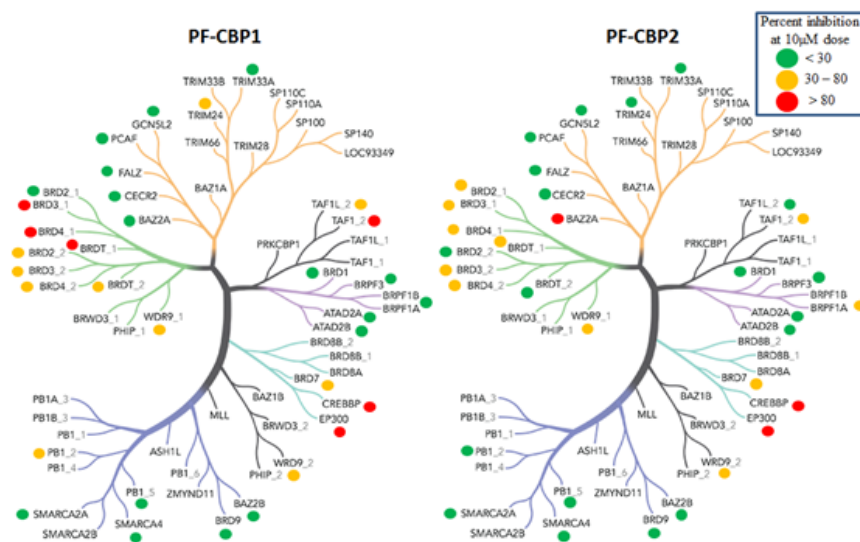
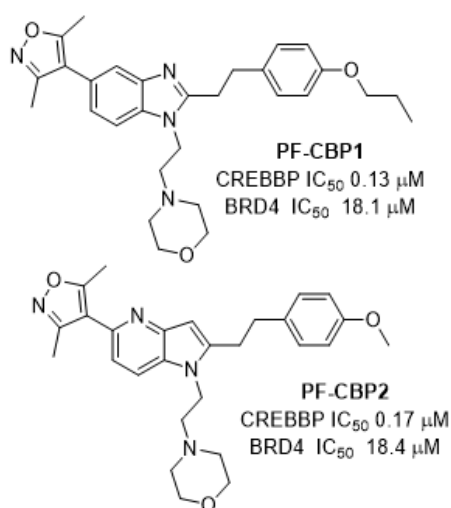


Structure-based design of highly selective inhibitors of the CREB binding protein bromodomain

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ABSTRACT: Chemical probes are required for preclinical target validation to interrogate novel biological targets and pathways. Selective inhibitors of the CREB Binding Protein (CREBBP)/EP300 bromodomains are required to facilitate the elucidation of biology associated with these important epigenetic targets. Medicinal chemistry optimization, that paid particular attention to physiochemical properties, delivered chemical probes with desirable potency, selectivity and permeability attributes. An important feature of the optimization process was the successful application of rational structure-based drug design to address bromodomain selectivity issues (particularly against the structurally related BRD4 protein).



Introduction

Post-translational modifications of chromatin regulate cellular transcription. The dynamic processes of remodeling histone tails are performed using epigenetic proteins called ‘writers’, ‘readers’ and ‘erasers’. Epigenetic writers such as histone acetyltransferases, histone methyltransferases, protein arginine methyltransferases and kinases install one or more post-translational marks, including acetyl, methyl and phosphate groups, on the histone tail. Histone deacetylases, lysine demethylases and phosphatases are epigenetic erasers that remove these marks and proteins containing bromodomains, chromodomains and Tudor domains read epigenetic marks to trigger downstream signaling.

Epigenetic aberrations are recognized as key drivers for several human diseases and drug discovery efforts in this area have primarily focused on writer and eraser proteins.¹ Recently, there has been an increased interest in histone reader proteins for several indications.² Proteins that possess a bromodomain are able to read the acetyl lysine marks associated with epigenetic information transfer. Small molecule inhibitors that perturb acetyl lysine reading interactions are used to decipher their associated biology.¹ One such example that of BET bromodomain inhibitors, has also illustrated the therapeutic potential of this class.²⁻³

CREB Binding Protein (CREBBP or CBP) is a master transcriptional regulator primarily involved in the activation of gene transcription. It is a ubiquitously expressed target that plays a key role in transcriptional activation in human cells. CREBBP possesses many domains including a nuclear receptor interaction domain, the CREB and MYB interaction domain, the cysteine/histidine region, an interferon response binding domain and a single bromodomain. The bromodomain acetyl-lysine reader is responsible for the recruitment of the transcription factor, CREB, to its promoter binding sites. CREBBP is also known to interact with many proteins, including p53 to induce the transcription of p53 target genes. BET bromodomain biology has been significantly illuminated through the use of selective chemical probes.⁴ We reasoned that learnings from the medicinal chemistry of BET bromodomains could be applied to the development of selective CREBBP bromodomain inhibitors to further interrogate the biology and therapeutic potential of this target. We recently reported the culmination of our efforts towards this goal that included the development and transcriptional profiling of a selective CREBBP/EP300 inhibitor (PF-CBP1) that highlighted the opportunities for this pharmacological approach in neurodegenerative diseases.⁵ Here we detail our broader medicinal chemistry efforts that have successfully yielded multiple novel and selective lead

series that should considerably accelerate drug discovery in this area. A significant effort to discover CREBBP selective tools and to study them in various biological systems has been launched by several research groups.⁶ The Structural Genomics Consortium (SGC) team has recently published a 40-fold selective probe SGC-CBP30 and demonstrated an interesting CREBBP/p53 associated biology (Figure 1).^{6c, 7} Working concurrently with the SGC team, it became our goal to obtain highly selective molecules to help further elucidate differences in the biology of CREBBP and BET bromodomains.

Achieving selectivity across the bromodomain family is a challenging task, but a recent publication by the SGC demonstrated the possibility of designing CREBBP chemical tools.⁷ The SGC had previously disclosed the isoxazole CREBBP inhibitor **1** which was used by our group as a starting point to design novel CREBBP selective probe molecules (Table 1).^{3c, 6c, 7-8} The project's objectives were to identify potent CREBBP tool compounds with 100-fold selectivity over other bromodomains, particularly BRD4. Here we report the design of selective CREBBP inhibitors that are structurally distinct from the recently reported compounds GNE272^{3c} and CPI637.^{3b} The myriad of small molecule-BRD4 crystal structures, and our own work to generate crystal structures with CREBBP, provided confidence that structure-based design could be used to achieve the desired selectivity profile in lead-like series for the project.

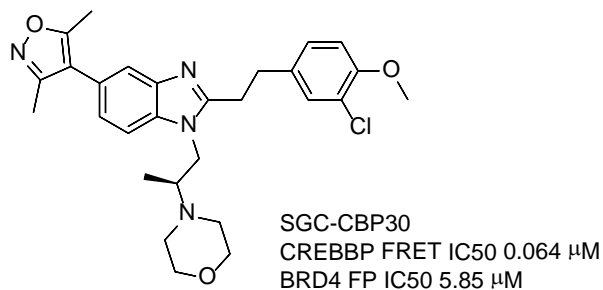


Figure 1. Chemical structure of SGC chemical probe SGC-CBP30.

Bromodomains are comprised of 4 helix bundles that can arrange either in a right or a left handed fashion.⁹ The helices are labeled as Z, A, B and C in going from the *N*- to *C*- terminus (Figure 2A). The spatial orientation of these 4 helix bundles selectively recognizes the monoacetylated terminal lysine in histones. The conserved Asn1168 residue in the α B helix makes a key hydrogen bond interaction with the acetyl lysine. The loops connecting ZA and BC bundles surround the binding site of the acetyl lysine binding pocket. A network of 5 structural water molecules provides

opportunities for hydrophilic interactions in the deep end of the binding pocket as shown in Figure 2A. The isoxazole headpiece oxygen in **1** makes a hydrogen bonding interaction with the Asn1168 amide moiety and the isoxazole nitrogen points towards water A which bridges with the Tyr1125 (Figure 2B). The benzimidazole connected to the isoxazole core acts as a spacer and the phenethyl substitution makes a key cation- π interaction with Arg1173. The morpholine substitution on the benzimidazole acts as a solubilizing group and it points towards solvent. A comparison of the Arg1173 orientation in the compound **1** crystal structure and crystal structures of other ligands (PDB IDs 3SVH and 3P1C)¹⁰ identified a different orientation of this critical binding residue. When CREBBP is crystallized with small ligands that are incapable of making cation- π interactions (Figure 3A) the guanidine moiety of Arg1173 points in the North-South orientation and all the carbon atoms of the Arg1173 side chain exist in the low energy staggered orientation.¹¹ This is an expected orientation in the apo-protein crystal structure (such as 3DWY).¹⁰ However, compound **1** induces a conformational change to rotate the guanidine in Arg1173 to an East-West orientation, and also introduces a higher energy gauche conformation along the C γ – C δ bond as shown in Figure 3B. An additional illustration of CREBBP and BRD4 binding pockets is shown in the supplemental material section (Figure S1 and Table S1). This ligand-induced residue rotation enables the cation- π interaction to be made. In addition, a careful investigation of the binding orientation of the phenethyl substituent in compound **1** reveals a higher energy gauche orientation along the ethyl bond. This observation emphasizes the importance of capturing an energetically favorable cation- π interaction in the design of new ligands that compensates for two unfavorable gauche orientations observed in the protein-ligand complex. The enhancement of this cation- π interaction thus constitutes a plausible design strategy to achieve requisite potency and selectivity in novel CREBBP bromodomain inhibitors.

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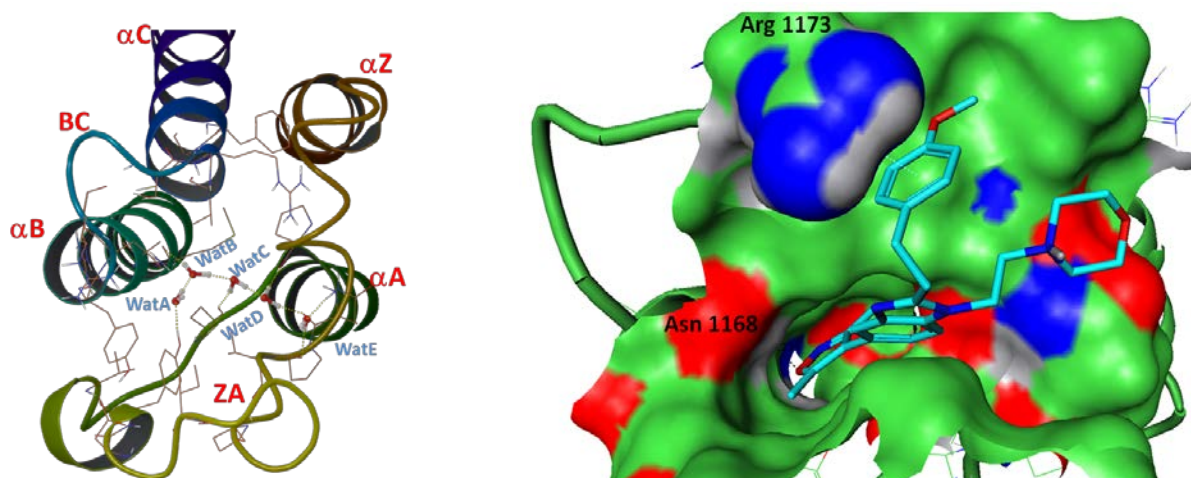


Figure 2. A: CREBBP protein domains with deep pocket water molecules labelled Wat A to E are shown; B: Crystal structure of **1** with CREBBP (PDB Code 5CGP). Cation-pi interaction between the Arg1173 and (4-methoxy)phenethyl moiety is shown.

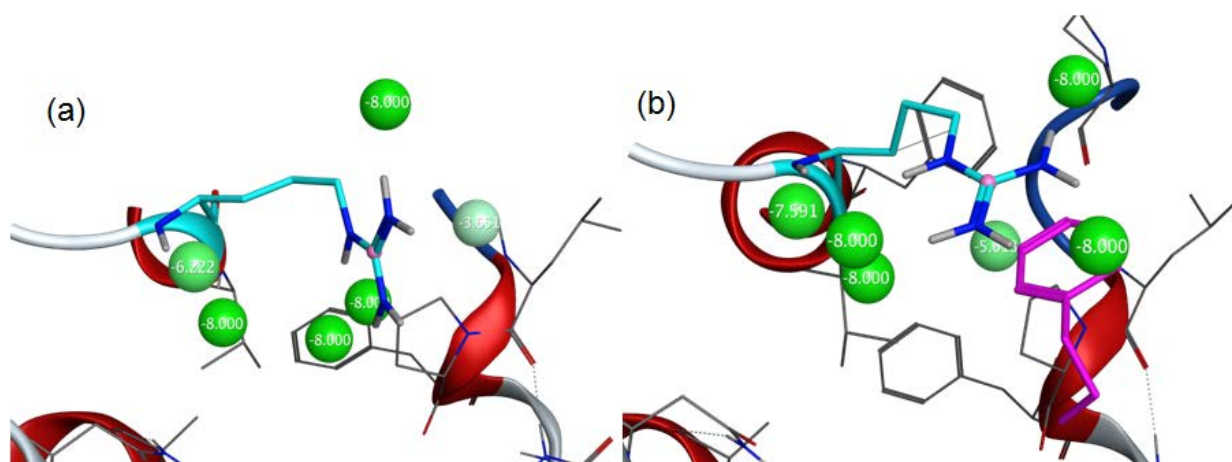


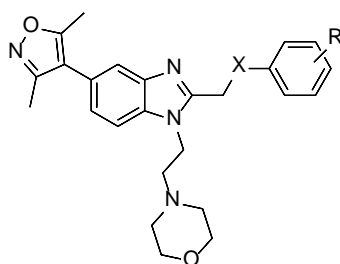
Figure 3. Water molecules around Arg1173 as predicted by 3D RISM.¹¹ **A:** Crystal structure with Arg in the staggered orientation (PDB IDs 3SVH and 3P1C) with predicted bulk water molecules around them (green sphere); **B:** Arg1173 is in a gauche orientation making an Arg-phenyl interaction. Predicted ΔG values for displacing water molecules are depicted on the green sphere.

