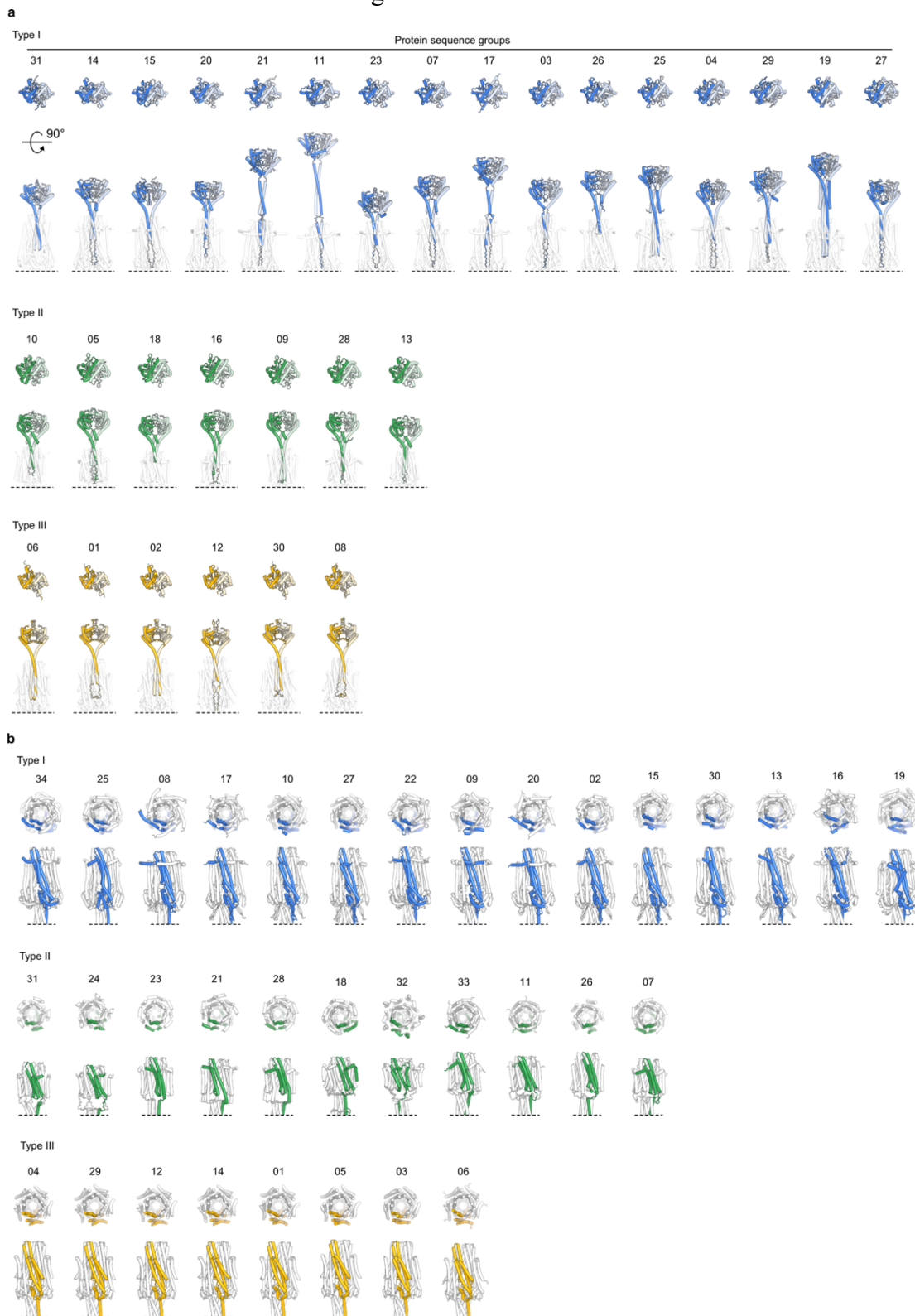


1 **Supplementary Discussion**

2 In addition to the mechanistic insights supported by our data, there are several aspects worthy of
3 future investigations. First, future studies will need to determine whether Zorya functions through
4 a true ‘localized-response’ model, where multiple ZorAB complexes diffuse to the site of phage
5 penetration of the cell envelope to amplify the recruitment of ZorC/ZorD in a ZorAB rotation-
6 dependent manner, or a more ‘global’ activation of ZorAB via perturbation of the IM-PG distance
7 throughout the cell envelope. Interestingly, our model predicts that the ‘reach’ of the ZorB PGBD
8 should be evolutionarily diversified to adapt to different bacterial cell envelope architectures,
9 which appears to be the case (**Supplementary Discussion Figure 1**). Accumulated knowledge
10 about phages and the mechanics phage-host physical interactions during adsorption and genome
11 injection would deepen our understanding of the proposed role of ZorAB as a ‘sensor’ during
12 phage infection, which is likely to be a universal mechanism of the Zorya defense system across
13 diverse bacteria.

