

A framework for low intensity low frequency ultrasound neuromodulation sonication parameters identification from micromechanical flexoelectricity modelling

Haoyu Chen^a, Antoine Jérusalem^{a,*}

^a*Department of Engineering Science, University of Oxford, Oxford, OX1 3PJ, UK*

Abstract

Low intensity, low frequency ultrasound (LILFU) has recently emerged as a promising technique to modulate non-invasively nerve activities at lower cost than other traditional and more-invasive neuromodulation methods. However, there is currently no consensus on the optimum sonication parameters to be used in LILFU applications, and most of the accepted ranges have arisen from trial-and-error approaches. Here we utilise a recently proposed micromechanics model of membrane flexoelectricity, a potential candidate for neuromodulation and simulate action potentials/membrane polarisation triggered by acoustic pulses of different pulse frequencies, pulse magnitudes and duty cycles. Results show that at constant duty cycles, increasing the transmit frequency increases the thresholds of both the pulse magnitude and the elastic energy rate density required to mechanically trigger an action potential, whereas at constant frequencies, increasing the duty cycle reduces both. The influence of transmit frequency is weakened at lower duty cycles. Our simulation results offer some guidance on the selections of sonication pa-

*Corresponding Author: Antoine Jérusalem, antoine.jerusalem@eng.ox.ac.uk

rameters used in LILFU for neurological disorder treatments in the context of the flexoelectricity hypothesis.

Keywords: Flexoelectricity, Neuromodulation, Low intensity low frequency ultrasound, Sonication parameters, Computational model

1 **Introduction**

2 In the past decade, low intensity, low frequency ultrasound (LILFU) has
3 emerged as a new technique to alter and modulate nerve activities with high
4 spatial precision and penetration depth (Tyler et al., 2008; Baek et al., 2017;
5 Jerusalem et al., 2019). Compared with traditional neuromodulation ap-
6 proaches, LILFU is non-invasive, low-cost and bi-modal, and possessing the
7 capability to both excite and suppress neuronal activity (Baek et al., 2017).
8 Compelling evidence of LILFU-mediated neuromodulatory effects through,
9 e.g., nerve electroencephalography or behavioural responses have been pub-
10 lished in both human and animal studies (Kubanek, 2018; Feng et al., 2019;
11 Blackmore et al., 2019). Paradoxically, the underlying micromechanisms
12 through which LILFU neuromodulation operates are still unclear. Different
13 theories have already been proposed as candidates: e.g., acoustic radiation
14 force, sonoporation and cavitation, flexoelectricity, among others (Sassaroli
15 and Vykhodtseva, 2016; Jerusalem et al., 2019; Blackmore et al., 2019). In
16 addition to this, sonication parameters used in LILFU such as transducer
17 frequency and intensity, duty cycle (DC), pulse repetition frequency (PRF)
18 and sonication duration vary dramatically across literature studies. A con-
19 sensus on the identification of optimum parameters thus still remains elusive
20 (Fomenko et al., 2018; Lozano and Fomenko, 2019).

21 Recently, several numerical models of mechano-electrophysiological cou-
22 pling in neurons have been proposed (Hady and Machta, 2015; Plaksin et al.,
23 2016; Cinelli et al., 2018; Kwong et al., 2019; Chen et al., 2019) with the
24 purpose of helping to understand how mechanical waves may interact or are
25 coupled with the electrophysiological pulses and, for some, how LILFU may

26 modulate neuronal activities. The study proposed here uses MECAXON, the
27 numerical model based on the direct flexoelectricity hypothesis previously
28 proposed by the authors (Chen et al., 2019), to simulate ultrasonic stimula-
29 tion of the neuronal axon. In particular, this work investigates how action
30 potentials (AP) can be triggered under different combinations of stimulation
31 parameters, eventually providing some insight on the selection of sonication
32 parameters used in LILFU neuromodulation.