Electron donating groups on the phenyl ring are expected to enhance cation- π interactions according to quantum mechanics calculations.¹² The unsubstituted phenyl analog **2** is 2-fold less active than the *para*-substituted compound **1** which was first disclosed by the SGC.^{3a, 6c, 13} A corresponding *meta*-compound **3** turned out to be even less active which is consistent with quantum

mechanics calculations for cation- π interactions.^{12d} However, the *ortho*-substituted analog **4** showed a significant drop in potency which may indicate an effect of *ortho*-substitution on the orientation of the phenethyl chain. *Ortho*- and *meta*- polar substitution may also lead to undesired solvation effects. Polar substituents on the phenyl ring must interact with the water molecules that solvate the Arg1173 residue. *Ortho*- and *meta*-methoxy groups may hinder water molecules from solvating Arg1173, thus creating unfavorable interactions (Figure 3). 3,4-Dimethoxy analog **5** is comparable to **1** in terms of lipophilic efficiency ($\text{LipE} = \text{pIC}_{50} - \text{LogP}$)¹⁴ which is a useful parameter in drug design as it describes desirable attributes related to potency and lipophilicity (high quality drug candidates possess a high LipE). 2,4-Dimethoxy compound **6** demonstrated even greater selectivity but with some loss of CREBBP potency. Another approach to enrich electron density within the aromatic system was to install an ether linkage between the benzimidazole and benzene ring, but the corresponding derivatives **7** and **8** lost CREBBP potency either due to a suboptimal tether geometry or the abovementioned solvation effect. The most successful approach to improve selectivity within this scaffold was to expand on the substitution at the *para*-position. While the hydroxyl-containing derivative **9** showed no change in its selectivity profile, ethoxy and propoxy analogs **10** and **11** had a remarkable improvement in selectivity (51 and 62-fold respectively). These compounds were followed up in an isothermal titration calorimetry (ITC) assay to confirm their selectivity. Propoxy analog **11** showed no thermal signature in the BRD4 ITC assay which correlates with the weak biochemical BRD4 potency (IC_{50} 18 μM , Table S2 in supplemental material section). Compound **11** (that we named PF-CBP1) was the most selective and potent CREBBP tool compound discovered at that time, and was used as a chemical probe for transcriptional profiling experiments.¹⁵ Additional *para*-position modifications **12-20** demonstrated either a loss of CREBBP potency or selectivity. Changing the propoxy group in **11** to *i*-Pr substituent (**12**) led to a loss of selectivity, and a similar effect was observed by extending the alkyl chain to butyl (**13**) or pentyl (**14**) groups. A remarkable loss in CREBBP potency was observed for the

electron withdrawing trifluoromethoxy analog **15**, while the retention of activity for the nitrile-containing compound **16** indicates a more complex picture than a simple cation- π interaction paradigm. The loss in potency of **15** can be attributed to the torsional preference of trifluoroanisole, where the trifluoromethoxy moiety orients 90° from the plane of the phenyl ring¹⁶ which creates a van der Waals clash with the side chain residues. The influence of an electron donating group on the SAR was further explored with *para*-amino derivatives **17-19** but the activity and/or selectivity suffered significantly. In the course of the SAR exploration, compound **20** was found to possess dual CREBBP/BRD4 activity, which served as a useful chemical probe to help delineate differences in inhibiting these bromodomains in our chemical biology studies.⁵ The compounds within this series display acceptable cellular permeability, thus making them suitable for cell-based studies. For example, the passive permeability rate for the chemical probes **11** and **20** are 7.9×10^{-6} cm/sec and 4.5×10^{-6} cm/sec at 2 μ M, respectively. However, the microsomal clearance for the majority of the compounds from the series is relatively high limiting their applications to predominantly in vitro work. Intrinsic clearance in human microsomal assay for compounds **11** and **20** are >300 μ L/min/mg and 206 μ L/min/mg at 1 μ M dose, respectively. The tool molecule **11** also demonstrated a relatively clean polypharmacology profile determined in a broad CEREP panel where the activity was observed only for the adrenergic A1a receptor (antagonist, IC_{50} 0.62 μ M, Supporting Information Table S3) and L-type calcium channel (antagonist, IC_{50} 3.0 μ M).

Table 1. Potency and physicochemical data of the 4-(2-(5-(3,5-dimethylisoxazol-4-yl)-2-phenethyl-1H-benzo[d]imidazol-1-yl)ethyl)morpholine chemical class.^a



Compound	R	X	CREBBP FRET IC ₅₀ (μM)	BRD4 FP IC ₅₀ (μM)	CREBBP/BRD 4 fold selectivity	LipE (CREBBP FRET) ^a
1	4-OMe	CH ₂	0.12	2.4	20	3.6
2	H	CH ₂	0.24	4.1	17	3.5
3	3-OMe	CH ₂	0.31	4.6	15	2.9
4	2-OMe	CH ₂	0.98	13.9	14	2.2
5	3,4-(OMe) ₂	CH ₂	0.47	9.4	20	3.6
6	2,4-(OMe) ₂	CH ₂	0.24	8.3	35	2.8
7	4-OMe	O	0.92	9.2	10	2.8
8	H	O	0.40	14.4	36	3.7
9	OH	CH ₂	0.24	4.4	18	3.6
10	4-OEt	CH ₂	0.13	6.6	51	2.8
11 (PF-CBP1)	4-OPr	CH ₂	0.13	18.1	62	3.5
12	4-O- <i>i</i> -Pr	CH ₂	0.20	4.5	23	2.6
13	4-OBu	CH ₂	0.26	11.2	43	1.9
14	4-O-Pn	CH ₂	0.69	6.9	10	1.5
15	4-OCF ₃	CH ₂	ND	18.5	ND	0.5
16	4-CN	CH ₂	0.30	4.9	16	3.2
17	4-NH ₂	CH ₂	0.09	3.4	38	3.5

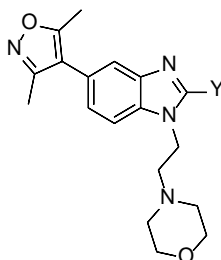
18	4-NMe ₂	CH ₂	0.21	4.3	20	3.6
19	4-NHAc	CH ₂	0.20	1.4	7	4.2
20	(CH ₂) ₃ NMe ₂	CH ₂	0.65	1.5	3	2.6

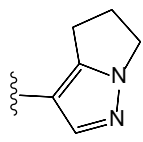
^aExperimental shake-flask LogD values were used.

One of our strategies to improve CREBBP potency was to establish ligand interactions such as a shielded salt bridge or a hydrogen bond with the Arg145 residue in the CREBBP binding pocket. This residue appeared to be accessible from the phenethyl tail present in the chemotype. Various heterocyclic derivatives were prepared to probe hydrogen bonding interactions with the Arg145 residue. Complicated structure-activity relationships in the phenethyl region of the molecule were further demonstrated by exploring heterocyclic substituents in place of the phenyl ring (Table 2). The 2-fold loss of CREBBP activity in 3-pyridyl analog **22** compared to 2-pyridyl (**21**) and 4-pyridyl (**23**) derivatives is consistent with the lone pair of **22** making unfavorable interactions with the water molecule, similar to that seen for *meta*-substituted derivative **3**. Another BRD4/CREBBP selectivity approach was to take advantage of the residue differences between these two bromodomain binding sites (Table S1, supplemental material section). Trp81 in BRD4 (Figure 4A) is significantly larger than the corresponding Leu81 in CREBBP, suggesting that adding appropriate substitution to the ligand template in this region may establish a van der Waals clash. Indolyl compound **24** was intended to support the hypothesis that electron rich heterocycles may improve CREBBP binding, as well as decrease BRD4 potency by increasing steric bulk, but the secondary binding effects besides the cation- π interaction, likely played a role in decreasing CREBBP activity, and the drop in BRD4 potency was not significant. Quinolinyln analogs **25** and **26** represent an interesting example of the complexity of the SAR in this region. The CREBBP potency of **25** is more than 10-fold higher than the regioisomer **26** which cannot be explained by stereoelectronic considerations related to the key cation- π interactions. The BRD4 activity of **25** turned out to be similar to the parent molecule **2** thus indicating a significant flexibility of the BRD4 binding pocket around the Trp81 residue. A substitution in this region was further explored by preparing a library of alkyl and aryl derivatives with a direct bond to the benzimidazole core. Compound **27** was discovered as a hit with appreciable CREBBP potency (and impressive LipE) even though CREBBP/BRD4 selectivity did not improve over the phenethyl chemotype. The binding mode of **27** is not able to adopt a cation- π interaction similar to the phenethyl

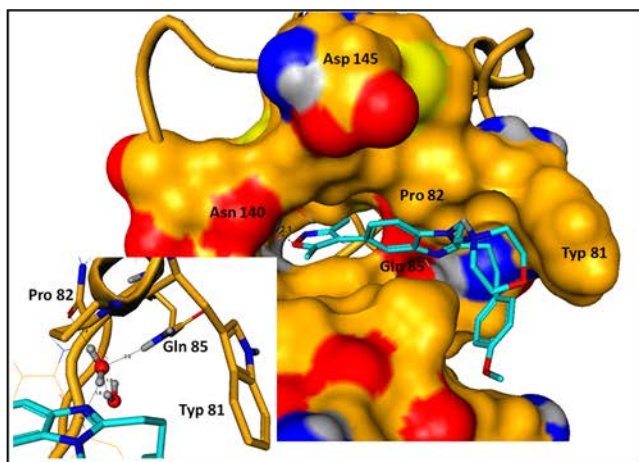
derivatives, thus indicating a potentially novel binding mode that would need to be further elucidated using X-ray crystallography.

Table 2. Structure-activity relationships relating to modifications of the phenethyl substituent.



Compound	Y	CREBBP FRET IC ₅₀ (μM)	BRD4 FP IC ₅₀ (μM)	CREBBP /BRD4 fold selectivity	LipE
21	CH ₂ CH ₂ -2-pyridyl	0.33	3.2	10	4.7
22	CH ₂ CH ₂ -3-pyridyl	0.78	5.8	7.4	4.3
23	CH ₂ CH ₂ -4-pyridyl	0.40	4.1	10	4.6
24	CH ₂ CH ₂ -3-indolyl	0.74	10.1	14	2.9
25	CH ₂ CH ₂ -2-quinolinyl	0.16	4.9	31	3.3
26	CH ₂ CH ₂ -4-isoquinolinyl	2.5	16.8	7	2.7
27		0.13	1.6	12	5.3

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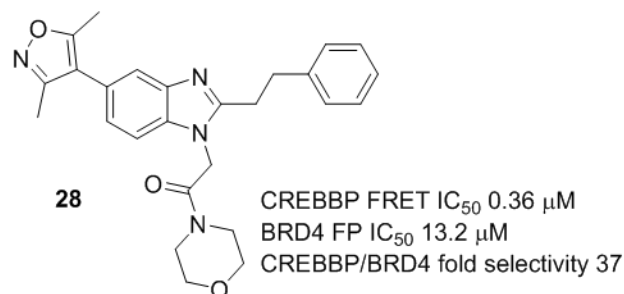


Figure 4. A: Crystal structure of **1** with BRD4 (PDB Code 5CFW). B: Compound **28**;

Crystal structure of **1** determined that the morpholine group points towards the Trp81 region of the BRD4 binding pocket (Figure 4A). Although the keto group of **28** (Figure 4B) decreased BRD4 activity, CREBBP potency generally suffered upon changes in this region. However, a single methyl group with the right stereochemistry in the SGC probe **SGC-CBP30** apparently provided a better compromise with respect to both target activities. A crystal structure of BRD4 with compound **1** demonstrated that Trp81 of BRD4 is rotated around the C_{α} - C_{β} bond by about 130 degrees and also C_{β} is shifted by about 1 Å (Figure 4A). This movement and rotation makes the pocket wider, thus providing a hydrophobic enclosure for compounds **1**, **2** and their analogs. The phenethyl moiety occupies a small pocket in the BRD4 active site and the morpholine group points straight out towards solvent. It is important to note that this orientation is a different binding mode compared to the CREBBP binding pose shown in Figure 2B. This observation provides opportunities for further selectivity enhancements through the development of constrained derivatives (Figure 5). For instance, the selectivity was improved to 67-fold for compound **29**. Conformational constraints in the ethyl tether were further explored (Figure 6). According to the dihedral computation, it was proposed that the 5- and 6-membered ring systems are predicted to have less strain than other ring sizes, and are closer to the crystal structure dihedral angle as shown in Figure 6. The remarkable drop in selectivity for compound **30** supports the hypothesis that a specific orientation is crucial to retain binding potency and selectivity. Further exploration of this strategy to provide **31** and **32** led to a loss of CREBBP potency likely due to the change in orientation of the morpholine ring.

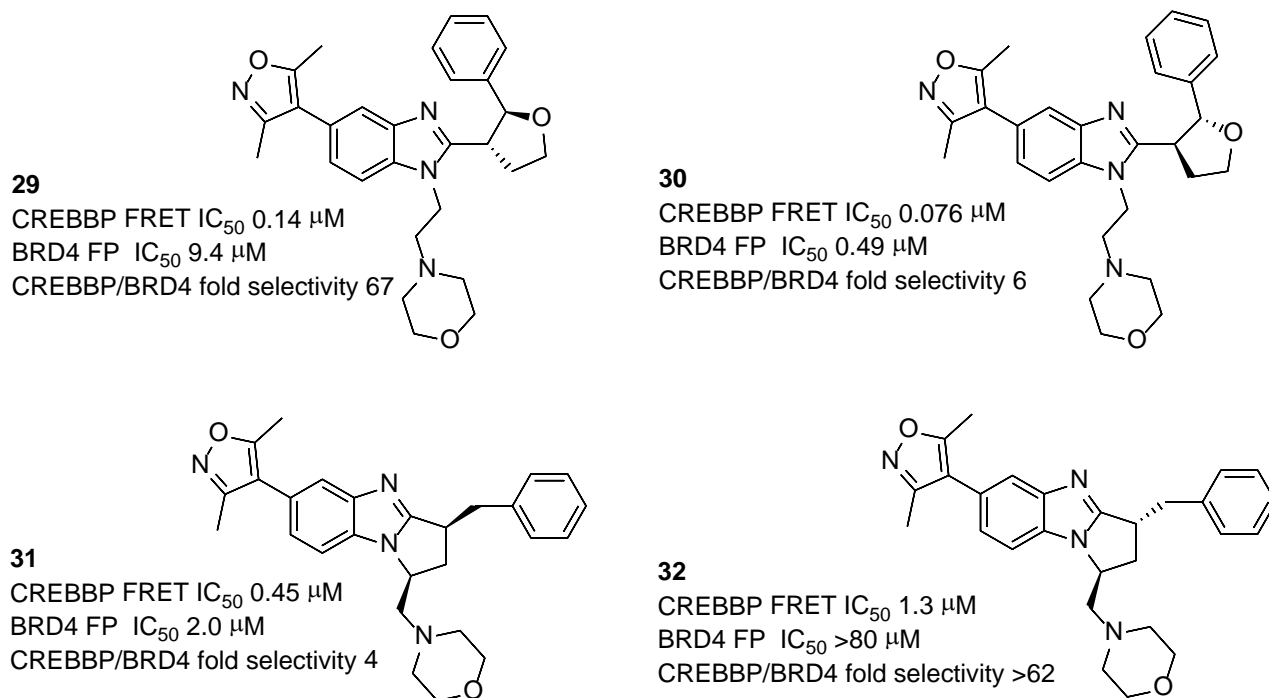


Figure 5. Constrained analogs at both morpholine and phenethyl chains (absolute stereochemistry arbitrarily assigned).

