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Island species–area relationships and species–accumulation curves are not equivalent: an analysis of habitat island datasets

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Abstract

Aim The relationship between species number and area is of fundamental importance within macroecology and conservation science. Yet, the implications of different means of quantitative depiction of the relationship remain contentious. We set out (i) to establish the variation in form of the relationship between two distinct methods applied to the same habitat island datasets, (ii) to explore the relevance of several key dataset properties for variation in parameters of these relationships, and (iii) to assess implications for applications of the resulting models.

Locations Global

Methods Through literature search we compiled 97 habitat island datasets. For each we analysed the form of the island species–area relationship (ISAR) and several versions of species accumulation curve (SAC), giving priority to a randomized form (Ran-SAC). Having established the validity of the power model, we compared the slopes (z-values) between the ISAR and the SAC for each dataset. We used boosted regression tree and simulation analyses to investigate the effect of nestedness and other variables in driving observed differences in z values between ISARs and SACs.

Results The Ran-SAC was steeper than the ISAR in 77% of datasets. The differences were primarily driven by the degree of nestedness, although other variables (e.g. number of islands in a dataset) were also important. The ISAR was often a poor predictor of archipelago species richness.

Main conclusions Slopes of the ISAR and SAC for the same data set can vary substantially, revealing their non-equivalence, with implications for applications of species–area curve parameters in conservation science. For example, the ISAR was a poor predictor of

archipelagic richness in datasets with a low degree of nestedness. Caution should be employed when using the ISAR for extrapolation and prediction purposes in habitat island systems.

Keywords Boosted regression trees, conservation biogeography, fragmentation, habitat islands, island biogeography, island species–area relationship, macroecology, nestedness, species–accumulation curve, species–area relationship

INTRODUCTION

The increase in number of species with area is one of the few laws of ecology (Scheiner, 2003). While those working with species–area curves have long recognised different forms of curve, confusion in terminology and usage has persisted (for use herein see Table 1), as evidenced by an exchange of papers in this journal disputing the number of fundamental types of relationship and the key distinctions between them (Scheiner, 2003, 2004; Gray et al., 2004a,b). In their contributions, Gray et al. (2004a,b) argued that the most important distinction was between species accumulation curves (SACs), which present cumulative counts of increased species number with sampling area, and island-type species–area relationships (herein termed ISARs) in which the function fitted is based on how many species are found in each sampled area (or island). By virtue of their mode of construction, SACs (which can be constructed in several distinct ways; see Appendix S1) must be rising functions, whereas in principle an ISAR can have a negative slope: a large island may have fewer species than a smaller one (Scheiner, 2003), suggesting the possibility that SAC and ISAR form may vary substantially. However, in the island literature and in its application in conservation science, it is sometimes assumed that SAC form and ISAR form are directly comparable (as noted by Ulrich & Buscko, 2007, p. 55). Two classic illustrations of this are

to be found in the seminal works of MacArthur and Wilson (1967) and Rosenzweig (1995, 2003).

MacArthur and Wilson (1967) in their Figure 3 provide two data series, one a cumulative species–area curve of contiguous sampling areas on mainland New Guinea, and the other a set of points representing the richness of each island in the region. They comment (p10) that “[t]he line and the cluster of points illustrate the principle that the increase in number of species with area is more rapid in the case of isolated islands or archipelagos than in expanding sample areas on a single land mass.” Similarly, Rosenzweig notes (1995, p10) in developing the three-scales of species–area relationship model on which his reconciliation ecology (2003) is largely based, that if scattered areas (or islands) are used to construct a species–area curve, this will produce a steeper slope (higher z -value) than if using contiguous sub-plots: moreover, the greatest disparity occurs between the richness estimated for the smallest areas, while with increasing area of sample units, what we term the SAC and ISAR converge towards the regional richness value. Rosenzweig (1995, p19) poses the question as to whether the difference in z -values might be a consequence of the different mode of construction of the two forms of function, but having provided a single demonstration of similarity of values he goes on to state (p19) that “The data suggest that we do not have a problem here.” The question of equivalence of SACs and ISARs is not trivial: if we assume that the different mode of construction of SACs and ISARs is of no great consequence, it follows that we can, for example, base projections of the consequences of habitat loss on comparisons of z -values for contiguous habitat (assessed by SAC models) with z -values for islands or habitat islands (assessed by ISARs). That this might be a dangerous assumption has previously been argued by several authors (e.g. Ulrich & Buszko, 2007; Whittaker & Fernández-Palacios, 2007; Hui, 2008; Dengler, 2009), while the significance of how SACs are constructed for projections of species extinctions has also seen lively recent debate (e.g.

He & Hubbell, 2011, 2013; Axelsen *et al.*, 2013), revealing that our understanding of the behaviour of different forms of species–area curve remains incomplete.

Our purpose herein is to advance understanding of the empirical differences between SAC and ISAR data structures by means of a systematic comparison for a large set of habitat island datasets. Specifically, we use a randomization procedure to construct a Scheiner type IIIb curve (termed Ran-SAC, Table 1), and compare the z -values with a Scheiner type IV curve (the ISAR) based on fits using the power model to test the hypothesis that they are non-equivalent. Strictly speaking, our Ran-SAC is a variant of a Scheiner type IIIb curve as, although the Ran-SAC is constructed using mean richness obtained by randomisation, the observational units are habitat islands of varying area rather than non-contiguous plots of equal size. However, despite this difference our Ran-SAC is still clearly a SAC, allowing us to compare SAC and ISAR data structures. Next, we explore the relevance of several key dataset properties for variation in parameters of these relationships, testing the hypothesis that compositional nestedness (see Table 1) will be a significant determinant of variation in z (slope), such that in anti-nested systems (Table 1) the Ran-SAC will be steeper than the ISAR and as the degree of nestedness increases this situation will gradually reverse. While it has long been appreciated that the form of species–area curves from non-contiguous samples is a consequence of both alpha (local) and beta (differentiation) diversity, this is to our knowledge, the first systematic attempt to quantify the interrelationship between ISAR and SAC form and system nestedness across a series of habitat islands.

MATERIALS AND METHODS

Data collection, formatting and species–area curve construction

Datasets were sourced via a comprehensive search of the literature following steps and criteria for evaluating suitability set out (in Appendix S2; see also Matthews, 2015). For each

selected dataset we recorded: geographic location, taxon, habitat island type (forested or non-forested), latitude and longitude of the study extent (for some datasets this was an estimate as the data were not presented in the source papers), range of species richness and island sizes, and a classification of the predominant matrix type (as per Appendix S2).

For each dataset we constructed (i) the standard ISAR using the raw island area and richness values, (ii) several forms of species accumulation curve (SAC), using different rules for combining islands into the sequence, namely small–large, large–small, poor–rich, rich–poor, random (Table S1). Except where stated, we present results for the smoothed random form of SAC (Ran-SAC) on the grounds that this is now standard practice for constructing such curves (e.g. Ugland *et al.*, 2003)(But see Fig. S1 in Appendix S1 for exemplification of the variation in form that can be observed using the full set of SAC rules). The Ran-SAC was constructed using a simple bootstrap procedure, randomly selecting the order of island addition into the SAC at each iteration. For example, for the second data point (i.e. two islands) of the SAC of a dataset, we randomly selected two islands, noting down the combined area and combined richness of this pair. We repeated this 5000 times, and used the mean of the 5000 x and y values as the data point representing two islands in the analysis.