33 **Materials and Methods**

34 The Mechano-Electrophysiological Coupling of Axon program (MECAXON)
35 was recently proposed as a 1D axisymmetric finite element model simulating
36 the flexoelectricity-induced electrical excitation of neuron/axon arising from
37 external mechanical stimuli Chen et al. (2019). In this section, we briefly
38 summarise the model formulation with emphasis on its further development
39 for this study. A detailed description of the model formulation including all
40 mathematical formulae and schematic diagrams can be found in the work of
41 Chen et al. (2019).

42 In MECAXON, the axon is modelled as a 1D axisymmetric cylindrical
43 tube, where the thin wall represents the membrane with a constant thick-
44 ness. It is observed that lipid bilayers usually exhibit both elastic and viscous
45 properties, and their mechanical properties may influence the propagation
46 (elastic) or attenuation (viscous) of mechanical signals across the cell mem-
47 brane (Zubko et al., 2013). Therefore, the axon mechanics is modelled by
48 means of viscoelasticity in a dynamic framework, governed by the balance
49 of linear momentum. A two-branch generalised Maxwell model is adopted

50 to model the membrane viscoelasticity. Material parameters in the model
51 are calibrated by simultaneously nonlinear-least-square fitting the equivalent
52 storage and loss moduli to the frequency-dependent rheological properties
53 of neurons experimentally measured in Ayala et al. (2016) using Prony se-
54 ries. Mechanical deformations of the axon membrane alter the electrical
55 properties of the membrane through geometrical (i.e., areal) changes, but
56 also redistributes the membrane dipole moments. This results in the devel-
57 opment of a self-polarised electrical field altering the membrane potential
58 Petrov (2002), following the direct flexoelectricity theory. This is adopted as
59 the main hypothesis that couples axon mechanics to cell electrophysiology:
60 a local polarisation density vector is defined and linked to the membrane
61 strain gradient field via a fourth-order direct flexoelectric tensor (Gharbi
62 et al., 2011; Yudin and Tagantsev, 2013), in turn used to compute the po-
63 larisation current density through the membrane. To update the membrane
64 potential, the axonal electrophysiology is modelled using the cable theory
65 for the internodal regions combined with the classic Hodgkin-Huxley (H-H)
66 model (Hodgkin and Huxley, 1952) for the Nodes of Ranvier.

67 Here, in order to model the mechanical stimuli as acoustic waves, the
68 axon boundary is displaced following a prescribed periodic function resem-
69 bling the pulsed ultrasound to achieve a propagating vibration in the radial
70 and longitudinal direction. Accordingly, the ultrasound is modelled as a
71 pulsed wave of 3 ms duration and 1 ms fixed pulse repetition period, i.e.,
72 three full repetitions per simulation with 1 kHz PRF. Doing so, we simplify
73 our investigation by solely focusing on examining the effects of the other three
74 major parameters which conjointly specify an acoustic wave, i.e., transmit

75 frequency, DC and pulse magnitude. The threshold of the mechanical wave
 76 for AP excitation is then identified at various combinations of transmit fre-
 77 quency (50–850 kHz) and DC (10%–100%). Here, we define the threshold for
 78 neuromodulation (TN) as the threshold for ultrasound conditions at which
 79 a H-H driven membrane depolarisation is triggered through flexoelectricity,
 80 taken here when the depolarisation reaches -42 mV (Kole and Stuart, 2008).
 81 We also calculate the pulse averaged elastic energy rate density (PAPEERD)
 82 of the axon during one pulse period T to quantify the mechanical effect of
 83 acoustic wave by conjugating the elastic stress and strain tensors:

$$\text{PAPEERD} = \frac{1}{T} \int_0^T \boldsymbol{\sigma}_e(t) : \dot{\boldsymbol{\epsilon}}_e(t) dt \quad (1)$$

84 where $\boldsymbol{\sigma}_e$ and $\boldsymbol{\epsilon}_e$ are the elastic stress and strain tensors, respectively Chen
 85 et al. (2019). PAPEERD has the unit of W/cm^2 , which is same as the most
 86 common unit used for ultrasound intensity. Physically, PAPEERD is re-
 87 lated to the spatial peak pulse average intensity (I_{SPPA}) of ultrasound that
 88 is locally transformed into axon vibration. In each simulation, the transmit
 89 frequency and DC are fixed at different combinations, while the pulse magni-
 90 tude is gradually varied to identify the TN and the PAPEERD. The transmit
 91 frequency is varied from 50 kHz to 850 kHz with an increment of 200 kHz,
 92 whereas DC is varied from 10% to 90% with an increment of 20%.

