.25 M and 0.50 M, with yields of 43-61%. *Note: The resulting propellane*
365 *stock solution contains bromobenzene which does not influence the reactions presented herein.*

366

367 **Reactions of [3.1.1]propellane**

368 ***General procedure: Photoredox-catalyzed ATRA³¹ (Figure 4b, conditions a)***

369 To a flame dried, screw-capped vial equipped with a stirrer bar was added *fac*-Ir(ppy)₃ (2.5 mol%),
370 alkyl or aryl halide (1.0 equiv.), *t*-BuCN (0.1 M) and [3.1.1]propellane (1.1 to 2.0 equiv. of a
371 solution in *n*-Bu₂O). The vial was placed under nitrogen, and the solution was degassed *via* a
372 modified freeze-pump-thaw cycle (vacuum was only applied while the reaction mixture was frozen

373 due to TCH volatility). The stirred mixture was irradiated with blue LEDs (Kessil PR160 456 nm)
374 with fan cooling for the indicated time. The reaction mixture was concentrated, and the residue was
375 purified by column chromatography.

376 **General procedure: ATRA with BEt_3 ³⁰ (Figure 4b, conditions b)**

377 Under air, a solution of alkyl iodide (1.0 equiv.) in [3.1.1]propellane (1.1 to 1.5 equiv. of a solution
378 in *n*-Bu₂O) was cooled to 0 °C and Et₃B (10 mol%, 1.0 M in hexane) was added *via* syringe (needle
379 tip in the solution). The mixture was stirred until the reaction reached completion as monitored by
380 thin layer chromatography. The reaction mixture was then concentrated and the residue purified by
381 column chromatography.

382 **General procedure: Ir/Cu catalysed additions to [3.1.1]propellane³² (Figure 4c, conditions d)**

383 To a flame dried, screw-capped vial equipped with a stirrer bar was added *fac*-Ir(ppy)₃ (2.0 mol%),
384 amine starting material (1.0 equiv.), iodomesitylene bismethoxylic acid (2.0 equiv.), Cu(acac)₂ or
385 Cu(TMHD)₂ (0.60 equiv), 2-*tert*-butyl-1,1,3,3-tetramethylguanidine BTMG (3.0 equiv.) and
386 anhydrous 1,4-dioxane (0.03 M). The solution was sparged with Ar for 10 minutes then
387 [3.1.1]propellane (1.5 equiv. of a solution in *n*-Bu₂O) was added, the vial capped and sealed with
388 parafilm. The mixture was stirred and irradiated with blue LEDs (Kessil PR160 456 nm) with fan
389 cooling for 16 h. The reaction mixture was diluted with EtOAc and washed with 30% aqueous
390 ammonia solution. The phases were separated, and the aqueous phase was extracted with EtOAc (3
391 ×). The combined organics were dried (Na₂SO₄), filtered and concentrated *in vacuo*. The residue
392 was purified by column chromatography.

393 **General procedure: Addition of thiols (Figure 4d, conditions g)**

394 To a solution of thiol (1.1 equiv.) in anhydrous diethyl ether was added [3.1.1]propellane (1.0
395 equiv., of a solution in *n*-Bu₂O) dropwise. The mixture was stirred for 1 h then the mixture was
396 diluted with diethyl ether, washed with 1 M aqueous NaOH solution (3 ×). The organic layer was

397 dried over Na₂SO₄, filtered and concentrated *in vacuo*. The obtained crude product was either
398 purified by column chromatography or trituration.

399 ***General procedure: Disulfide addition***³⁷ (Figure 4d, conditions g)

400 To a flame dried, screw-capped vial was added disulfide (3.0 equiv.) and [3.1.1]propellane (1.0
401 equiv. of a solution in *n*-Bu₂O). The mixture was irradiated with a LED lamp (HepatoChem
402 Evoluchem HCK1012-01-011 365 nm) with fan cooling for 20 h. The solvents were removed *in*
403 *vacuo* and the resulting residue was purified by column chromatography.

404 ***General procedure: Addition of Sulfonylthionates***³⁵ (Figure 4d, conditions g)

405 To a solution of the specific thiosulfonate (1.0 equiv.) in anhydrous MeCN was added
406 [3.1.1]propellane solution (1.5 equiv.). The flask was sealed and heated to 60 °C for 16 h, then
407 cooled to room temperature and concentrated *in vacuo*. The crude product was purified by column
408 chromatography.

409 ***General procedure: Iron-catalyzed Kumada coupling***³⁹ (Figure 5a)

410 To a flame-dried vial was added BCHeP iodide (1.0 equiv.) and Fe(acac)₃ (20 mol%). The vial was
411 then evacuated and refilled with N₂ (g) three times. To this was added anhydrous THF (0.2 mL) and
412 TMEDA (40 mol%), and the resulting mixture was stirred for 5 minutes. The Grignard reagent (1.6
413 equiv.) was then added *via* syringe pump over approximately 1 h at room temperature. The reaction
414 was stirred for a further 1 h, then quenched by addition of aqueous NH₄Cl (2 mL, saturated). The
415 layers were separated, and the aqueous layer was extracted with Et₂O (3 × 1 mL). The combined
416 organic layers were washed with brine, dried over MgSO₄, filtered and concentrated *in vacuo*. The
417 crude product was purified by column chromatography.

418 **Author Information**

419 The authors declare no competing interests. Correspondence and requests for materials should be
420 addressed to Edward A. Anderson (edward.anderson@chem.ox.ac.uk).