Our aim was to compare the form of the ISAR with that of Ran-SAC, for the same dataset. The simplest way to do this is to compare the z values of the power model ($S = cA^z$) fitted to both sets of data. We focus principally on z as this parameter has attracted far more attention and application than has c (e.g. Rosenzweig, 1995; Tjørve & Tjørve, 2008; Triantis *et al.*, 2012). However, this method is arguably only appropriate if the power model provides a reasonably good fit to both sets of data. Thus, we devised a set of criteria to select suitable datasets for analysis. First, we fitted the power (non-linear) model to the two different data types for each dataset using non-linear regression and the ‘mmSAR’ R package (Guilhaumon

et al., 2010), and recorded the two parameters (c , z), the R^2 , and whether the z value was significantly different from zero. A dataset was deemed satisfactory according to this process if the z value of the power model was significant for both the SAC and ISAR structures, and the R^2 was ≥ 0.5 in both cases. This threshold was chosen arbitrarily to eliminate datasets in which the power model explained only a small amount of variation in the SAR. Second, while the observed shape of the power (non-linear) model is generally convex, we wished to determine whether a model with a different shape provided a better fit to our data. Thus, we fitted a set of 20 ISAR models (including the power model) to the ISAR and SAC data from each dataset using a modified version of the fitting algorithm within the mmSAR R package (see Triantis *et al.*, 2012; Matthews *et al.*, 2015b). We considered the model with the lowest Akaike's information criterion corrected for small sample size (AIC_c), as providing the best fit (Burnham & Anderson, 2002). AIC_c weights were computed for each model fit. Because of the non-nullity constraint on the denominator of the formula, AIC_c could not be calculated for datasets with fewer than seven islands and thus these datasets were discarded as were models with inadequate fits (e.g. due to non-normal residuals; cf. Triantis *et al.*, 2012). We then determined the observed shape of the best model fit (convex, sigmoid or linear) according to the algorithm outlined in Triantis *et al.* (2012; and see Appendix S3). For each dataset, if the observed shape of either of the best model fits (i.e. to the ISAR and SAC data) was linear or sigmoid (a check was made for linear power model fits and none were observed), the dataset was discarded as the inclusion of z values from these fits may bias our comparisons. In sum, to be classified as satisfactory, both the ISAR and SAC forms of each dataset needed to have: (a) significant z values from the power model fits, (b) R^2 values ≥ 0.5 from the power model fits, and (c) observed convex fits, when considering the best model from a set of twenty competing models.

For these satisfactory datasets, the difference between the Ran-SAC z value and the ISAR z value ($zDif$; Table 1) was used as our response variable. We also re-ran our analyses using the $zDif$ values from all datasets (i.e. those deemed satisfactory and unsatisfactory).

Relative influence of the explanatory variables

To determine factors potentially accounting for between-dataset differences in $zDif$, we fitted a boosted regression trees model (BRT; Table 1) using the functions provided in Elith *et al.* (2008). In contrast to traditional regression, for which inference is drawn from a single best model, BRT combines a large number of tree models in order to increase predictive performance (Elith *et al.*, 2008). Recent work has shown that using weight of evidence values (Burnham & Anderson, 2002) within a linear modelling framework to determine the relative importance of predictor variables is flawed (Galipaud *et al.*, 2014). BRT offers a useful intuitive and robust alternative (see: Elith *et al.*, 2008).

In the BRT analysis, $zDif$ was used as the response variable and the dataset characteristics listed above were used as predictor variables. As a further predictor variable, we also calculated the degree of nestedness in each dataset using the NODF metric (Almeida-Neto *et al.*, 2008), implemented in the ‘vegan’ R package (Oksanen *et al.*, 2013). NODF was calculated using the maximally packed matrix. Sites containing zero species were removed prior to calculation, as by definition such sites have zero nestedness, and act to depress the NODF value for the full set of sites (see Matthews *et al.*, 2015a). Cross-validation functionality was used to determine the optimum number of trees. We experimented with varying tree complexity, learning rate and bag fraction parameter values in order to minimise the minimum predictive errors, selecting final values of 5 (tree complexity), 0.001 (learning rate; 0.01 when using all datasets) and 0.5 (bag fraction). These values resulted in an optimal number of 7050 fixed trees using only the satisfactory datasets, and 4500 trees when all

datasets were used. The error structure was considered to be Gaussian. No outliers were removed when running the BRT analysis using the satisfactory datasets, but two outliers had to be removed when using all datasets to induce normality in z Dif. The predictive power of the model was assessed through cross-validation, whereby the fitted model was tested on withheld portions of the data (10 subsets were used).

The relative influence of the predictor variables was again calculated using the functions in Elith *et al.* (2008), which weigh the number of times a predictor is chosen for splitting by the squared improvement to the model due to each split. Partial dependence plots were also used to assess and visualize the effect of a predictor after accounting for the effects of all other model predictors (Elith *et al.*, 2008). In partial dependence plots the y-axis shows the effect of a predictor variable on the response variable after accounting for the effects of the other model predictors (i.e. the marginal effect of the predictor).

Simulation analyses

As the BRT analyses revealed NODF to be the most important variable (see Results), we undertook a simulation analysis to further examine the relevance of nestedness for the difference between the z values of the two forms of dataset. First, we simulated a set of six islands with area and species richness values conforming roughly to a convex ISAR (area= 1, 5, 10, 15, 20, and 30; containing 10, 12, 14, 16, 17 & 18 species respectively; herein ‘Sim1’). We fitted the power model (non-linear) to the ISAR-structured form of Sim1 and recorded the model parameters. We then simulated a perfectly nested (i.e. NODF=100) presence/absence matrix using the data characteristics of Sim1 (i.e. island number and species richness) and used this matrix to construct the SL (small–large) SAC (see Appendix S1), fitting the power model to the resulting curve. We used the SL-SAC in this analysis, as constructing the Ran-SAC for this many matrices would have been computationally

intensive. We then altered the presence/absence matrix (without changing the area and richness values) to change the degree of matrix nestedness, by randomly shuffling the presences along the sites (i.e. matrix rows) using the “commsimulator” function in the vegan R package and discarding any matrix which had already been simulated. The total number of species in the species pool (i.e. the total number of possible columns in the presence-matrix) was set to 40, and we allowed gamma diversity to change between matrices, while the alpha diversity of each island remained constant. This permitted us to create matrices with substantially different levels of nestedness. However, as this meant that the number of columns (total number of species) varied between matrices we used the NODF-by-rows value as our measure of nestedness. For each accepted matrix, we constructed the SAC curve and fitted the power model. We started the random shuffling from different starting points (i.e. different initial presence–absence matrices), used a variety of community simulation algorithms (i.e. R0, R1 and R2) and repeated this process iteratively for 12,000 runs, to cover a wide range of NODF values. We then examined $zDif$ values for each iteration, plotting $zDif$ as a function of nestedness.

Determining the degree of deviation of the archipelagic point

To determine whether the archipelagic point (‘regional richness’) of a dataset deviated from the ISAR of the constituent islands, we followed the method and nomenclature outlined by Santos *et al.* (2010). We also use ‘archipelagic point’ to refer to the total area and richness of the corresponding set of habitat islands (i.e. the archipelago). For each dataset, we fit the power (log–log; base 10) SAR model to the ISAR structure and derived the fitted values of the model for each island (‘SIpred’). The log–log version of the power model was used in this analysis as it was the model used by Santos *et al.* (2010) and thus allowed us to compare our results with theirs. Also following Santos *et al.* (2010), we added 0.01 to each species richness value to avoid zero values, although we acknowledge that other constants could also

be used. We then calculated the archipelagic point as the cumulative total area of all habitat islands in the dataset plotted against the cumulative species richness total ('SAobs'); and predicted the number of species in the archipelagic point ('SApred') using the ISAR model of the constituent islands. We followed Santos *et al.* (2010) and calculated the archipelagic residual ('ArcRes'; Table 1) as the absolute difference between $\log(\text{SAobs})$ and SApred, standardised by $\log(\text{SAobs})$ (see Appendix S3 for details). We noted whether the ISAR over- or under- predicted richness in the archipelagic point for each dataset. This ArcRes methodology represents a simple metric with which to describe how well the ISAR predicts the archipelagic richness.