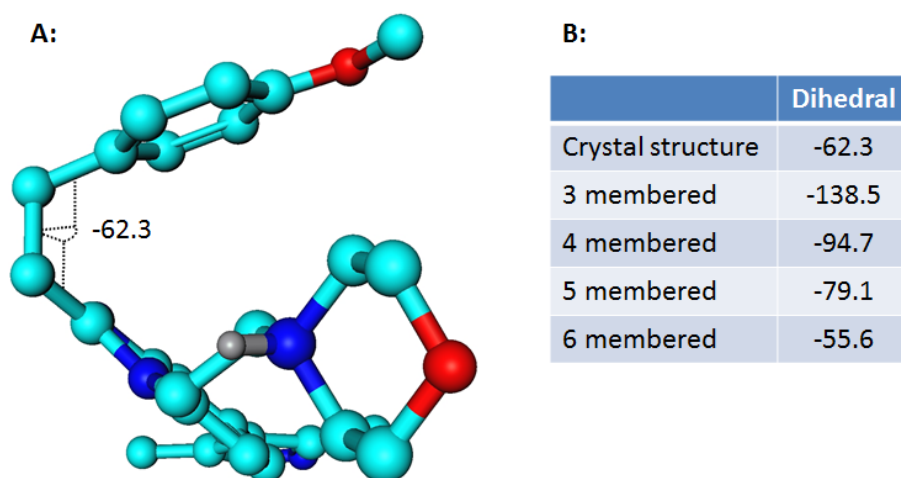


Figure 6. A: The dihedral angle of **1**; B: The dihedral angle table for the lowest energy conformation in the crystal structure of **1** and cyclization to provide the 3- to 6-membered ring systems at the phenethyl group.

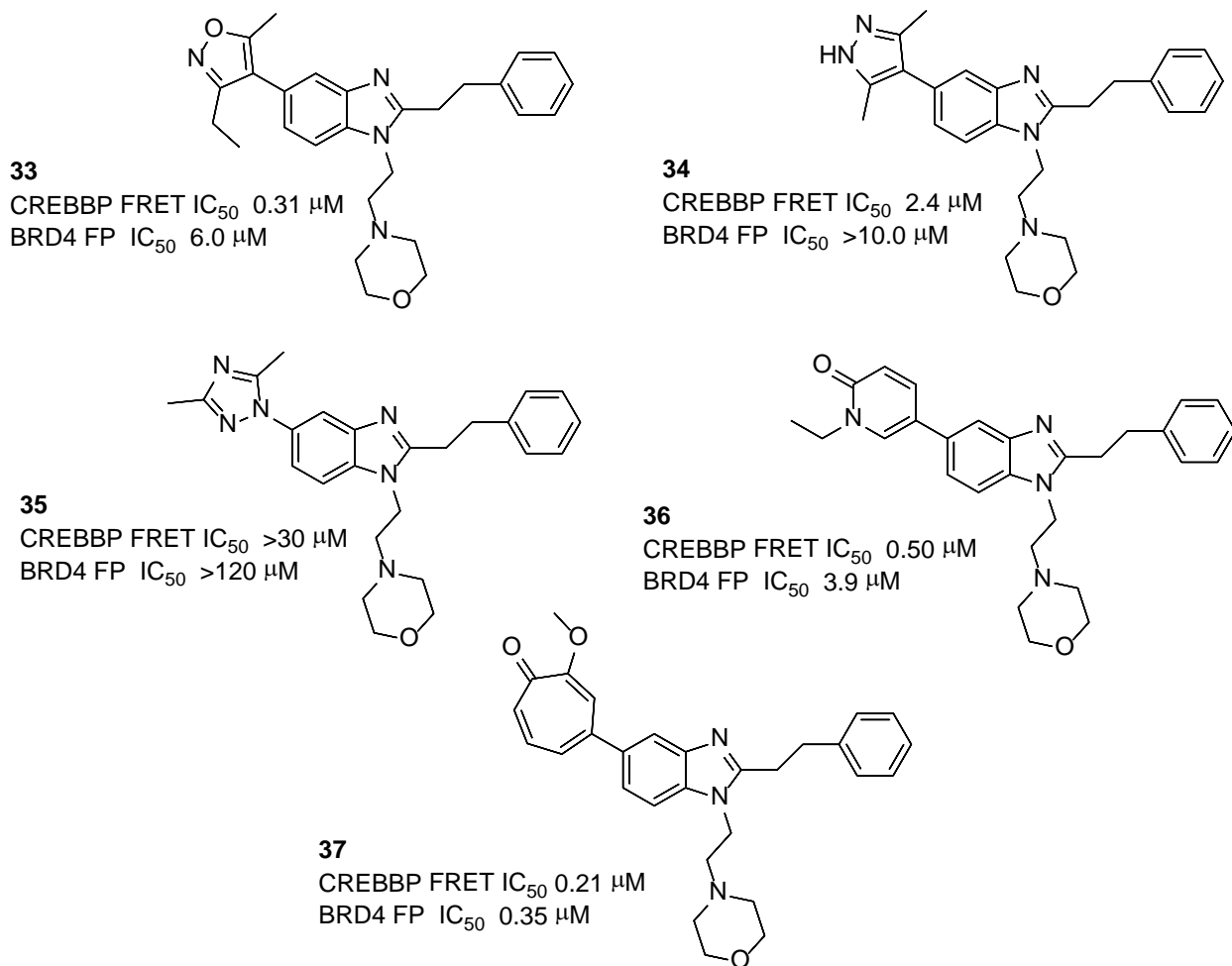


Figure 7. Modifications to the acetyl lysine mimic headpiece.

The dimethylisoxazole head group is a common acetyl lysine mimic used in various bromodomain inhibitors,^{13, 17} and it became our goal to replace this fragment with alternative head pieces. Pyrazole derivative **34** demonstrated that one of the hydrogen bond acceptors of the isoxazole ring cannot be substituted with a hydrogen bond donor to reverse an interaction with water A (Figure 7). Compound **35** suggested that a single hydrogen bond acceptor atom is also not sufficient for potent binding to either BRD4 or CREBBP. *N*-ethylpyridone analog **36** and tropolone derivative **37** were prepared as part of a library with various heterocyclic moieties. These compounds prove that the dimethylisoxazole group can be substituted with alternative head pieces though with some loss of potency. We have previously described the utility of tropolone bromodomain inhibitors from this work as photoaffinity probes of cell-based target occupancy.^{6a}

An additional structure-based drug design strategy to improve selectivity was to restrict the inhibitors into their desired conformation by forming an atropisomer. To determine whether a compound would exist as an atropisomer we performed a dihedral scan using molecular mechanics and quantum mechanics calculations. A systematic dihedral scan of the rotatable bond was performed in MacroModel¹⁸ using OPLS 2005 force field and energy barriers were calculated. If the energy barrier for the dihedrals were found to be <15 kcal/mol, the compound was classified as not being atropisomeric and no further QM calculations were performed. If the energy barrier is calculated to be above 15 kcal/mol, additional dihedral scans were performed using quantum mechanics using *Terachem Program* from *Petachem LLC*.¹⁹

Quantum mechanics dihedral scans were performed at the B3LYP/6-31G level of theory. Torsional scan around the bond were performed using a rotational step of 30 degrees. After each rotation, the resulting structure was optimized using a relaxed termination criterion with $E_{\text{tolerance}}$ of 5×10^{-4} hartree. During optimization, coordinates of the four atoms that define the given dihedral angle were kept fixed and all other atoms were allowed to move. On the optimized structures, single point energies were calculated. If the barrier height based on single point energies was calculated to be > 20 kcal/mol then the compound was classified as very likely to exhibit atropisomerism. The torsional profile scan using molecular mechanics and QM calculations showed an energy barrier of 24.2 and 22.8 kcal/mol respectively for the rotation around the isoxazole/benzimidazole bond by adding methyl substituents to positions 4 and 6 of the benzimidazole **38** (ISOX-INACT, Figure 8).⁵ This atropisomer rigidifies the benzimidazole ring at 90° from the isooxazole ring there by clashing with Trp81 in BRD4 in both its orientations. This negative interaction with Trp81 explains the reduced potency of binding to BRD4 ($IC_{50} > 120 \mu M$) as expected. The loss of CREBBP potency was significant due to poor conserved interactions with the Asn1168 residue and cation- π interactions with the Arg1173 residue. This compound in fact served as a useful negative control for this chemotype in our previously described cell-based experiments.⁵

Finally, a successful selectivity approach was discovered by converting the benzimidazole core into an indole moiety (Figure 8). The benzoimide N-3 nitrogen, as seen in the BRD4 crystal structure acts as a hydrogen bond acceptor to Water E, thus mediating an interaction with Pro82 (Figure 4B). Our strategy was to convert the benzimidazole into an indole group therefore preventing the hydrogen bonding interaction. Indole analog **39** showed a significant decrease in BRD4 potency. However, the

CREBBP binding mode would cause there to be an unfavorable orientation of the indole position 3 hydrogen towards the solvent exposed region of the pocket. Therefore, a more polar substituent might be favored in this area of the molecule and cyano group in **40** was expected to be pointing to the solvent exposed region in CREBBP. Incorporation of a nitrile moiety to yield derivative **40** resulted in a complete loss of BRD4 activity i.e., greater than 240-fold selectivity, with CREBBP potency remaining within several fold of compound **1** activity. To accommodate a more polar substitution in this area of the indole chemotype, we converted indole analog **39** into Azaindole **41** where the additional nitrogen atom would be pointing towards solvent. Azaindole **41** showed an improvement in CREBBP potency to a level comparable to compound **1** activity (170 nM). BRD4 potency remained weak (18.4 μ M) resulting in >100-fold selectivity in the biochemical assays. Lead molecule **41** also possessed a clean polypharmacology profile ($IC_{50} > 5 \mu$ M in a broad CEREP panel, Supporting Information Table S3), good cellular permeability (6.8×10^{-6} cm/sec at 2 μ M), and shake flask LogD (3.29 at pH 7.4). The discovery of this very selective and potent chemotype using structure-based design provides confidence in the future medicinal chemistry development of this series.

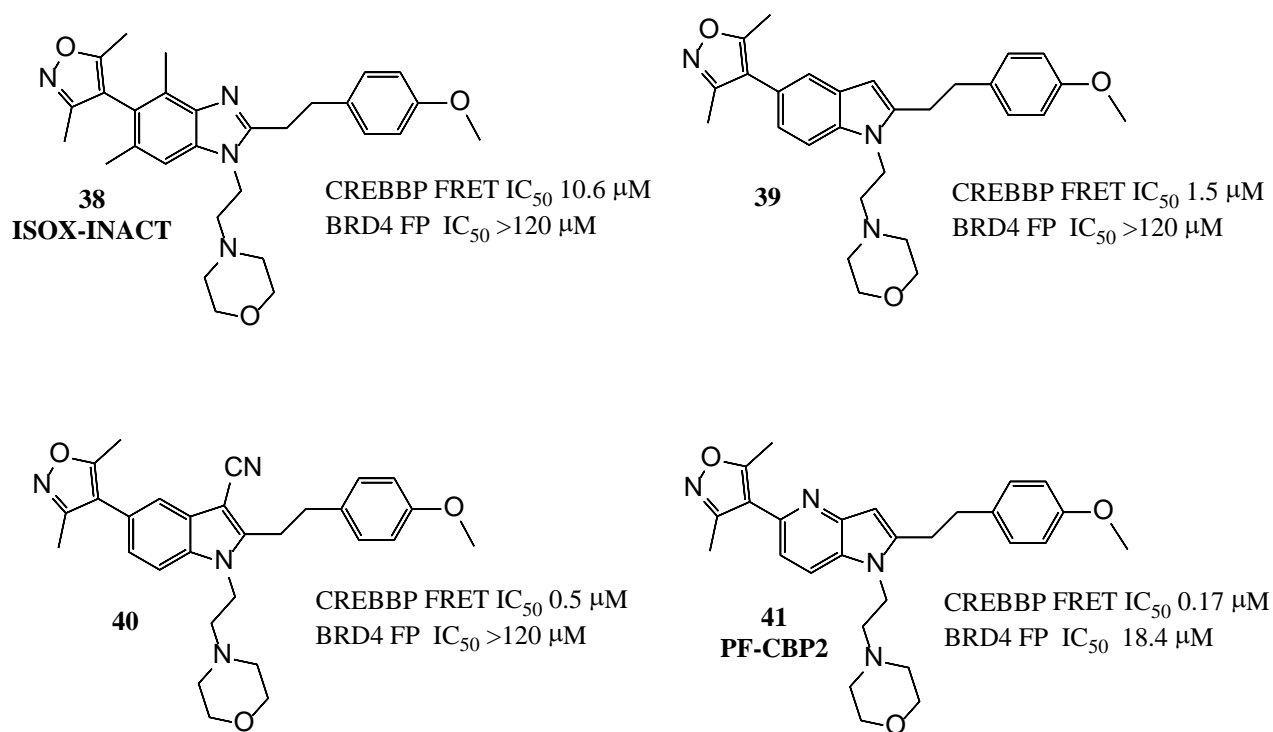


Figure 8. Benzimidazole core modifications

The bromodomain selectivity of lead compounds was assessed using the DiscoverX BROMOScan platform.¹⁵ A screening panel of more than 30 bromodomains was utilized. The ultra-

sensitive qPCR based technology demonstrated a correlation with internally generated data. Even though the assay formats of BROMOScan and biochemical/biophysical assays may account for the differences in potency values, general trends support the selectivity of the isoxazole-benzimidazole compound class (Figure 9). Both compounds **11** and **41** demonstrated activity for the closely related target EP300 that possesses significant homology to CREBBP with respect to the bromodomain binding pocket. It is interesting to note that the bromodomain selectivity profile of compound **41** surpasses that of compound **11**, thus suggesting **41** would serve as a useful lead series for future drug discovery efforts.