93 Results

94 Figure 1 shows the temporal evolutions of membrane potential at the
 95 Node of Ranvier at the axon end subjected to four different conditions of
 96 acoustic stimuli. In the figure, three unsuccessful cases of AP induction in

97 which the TN is not achieved are compared to one successful case where H-H
98 driven membrane depolarisation occurs. In these three unsuccessful cases,
99 the values of membrane potential did not reach the threshold (-42 mV) for
100 triggering an AP during the full simulation of 3ms. Therefore, only perturba-
101 tions of the membrane potential could be observed. In the successful case,
102 the membrane potential depolarises and peaks after 1.2 ms, and repolarises
103 afterwards, i.e., the typical features of an AP.

104 Figure 2 shows the thresholds of the axial strain magnitudes above which
105 an AP can be induced by acoustic stimulation at different combinations of
106 pulse DCs and frequencies. It can be seen that at a constant fixed transmit
107 frequency, a larger magnitude is needed to excite the membrane enough if the
108 pulse DC is reduced. On the other hand, increasing the transmit frequency
109 generally requires a larger pulse magnitude for an AP initiation.

110 Figure 3 shows the PAPEERD at TN above which an AP is acoustically
111 triggered for different DCs and frequencies. Increasing the DC reduces the
112 PAPEERD threshold, and this trend is more notable at lower frequencies.
113 At constant DC, when the transmit frequency is increased, the PAPEERD
114 threshold is also elevated. However, at lower DCs, the influence of transmit
115 frequency on the PAPEERD at TN is weaker.

116 Discussion

117 Our simulation results demonstrate a clear trend of the combination of
118 the sonication parameters that can successfully induce an AP, i.e., higher fre-
119 quencies and lower DCs require higher pulse magnitudes and PAPEERDs,
120 whereas lower frequencies and higher DCs are accompanied by lower pulse

121 magnitudes and PAPEERDs. These trends generally agree with the simula-
122 tion results predicted from other pioneering numerical models (Plaksin et al.,
123 2016; Lemaire et al., 2019) which explicitly simulate local APs resulting from
124 acoustically induced intramembrane cavitation, despite the fact that the
125 underlying modulation mechanisms between our model (MECAXON) and
126 theirs (NICE and SONIC) are different (direct flexoelectricity in MECAXON
127 model vs. intramembrane cavitation excitation in NICE and SONIC mod-
128 els). This further highlights the need to acquire a deeper understanding of
129 the underlying natural principles behind LILFU neuromodulation. However,
130 it should be noted that a success in inducing an AP cannot directly imply
131 a good response rate. In fact, experimental findings suggest that high re-
132 sponse rates usually occurs at intermediate levels of DC (50% – 75%), and
133 are affected by other sonication parameters such as PRF and total stimula-
134 tion duration (Kubanek et al., 2018; Yoon et al., 2019; Wang et al., 2020).
135 Additionally, there is still no consensus on whether higher or lower intensity
136 and pressure amplitude can result in a better modulation effect. More work
137 is clearly needed to find an optimised combination of sonication parameters
138 to achieve the best modulation outcome.