To determine if any dataset characteristics (above) could explain variation in ArcRes between datasets we repeated our BRT analyses (learning rate of 0.01) using ArcRes as the response variable. We only used ArcRes values from datasets in which the power (log-log) provided a significant fit (cf. Santos *et al.*, 2010). Nine outlying points required removing to induce normality, as there was a long left-hand tail in the distribution of ArcRes values. The optimal number of fixed trees was 2200. All analyses and simulations were conducted in R (version 3.1.1.; R Development Core Team, 2014). A significance level of 0.05 was employed in all analyses.

RESULTS

We screened over 1000 published articles, of which 97 were deemed suitable for analysis (Table S2 in Appendix S2): 69 vertebrate, 20 invertebrate and 8 plant datasets. For the majority of both the ISAR and Ran-SAC datasets the observed best-fitting model shape was convex (for the ISARs the mean $wAIC_c$ of convex model fits = 0.86, linear = 0.09, sigmoid = 0.09; for Ran-SAC the equivalent results were, convex = 0.99, linear = 0.00, sigmoid = 0.02). The power model was within the set of six best models (i.e. was ranked as one of the top six

models according to AIC_c) for 67 of the ISAR datasets, and nine of the Ran-SAC datasets.

According to our three additional dataset acceptance criteria, 50 datasets were deemed satisfactory for the z Dif analyses.

Differences in the z value of ISARs and Ran-SACs

Considering only the 50 satisfactory datasets, the Ran-SAC z was greater than the ISAR z for 32 (e.g. see Fig. 1) and was smaller for the remaining 18 cases. The power model explained a larger amount of variance for the Ran-SAC (mean $R^2 = 0.97$) than for the ISAR (mean $R^2 = 0.76$); although this result is easily explained by the smoothing process involved in calculating the Ran-SAC values. Considering all 97 datasets, the Ran-SAC z value was larger in 75 cases. The z and c values are provided in Table S3 in Appendix S4, as are values from the power model fitted to the other SAC data structures (e.g. the Small-Large SAC).

Boosted regression tree results

When z Dif values from satisfactory datasets were used as the response variable, NODF was the most important explanatory variable (relative influence value of 24.6; see Table 2). Both the minimum number of species and the number of islands also had relatively high influence values (Table 2, Fig. 2a, b). In terms of the overall predictive performance of the model, the mean coefficient of the correlation between the observed and predicted response values was 0.63 when based on the cross-validation data. Results were qualitatively similar when considering z Dif values from all datasets (mean correlation coefficient = 0.68).

As predicted, further analysis revealed that in highly nested systems the z values for ISARs were greater than those for Ran-SACs, and the reverse for systems with little nesting (see Fig. 2a and Fig. 3a). In addition, the z value of the Ran-SAC was significantly related to

the NODF value (Fig. 3b), whereas there was no relationship between the z of the ISAR and NODF (Fig. 3c).

Nestedness simulations

The simulation analyses revealed further evidence for the importance of nestedness in explaining the difference in z value between ISARs and SACs. In the case of Sim1, when the system was anti-nested (low NODF values), the z value of the SL-SAC was greater than that of the ISAR. As the degree of nestedness was increased, the difference in z values declined until the ISAR z exceeded that of the SL-SAC (Fig. 4a). Figure 4b illustrates this effect for two SL-SAC curves, constructed from the most nested and anti-nested Sim1 iterations, respectively (we used the SL-SAC curve type as the area range of the ISAR and SAC are similar, making the plot easier to interpret).

The fit of the archipelagic data point

Considering only datasets in which the power (log–log) model provided a significant fit ($n=73$), the archipelagic point deviated substantially (following the rule of thumb used by Santos *et al.*, 2010) from the prediction of the constituent ISAR in 14 datasets (see Fig. 5 for an example of both scenarios). The ISAR under-predicted the archipelagic richness in 45 cases (Table S3 in Appendix S4). Considering 64 datasets (the 75 significant fits minus the nine outliers), when ArcRes was used as a response variable in a BRT analysis, NODF was the variable with the highest relative influence value (Table 2). The number of species and the minimum number of species in a dataset were also important variables (Table 2). The mean coefficient of the correlation between the observed and predicted response values was 0.35.

DISCUSSION

To evaluate the impact of confounding ISAR and SACs in SAR analyses and syntheses and to understand their differing emergent properties we compared the z values of ISAR and Ran-SAC curves using a large compilation of habitat island datasets. Constructing SACs from island data has previously been attempted (e.g. Quinn & Harrison, 1988; Rosenzweig, 1995; Fischer & Lindenmayer, 2002), although previous studies have largely focused on small-large and large-small SACs. We decided to focus on a Ran-SAC as it represents an average of the various SAC construction permutations (see Appendix S1) and thus seems a sensible choice if only one type of SAC is to be used in comparative analyses. As noted by previous authors (e.g. Ulrich & Buszko, 2007), critical tests of how different types of SAR affect model shape and parameters are lacking. We found that the z values and form of the ISAR and SAC curves varied considerably within datasets, and that this variation was primarily explicable as a function of the degree of nestedness in the archipelago. We also observed that the ISAR was often a poor predictor of SAobs, demonstrating the dangers of extrapolating the ISAR, for example, to predict the richness of an archipelago of fragments.

Why are SACs generally steeper than ISARs?

In accordance with our first hypothesis we found considerable differences between the z values of ISARs and SACs (e.g. Figs 1 and S1). Hence, our results illustrate that different inferences might be drawn depending on which data structure is used in an analysis (and see Ulrich & Buszko, 2007, for an analysis based on a different SAC type). Consistent with our second hypothesis, the difference in the z values of SACs and ISARs is best explained by variation in compositional nestedness, a pattern which is to be expected as a function of the procedures used in constructing the two types of curves (Gray *et al.*, 2004a, b); a fact highlighted by both empirical and simulation analyses (Figs 2, 4).

A number of factors have been argued to underpin the z value of SACs, through an effect on species overlap and/or nestedness (e.g. Quinn & Harrison, 1988; Chase & Knight, 2013). Two of these factors are particularly relevant to habitat island systems. First, a large species pool means that, all else being equal, there is a smaller probability that the same species will succeed in occupying each island, and thus species overlap between islands is reduced. Furthermore, if islands within an archipelago (of real or habitat islands) draw species from different species pools the likelihood is that species overlap across all islands will be reduced (Whittaker & Fernández-Palacios, 2007). Second, habitat islands are generally disturbed systems, and disturbance usually has a disproportionate effect in smaller islands (Whittaker & Fernández-Palacios, 2007). This means that matrix and generalist species are relatively over-represented in smaller fragments (Matthews *et al.* 2014b), again resulting in reduced overlap. In previous work we have shown that the relatively high incidence of generalist species in smaller fragments reduces the slope of a multimodel ISAR curve for a number of habitat island datasets (Matthews *et al.*, 2014b). This reduction in ISAR slope coupled with the reduced species overlap across islands provides one explanation for the many positive $zDif$ values (Ran-SAC steeper than ISAR) observed.

Our analyses indicated that other variables are also important, independent of nestedness (Table 2), with the minimum number of species (Min.) on an island in a dataset being the second most influential variable explaining variation in $zDif$ (Table 2). The partial dependence plot of this variable (Fig. 2b) indicates that low Min. values have a negative effect on $zDif$; in this case, in many datasets with low Min. values the ISAR is steeper than the Ran-SAC (i.e. $zDif$ is negative). Interestingly, the maximum number of species and the total number of species in the archipelago had low relative influence values (Table 2). It is possible that this finding could be due to an indirect effect of additional variables not

included in our analyses that are correlated with Min. Further research is needed to fully explore the implications of these results.