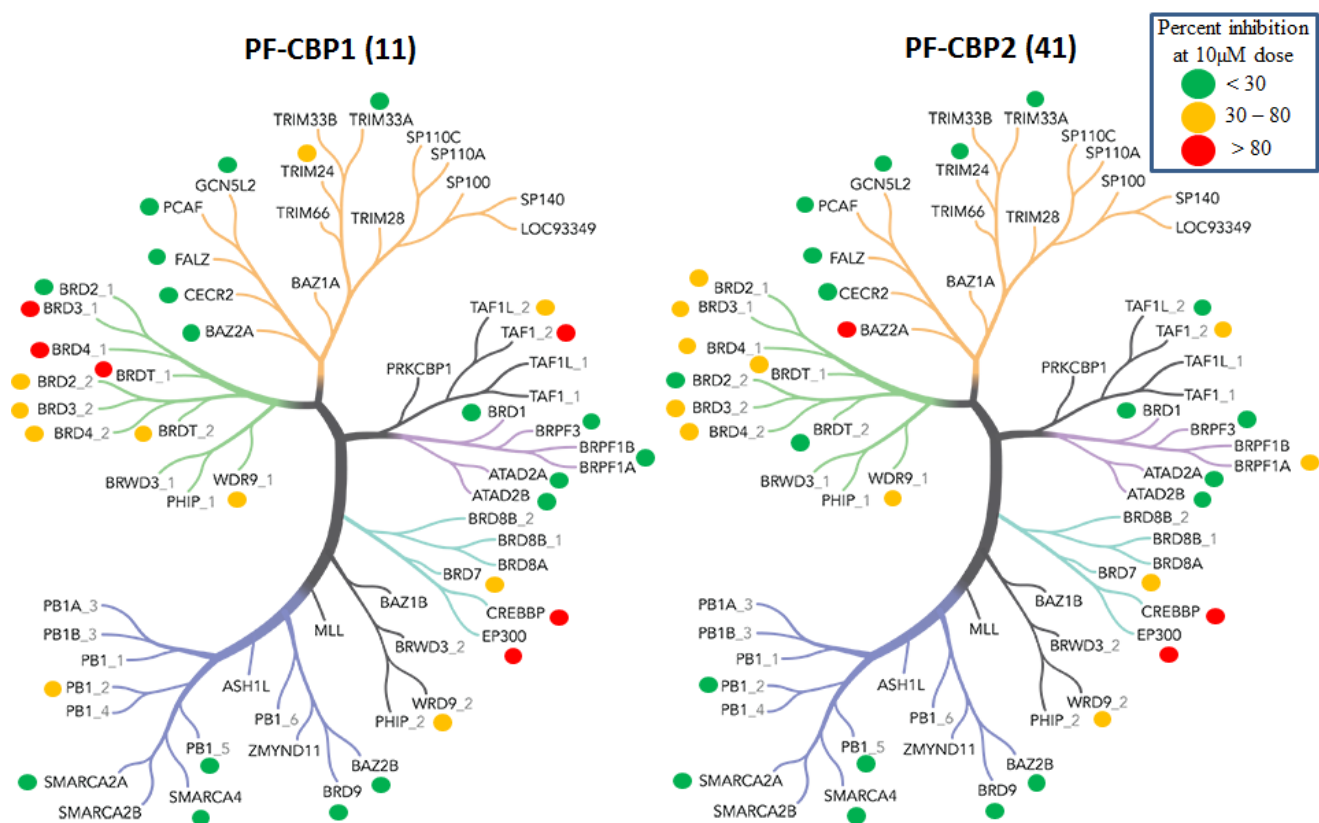


Figure 9. Bromodomain selectivity profile of lead molecules **PF-CBP1 (11)** and **PF-CBP2 (41)**. Green, yellow and red dots indicate the inhibition <30%, 30-80% and >80% respectively in the DiscoverRX assay at 10µM dose.

In conclusion, we have developed highly selective inhibitors of the CREBBP/EP300 bromodomains using structure-based drug design. These compounds will serve as useful lead series for future drug discovery efforts that should focus on addressing other important features such as metabolic stability. Through this work we identified chemical probes PF-CBP1 and PF-CBP2 that have

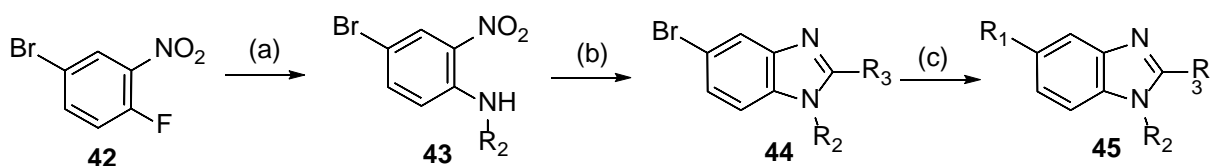
found utility in cell biology and transcriptional profiling experiments that furthers our understanding of CREBBP bromodomain pharmacology.⁵ Novel acetyl lysine mimics were also identified that should be of broad interest to those working in this area.

Experimental Section

Synthesis

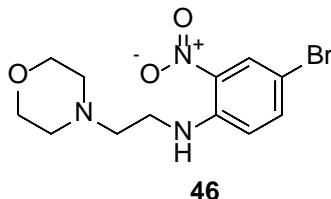
Starting materials and other reagents were purchased from commercial suppliers and were used without further purification unless otherwise indicated. All reactions were performed under a positive pressure of nitrogen, argon, or with a drying tube, at ambient temperature (unless otherwise stated), in anhydrous solvents, unless otherwise indicated. Analytical thin-layer chromatography was performed on glass-backed Silica Gel 60_F 254 plates (Analtech (0.25 mm)) and eluted with the appropriate solvent ratios (v/v). The reactions were assayed by high-performance liquid chromatography (HPLC) or thin-layer chromatography (TLC) and terminated as judged by the consumption of starting material. The TLC plates were visualized by UV, phosphomolybdic acid stain, or iodine stain. Microwave assisted reactions were run in a Biotage Initiator. ¹H NMR spectra were recorded on a Bruker instrument operating at 400 MHz unless otherwise indicated. ¹H NMR spectra are obtained as DMSO-*d*₆ or CDCl₃ solutions as indicated (reported in ppm), using chloroform as the reference standard (7.25 ppm) or DMSO-*d*₆ (2.50 ppm). Other NMR solvents were used as needed. When peak multiplicities are reported, the following abbreviations are used: s = singlet, d = doublet, t = triplet, m = multiplet, br = broadened, dd = doublet of doublets, dt = doublet of triplets. Coupling constants, when given, are reported in Hertz. The mass spectra were obtained using liquid chromatography mass spectrometry (LC-MS) on an Agilent instrument using atmospheric pressure chemical ionization (APCI) or electrospray ionization (ESI). High resolution mass measurements were carried out on an Agilent TOF 6200 series with ESI. All test compounds showed > 95% purity as determined by either combustion analysis, high-performance liquid chromatography (HPLC), liquid chromatography/mass spectrometry analysis (LCMS), or ¹HNMR integration. HPLC conditions were as follows: XBridge C18 column @ 80 °C, 4.6 mm x 150 mm, 5 μm, 5%-95% MeOH/H₂O buffered with 0.2% formic acid/0.4% ammonium formate, 3 min run, flow rate 1.2 mL/min, UV detection (λ = 254, 224 nm). Combustion analyses were performed by Atlantic Microlab, Inc. (Norcross, Georgia).

General Approach to substituted benzimidazoles:

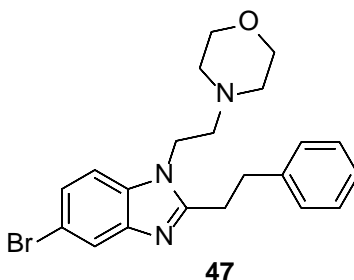


(a) R_2NH_2 , Et_3N , DMSO, 80 °C, 18 h; (b) R_3CHO , $Na_2S_2O_4$, 4:1 EtOH/H₂O, 70 °C, 18 h; (c) $R_1B(OH)_2$, $PdCl_2(dppf)CH_2Cl_2$, Cs_2CO_3 , *p*-diox. 100 °C, 18 h;

General procedure A (Benzimidazole formation):



4-Bromo-N-(2-morpholinoethyl)-2-nitroaniline (46): A mixture of 5-bromo-2-fluoronitrobenzene (**42**) (81.4 g, 0.37 mol), *N*-(2-aminoethyl)morpholine (60.0 g, 0.46 mol) and triethylamine (46.5 g, 0.46 mol) in DMSO (1.2 L) was heated to 80 °C and stirred for 2 hours, when TLC (petroleum ether/EtOAc = 1:1) showed the reaction was complete. To the cooled reaction was added water (500 mL) and the mixture was extracted with ethyl acetate (500 mL \times 4). The combined organic layer was washed with water (500 mL) and brine (500 mL \times 2), dried over Na_2SO_4 , filtered, and concentrated *in vacuo*. The residue was purified by flash chromatography (0-50% ethyl acetate in petroleum ether) to afford the titled 4-Bromo-N-(2-morpholinoethyl)-2-nitroaniline (**43**) (145 g, 95%) as an orange solid. ¹H NMR (400 MHz, $CDCl_3$): δ 2.46-2.58 (m, 4H), 2.69-2.76 (m, 2H), 3.30-3.38 (m, 2H), 3.71-3.80 (m, 4H), 6.73 (d, J = 9.0 Hz, 1H), 7.50 (dd, J = 9.0, 1.6 Hz, 1H), 8.33 (d, J = 2.4 Hz, 1H), 8.51 (br s, 1H); LC-MS m/z 330 (M+1)⁺.



4-(2-(5-bromo-2-phenethyl-1H-benzod[imidazol-1-yl)ethyl)morpholine (47): To a mixture of 4-Bromo-N-(2-morpholinoethyl)-2-nitroaniline (**46**) (2.01 g, 6.09 mmol) and 3-phenylpropionaldehyde (2.50 g, 19.0 mmol) in 4:1 ethanol/H₂O (60 mL, 0.10M) was added $Na_2S_2O_4$ (3.95 g, 19.0 mmol). The mixture was stirred at 70 °C for 18 hours under nitrogen. The resulting mixture was diluted with saturated sodium bicarbonate (50 mL) and extracted with ethyl acetate (50 mL \times 3). The combined organic layers were washed with brine (50 mL), dried over Na_2SO_4 , filtered and concentrated *in vacuo*. The residue was purified by flash silica chromatography (0-100% heptane in ethyl acetate) and the major fraction corresponding to the desired product was collected and concentrated to give the titled 4-(2-(5-bromo-2-phenethyl-1H-benzod[imidazol-1-yl)ethyl)morpholine (1.41 g, 56%) as a light yellow oil: ¹H NMR(400 MHz, $CDCl_3$) δ 2.40-2.46 (m, 4H), 2.51 (t, J = 7.0 Hz, 2H), 3.16-3.20 (m, 2H), 3.24-

3.29 (m, 2H), 3.64-3.69 (m, 4H), 4.06 (t, $J = 6.6$ Hz, 2H), 7.17-7.24 (m, 4H), 7.26-7.30 (m, 2H), 7.35 (dd, $J = 8.6, 2.0$ Hz, 1H), 7.89 (d, $J = 1.6$ Hz, 1H).

4-(2-(5-(3,5-dimethylisoxazol-4-yl)-2-phenethyl-1H-benzo[d]imidazol-1-yl)ethyl)morpholine (2). To a RB flask charged with 4-(2-(5-bromo-2-phenethyl-1H-benzo[d]imidazol-1-yl)ethyl)morpholine (**47**) (506 mg, 1.22 mmol), was added sequentially (3,5-dimethylisoxazol-4-yl)boronic acid (260 mg, 1.28 mmol), palladium acetate (17 mg, 0.08 mmol), RuPHOS (CAS 787618-22-8; 73 mg, 0.15 mmol), sodium carbonate (262 mg, 2.47 mmol), and ethanol (6.7 mL, 0.18 M). The mixture was sparged with argon for 5 mins, then heated to 85 °C for 1.5 hours under nitrogen. The mixture was then cooled to rt and then filtered through a pad of celite and rinsed with ethanol. The filtrates were then concentrated and then purified by flash silica chromatography (0-100% heptane in ethyl acetate) and the major fraction corresponding to the desired product was collected and concentrated to give the titled compound **2** (333 mg, 63%) as a viscous yellow oil: ^1H NMR(400 MHz, CDCl_3) δ 2.30 (s, 3H), 2.43 (s, 3H), 2.45-2.61 (m, 6H), 3.24-3.35 (m, 4H), 3.63-3.81 (m, 4H), 4.07-4.25 (m, 2H), 7.17 (dd, $J = 8.2, 1.2$ Hz, 1H), 7.22-7.26 (m, 3H), 7.29-7.33 (m, 2H), 7.43 (br s, 1H), 7.67 (s, 1H).

The following compounds were prepared according to general procedure A:

5-(3,5-dimethylisoxazol-4-yl)-2-[2-(4-methoxyphenyl)ethyl]-1-(2-morpholin-4-ylethyl)-1H-benzimidazole (1): (1.3 g, 60%) as yellow liquid. ^1H NMR (400 MHz, CDCl_3) δ 2.30 (s, 3H), 2.43 (s, 3H), 2.44-2.47 (m, 4H), 2.58 (t, $J = 6.8$ Hz, 2H), 3.15-3.26 (m, 4H), 3.66-3.68 (m, 4H), 3.79 (s, 3H), 4.11 (t, $J = 6.8$ Hz, 2H), 6.84 (d, $J = 8.0$ Hz, 2H), 7.11-7.16 (m, 3H), 7.34 (d, $J = 8.0$ Hz, 1H), 7.63 (d, $J = 0.8$ Hz, 1H); LC-MS m/z 461 ($\text{M}+1$)⁺.

5-(3,5-Dimethylisoxazol-4-yl)-2-[2-(3-methoxyphenyl)ethyl]-1-(2-morpholin-4-ylethyl)-1H-benzimidazole (3). Prepared according to general procedure A using 4-(3,5-Dimethylisoxazol-4-yl)-*N*-(2-morpholinoethyl)-2-nitroaniline (100 mg, 0.29 mmol) and 3-(3-methoxyphenyl)propionaldehyde¹ giving **3** (10.6 mg, 8%) as yellow oil. ^1H NMR (400 MHz, CDCl_3): δ 2.29 (s, 3H), 2.42 (s, 3H), 2.44-2.47 (m, 4H), 2.59 (t, $J = 6.8$ Hz, 2H), 3.18-3.22 (m, 2H), 3.25-3.30 (m, 2H), 3.66-3.68 (m, 4H), 3.74 (s, 3H), 4.11 (t, $J = 6.8$ Hz, 2H), 6.77-6.79 (m, 2H), 6.84 (d, $J = 7.6$ Hz, 1H), 7.11 (dd, $J = 8.0, 1.6$ Hz, 1H), 7.21-7.25 (m, 1H), 7.35 (d, $J = 8.0$ Hz, 1H), 7.63 (d, $J = 1.2$ Hz, 1H).