139 The transmit frequency of focused low frequency ultrasound for neuro-
140 modulation typically lies in the sub-MHz range (Kubanek, 2018; Wang et al.,
141 2019). In this study, we limit the ultrasound transmit frequency in all sim-
142 ulations to be within 50 – 850 kHz. In finite element simulations, modelling
143 dynamic response of materials under high frequency vibrations requires very
144 small time steps. This results in an enormous increase in the computational
145 cost and limits the total duration for which the dynamic event can be simu-

146 lated. This situation is further exacerbated if the geometry of the modelling
147 object is complex, or if the material itself is heterogeneous, both of which
148 potentially requiring finer space discretisation. For this reason, we limit the
149 duration of our simulations to be 3 ms, i.e., three full pulse repetitions. A
150 simulation of 50 kHz transmit frequency runs for roughly 20 hours on a sin-
151 gle core processor on a 64GB RAM, Intel Core i7-7700 desktop, and run for
152 roughly more than two hours on a 2-node, 32-core cluster, including the time
153 to transfer simulation data back to local desktop from the server. Compar-
154 atively, a simulation of 850 kHz transmit frequency will need more than two
155 days to finish on the same cluster. A typical LILFU sonication duration usu-
156 ally lasts for hundreds of milliseconds, with up to 1,000 tone bursts (Wang
157 et al., 2019). There are indications that it is in fact the longer temporal effect
158 of the acoustic wave that has the neuromodulatory effect and in this case, a
159 longer simulation that can depict a full neural response will be more appro-
160 priate. For achieving this, high performance computing with more advanced
161 hardware or the utilisation of graphics processing unit computing is clearly
162 indispensable. Adoption of another faster numerical methods to solve the
163 mathematical equations, such as the k-space pseudospectral method, could
164 also be one alternative option to speed up a simulation of a full LILFU neu-
165 romodulation duration.

166 Currently, the mechanisms at play during LILFU neuromodulation are
167 still unclear, and a few hypotheses have been proposed as promising candi-
168 dates to explain this mechanism (Sassaroli and Vykhodtseva, 2016; Jerusalem
169 et al., 2019; Blackmore et al., 2019). The direct flexoelectricity theory, be-
170 ing one of them, suggested that ultrasound may affect neuron electrophys-

171 iological activities through the process of redistributing and reorientating
172 membrane dipole molecules, inducing a self-polarised electrical field altering
173 membrane potential. This hypothesis is adopted as the main bridging mech-
174 anism linking axon electrophysiology to its mechanics. **Note, however, in the**
175 **model, this hypothesis could be substituted by other proposed mechano-**
176 **electrophysiological coupling hypotheses, such as membrane sonoporation**
177 **and cavitation, as well as interference with membrane thermodynamic waves.**
178 **When additional experimental data become available, one can then compare**
179 **the results from different hypotheses, and select the most appropriate one to**
180 **be used within the future framework.**

181 In classical ultrasound neuromodulation studies, intensity is a quantity as-
182 sociated with the ultrasound transducer, normally measured and calibrated
183 together with the transducer using a hydrophone in a water tank (Civale
184 et al., 2018). The intensity of an ultrasound beam usually attenuates expo-
185 nentially with the propagation distance due to both absorption and scattering
186 (Feng et al., 2019). Ultrasound attenuates fast when penetrating the skull as
187 much more scattering and absorption occurs in bone than in soft tissue. As a
188 result, when reaching the targeted neurons, the actual intensity of the ultra-
189 sound beam might be much lower than the one specified by the transducer.
190 **In this study, we use the PAPEERD to quantify the strength of the acoustic**
191 **stimulus. This parameter could potentially be directly related to the I_{SPPA} of**
192 **ultrasound by an empirical power law (Naor et al., 2016), depending on both**
193 **the viscous property of the brain tissue and the distance that the ultrasound**
194 **has travelled. Therefore, values of PAPEERD evaluated in our study should**
195 **be used as a reference with caution when compared with typical values of**