Landscape context variables (e.g. the habitat matrix, and island type) were relatively unimportant (Table 2). This is surprising as the habitat matrix is considered to be important in determining differences in z value between ISARs and SACs due to the influence of matrix properties on species turnover (see Crist & Veech, 2006). The lack of an effect of these variables is likely due, at least in part, to the coarse matrix and island type classifications utilised in our analyses.

Interpreting variation in ArcRes

Using the simple descriptive metric adopted by Santos *et al.* (2010) we found that the archipelagic point deviated substantially from the ISAR prediction in 19% of datasets with a significant ISAR, compared to 12% in Santos *et al.*'s analyses of true island datasets.

Consistent with our findings for slope differences (i.e. $zDif$; above), NODF had the largest effect on ArcRes (Table 2), while the ISAR more frequently under-predicted than over-predicted the richness of the archipelagic point (Fig. 5a). A role for nestedness in under-prediction of system richness may reflect the fact that habitat islands often contain a large number of singletons (i.e. species that are only sampled in one habitat island in a dataset and whose presence reduces the nestedness of the full data matrix) as a result of factors such as source–sink dynamics and transient species that may be using suitable, but unsampled patches of habitat within the study area, thus enabling their persistence despite low frequencies in the dataset. A recent meta-analysis, using many of the same datasets, has shown that contrary to earlier work, the majority of habitat island systems described in the literature have low levels of nestedness, and indeed that significant anti-nestedness is more common than significant nestedness (Matthews *et al.*, 2015a). Taken together, these findings

bring into question the extrapolation of ISARs (see also Fig. 1b) and certain other species–area curves of similar or composite construction: a common applied use of the ISAR (cf. Whittaker *et al.*, 2005; Ulrich & Buszko, 2007; Hui, 2008).

Implications and conclusions

The choice of species–area curve construction and method of analysis are important considerations when using the SAR for applied purposes (Whittaker *et al.*, 2005; Ulrich & Buszko, 2007; Halley *et al.*, 2013; Matthews *et al.*, 2015b), but conservation biogeographic studies are often unclear about the type of SAR employed and indeed, terminology is inconsistent and disputed across the literature (see e.g. Scheiner, 2003, 2004; Gray *et al.*, 2004a, b; Whittaker & Fernández-Palacios, 2007). Our analyses have provided empirical proof that ISAR and SAC are not equivalent and may differ substantially in fitted parameters (contrary to assumptions in e.g. MacArthur & Wilson, 1967, p10; Rosenzweig, 2005, p19). This distinction has not always been recognised and greater attention should therefore be paid to explicitly identifying the type of species–area curve and method of fit used in future ecological analyses (see also Scheiner, 2003; Ulrich & Buszko, 2007). The present analysis does not argue in favour of one specific species–area function being universally preferable; rather, the choice depends on the aim of study and the data available. For example, SACs are arguably more useful than ISARs for assessing the contribution of different sized islands to a protected area network (Fischer & Lindenmayer, 2002; Watson *et al.*, 2009).

Based on the self-similar scaling properties of the power model, Tjørve & Tjørve (2008; see also Harte *et al.*, 1999) used a form of SAC to show mathematically that the proportional overlap between two areas of the same size can be plotted as a function of z : as z increases, the degree of species overlap decreases. For example, a z of 0.58 equates to 50% of the species being shared between the two areas (Tjørve & Tjørve, 2008). Our results provide

an empirical illustration of this point (e.g. Fig. 3b) and indicate that (a) the SAC is often steeper than the ISAR when the curves are constructed using habitat island data, and (b) the z value of the ISAR only becomes consistently steeper than the z of the SAC when the data are highly nested i.e., the NODF value is approximately 70 or above (e.g. Fig. 3a & Fig. 4b).

It is also evident that for a number of habitat island systems the ISAR is a poor predictor of the overall number of species in an archipelago (e.g. Fig. 5a). Thus, caution should be employed when using the ISAR for extrapolation purposes in fragmented systems, particularly when species overlap is thought to be low (Crist & Veech, 2006). The SAC is likely to provide more accurate results in such contexts as it incorporates information on the degree of nestedness/overlap in the region (Quinn & Harrison, 1988; Hui, 2008). It is already acknowledged that the choice of sampling design is an important consideration in SAR studies, and our results provide more evidence for those who have recently argued for a more consistent SAR theory applicable to general SAR sampling designs (e.g. Whittaker *et al.*, 2005; He & Hubbell, 2013).

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SUPPORTING INFORMATION

Appendix S1 The different types of species–area relationship

Appendix S2 Source paper information

Appendix S3 Expanded study methods

Appendix S4 Supplementary results

BIOSKETCHES

Tom Matthews is a research associate at the University of Oxford. He is interested in the study of macroecological patterns in habitat islands, and the application of island theory to fragmented landscapes.

Author contributions: T.J.M., K.A.T. and R.J.W. conceived the ideas; T.J.M. collected the data; T.J.M. ran the analyses; and T.J.M. and R.J.W. wrote the paper with a significant contribution from K.A.T, M.K.B., F.R. and F.G. All authors discussed the results and commented on the manuscript.

Appendix 1 - Data sources

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TABLES

Table 1 A glossary of the terms used in this study.

Abbreviation	Full term	Definition
.....	Species–area curve/relationship	Here used as general terms for the relationship between sample area and species richness/number.
ISAR	Island species–area relationship	Whereby the number of species occurring within each of a set of islands is analysed as a function of the area of each island. Equivalent to Scheiner (2003) type IV curves.
SAC	Species accumulation curve	Plots of increasing cumulative species number with increasing sampling effort/area
SL SAC	Small-Large SAC	The order of island incorporation into the SAC starts with the smallest island and increases up to the largest.
Ran-SAC	Random SAC	The order of island accumulation when constructing the SAC is random.
z		The slope parameter of the power SAR model.
z_{Dif}		The difference between the SAC z value and the ISAR z value.
BRT	Boosted regression tree analysis	A regression method which combines a large number of tree models.
Nestedness	The situation in which depauperate island faunas constitute proper subsets of the species in richer islands (see Matthews <i>et al.</i> , 2015a). In the current study, we are interested in species nestedness between isolates.
Anti-nestedness	A dataset which is significantly less nested than expected by chance (Matthews <i>et al.</i> , 2015a).
NODF	Nestedness metric based on overlap and decreasing fill	A nestedness metric based on the twin properties of standardized differences in row and column fills and the overlap of presences in two adjacent columns.
ArcRes	Archipelagic residual	The standardised absolute difference between the observed number of species across all islands in a dataset, and the number of species predicted by the log–log power model (log transformed).