5-(3,5-Dimethylisoxazol-4-yl)-2-[2-(2-methoxyphenyl)ethyl]-1-(2-morpholin-4-ylethyl)-1H-benzimidazole (4). Prepared according to general procedure A using 4-(3,5-Dimethylisoxazol-4-yl)-*N*-(2-morpholinoethyl)-2-nitroaniline (125 mg, 0.36 mmol) and 3-(2'-methoxyphenyl)propionaldehyde² giving **4** (5.3 mg, 4%) as a brown oil. ^1H NMR (400 MHz, CD_3OD): δ 2.27 (s, 3H), 2.43 (s, 3H), 2.47

¹ Browder, C.C.; Marmsäter, F.P.; West, F.G. *Org. Lett.*, **2001**, 3, 3033 DOI: 10.1021/ol010159w.

² Gangjee, A.; Qiu, Y.; Kisliuk, R.L. *J. Heterocyclic Chem.* **2004**, 41, 941 DOI: 10.1002/jhet.5570410613.

(t, $J = 4.8$ Hz, 4H), 2.63 (d, $J = 6.8$ Hz, 2H), 3.17 (s, 2H), 3.24 (d, $J = 8.0$ Hz, 2H), 3.63 (t, $J = 4.4$ Hz, 4H), 3.81 (s, 3H), 4.21 (d, $J = 6.4$ Hz, 2H), 6.84 (d, $J = 0.8$ Hz, 1H), 6.97 (d, $J = 8.4$ Hz, 1H), 7.08 (s, 1H), 7.22 (dd, $J = 4.0, 0.8$ Hz, 2H), 7.54-7.58 (m, 2H); LC-MS m/z 461 (M+1)⁺.

2-[2-(3,4-Dimethoxyphenyl)ethyl]-5-(3,5-dimethylisoxazol-4-yl)-1-(2-morpholin-4-ylethyl)-1H-benzimidazole (5). Prepared according to general procedure A using 4-(3,5-Dimethylisoxazol-4-yl)-*N*-(2-morpholinoethyl)-2-nitroaniline (52 mg, 0.15 mmol) and 3-(3,4-dimethoxyphenyl)propionaldehyde³ giving **5** (21.2 mg, 29%) as a brown solid. ¹H NMR (400 MHz, CDCl₃): δ 2.30 (s, 3H), 2.43 (s, 3H), 2.45-2.47 (m, 4H), 2.57 (t, $J = 6.8$ Hz, 2H), 3.18-3.25 (m, 4H), 3.67-3.69 (m, 4H), 3.72 (s, 3H), 3.86 (s, 3H), 4.08 (t, $J = 6.8$ Hz, 2H), 6.67 (s, 1H), 6.77-6.82 (m, 2H), 7.12 (d, $J = 8.4$ Hz, 1H), 7.34 (d, $J = 8.4$ Hz, 1H), 7.64 (s, 1H).

2-[2-(2,4-Dimethoxyphenyl)ethyl]-5-(3,5-dimethylisoxazol-4-yl)-1-(2-morpholin-4-ylethyl)-1H-benzimidazole (6). Prepared according to general procedure A using 4-(3,5-Dimethylisoxazol-4-yl)-*N*-(2-morpholinoethyl)-2-nitroaniline (107 mg, 0.31 mmol) and 3-(2',4'-dimethoxyphenyl)propionaldehyde⁴ giving **6** (6.3 mg, 5%) as a brown oil. ¹H NMR (400MHz, CD₃OD): δ 2.28 (s, 3H), 2.45 (s, 3H), 3.22 (s, 4H), 3.43 (s, 6H), 3.57 (s, 4H), 3.77 (s, 4H), 4.02 (s, 4H), 6.44-6.52 (m, 2H), 7.19 (s, 1H), 7.65-7.73 (m, 2H), 8.20 (s, 1H); LC-MS m/z 491 (M+1)⁺.

5-(3,5-Dimethylisoxazol-4-yl)-2-[(4-methoxyphenoxy)methyl]-1-(2-morpholin-4-ylethyl)-1H-benzimidazole (7). Prepared according to general procedure A using 4-(3,5-Dimethylisoxazol-4-yl)-*N*-(2-morpholinoethyl)-2-nitroaniline (93 mg, 0.27 mmol) and (4-methoxyphenoxy)acetaldehyde⁴ giving **7** (9 mg, 7%) as a colorless solid. ¹H NMR (400MHz, CD₃OD): δ 2.27 (s, 3H), 2.43 (s, 3H), 2.51 (m, 4H), 2.79-2.83 (m, 2H), 3.64 (m, 4H), 3.75 (s, 3H), 4.53-4.56 (m, 2H), 5.42 (s, 2H), 6.88 (d, $J = 8.8$ Hz, 2H), 7.04 (d, $J = 8.4$ Hz, 2H), 7.32 (d, $J = 8.0$ Hz, 1H), 7.60 (s, 1H), 7.69 (d, $J = 8.4$ Hz, 1H); LC-MS m/z 463 (M+1)⁺.

5-(3,5-Dimethylisoxazol-4-yl)-2-phenoxyethyl-1-(2-morpholin-4-ylethyl)-1H-benzimidazole (8). Prepared according to general procedure A using 4-(3,5-Dimethylisoxazol-4-yl)-*N*-(2-morpholinoethyl)-2-nitroaniline (127 mg, 0.37 mmol) and phenoxyacetaldehyde giving **8** (9 mg, 6 %) as a light brown solid. ¹H NMR (400MHz, CD₃OD): δ 2.29 (s, 3H), 2.46 (s, 3H), 3.48 (m, 4H), 3.76-3.80 (m, 2H), 4.04 (m, 4H), 5.12-5.15 (m, 2H), 5.83 (s, 2H), 7.11 (t, $J = 7.5$ Hz, 1H), 7.28-7.30 (m, 2H), 7.40-7.44 (m, 2H), 7.66 (d, $J = 8.0$ Hz, 1H), 7.80 (s, 1H), 8.24 (d, $J = 8.4$ Hz, 1H); LC-MS m/z 433 (M+1)⁺.

³ Ermolae'ev, D.S.; Alifanov, V.L.; Rybakov, V.B.; Babaev, E.V.; Van der Eycken, E.V. *Synthesis*, **2008**, 2083 DOI: 10.1055/s-2008-107844.

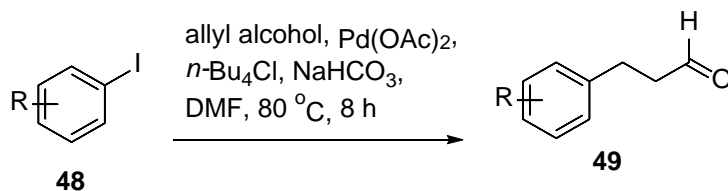
⁴ Piggott, M.J.; Wege, D. *Aust. J. Chem.* **2003**, 56, 691 DOI: 10.1071/CH02253.

4-{2-[5-(3,5-Dimethylisoxazol-4-yl)-1-(2-morpholin-4-ylethyl)-1H-benzimidazol-2-yl]ethyl}-phenol (**9**). A mixture of 4-(2-(5-(3,5-Dimethylisoxazol-4-yl)-2-(4-((tetrahydro-2H-pyran-2-yl)oxy)phenethyl)-1H-benzo[d]imidazol-1-yl)ethyl)morpholine (80 mg, 0.15 mmol) in ethanol (1.3 mL) was added concentrated HCl (0.76 mL). The mixture was stirred at ambient temperature for 4 hours. The resulting mixture was concentrated *in vacuo* and purified by prep-TLC to give **9** (55.8 mg, 83%) as a yellow solid. ¹H NMR (400MHz, CDCl₃): δ 2.27 (m, 3H), 2.40 (s, 3H), 2.49 (m, 4H), 2.64-2.65 (m, 2H), 3.18 (m, 4H), 3.69 (m, 4H), 4.15 (m, 2H), 6.73 (d, *J* = 7.6 Hz, 2H), 7.01 (d, *J* = 8.0 Hz, 2H), 7.12 (d, *J* = 7.6 Hz, 1H), 7.36 (d, *J* = 7.6 Hz, 1H), 7.59 (s, 1H), phenolic proton not observed; LC-MS *m/z* 447 (M+1)⁺.

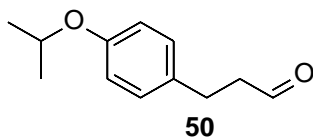
5-(3,5-Dimethylisoxazol-4-yl)-2-[2-(4-ethoxyphenyl)ethyl]-1-(2-morpholin-4-ylethyl)-1H-benzimidazole (**10**). A mixture of **9** (45 mg, 0.10 mmol) and iodoethane (19 mg, 0.12 mmol) and K₂CO₃ (41 mg, 0.30 mmol) in acetonitrile (2 mL) was stirred at 50 °C for 5 hours. The reaction mixture was filtered and purified by prep-HPLC to give **10** (1.6 mg, 3%) as a light yellow solid. ¹H NMR (400MHz, CDCl₃): δ 1.42 (t, *J* = 7.2 Hz, 3H), 2.31 (s, 3H), 2.44 (s, 3H), 2.46-2.49 (m, 4H), 2.60 (t, *J* = 7.2 Hz, 2H), 3.18-3.24 (m, 4H), 3.67-3.69 (m, 4H), 4.02 (q, *J* = 7.2 Hz, 2H), 4.07-4.13 (m, 2H), 6.84 (d, *J* = 8.8 Hz, 2H), 7.12-7.16 (m, 3H), 7.35 (d, *J* = 8.0 Hz, 1H), 7.64 (s, 1H); LC-MS *m/z* 475 (M+1)⁺.

5-(3,5-Dimethylisoxazol-4-yl)-1-(2-morpholin-4-ylethyl)-2-[2-(4-propoxyphenyl)ethyl]-1H-benzimidazole (**11**). A mixture of **9** (45 mg, 0.10 mmol) and iodopropane (20 mg, 0.12 mmol) and K₂CO₃ (41 mg, 0.30 mmol) in acetonitrile (2 mL) was stirred at 50 °C overnight. The reaction mixture was filtered and purified by prep-HPLC to give **11** (4.9 mg, 10%) as a yellow solid. ¹H NMR (400MHz, CDCl₃): δ 1.04 (t, *J* = 7.6 Hz, 3H), 1.80-1.83 (m, 2H), 2.31 (s, 3H), 2.44 (s, 3H), 2.47-2.49 (m, 4H), 2.58-2.61 (m, 2H), 3.18-3.24 (m, 4H), 3.67-3.69 (m, 4H), 3.91 (t, *J* = 6.8 Hz, 2H), 4.10-4.13 (t, *J* = 7.2 Hz, 2H), 6.85 (d, *J* = 8.8 Hz, 2H), 7.12-7.15 (m, 3H), 7.35 (d, *J* = 8.4 Hz, 1H), 7.64 (s, 1H); LC-MS *m/z* 489 (M+1)⁺.

General procedure B (Synthesis of 3-arylpropionaldehydes (**49**)):



Allyl alcohol (2.0 eq) was added to a solution of the aryl iodide (1.0 eq), Pd(OAc)₂ (0.25 eq), Bu₄NCl (1.33 eq) and NaHCO₃ (3.0 eq) in DMF (0.2M). The mixture was allowed to stir at 80 °C for 8 hours. The mixture was filtered and the resulting filtrate was concentrated *in vacuo*. The residue was purified by flash chromatography giving the desired 3-arylpropionaldehydes as described below.



3-(4-Isopropoxyphenyl)propionaldehyde (50): Prepared according to general procedure B using 4-(isopropoxy)iodobenzene (200 mg, 0.76 mmol). Purification by flash column chromatography (5-10% ethyl acetate in petroleum ether) gave 3-(4-Isopropoxyphenyl)propionaldehyde (55 mg, 37%) as yellow liquid which exactly matched the previous data provided for this compound.⁵ The material was used directly in the next step.

5-(3,5-Dimethylisoxazol-4-yl)-2-[2-(4-isopropoxyphenyl)ethyl]-1-(2-morpholin-4-ylethyl)-1H-benzimidazole (12). Prepared according to general procedure A using 4-(3,5-Dimethylisoxazol-4-yl)-N-(2-morpholinoethyl)-2-nitroaniline (468 mg, 1.35 mmol) and 3-(4-Isopropoxyphenyl)propionaldehyde giving **12** (5 mg, 4%) as a yellow solid. ¹H NMR (400MHz, CDCl₃): δ 1.33 (d, *J* = 6.0 Hz, 6H), 2.30 (s, 3H), 2.43 (s, 3H), 2.44-2.48 (m, 4H), 2.58-2.61 (m, 2H), 3.15-3.25 (m, 4H), 3.65-3.69 (m, 4H), 4.09-4.12 (m, 2H), 4.51 (sept. *J* = 6.0 Hz, 1H), 6.83 (d, *J* = 8.5 Hz, 2H), 7.10-7.14 (m, 3H), 7.35 (d, *J* = 8.0 Hz, 1H), 7.63 (s, 1H); LC-MS *m/z* 489 (M+1)⁺.

5-(3,5-Dimethylisoxazol-4-yl)-1-(2-morpholin-4-ylethyl)-2-[2-(4-butyloxyphenyl)ethyl]-1H-benzimidazole (13). To a mixture of **9** (0.075 mmol) and potassium *tert*-butoxide (0.15 mmol) in DMA (0.7 mL) was added iodobutane (0.11 mmol). The mixture was allowed to stir at 60 °C for 1 hour. The reaction mixture was cooled to room temperature concentrated *in vacuo*. The residue was purified by prep-HPLC to give **13**. LC-MS *m/z* 502 (M+1)⁺.

5-(3,5-Dimethylisoxazol-4-yl)-1-(2-morpholin-4-ylethyl)-2-[2-(4-pentyloxyphenyl)ethyl]-1H-benzimidazole (14). To a mixture of **9** (0.075 mmol) and potassium *tert*-butoxide (0.15 mmol) in DMA (0.7 mL) was added bromopentane (0.11 mmol). The mixture was allowed to stir at 60 °C for 1 hour. The reaction mixture was cooled to room temperature concentrated *in vacuo*. The residue was purified by prep-HPLC to give **14**. LC-MS *m/z* 517 (M+1)⁺.