196 **ultrasound intensities used in literature experiments.**

197 According to literature studies, ultrasound has been proven to be bimodal,
198 with capabilities of both enhancing and suppressing nerve activities. How-
199 ever, it is still unclear under what circumstances ultrasounds can enhance or
200 suppress a nerve signal. Surprisingly, mixed stimulatory and inhibitory re-
201 sults can be often seen within the same study (Yoo et al., 2011; Yoon et al.,
202 2019). Experimental results show that the outcome depends on multiple
203 factors, including sonication duration, latency time between the pulse and
204 APs, neuron connectivity, and primarily, ultrasound parameters (Blackmore
205 et al., 2019). For instance, it is observed that ultrasounds with lower in-
206 tensities tend to enhance neuron activity, whereas ultrasounds with higher
207 intensities tend to have an inhibitory effect (Feng et al., 2019). In addition,
208 pulses with lower DCs are more prone to have an inhibitory effect, and a
209 stimulatory effect when the DC is increased (Yoo et al., 2011; Kim et al.,
210 2015; Plaksin et al., 2016). Our study utilises MECAXON to simulate APs
211 triggered by acoustic pulses. As implied from the model itself Chen et al.
212 (2019), enhancement or suppression depends on the directionality of the me-
213 chanical pulse: a mechanical pulse travelling in the same direction as the
214 AP will enhance the AP, whereas a mechanical pulse travelling in opposite
215 direction will suppress it. The divergence between experimental observa-
216 tions in the literature and our model predictions was previously attributed
217 to the fundamental assumptions of the model. The H-H theory is adopted to
218 model the ionic mechanisms underlying APs. As a result, AP enhancement
219 is due to voltage-controlled channel opening whereas suppression is linked to
220 cell refractory periods. A more comprehensive understanding of the working

221 mechanism of LILFU is thus needed so that our model can be improved to
222 better capture experimental observations.

223 Experimental observations show that myelinated and unmyelinated neu-
224 rites differ from each other, not only by the presence of myelin layers, but also
225 by the densities and even the types of ion channels expressed on the neu-
226 ronal membrane. For example, myelinated axons feature a highly structured
227 distribution of voltage-gated sodium ion channels, with a characteristic ac-
228 cumulation at the Nodes of Ranvier (Neishabouri and Faisal, 2014; Freeman
229 et al., 2015; Stadelmann et al., 2019). Different types of potassium ion chan-
230 nels are also found to possess distinct density distributions along the axon
231 (Waxman and Ritchie, 1985; Röper and Schwarz, 1989; Black et al., 1990).
232 In addition, it has been discovered that the mechanosensitive TREK-1 and
233 TRAAK channels are prone to be localised to the Nodes of Ranvier (Bro-
234 hawn et al., 2019; Kanda et al., 2019). Bearing in mind that the underlying
235 mechanism of ultrasound neuromodulation is still unclear, this difference in
236 membrane electrophysiological dynamics between neurites with and without
237 myelination may imply significant variations in the way these two types of
238 neurites are modulated by acoustic stimuli. Therefore, it is reasonable to in-
239 fer that the TN may also be different between different types of neurites. In
240 fact, evidence of the difference of threshold potentials for neuron excitation
241 between myelinated and unmyelinated fibres have already been documented
242 in the literature (Grill et al., 2002; Moffitt et al., 2004). This difference of
243 TN between these fibres could not be predicted by the MECAXON model
244 used in our study. This is mainly due to the mathematical formulation of
245 the model, as unmyelinated regions are assumed to have the same mechani-

246 cal properties as the myelinated segments (the assumption being made was
247 that the membrane could “vibrate” independently from the myelin layers),
248 and, in order to initiate an AP, the boundary elements of a myelinated axon
249 at which the mechanical stimulus is applied are set to be Nodes of Ranvier.
250 A successful induction is recorded when a mechanically induced flexoelec-
251 tric current perturbs the local membrane potential and raises the membrane
252 depolarisation above -42 mV. As such, this formulation cannot distinguish
253 between myelinated and unmyelinated axon, although the electrical pulse
254 propagation patterns will be distinct between these two types of neurites.
255 Another limitation of this research is the lack of a sensitivity study on the
256 influence of model dimensions on the TN. According to experimental find-
257 ings, fibre radii in the white matter of human brains range from 0.08 – 4.5 μm
258 (Liewald et al., 2014). The radius of the axon being modelled in the study
259 is set to be 2.5 μm , which is within this range. A systematic investigation
260 of the influence of model dimensions on the mechanical excitation thresholds
261 under different combinations of these sonication parameters is undeniably
262 worth being conducted in the future.