Table 2 The relative contributions (%) of predictor variables for boosted regression tree models developed using cross-validation. The model was fitted using two different response variables: z Dif (number of datasets = 50; 7050 fixed trees), and ArcRes (number of datasets = 66; 2300 fixed trees). z Dif is the difference between the z value of the SAC and the z value of the ISAR, for each dataset. ArcRes is the standardised difference between the cumulative species richness total of the archipelago and the predicted the number of species in the archipelagic point using the ISAR model of the constituent islands. A set of dataset characteristics were used as the predictor variables (see Materials and Methods): Min = minimum, Max = maximum, No = number of, Ar. = area, Isl. = island, Sp. = species. The nestedness of the archipelago was measured using the NODF metric (maximally packed matrix). Island type was a categorical variable indicating whether a dataset was a forested or a non-forest island, and taxon was a categorical variable indicating whether a dataset was a plant, vertebrate or invertebrate dataset. Matrix was a variable indicating the permeability of the surrounding matrix type. Variables are ordered by their relative influence (Rel. influence) in each case.

z Dif		ArcRes	
Variable	Rel. Influence	Variable	Rel. Influence
NODF	24.61	NODF	30.11
Min. Sp.	17.94	No. Sp.	16.62
No. Isl.	17.12	Min. Sp.	13.60
Ar. Ratio	13.15	Ar. Ratio	8.00
Max Ar.	9.80	Latitude	7.15
Longitude	4.53	Longitude	6.33
Latitude	4.52	Max Ar.	5.31
No. Sp.	2.78	Max Sp.	3.44
Max Sp.	2.14	No. Isl.	3.40
Matrix	1.43	Taxon	2.20
Min. Ar.	0.99	Min. Ar.	2.06
Isl. Type	0.88	Isl. Type	0.92
Taxon	0.11	Matrix	0.86

FIGURES

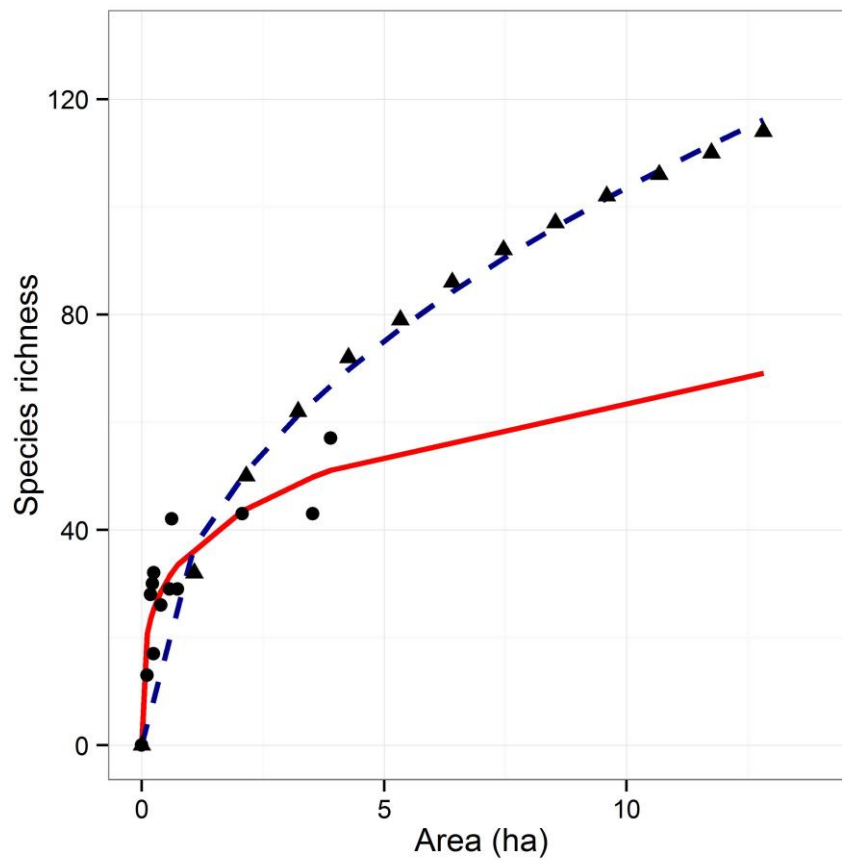


Figure 1 Power model fits constructed from the same habitat island dataset: Gavish *et al.* (2012; invertebrates in a study system from Israel; number of islands = 12; number of species = 114). The fits of the power model (non-linear) to the ISAR data (solid circles; solid line = fit of the model) and the Ran SAC data (solid triangles; dashed line) are shown. For the ISAR model fit, the power model has been extrapolated to the total cumulative area of all habitat islands in the system using the parameters derived from model fitting process. The z values of the models are 0.25 for the ISAR model, and 0.46 for the SAC model.

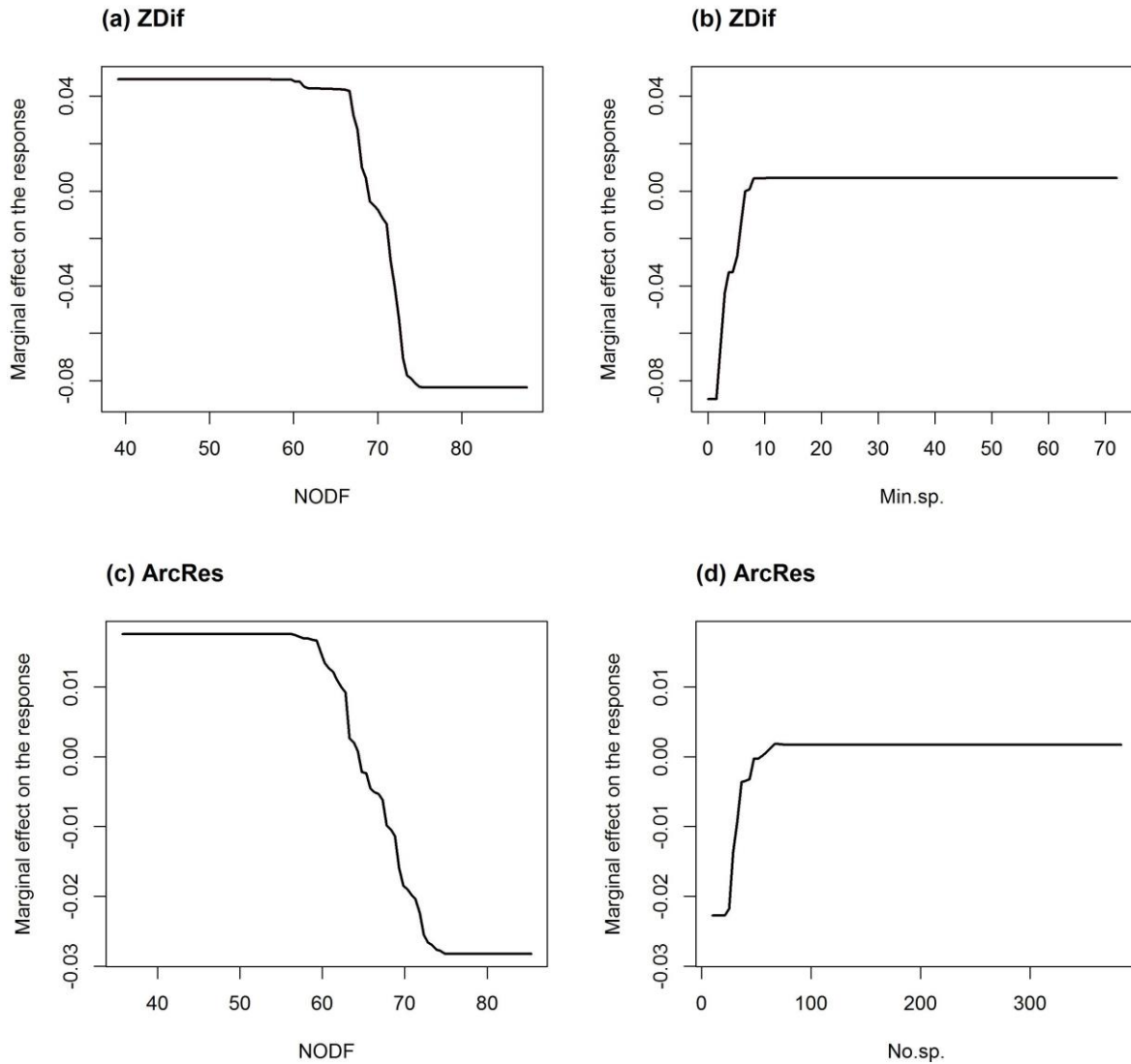


Figure 2 Partial dependence plots for two of the most influential variables in two boosted regression tree (BRT) models. The response variable was $zDif$ (a-b; the difference between the z value of the SAC and the z value of the ISAR, for each dataset), and in the second model the response was $ArcRes$ (c-d; the standardised difference between the cumulative species richness total of the archipelago and the predicted the number of species in the archipelagic point using the ISAR model of the constituent islands). Only $zDif$ values from satisfactory datasets were used ($n = 50$) in (a) and (b), and only $ArcRes$ values from datasets in which the fit of the power (log–log) model was significant ($n = 73$, reduced to 64 after removing nine outliers) were used in (c) and (d). The two predictor variables in the first model are (a) nestedness (NODF; for the relative contributions of each variable see Table 2) and (b) the minimum number of species in a dataset (Min. sp.). The two predictor variables in the second model are (c) nestedness (NODF), and (d) the number of species in a dataset (No. sp.). The y-axis shows the effect of a predictor variable (x-axis) on the response variable after