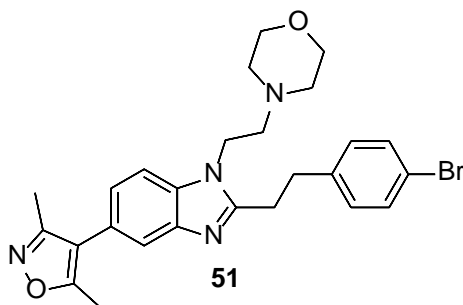
5-(3,5-Dimethylisoxazol-4-yl)-1-(2-morpholin-4-ylethyl)-2-[2-[4-(trifluoromethoxy)phenyl]-ethyl]-1H-benzimidazole (15). Prepared according to general procedure A using 4-(3,5-Dimethylisoxazol-4-yl)-N-(2-morpholinoethyl)-2-nitroaniline (72 mg, 0.21 mmol) and 3-(4-trifluoromethoxyphenyl)propionaldehyde⁶ giving **15** (18.2 mg, 17%) as yellow oil. ¹H NMR (400MHz,

⁵ Khan, A. M. *et al* PCT2004034999 29 Apr 2004

⁶ Kim, P.; Kang, S.; Boshoff, H.I.; Jiricek, J.; Collins, M.; Singh, R.; Manjunatha, U.H.; Niyomrattanakit, P.; Zhang, L.; Goodwin, M.; Dick, T.; Keller, T.H.; Dowd, C.S.; Barry, C.E. *J. Med. Chem.* **2009**, 52, 1329 DOI: 10.1021/jm801374t.

CDCl₃): δ 2.29 (s, 3H), 2.42 (s, 3H), 2.43-2.46 (m, 4H), 2.60 (t, J = 6.8 Hz, 2H), 3.17-3.20 (m, 2H), 3.29-3.33 (m, 2H), 3.64-3.66 (m, 4H), 4.12 (t, J = 6.8 Hz, 2H), 7.11-7.15 (m, 3H), 7.21 (s, 2H), 7.34 (d, J = 8.4 Hz, 1H), 7.62 (s, 1H).

4-{2-[5-(3,5-Dimethylisoxazol-4-yl)-1-(2-morpholin-4-ylethyl)-1H-benzimidazol-2-yl]ethyl}-benzonitrile (**16**). Prepared according to general procedure A using 4-(3,5-Dimethylisoxazol-4-yl)-N-(2-morpholinoethyl)-2-nitroaniline (42 mg, 0.12 mmol) and 4-(3-oxopropyl)benzonitrile⁷ giving **16** (20 mg, 36%) as a brown solid. ¹H NMR (400MHz, CDCl₃): δ 2.30 (s, 3H), 2.43 (s, 3H), 2.49 (s, 4H), 2.66 (t, J = 6.8 Hz, 2H), 3.22-3.26 (m, 2H), 3.38-3.42 (m, 2H), 3.67-3.69 (m, 4H), 4.18 (t, J = 6.8 Hz, 2H), 7.15 (d, J = 8.4 Hz, 1H), 7.37-3.40 (m, 3H), 7.60-7.64 (m, 3H).



2-[2-(4-Bromophenyl)ethyl]-5-(3,5-dimethylisoxazol-4-yl)-1-(2-morpholin-4-ylethyl)-1H-benzimidazole (**51**): Prepared according to general procedure A using 4-(3,5-Dimethylisoxazol-4-yl)-N-(2-morpholinoethyl)-2-nitroaniline (541 mg, 1.60 mmol) and 3-(4-bromophenyl)propionaldehyde⁸ giving 2-[2-(4-Bromophenyl)ethyl]-5-(3,5-dimethylisoxazol-4-yl)-1-(2-morpholin-4-ylethyl)-1H-benzimidazole (400 mg, 50%) as a light brown solid. ¹H NMR (400MHz, CDCl₃): δ 2.30 (s, 3H), 2.43 (s, 3H), 2.45-2.47 (m, 4H), 2.57-2.62 (m, 2H), 3.16-3.20 (m, 2H), 3.26-3.29 (m, 2H), 3.66-3.68 (m, 4H), 4.11-4.14 (m, 2H), 7.11-7.14 (m, 3H), 7.35 (d, J = 8.4 Hz, 1H), 7.42 (d, J = 8.4 Hz, 2H), 7.63 (s, 1H); LC-MS m/z 509, 511 (M+1)⁺.

General Procedure C (Synthesis of N-arylated systems, e. g. **18**):

4-{2-[5-(3,5-Dimethylisoxazol-4-yl)-1-(2-morpholin-4-ylethyl)-1H-benzimidazol-2-yl]ethyl}-N,N-dimethylaniline (**18**): A mixture of 2-[2-(4-Bromophenyl)ethyl]-5-(3,5-dimethylisoxazol-4-yl)-1-(2-

⁷ Colbon, P.; Ruan, J.; Purdie, M.; Mulholland, K.; Xiao, J. *Org. Lett.* **2011**, *13*, 5456 DOI: 10.102/ol202144z.

⁸ Liu, J.; He, S.; Jian, T.; Dobbelaar, P.H.; Sebhat, I.K.; Lin, L.S.; Goodman, A.; Guo, C.; Guzzo, P.R.; Hadden, M.; Henderson, A.J.; Pattamana, K.; Ruenz, M.; Sargent, B.J.; Swenson, B.; Yet, L.; Tamvakopoulos, C.; Peng, Q.; Pan, J.; Kan, Y.; Palyha, O.; Kelly, T.M.; Guan, X.-M.; Howard, A.D.; Marsh, D.J.; Metzger, J.M.; Reitman, M.L.; Wyvratt, M.J.; Nargund, R.P. *Bioorg. Med. Chem. Lett.* **2010**, *20*, 2074 DOI: 10.1016/j.bmcl.2010.02.076.

morpholin-4-ylethyl)-1*H*-benzimidazole (**51**) (26 mg, 0.05 mmol), Pd(OAc)₂ (1.1 mg, 0.005 mmol), Ru-Phos (3.7 mg, 0.008 mmol), dimethylammonium chloride (25 mg, 0.31 mmol) and sodium *tert*-butoxide (44 mg, 0.46 mmol) in THF (0.4 mL) was heated at 70 °C under nitrogen overnight. The reaction mixture was cooled to ambient temperature, diluted with THF and partitioned between ethyl acetate and ammonium chloride solution. The layers were separated and aqueous was extracted with ethyl acetate. The combined extracts were washed with brine. Purification of the crude residue by prep-HPLC gave **18** (12.8 mg, 54%) as a colorless solid. ¹H NMR (400MHz, CDCl₃): δ 2.30 (s, 3H), 2.43 (s, 3H), 2.46-2.49 (m, 4H), 2.58 (t, *J* = 6.8 Hz, 2H), 2.92 (s, 6H), 3.16-3.20 (m, 4H), 3.68-3.70 (m, 4H), 4.13 (t, *J* = 6.8 Hz, 2H), 6.70 (d, *J* = 8.2 Hz, 2H), 7.09-7.13 (m, 3H), 7.36 (d, *J* = 8.2 Hz, 1H), 7.64 (s, 1H); LC-MS *m/z* 474 (M+1)⁺.

5-(3,5-Dimethylisoxazol-4-yl)-2-[2-(4-aminophenyl)ethyl]-1-(2-morpholin-4-ylethyl)-1*H*-benzimidazole (**17**). Prepared according to general procedure C using ammonium chloride (500 mg, 94%) as a yellow solid: ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.68 – 7.57 (m, 5H), 7.31 (dd, *J* = 14.2, 8.1 Hz, 3H), 7.15 (s, 1H), 7.04 (d, *J* = 21.2 Hz, 3H), 4.60 (t, *J* = 7.7 Hz, 2H), 4.13 – 4.00 (m, 5H), 3.25 (d, *J* = 7.9 Hz, 2H), 3.13 (t, *J* = 7.9 Hz, 2H), 2.34 (s, 3H), 2.17 (s, 3H), 2.02 (s, 1H), 1.55 (q, *J* = 6.0 Hz, 2H), 1.35 – 1.14 (m, 21H), 0.85 – 0.72 (m, 15H).

N-(4-{2-[5-(3,5-Dimethylisoxazol-4-yl)-1-(2-morpholin-4-ylethyl)-1*H*-benzimidazol-2-yl]-ethyl}phenyl)acetamide (**19**). Prepared according to general procedure C. The reaction mixture was concentrated and the residue was purified by prep-HPLC to give **19** (34.7 mg, 65%) as a colorless solid. ¹H NMR (400 MHz, CDCl₃): δ 2.18 (s, 3H), 2.30 (s, 3H), 2.43 (s, 3H), 2.46-2.49 (m, 4H), 2.59-2.62 (m, 2H), 3.16-3.28 (m, 4H), 3.66-3.69 (m, 4H), 4.11-4.14 (m, 2H), 7.12 (dd, *J* = 8.5, 1.5 Hz, 1H), 7.18-7.20 (m, 3H), 7.36 (d, *J* = 8.0 Hz, 1H), 7.42 (d, *J* = 8.5 Hz, 2H), 7.63 (s, 1H); LC-MS *m/z* 488 (M+1)⁺.

3-(4-{2-[5-(3,5-Dimethylisoxazol-4-yl)-1-(2-morpholin-4-ylethyl)-1*H*-benzimidazol-2-yl]-ethyl}phenoxy)-*N,N*-dimethylpropan-1-amine (**20**). To a mixture of **9** (30 mg, 0.067 mmol) and potassium *tert*-butoxide (22.5 mg, 0.20 mmol) in DMA (2.5 mL) was added *N*-(3-bromopropyl)dimethylamine (16.6 mg, 0.10 mmol). The mixture was allowed to stir at 60 °C for 2 hours. The reaction mixture was cooled to room temperature and filtered. The filtrate was purified by prep-HPLC to give **20** (8.1 mg, 23 %) as colorless solid. ¹H NMR (400MHz, CD₃OD): δ 2.16-2.23 (m, 2H), 2.27 (s, 3H), 2.42 (s, 3H), 2.44-2.46 (m, 4H), 2.56 (t, *J* = 6.5 Hz, 2H), 2.91 (s, 6H), 3.15-3.27 (m, 4H), 3.31-3.34 (m, 2H), 3.61-3.63 (m, 4H), 4.07 (t, *J* = 5.5 Hz, 2H), 4.22 (t, *J* = 6.3 Hz, 2H), 6.86 (d, *J* = 8.5 Hz, 2H), 7.15 (d, *J* = 8.5 Hz, 2H), 7.22 (d, *J* = 8.0 Hz, 1H), 7.54-7.58 (m, 2H); LC-MS *m/z* 532 (M+1)⁺.

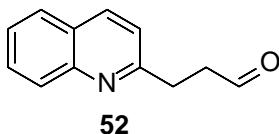
5-(3,5-Dimethylisoxazol-4-yl)-1-(2-morpholin-4-ylethyl)-2-(2-pyridin-2-ylethyl)-1*H*-benzimidazole (**21**). Prepared according to general procedure A using 4-(3,5-Dimethylisoxazol-4-yl)-*N*-(2-

morpholinoethyl)-2-nitroaniline (375 mg, 1.08 mmol) and 2-pyridinyl-propionaldehyde⁹ giving **21** (74 mg, 16%) as a yellow oil. ¹H NMR (400MHz, CDCl₃): δ 2.27 (s, 3H), 2.40 (s, 3H), 2.49-2.50 (m, 4H), 2.67 (t, *J* = 6.8 Hz, 2H), 3.38-3.48 (m, 4H), 3.65-3.67 (m, 4H), 2.24 (t, *J* = 6.8 Hz, 2H), 7.09-7.15 (m, 2H), 7.22 (d, *J* = 8.0 Hz, 1H), 7.34 (d, *J* = 8.4 Hz, 1H), 7.56-7.60 (m, 2H), 8.54 (d, *J* = 4.4 Hz, 1H); LC-MS *m/z* 432 (M+1)⁺.

5-(3,5-Dimethylisoxazol-4-yl)-1-(2-morpholin-4-ylethyl)-2-(2-pyridin-3-ylethyl)-1H-benzimidazole (**22**). Prepared according to general procedure A using 4-(3,5-Dimethylisoxazol-4-yl)-*N*-(2-morpholinoethyl)-2-nitroaniline (332 mg, 0.96 mmol) and 3-pyridinyl-propionaldehyde¹⁰ giving **22** (183 mg, 37%) as a yellow oil. ¹H NMR (400MHz, CDCl₃): δ 2.29 (3H), 2.42 (s, 3H), 2.44-2.46 (m, 4H), 2.61 (t, *J* = 6.8 Hz, 2H), 3.19-3.24 (m, 2H), 3.32-3.36 (m, 2H), 3.64-3.66 (m, 4H), 4.14 (t, *J* = 6.8 Hz, 2H), 7.11-7.14 (m, 1H), 7.22-7.26 (m, 1H), 7.35 (d, *J* = 8.0 Hz, 1H), 7.58 (d, *J* = 8.0 Hz, 1H), 7.62 (d, *J* = 0.8 Hz, 1H), 8.47-8.49 (m, 1H), 8.53 (d, *J* = 1.6 Hz, 1H); LC-MS *m/z* 432 (M+1)⁺.

5-(3,5-Dimethylisoxazol-4-yl)-1-(2-morpholin-4-ylethyl)-2-(2-pyridin-4-ylethyl)-1H-benzimidazole (**23**). Prepared according to general procedure A using 4-(3,5-Dimethylisoxazol-4-yl)-*N*-(2-morpholinoethyl)-2-nitroaniline (498 mg, 1.44 mmol) and 4-pyridinyl-propionaldehyde¹⁰ giving **23** (159 mg, 21%) as a yellow oil. ¹H NMR (400MHz, CDCl₃): δ 2.29 (s, 3H), 2.43 (s, 3H), 2.45-2.48 (m, 4H), 2.65 (t, *J* = 6.8 Hz, 2H), 3.21-3.25 (m, 2H), 3.33-3.37 (m, 2H), 3.65-3.67 (m, 4H), 4.15-4.18 (m, 2H), 7.13-7.15 (m, 1H), 7.20 (d, *J* = 6.0 Hz, 2H), 7.25-7.35 (m, 1H), 7.63 (d, *J* = 0.8 Hz, 1H), 8.53 (d, *J* = 6.0 Hz, 2H); LC-MS *m/z* 432 (M+1)⁺.