263 **Conclusions**

264 There is currently no consensus on the optimum values of sonication pa-
265 rameters to be used in low intensity LILFU. To this end, we leveraged a
266 recently proposed numerical model, MECAXON, to simulate flexoelectricity
267 driven AP/membrane polarisation triggered by acoustic pulses for multiple
268 combinations of transmit frequency and DCs. Simulation results show that
269 at constant DCs, increasing the transmit frequency increases the thresholds

270 of both pulse magnitude and elastic energy rate density needed to mechani-
271 cally trigger an AP, whereas at constant frequencies, increasing the DC has
272 opposite effect. At lower DCs, the influence of transmit frequency is weak-
273 ened. These simulation results may offer some guidance on the selections of
274 sonication parameters used in LILFU, and may help to gain further insights
275 into the mechanism of ultrasound neuromodulation.

276 **Acknowledgements**

277 The authors acknowledge funding from the EPSRC Healthcare Technolo-
278 gies Challenge Award No. EP/N020987/1.

279 **References**

- 280 Ayala YA, Pontes B, Ether DS, Pires LB, Araujo GR, Frases S, Romão
281 LF, Farina M, Moura-Neto V, Viana NB, Nussenzveig HM. Rheological
282 properties of cells measured by optical tweezers. *BMC Biophysics*, 2016;9.
- 283 Baek H, Pahk K, Kim H. A review of low-intensity focused ultrasound for
284 neuromodulation. *Biomedical Engineering Letters*, 2017;7:135–142.
- 285 Black JA, Kocsis JD, Waxman SG. Ion channel organization of the myeli-
286 nated fiber. *Trends in Neurosciences*, 1990;13:48–54.
- 287 Blackmore J, Shrivastava S, Sallet J, Butler CR, Cleveland RO. Ultrasound
288 neuromodulation: A review of results, mechanisms and safety. *Ultrasound
289 in Medicine & Biology*, 2019;45:1509–1536.
- 290 Brohawn SG, Wang W, Handler A, Campbell EB, Schwarz JR, MacKinnon
291 R. The mechanosensitive ion channel TRAAK is localized to the mam-
292 malian node of ranvier. *eLife*, 2019;8.
- 293 Chen H, Garcia-Gonzalez D, Jérusalem A. Computational model of the
294 mechano-eletrophysiological coupling in axons with application to neuro-
295 modulation. *Physical Review E*, 2019;In press.
- 296 Cinelli I, Destrade M, Duffy M, McHugh P. Electro-mechanical response of
297 a 3d nerve bundle model to mechanical loads leading to axonal injury.
298 *International Journal for Numerical Methods in Biomedical Engineering*,
299 2018;34:e2942.

300 Civale J, Rivens I, Shaw A, ter Haar G. Focused ultrasound transducer spatial
301 peak intensity estimation: a comparison of methods. *Physics in Medicine
302 & Biology*, 2018;63:055015.

303 Feng B, Chen L, Ilham SJ. A review on ultrasonic neuromodulation of the
304 peripheral nervous system: Enhanced or suppressed activities? *Applied
305 Sciences*, 2019;9:1637.

306 Fomenko A, Neudorfer C, Dallapiazza RF, Kalia SK, Lozano AM. Low-
307 intensity ultrasound neuromodulation: An overview of mechanisms and
308 emerging human applications. *Brain Stimulation*, 2018;11:1209–1217.