accounting for the effects of the other model predictors (i.e. the marginal effect of the predictor). The y-axis represents the effect of the predictor on the response, and is not an indicator of the value of the response at a given value of the predictor. A positive y-value indicates that at the given x-value, the effect (based on the model) on the y-value (the response) is positive, and *vice versa*. The ends of the curves represent areas of the plotting space with fewer data points and are relatively uninformative.

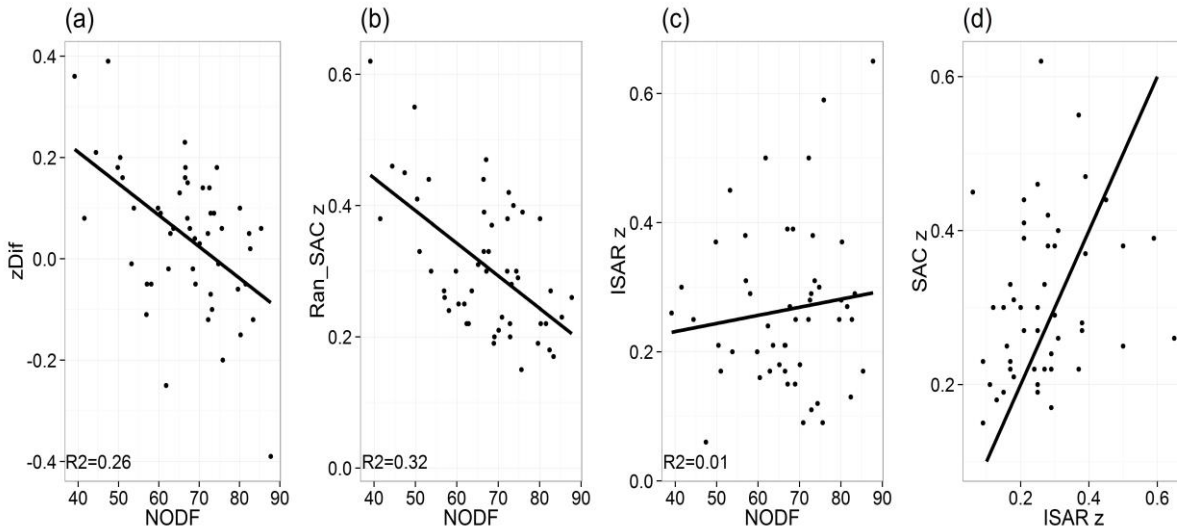


Figure 3 The relationship between nestedness (measured by the NODF metric) and (a) z_{Dif} (the difference between the Ran SAC z value and the ISAR z value, (b) the SAC z value, and (c) the ISAR z value. Only z values from the satisfactory datasets ($n=50$) were used to construct the plots. In each plot, the solid line represents the best line of a linear regression model, and the R^2 of this fit is given on each plot. The z value was significant in each plot apart from (c), which is shown for illustrative value only. Increasing NODF value indicates an increasing degree of nestedness, according to this metric. (d) The z value of the power model fitted using ISAR structured data plotted against z values derived using randomly constructed SAC data. The solid line in (d) represents a 1:1 fit line; points below the line represent datasets in which the ISAR z value was larger than the SAC z value, and vice versa. The z values in all plots relate to the non-linear power SAR model.

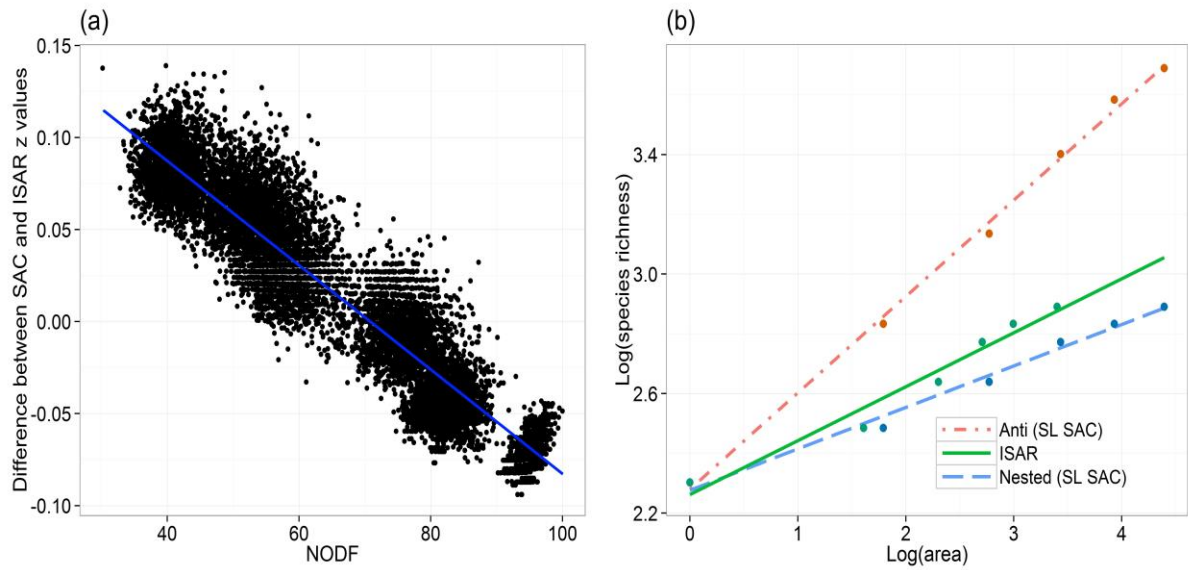


Figure 4 The relationship between nestedness and the variation in z values of ISARs and small-large (SL; the order of island incorporation into the SAC starts with the smallest island and increases up to the largest; see Appendix S1) constructed SACs. For (a), a perfectly nested presence/absence matrix for a set of six islands of varying area was simulated (area of each island = 1, 5, 10, 15, 20 & 30; species richness of each island = 10, 12, 14, 16, 17 & 18). First, the power model (non-linear) was fitted to the data matrix in ISAR form, and the z value recorded. The SL SAC was then constructed using the same data matrix and, again the power model was fitted. The presence/absence matrix was then rearranged to change the level of nestedness, with the constraint that the species richness of each island was kept constant (i.e. the ISAR remained unchanged), although the overall number of species in the archipelago was allowed to vary; the power model was then fitted to both the ISAR and SAC constructed using this new data matrix. This process was repeated iteratively along a gradient of NODF (by rows, i.e. sites) values. A total of 12,000 matrix permutations were simulated. The best fit linear regression line (solid line) through these points (dots) is also shown. (b) The power (log–log) model, fitted to ISAR structured data (solid line, $z = 0.18$), and to SL SAC structured data using the same simulated island data as for (a). The fit of SL SAC curves to two matrix permutations are shown: a perfectly nested set of isolates (dashed line, $z = 0.14$), and a perfectly anti-nested set of isolates (dashed and dotted line, $z = 0.32$).