5-(3,5-Dimethylisoxazol-4-yl)-2-[2-(1H-indol-3-yl)ethyl]-1-(2-morpholin-4-ylethyl)-1H-benzimidazole (**24**). Prepared according to general procedure A using 4-(3,5-Dimethylisoxazol-4-yl)-*N*-(2-morpholinoethyl)-2-nitroaniline (35 mg, 0.10 mmol) and 2-(1H-indol-3-yl)propionaldehyde¹⁰ giving **24** (23 mg, 49%) as a yellow solid. ¹H NMR (400MHz, CD₃OD): δ 2.30 (s, 3H), 2.47 (s, 3H), 2.70-3.20 (m, 6H), 3.52-3.54 (m, 2H), 3.72-3.74 (m, 2H), 3.91 (m, 4H), 4.61-4.63 (m, 2H), 7.04-7.07 (m, 2H), 7.18 (m, 1H), 7.44 (d, *J* = 8.4 Hz, 1H), 7.56 (d, *J* = 8.0 Hz, 1H), 7.61 (d, *J* = 8.8 Hz, 1H), 7.78 (s, 1H), 8.12 (d, *J* = 4.8 Hz, 1H), indole NH not observed; LC-MS *m/z* 470 (M+1)⁺.

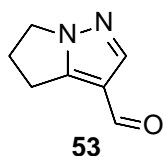


⁹ Kitbunnadaj, R.; Zuiderveld, O.P.; Christophe, B.; Hulscher, S.; Menge, W.; Gelens, E.; Snip, E.; Bakker, R.A.; Celanire, S.; Gillard, M.; Talaga, P.; Timmerman, H.; Leurs, R. *J. Med. Chem.* **2004**, *47*, 2414 DOI: 10.1021/jm049932u.

¹⁰ Hobley, G.; McKelvie, J.C.; Harmer, J.E.; Howe, J.; Oyston, P.C.F.; Roach, P.L. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 3079 DOI: 10.1016/j.bmcl.2012.03.072.

3-(*Quinolin-2-yl*)propionaldehyde (**52**): Prepared according to general procedure B using 2-bromoquinoline (1.7 g, 8.17 mmol). The crude aldehyde (500 mg, 33%) obtained as a yellow oil was used directly in the next reaction.

2-{2-[5-(3,5-Dimethylisoxazol-4-yl)-1-(2-morpholin-4-ylethyl)-1*H*-benzimidazol-2-yl]ethyl}quinoline (**25**). Prepared according to general procedure A using 3-(*Quinolin-2-yl*)propionaldehyde (500 mg, 2.70 mmol) giving **25** (447 mg) as a brown oil. LC-MS *m/z* 482 (*M*+1)⁺.



5,6-Dihydro-4*H*-pyrrolo[1,2-*b*]pyrazole-3-carboxaldehyde (**53**): To a mixture of 5,6-dihydro-4*H*-pyrrolo[1,2-*b*]pyrazole¹¹ (55.5 g, 0.51 mol), DMF (76 g, 1.04 mol) in 1,2-dichloroethane (550 mL) was added POCl₃ (160 g, 1.05 mol) dropwise at 4~7 °C. The reaction was heated to reflux for 4 hours, and then allowed to stir at room temperature overnight. The mixture was poured into a mixture of ice water (2.4 L) and anhydrous sodium acetate (267 g) and stirred at 25 °C for 1 hour. The organic layer was separated and the aqueous layer was extracted with 1,2-dichloroethane (6 × 150 mL). The combined organic layers were washed with saturated NaHCO₃ (2 × 200 mL), dried over Na₂SO₄ and concentrated *in vacuo*. Pre-purification by column chromatography (10-50% ethyl acetate/petroleum ether), followed by re-crystallization from *i*-PrOH at -35 °C afforded pure 5,6-Dihydro-4*H*-pyrrolo[1,2-*b*]pyrazole-3-carboxaldehyde (35 g, 45.5%) as a yellow liquid at ambient temperature. ¹H NMR (300 MHz, CDCl₃): δ 2.71 (m, 2H), 3.12 (t, *J* = 7.2 Hz, 2H), 4.18 (t, *J* = 7.2 Hz, 2H), 7.93 (s, 1H); LC-MS *m/z* 137 (*M*+1)⁺.

4-(2-(5-(3,5-dimethylisoxazol-4-yl)-2-(2-(*isoquinolin-4-yl*)ethyl)-1*H*-benzo[*d*]imidazol-1-yl)ethyl)morpholine (**26**). Prepared according to general procedure A giving **26** (55 mg, 78%) as a yellow oil. ¹H NMR (400 MHz, CDCl₃) 2.31 (s, 3H), 2.33-2.35 (m, 4H), 2.44 (s, 3H), 2.52 (dd, *J* = 7.0, 6.6 Hz, 2H), 3.34-3.38 (m, 2H), 3.57-3.59 (m, 4H), 3.73-3.77 (m, 2H), 4.04 (t, *J* = 6.0 Hz, 2H), 7.13 (dd, *J* = 8.2, 1.6 Hz, 1H), 7.33 (d, *J* = 8.2 Hz, 1H), 7.63-7.67 (m, 2H), 7.75 (ddd, *J* = 8.2, 7.0, 1.3 Hz, 1H), 8.03 (d, *J* = 8.2 Hz, 1H), 8.12 (d, *J* = 8.2 Hz, 1H), 8.43 (s, 1H), 9.18 (s, 1H); HR-MS (*M*+H)⁺ *m/z* calc'd. 482.2551; found 482.2550.

2-(5,6-dihydro-4*H*-pyrrolo[1,2-*b*]pyrazol-3-yl)-5-(3,5-dimethylisoxazol-4-yl)-1-(2-morpholin-4-ylethyl)-1*H*-benzimidazole (**27**). Prepared according to general procedure A using 4-(3,5-Dimethylisoxazol-4-yl)-*N*-(2-morpholinoethyl)-2-nitroaniline (2.42 g, 7.0 mmol) and 5,6-Dihydro-4*H*-

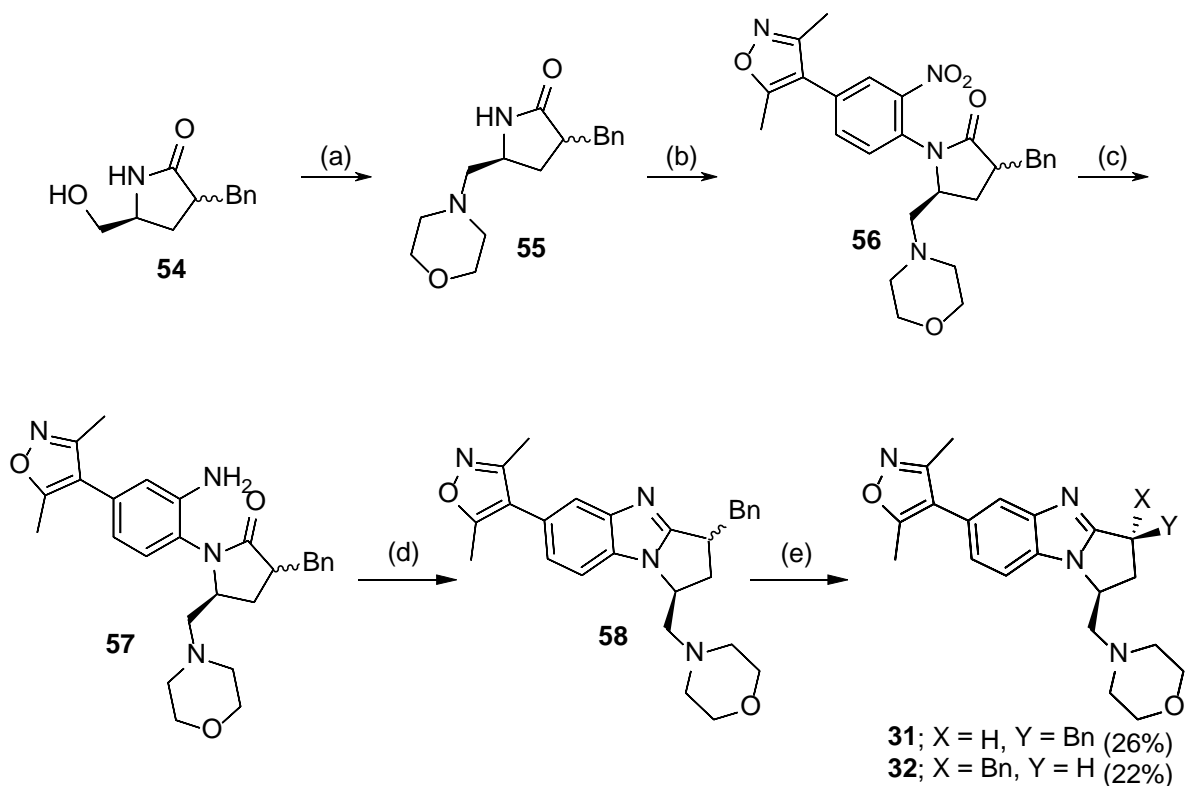
¹¹ Kulinkovich, O.; Masalov, N.; Tyvorskii, V.; De Kimpe, N.; Keppens, M. *Tetrahedron Lett.* **1996**, 37, 1095 DOI: 10.1016/0040-4039(95)02313-5.

pyrrolo[1,2-*b*]pyrazole-3-carboxaldehyde (2.95 g, 21.7 mmol) giving **27** (1.2 g, 40%) as a colorless solid. ¹H NMR (400MHz, CDCl₃): δ 2.31 (s, 3H), 2.44 (s, 3H), 2.51-2.54 (m, 4H), 2.72-2.76 (m, 2H), 2.84 (t, *J* = 8.2 Hz, 2H), 3.26 (t, *J* = 7.2 Hz, 2H), 3.68-3.71 (m, 4H), 4.27 (t, *J* = 7.2 Hz, 2H), 4.46 (t, *J* = 7.2 Hz, 2H), 7.14 (dd, *J* = 8.4, 1.6 Hz, 1H), 7.44 (d, *J* = 8.0 Hz, 1H), 7.62 (d, *J* = 0.8 Hz, 1H), 8.00 (s, 1H).

2-(5-(3,5-dimethylisoxazol-4-yl)-2-phenethyl-1H-benzo[d]imidazol-1-yl)-1-morpholinoethan-1-one (**28**). Prepared according to general procedure A giving **28** (13 mg, 21%) as a white solid. ¹H NMR (400MHz, CDCl₃): δ 2.31 (s, 3H), 2.44 (s, 3H), 3.26 (t, *J* = 7.2 Hz, 2H), 3.18-3.71 (m, 4H), 4.27 (t, *J* = 7.2 Hz, 2H), 4.46 (t, *J* = 7.2 Hz, 2H), 7.14 (dd, *J* = 8.4, 1.6 Hz, 1H), 7.44 (d, *J* = 8.0 Hz, 1H), 7.62 (d, *J* = 0.8 Hz, 1H), 8.00 (s, 1H).

*4-(2-(5-(3,5-dimethylisoxazol-4-yl)-2-((2*S*,3*R*)-2-(4-methoxyphenyl)tetrahydrofuran-3-yl)-1H-benzo[d]imidazol-1-yl)ethyl)morpholine* (**29**). Prepared according to general procedure A as a racemate which was then separated using chiral SFC conditions (Lux cellulose-3 250 mm x 21.2 mm, 5 μm, 80 mL/min, first eluting peak, stereochemistry arbitrarily assigned) giving **29** (45 mg, 17%) as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ ppm 2.14 (d, *J*=7.02 Hz, 1 H) 2.21 - 2.34 (m, 8 H) 2.41 (s, 4 H) 3.45 (t, *J*=4.49 Hz, 4 H) 3.56 - 3.66 (m, 1 H) 3.71 (s, 3 H) 3.92 - 4.05 (m, 1 H) 4.05 - 4.29 (m, 3 H) 5.14 (d, *J*=8.59 Hz, 1 H) 6.87 (d, *J*=8.98 Hz, 2 H) 7.18 (dd, *J*=8.20, 1.56 Hz, 1 H) 7.24 (d, *J*=8.98 Hz, 2 H) 7.55 (d, *J*=8.59 Hz, 1 H) 7.63 (d, *J*=1.56 Hz, 1 H); the second eluting peak (**30**) under these chiral SFC conditions was recovered as 30 mg (11%) as a white solid having an identical ¹H NMR signature.

General Approach to compounds 31-32



(a) $\text{PPh}_3(\text{resin})$, I_2 , morpholine, TEA, 50 °C, 16 h; 56% (b) 4-(4-fluoro-3-nitrophenyl)-3,5-dimethylisoxazole, Cs_2CO_3 , acetone, 90 °C, 20 h; 11% (c) $\text{Na}_2\text{S}_2\text{O}_4$, aq. EtOH, 80 °C, 16 h; 99% (d) AcOH, DMF, 150 °C, 2 h; 56% (e) SFC separation

Step 1. (5S)-3-benzyl-5-(morpholinomethyl)pyrrolidin-2-one (55): To a solution of benzyl-5-(hydroxymethyl)pyrrolidin-2-one¹² (**54**) (1.2 g, 5.9 mmol) in THF (25 mL) was added solid-supported triphenylphosphine (70.8 g, 23.6 mmol, 3 mmol/g loading), I_2 (3 g, 11.8 mmol), triethylamine (3 g, 29.5 mmol) and morpholine (5.1 g, 59 mmol) and the reaction was stirred for 16 hours at 50 °C under nitrogen atmosphere. The reaction was allowed to cool to room temperature and filtered. The filtrate was concentrated in vacuum and the residue was purified by flash chromatography (0~7 % of MeOH in DCM) to give the titled compound (900 mg, 56%) as a colorless oil: LCMS m/z [$\text{M} + \text{H}^+$]: 274.9.

Step 2. (5S)-3-benzyl-1-(4-(3,5-dimethylisoxazol-4-yl)-2-nitrophenyl)-5-(morpholinomethyl)pyrrolidin-2-one (56): To a mixture of 3-benzyl-5-(morpholinomethyl)pyrrolidin-2-one (600 mg, 2.18 mmol), 4-(4-fluoro-3-nitrophenyl)-3,5-dimethylisoxazole¹³ (600 mg, 2.54 mmol) and cesium carbonate (3.6 g,

¹² Beard, M. J.; Bailey, J. H.; Cherry, D. T.; Moloney, M. G.; Shim, S. B.; Statham, K. A.; Bamford, M. J.; Lamont, R. B. *Tetrahedron*, **1996**, 52, 3719-40.

¹³ Hay, D. A.; Fedorov, O.; Martin, S.; Singleton, D. C.; Tallant, C.; Wells, C.; Picaud, S.; Philpott, M.; Monteiro, O. P.; Rogers, C. M., *et al. J. Am. Chem. Soc.* **2014**, 136, 9308-19.