14 Another open question is the role of two striking features of the ZorAB structure: the rotary motor
15 and the conserved long tail. One hypothesis for the role of rotation is in signal transduction from
16 phage ingress at the cell envelope to activation of ZorC and ZorD. If the direction of the ZorAB
17 motor is the same as for MotAB (clockwise rotation of A around B, as viewed from the outside of
18 the cell), rotation could potentially induce untwisting of the pentameric ZorA tail super-helix,
19 thereby altering the conformation of the tail to facilitate recruitment or activation of ZorC and/or
20 ZorD (**Supplementary Discussion Figure 2a, b**). A somewhat similar sensory transmission
21 mechanism occurs with bacterial chemosensory arrays and the long ZorA tail might serve to
22 transmit the signal in a manner analogous to the long coiled-coil cytoplasmic signaling domains
23 of methyl-accepting chemotaxis proteins²³. A second hypothesis accounting for both rotation and
24 a long ZorA tail is that phage DNA might be ‘reeled in’ around the tail, trapping the foreign DNA
25 in an inactive state (e.g. preventing RNA polymerase access) prior to degradation by ZorD
26 (**Supplementary Discussion Figure 2c**). A ‘reeled in’ mechanism could greatly enhance phage
27 genome localization and sequester it from interactions required for host infection. A typical
28 double-stranded DNA phage genome is 10s of μm long and would form a random coil with a
29 radius of gyration similar to the size of the entire cell in the absence of constraints – thus negating
30 any advantage of a localized nuclease defense response at the site of entry. Binding to multiple
31 Zor complexes, perhaps via ZorC and/or D might contribute to localizing phage DNA to the entry
32 site. Unless and until tail rotation is resisted, an almost inevitable consequence of tail rotation
33 combined with DNA binding is that the DNA will wind around the tail like wire on a reel. If
34 ‘reeled in’ of the phage DNA were an essential feature of the Zorya defense mechanism, this
35 would explain the need for both rotation and a long ZorA tail. The length of the ZorA tail is similar
36 to the diameter of a typical phage capsid into which an entire phage genome can be tightly packed,
37 indicating that a single Zor complex may be sufficient to capture an entire genome. Rough
38 calculations indicate that 100s of turns would be required to wind a 60 kb Bas24 genome onto a
39 70 nm tail, allowing the rotary ZorAB motor cumulatively to supply the necessary energy to wind

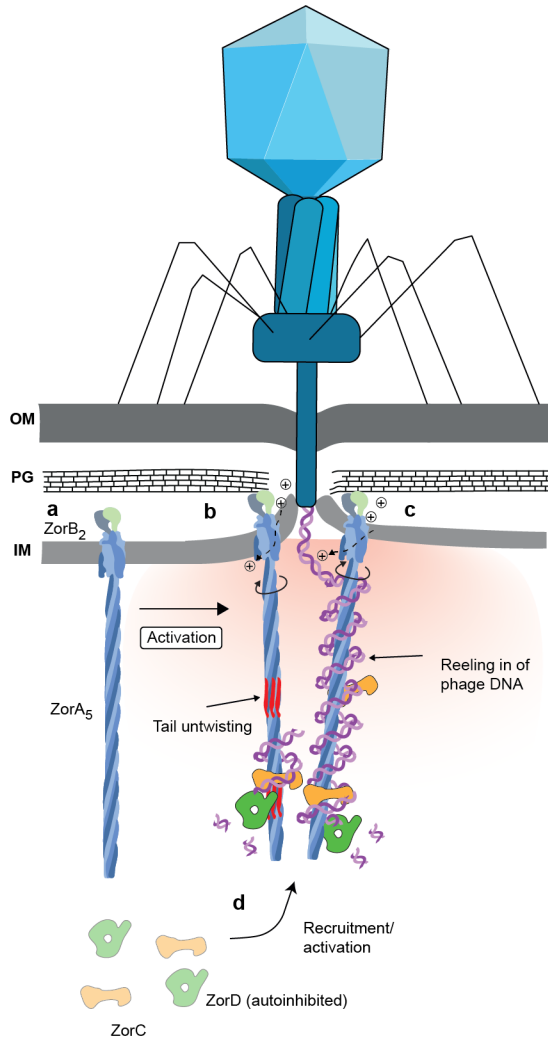
40 and compact the phage DNA. Reeling would also tighten any loops that might form between DNA
 41 sites bound to different ZorA tails, removing any freedom for further ZorA rotation - as required
 42 by the model of activation via untwisting of the ZorA tail.



43

44 **Supplementary Discussion Figure 1. Structural prediction of the representative ZorAB**
45 **complexes form different Zorya system types.**

46 Representative predicted structures of ZorA₅B₂ complexes (omitting the ZorA tail region) for each
47 ZorA and ZorB sequence family (protein sequence groups numbered above each predicted
48 structure) for type I, II, and III Zorya systems. **a**, The predicted dimerized ZorB PGBDs. One ZorB
49 subunit is colored in each structure. **b**, The predicted ZorAB transmembrane motor complex. One



50 ZorA subunit is colored in each structure.

51 **Supplementary Discussion Figure 2: Putative ZorA tail conformational change and phage**
52 **DNA ‘reeling in’ mechanism in the activated Zorya defense system.**

53 **a**, An inactive ZorAB embedded in the inner membrane. **b**, Phage invasion triggers ZorAB
54 activation. The rotation of ZorA and its long intracellular tail around ZorB causes ZorA tail tip
55 conformational change, which would recruit ZorC and ZorD. **c**, Reeling in of phage DNA
56 around the long ZorA tail in the activated Zorya defense system.