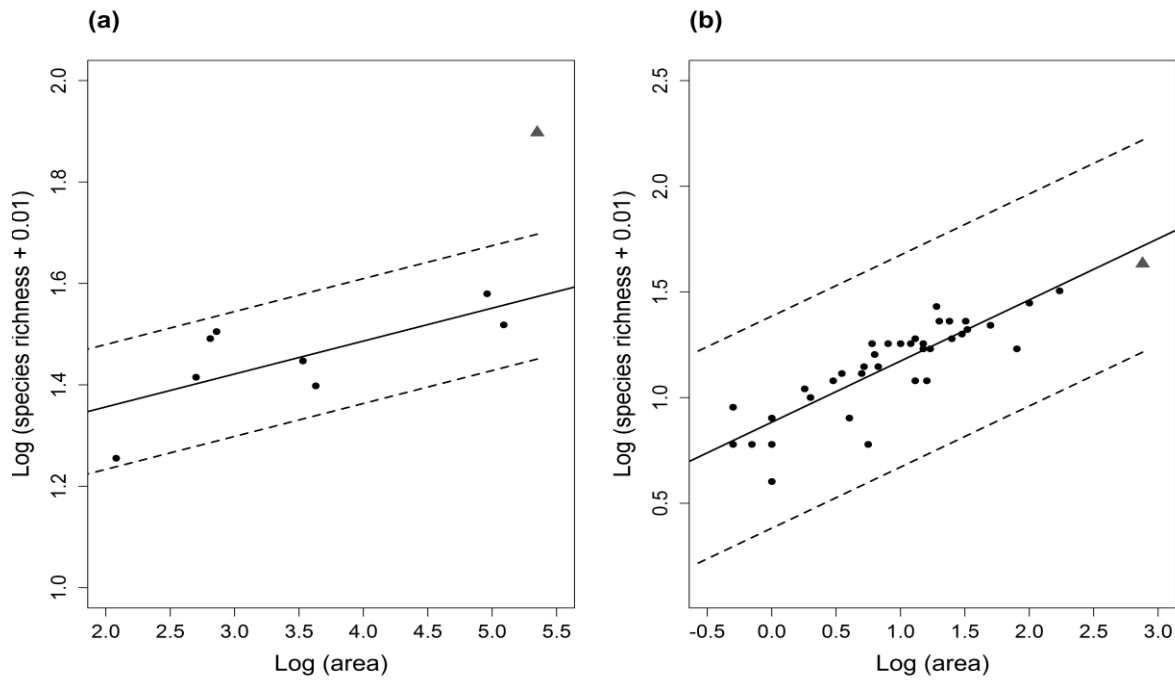


Figure 5 The island species–area relationship (power log–log model) for two habitat island datasets: (a) Benedick *et al.* (2006; invertebrates in Malaysia; number of islands = 8), and (b) this study (birds in the UK; number of islands = 37). The dots represent individual habitat islands and the fit of the power (log–log) model is indicated by the solid line. In both plots the archipelagic point, calculated as the total cumulative area and species richness of all habitat islands in the dataset, and is illustrated as a triangle. The intervals defined by the maximum residual criterion are shown as the dashed line: this criterion relates only to the difference between the ISAR prediction and the archipelagic point, the dashed lines have then just been continued down the y-axis. If the archipelagic point lies outside these bounds (e.g. (a)) it is said to deviate from the prediction of the ISAR calculated using the constituent habitat islands; and *vice versa* (e.g. b; cf. Santos *et al.*, 2010).

SUPPORTING INFORMATION

Island species–area relationships and species–accumulation curves are not equivalent data structures: a synthetic analysis of habitat island datasets

Thomas J. Matthews, Kostas A. Triantis, François Rigal, Michael K. Borregaard and Robert J. Whittaker

Global Ecology and Biogeography

Appendix S1 The different types of species–area relationship

Whilst a number of different types of SAR have been formulated (e.g. Scheiner, 2003), in effect these different forms can be dichotomised into island species–area relationships (‘ISARs’) and species accumulation curves (‘SACs’). The ISAR represents a curve whereby the number of species occurring within each of a set of islands is plotted as a function of the area of each island. The SAC refers to plots of increasing cumulative species number with increasing sampling effort/area. The SAC can be constructed in multiple ways that vary the order in which samples are accumulated to build the curve: in order of increasing or decreasing area or of increasing or decreasing species richness. Finally, the SAC can be constructed in a random manner, whereby the final curve represents an average of multiple permutations of random SAC construction. In our analysis, we construct the random SAC using a simple bootstrap procedure in which the data points were generated by randomly selecting the order of island addition into the SAC at each iteration, and taking the average data point values of 5000 iterations. While a variety of other methods have been proposed to calculate the random SAC (e.g. Colwell & Coddington, 1994; Ugland *et al.*, 2003), we used our bootstrap method as it was (a) relatively simple to compute, (b) enabled easy comparison and integration with our methods for calculating the other SAC structures, (c) has been shown to work in a similar context (Flather, 1996), and (d) does not require abundance data. The order in which samples are combined to construct the SAC determines the shape of the resultant curve, due to e.g. turnover of species between samples and sampling error (Gray *et al.*, 2004). These five types of SAC, in addition to the ISAR, are summarised in Table S1 below. Figure S1 illustrates how these different data structures can result in different curve shapes, when constructed from the same underlying data.

Table S1 The different types of species–area relationship data structures compared in this study. The acronyms are used throughout the paper. SAC refers to the species–accumulation curve.

Data structure	Acronym	Explanation
Island species–area relationship	ISAR	The number of species occurring within each of a set of islands is plotted as a function of the area of each isolate
Small–large SAC	SL	The order of island incorporation into the SAC starts with the smallest island and increases up to the largest
Large–small SAC	LS	The order of island incorporation into the SAC starts with the largest area and works down to the smallest island.
Poor–rich SAC	PR	The order of incorporation starts with the most species-poor island and, and increases up to the most species-rich island.
Rich–poor SAC	RP	The order of incorporation starts with the most species-rich island and up to the most species rich island.
Random SAC	Ran	The order of island accumulation is random.