309 Freeman SA, Desmazières A, Fricker D, Lubetzki C, Sol-Foulon N. Mech-
310 anisms of sodium channel clustering and its influence on axonal impulse
311 conduction. *Cellular and Molecular Life Sciences*, 2015;73:723–735.

312 Gharbi M, Sun Z, Sharma P, White K, El-Borgi S. Flexoelectric properties
313 of ferroelectrics and the nanoindentation size-effect. *International Journal
314 of Solids and Structures*, 2011;48:249–256.

315 Grill W, Richardson A, McIntyre C. Influence of the myelin sheath on ex-
316 citation properties of nerve fibers. In: *Proceedings of the 22nd Annual
317 International Conference of the IEEE Engineering in Medicine and Biol-
318 ogy Society (Cat. No.00CH37143)*. IEEE, 2002.

319 Hady AE, Machta BB. Mechanical surface waves accompany action potential
320 propagation. *Nature Communications*, 2015;6:1–17.

321 Hodgkin A, Huxley A. A Quantitative Description of Membrane Current and

322 its Application to Conduction and Excitation in Nerves. *The Journal of*
323 *Physiology*, 1952;117:500–544.

324 Jerusalem A, Al-Rekabi Z, Chen H, Ercole A, Malboubi M, Tamayo-Elizalde
325 M, Verhagen L, Contera S. Electrophysiological-mechanical coupling in
326 the neuronal membrane and its role in ultrasound neuromodulation and
327 general anaesthesia. *Acta Biomaterialia*, 2019.

328 Kanda H, Ling J, Tonomura S, Noguchi K, Matalon S, Gu JG. TREK-1 and
329 TRAAK are principal K⁺ channels at the nodes of ranvier for rapid action
330 potential conduction on mammalian myelinated afferent nerves. *Neuron*,
331 2019;104:960–971.e7.

332 Kim H, Park MY, Lee SD, Lee W, Chiu A, Yoo SS. Suppression of EEG
333 visual-evoked potentials in rats through neuromodulatory focused ultra-
334 sound. *NeuroReport*, 2015;26:211–215.

335 Kole MHP, Stuart GJ. Is action potential threshold lowest in the axon?
336 *Nature Neuroscience*, 2008;11:1253–1255.

337 Kubanek J. Neuromodulation with transcranial focused ultrasound. *Neuro-*
338 *surgical Focus*, 2018;44:E14.

339 Kubanek J, Shukla P, Das A, Baccus SA, Goodman MB. Ultrasound
340 elicits behavioral responses through mechanical effects on neurons and
341 ion channels in a simple nervous system. *The Journal of Neuroscience*,
342 2018;38:3081–3091.

343 Kwong MT, Bianchi F, Malboubi M, García-Grajales JA, Homsí L, Thomp-
344 son M, Ye H, Noels L, Jérusalem A. 3d finite element formulation for me-

345 chanical–electrophysiological coupling in axonopathy. *Computer Methods*
346 in *Applied Mechanics and Engineering*, 2019;346:1025–1050.

347 Lemaire T, Neufeld E, Kuster N, Micera S. Understanding ultrasound neu-
348 romodulation using a computationally efficient and interpretable model of
349 intramembrane cavitation. *Journal of Neural Engineering*, 2019;16:046007.

350 Liewald D, Miller R, Logothetis N, Wagner HJ, Schüz A. Distribution of axon
351 diameters in cortical white matter: an electron-microscopic study on three
352 human brains and a macaque. *Biological Cybernetics*, 2014;108:541–557.

353 Lozano A, Fomenko A. Neuromodulation and ablation with focused ultra-
354 sound – toward the future of noninvasive brain therapy. *Neural Regenera-*
355 *tion Research*, 2019;14:1509.

356 Moffitt M, McIntyre C, Grill W. Prediction of myelinated nerve fiber stim-
357 ulation thresholds: Limitations of linear models. *IEEE Transactions on*
358 *Biomedical Engineering*, 2004;51:229–236.