11 mmol) in acetone (20 mL) was stirred for 20 hours at 90 °C. The reaction was then allowed to cool to room temperature and the mixture was filtered through a pad of silica. The filtrate was concentrated under reduced pressure to give the titled compound (120 mg, 56 %) as colorless oil which was taken forward without any further purification or characterization.

Step 3. (5*S*)-1-(2-amino-4-(3,5-dimethylisoxazol-4-yl)phenyl)-3-benzyl-5-(morpholinomethyl)pyrrolidin-2-one (**57**). A mixture of (5*S*)-3-benzyl-1-(4-(3,5-dimethylisoxazol-4-yl)-2-nitrophenyl)-5-(morpholinomethyl)pyrrolidin-2-one (100 mg, 0.2 mmol) and Na₂S₂O₄ (174 mg, 1 mmol) in EtOH/H₂O (4 mL/1 mL) was stirred at 80 °C for 16 hours. The reaction mixture was then concentrated under reduced pressure and the crude residue (100 mg) was taken forward without any further purification or characterization.

Step 4. 4-(((1*S*)-3-benzyl-6-(3,5-dimethylisoxazol-4-yl)-2,3-dihydro-1*H*-benzo[*d*]pyrrolo[1,2-*a*]imidazol-1-yl)methyl)morpholine (**58**). A mixture of (5*S*)-1-(2-amino-4-(3,5-dimethylisoxazol-4-yl)phenyl)-3-benzyl-5-(morpholinomethyl)pyrrolidin-2-one (100 mg, 0.2 mmol) in DMF/AcOH (10 mL / 0.5 mL) was heated in microwave vessel for 2 hours at 150 °C. The mixture was then diluted with water (10 mL) and extracted three times with ethyl acetate (20 mL). The combined organic layer was dried over Na₂SO₄, filtered and concentrated under reduced pressure. The crude residue was purified by flash silica chromatography (1:1 petroleum ether:EtOAc) to give a mixture of the titled compounds (50 mg, 56%) as a yellow solid: LCMS *m/z* [M + H⁺]: 443.2.

Step 5: 4-(((1*S*,3*S*)-3-benzyl-6-(3,5-dimethylisoxazol-4-yl)-2,3-dihydro-1*H*-benzo[*d*]pyrrolo[1,2-*a*]imidazol-1-yl)methyl)morpholine (**31**) and 4-(((1*S*,3*R*)-3-benzyl-6-(3,5-dimethylisoxazol-4-yl)-2,3-dihydro-1*H*-benzo[*d*]pyrrolo[1,2-*a*]imidazol-1-yl)methyl)morpholine (**32**): The mixture of titled compounds (50 mg) was purified by SFC chromatography using the following conditions: Chiralpak-AD (250x30mm) 20 mm, 25% MeOH (NH₄OH), 80 mL/min. Two peaks eluted which were resolved using the following conditions: Chiralpak AD-3 (50x4.6mm), 3mm, MeOH (0.05% DEA) in CO₂ (5-40%), 4 mL/min, 4 min.

The first peak eluted at 1.51 min and was assigned as 4-(((1*S*,3*R*)-3-benzyl-6-(3,5-dimethylisoxazol-4-yl)-2,3-dihydro-1*H*-benzo[*d*]pyrrolo[1,2-*a*]imidazol-1-yl)methyl)morpholine (**7**): LCMS *m/z* [M + H⁺]: 443.2; ¹HNMR (CDCl₃) δ 7.67-7.65 (m, 1H), 7.61 (s, 1H), 7.32-7.29 (m, 2H), 7.26-7.21 (m, 3H), 7.10-7.08 (m, 1H), 4.45 (br s, 1H), 3.75-3.65 (m, 5H), 3.56-3.53 (m, 1H), 2.84-2.71 (m, 1H), 2.55-2.40 (m, 10H), 2.27 (s, 3H).

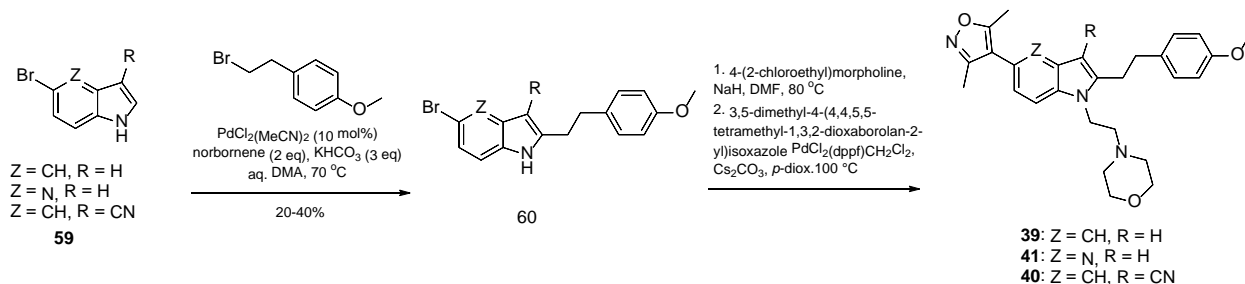
The second peak eluted at 1.66 min and was assigned as 4-(((1*S*,3*S*)-3-benzyl-6-(3,5-dimethylisoxazol-4-yl)-2,3-dihydro-1*H*-benzo[*d*]pyrrolo[1,2-*a*]imidazol-1-yl)methyl)morpholine (**6**): LCMS *m/z* [M + H⁺]: 443.2; ¹HNMR (CDCl₃) δ 7.89-7.87 (m, 1H), 7.61 (s, 1H), 7.30-7.22 (m, 5H), 7.10-7.09 (m, 1H), 4.52-4.49 (m, 1H), 3.79-3.77 (m, 4H), 3.57-3.53 (m, 2H), 2.98-2.97 (m, 1H), 2.77-2.66 (m, 1H), 2.65-2.63 (m, 1H), 2.59-2.57 (m, 3H), 2.47-2.40 (m, 6H), 2.30 (s, 3H).

Preparation of compounds 33-38

Derivatives **33-38** prepared in library format using General Procedure A (benzimidazole formation) substituting the appropriate boronic acid/pinacol borane, arylpropionic aldehyde, and o-nitrofluoroarene within the specified sequence to produce compounds **33-38** as exemplified by structure **33** below:

4-(2-(5-(3-ethyl-5-methylisoxazol-4-yl)-2-phenethyl-1H-benzo[d]imidazol-1-yl)ethyl)morpholine (**33**)
LCMS m/z [M + H]⁺: 445.2.

General Approach to the Construction of indoles/azaindoles 39-41.



Step 1: 5-bromo-2-(4-methoxyphenethyl)-1H-pyrrolo[3,2-b]pyridine.¹⁴ (**60**): To a RB flask charged with 5-bromo-4-azaindole (250 mg, 1.27 mmol), norbornene (239 mg, 2.54 mmol), potassium bicarbonate (381 mg, 3.81 mmol), and bis(acetonitrile)dichloropalladium (ii) (65.9 mg, 0.254 mmol) was added 4 mL of aq DMA (1N). The mixture was sparged with nitrogen gas for 2-3 minutes prior to the dropwise addition of 1-(2-bromoethyl)-4-methoxybenzene (409 mg, 1.90 mmol) over the course of 1 minute. The mixture was then swept with nitrogen gas and then heated to 70 °C for 20 h. The mixture was then cooled to rt, diluted with ethyl acetate (10 mL) and then washed three times with water. The organics were collected, washed with brine, dried with sodium sulfate, filtered, and concentrated under reduced pressure to give a brown oily residue which was purified by flash chromatography (0-100% EtOAc in Heptane over 25 mins) to give 84 mg (20%) of the titled compound as a brown oil: ¹H NMR (400 MHz, Chloroform-*d*) δ 7.61 (d, *J* = 8.3 Hz, 1H), 7.16 (d, *J* = 8.3 Hz, 2H), 7.07 (d, *J* = 8.3 Hz, 1H), 6.89 – 6.82 (m, 2H), 6.54 (s, 1H), 4.15 (t, *J* = 7.1 Hz, 2H), 3.80 (s, 3H), 3.68 (t, *J* = 4.5 Hz, 4H), 3.09 (s, 4H), 2.57 (d, *J* = 21.1 Hz, 5H), 2.44 (d, *J* = 16.9 Hz, 7H), 1.91 (s, 2H), 1.26 (d, *J* = 5.7 Hz, 2H), 1.24 (s, 13H). LCMS m/z [M + H]⁺: 332.9.

Steps 2 & 3: 4-(2-(5-(3,5-dimethylisoxazol-4-yl)-2-(4-methoxyphenethyl)-1H-pyrrolo[3,2-b]pyridin-1-yl)ethyl)morpholine (**41**): To an oven-dried RB flask charged with DMF (2 mL) was added 60% sodium hydride in dispersion oil (29.0 mg, 0.725 mmol) at RT. To this mixture was then dropwise added a solution of 5-bromo-2-(4-methoxyphenethyl)-1H-pyrrolo[3,2-b]pyridine (40 mg, 0.12 mmol) in 1 mL of DMF at rt, and subsequently 4-(2-chloroethyl)morpholine (67.4 mg, 0.362 mmol). The mixture was allowed to stir at rt for five minutes prior to heating to 80 °C for 45 minutes. The mixture was then cooled to rt and poured into water. The mixture was then extracted three times with ethyl acetate and the combined organics were washed with water, brine, dried with sodium sulfate, filtered, and concentrated under reduced pressure to give 12 mg (22%) of 4-(2-(5-bromo-2-(4-methoxyphenethyl)-1H-pyrrolo[3,2-b]pyridin-1-yl)ethyl)morpholine as a yellow oil: ¹H NMR (400 MHz, Chloroform-*d*) δ 7.95 (s, 1H), 7.35 (d, *J* = 8.5 Hz, 3H), 7.14 – 7.01 (m, 9H), 6.80 – 6.72 (m, 6H), 6.39 (s, 3H), 4.01 (s, 1H), 3.72 (s, 8H), 3.58 (s, 11H), 2.98 (q, *J* = 3.3 Hz, 12H), 2.88 (s, 3H), 2.81 (s, 3H), 2.47 (t, *J* = 6.9 Hz, 6H), 2.33 (s, 11H). This product was immediately carried forward to the next step in the synthetic sequence without further purification or characterization.

¹⁴ For studies related to the C-H insertion chemistry employed to furnish these systems, see: Jiao, L.; Bach, T. *J. Am. Chem. Soc.* **2012**, *134*, 14563-72.

To a RB flask charged with 8 mg (0.02 mmol) of 4-(2-(5-bromo-2-(4-methoxyphenethyl)-1H-pyrrolo[3,2-b]pyridin-1-yl)ethyl)morpholine from the previous synthetic operation was added 0.5 mL of 1:1 ethanol/water. To this mixture was sequentially added 3,5-dimethyl-4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)isoxazole (6.00 mg, 0.027 mmol), palladium acetate (0.40 mg, 0.002 mmol), 2-dicyclohexylphosphino-2',6'-diisopropoxybiphenyl (RUPHOS ®, 1.40 mg, 0.003 mmol), sodium carbonate (5.70 mg, 0.054 mmol), and 0.5 mL *p*-dioxane. The mixture was then heated to 100 C for 2 h, then cooled to rt and poured into water. The mixture was then extracted with ethyl acetate and then the combined organics were washed with water, brine, then dried with sodium sulfate, filtered, and concentrated to give an oily residue which was purified by flash chromatography (0-100% EtOAc in Heptane over 12 mins). The major fraction was collected and concentrated to give 2.6 mg (30%) the titled compound (**41**) as a pale yellow solid: ¹H NMR (400 MHz, Chloroform-*d*) δ 7.61 (d, *J* = 8.3 Hz, 1H), 7.16 (d, *J* = 8.3 Hz, 2H), 7.07 (d, *J* = 8.3 Hz, 1H), 6.89 – 6.82 (m, 2H), 6.54 (s, 1H), 4.15 (t, *J* = 7.1 Hz, 2H), 3.80 (s, 3H), 3.68 (t, *J* = 4.5 Hz, 4H), 3.09 (s, 4H), 2.57 (d, *J* = 21.1 Hz, 5H), 2.44 (d, *J* = 16.9 Hz, 7H), 1.91 (s, 2H), 1.26 (d, *J* = 5.7 Hz, 2H), 1.24 (s, 13H). LCMS *m/z* [*M* + *H*⁺]: 461.2.

4-(2-(5-(3,5-dimethylisoxazol-4-yl)-2-(4-methoxyphenethyl)-1H-indol-1-yl)ethyl)morpholine (**39**) was prepared in a fashion similar to that of **41**: ¹H NMR (400 MHz, Chloroform-*d*) δ 7.37 – 7.27 (m, 1H), 7.17 – 7.05 (m, 1H), 6.95 (dt, *J* = 8.4, 3.1 Hz, 1H), 6.83 – 6.73 (m, 1H), 6.27 (s, 1H), 4.13 (t, *J* = 7.5 Hz, 2H), 3.73 (s, 3H), 3.64 (t, *J* = 4.7 Hz, 1H), 2.98 (s, 3H), 2.57 (t, *J* = 7.4 Hz, 1H), 2.44 (t, *J* = 4.7 Hz, 1H), 2.33 (s, 0H), 2.21 (s, 3H), 2.00 – 1.92 (m, 1H), 1.52 (t, *J* = 8.2 Hz, 2H), 1.18 (d, *J* = 9.0 Hz, 1H), 1.14 – 1.00 (m, 1H), 0.96 (dd, *J* = 18.3, 6.0 Hz, 1H).

5-(3,5-dimethylisoxazol-4-yl)-2-(4-methoxyphenethyl)-1-(2-morpholinoethyl)-1H-indole-3-carbonitrile (**40**) was prepared in a fashion similar to that of **41**: ¹H NMR (400 MHz, Chloroform-*d*) δ 7.58 (d, *J* = 1.5 Hz, 1H), 7.40 (d, *J* = 8.5 Hz, 1H), 7.21 – 7.07 (m, 3H), 6.86 (d, *J* = 8.5 Hz, 2H), 4.06 (t, *J* = 7.0 Hz, 2H), 3.82 (s, 3H), 3.71 (t, *J* = 4.6 Hz, 4H), 3.29 (t, *J* = 7.6 Hz, 2H), 3.09 (t, *J* = 7.6 Hz, 2H), 2.60 (t, *J* = 7.0 Hz, 2H), 2.47 (d, *J* = 5.4 Hz, 7H), 2.32 (s, 3H).

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