359 Naor O, Krupa S, Shoham S. Ultrasonic neuromodulation. *Journal of Neural*
360 *Engineering*, 2016;13.

361 Neishabouri A, Faisal AA. Saltatory conduction in unmyelinated axons: clus-
362 tering of Na⁺ channels on lipid rafts enables micro-saltatory conduction
363 in c-fibers. *Frontiers in Neuroanatomy*, 2014;8.

364 Petrov AG. Flexoelectricity of model and living membranes. *Biochimica et*
365 *Biophysica Acta* ({BBA}) - *Biomembranes*, 2002;1561:1–25.

- 366 Plaksin M, Kimmel E, Shoham S. Cell-type-selective effects of intramembrane
367 cavitation as a unifying theoretical framework for ultrasonic neuromodu-
368 lation. *eneuro*, 2016;3:ENEURO.0136–15.2016.
- 369 Röper J, Schwarz JR. Heterogeneous distribution of fast and slow potas-
370 sium channels in myelinated rat nerve fibres. *The Journal of Physiology*,
371 1989;416:93–110.
- 372 Sassaroli E, Vykhodtseva N. Acoustic neuromodulation from a basic science
373 prospective. *Journal of Therapeutic Ultrasound*, 2016;4:17.
374 URL <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4875658/>
- 375 Stadelmann C, Timmler S, Barrantes-Freer A, Simons M. Myelin in the cen-
376 tral nervous system: Structure, function, and pathology. *Physiological Re-*
377 *views*, 2019;99:1381–1431.
- 378 Tyler WJ, Tufail Y, Finsterwald M, Tauchmann ML, Olson EJ, Majestic C.
379 Remote excitation of neuronal circuits using low-intensity, low-frequency
380 ultrasound. *PLoS ONE*, 2008;3.
- 381 Wang P, Zhang J, Yu J, Smith C, Feng W. Brain modulatory effects by low-
382 intensity transcranial ultrasound stimulation (TUS): A systematic review
383 on both animal and human studies. *Frontiers in Neuroscience*, 2019;13.
- 384 Wang X, Yan J, Wang Z, Li X, Yuan Y. Neuromodulation effects of ul-
385 trasound stimulation under different parameters on mouse motor cortex.
386 *IEEE Transactions on Biomedical Engineering*, 2020;67:291–297.
- 387 Waxman S, Ritchie J. Organization of ion channels in the myelinated nerve
388 fiber. *Science*, 1985;228:1502–1507.

- 389 Yoo S, Bystritsky A, Lee J, Zhang Y, Fischer K, Min B, McDannold N,
390 Pascual-Leone A, Jolesz F. Focused ultrasound modulates region-specific
391 brain activity. *NeuroImage*, 2011;56:1267–1275.
- 392 Yoon K, Lee W, Lee JE, Xu L, Croce P, Foley L, Yoo SS. Effects of soni-
393 cation parameters on transcranial focused ultrasound brain stimulation in
394 an ovine model. *PLOS ONE*, 2019;14:e0224311.
- 395 Yudin PV, Tagantsev AK. Fundamentals of flexoelectricity in solids. *Nan-*
396 *otechnology*, 2013;24:432001.
- 397 Zubko P, Catalan G, Tagantsev AK. Flexoelectric Effect in Solids. *Annual*
398 *Review of Materials Research*, 2013;43:387–421.

399 **Figure Captions**

400 **Figure 1:** Four cases of membrane potential evolution at the Node of Ran-
401 vier at the axon end subjected to different combinations of sonication
402 parameters. Red solid curve: successful; Other curves: unsuccessful.
403 TF: Transmit Frequency, AS: Axial Strain, DC: Duty Cycle.

404 **Figure 2:** Thresholds of the axial strain magnitudes for an acoustically-
405 triggered action potential at different pulse duty cycles and frequencies;
406 full line: exponential fit.

407 **Figure 3:** Thresholds of the pulse averaged elastic energy rate density for
408 an acoustically-triggered action potential at different pulse duty cycles
409 and frequencies; full line: exponential fit.