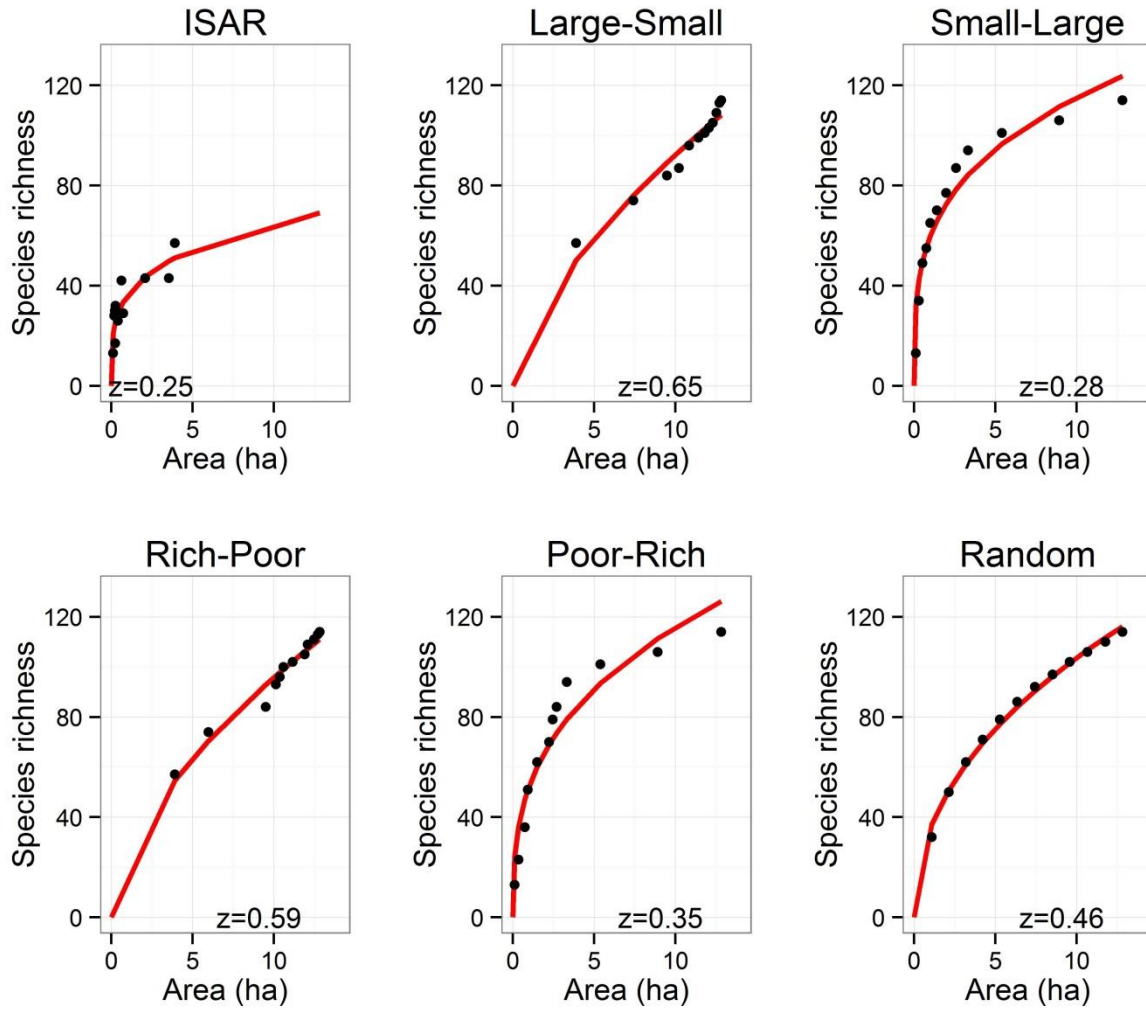


Figure S1 Exemplar fits of the power (non-linear) species–area model to six different data structures (the ISAR and five versions of SAC), constructed from the same habitat island dataset: Gavish *et al.* (2012; invertebrates in a study system from Israel; number of islands = 12; number of species = 114). For each plot data for each of the individual habitat islands (black circles) are shown, alongside the fit of the power model (solid red line). The z value of the model is given in each instance. In the ISAR plot (top left), the fit of the power model has been extrapolated to the total cumulative area of all habitat islands in the system using the parameters derived from the model fitting process. That is, the extrapolated area data point was not included in the model fitting process.

References

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Appendix S2 Source paper information

Data collection

We searched three abstracting services for relevant habitat island studies and datasets between May 2010 and August 2013: JSTOR (1913–2003); ISI Web of Knowledge (1980–2013); and BIOSIS Biological Abstracts (1980–2003). We used search keywords encompassing different combinations of ‘habitat islands’, ‘species richness’, and ‘forest fragments’. We searched through the reference lists of papers found in these abstracting services for additional studies. Certain datasets were obtained from the authors of the source papers, whilst others were supplemented with additional data from the source paper authors. Following Matthews et al. (2014), our criteria for inclusion of source data were as follows: (i) habitat islands were defined as discrete patches of habitat surrounded by contrasting matrix habitat (we also included a small number of datasets consisting of protected areas in which the contrast between the islands and the intervening matrix was not so pronounced); (ii) the number of habitat islands within each dataset was at least five; and (iii) the area of each habitat island was listed, and the species lists provided. For the purposes of this study, we considered a few island systems within an aquatic matrix (e.g. forest fragments within a reservoir) as habitat islands, as the range of island areas and dominant assembly processes are more similar to habitat islands than oceanic islands (see Matthews, 2015).

Classifying the matrix

For each dataset we recorded the predominant land-use type of the matrix surrounding the habitat islands using information in the source papers. Where possible we also checked this information using Google Earth, although this check was undertaken tentatively as many datasets were compiled decades ago and it is likely that the land-use has changed in the interim. These land-use categories were then converted into a 5-point scale of matrix permeability in which 1 represented the most permeable matrices, and 5 represented the less permeable matrices. This classification scale was determined using the results of Matthews *et al.* (2015), a study which examined how the z value of the island species–area relationship varied across multiple habitat island types. Obviously, this is a coarse approach, but obtaining fine scale matrix data for such a large number of studies is infeasible; particularly as such information is rarely, if ever, presented in the source papers. The classification was as follows:

1 – Secondary/re-growth forest (if the island type was forest) or scrub (if the island type was grassland)

2 – Scrub/grassland (if the island type was forest)

3 – Agriculture (e.g. crops, pasture)

4 – All mountaintop islands were included in this category as the matrix between them is known to be relatively impermeable (see Matthews *et al.*, 2015).

5 – Urban land-uses and water (i.e. flooded forest fragment archipelagos)

Table S2 Description of the source papers, including the taxon, number of islands, the minimum, maximum and total cumulative island area in a dataset, and the number of species. Inver. = Invertebrates; Ver. = Vertebrates; and Pla. = Plants. The full references are provided in Appendix 1.

Source	Taxon	No. Islands	Min. Area (Ha)	Max. Area (Ha)	Tot. Area (Ha)	No. Sp.
Báldi & Kisbenedek (1999)	Inver.	27	0.018	40	84.7	36
Behle (1978)	Ver.	14	2849	95312	391349	75
Benedick <i>et al.</i> (2006)	Inver.	8	120.2	123026.9	223873.7	79
Blake & Karr 1984	Ver.	12	1.8	600	955.9	26
Brotons & Herrando (2001)	Ver.	51	0.1	38.4	325.6	37
Brown (1971)	Ver.	17	3108	305102	1455062	13
Brown (1978)	Ver.	13	12173	191142	795648	11
Brown (1978)	Ver.	19	3107.988	305100.8	1490539	16
Cabrera-Guzmán & Reynoso (2012)	Ver.	6	1.4	17.4	38	35
Castelletta <i>et al.</i> 2005	Ver.	6	7	935	1640	132
Charles & Ang (2010)	Ver.	9	0.01	149	210	18
Cieślak & Dombrowski (1993)	Ver.	50	0.04	15.4	109.3	50
Crooks (2002)	Ver.	29	2	102	458	7

Crowe (1979)	Pla.	26	0.011	0.7	3.2	128
Daily & Ehrlich (1995)	Inver.	8	3	227	318	38
Dalecky <i>et al.</i> (2002)	Ver.	38	0.1	67	252.5	28
Darlington <i>et al.</i> (2001)	Inver.	25	0	0.4	0.9	23
Davies <i>et al.</i> (2003)	Inver.	10	0.2	80	158.6	95
Davis <i>et al.</i> (1988)	Ver.	12	800	2181300	3305300	12
Dickman (1987)	Ver.	9	0.23	2.4	8.8	16
Dickman (1987)	Ver.	12	0.21	7.4	15.1	20
Dinesen <i>et al.</i> (2001)	Ver.	18	100	52200	183000	9
dos Anjos & Boçon 1999	Ver.	12	0.5	840	962.5	188
dos Santos <i>et al.</i> (2007)	Pla.	13	12.4	244.9	977.5	248
Edenius & Sjöberg (1997)	Ver.	18	2	178	774	23
Essl & Dirnböck (2012)	Inver.	60	0.032	7	52.7	33
Feeley (2003)	Ver.	26	0.2	180	289.1	43
Fernández-Juricic (2000)	Ver.	25	1	118.2	486.1	32
Filgueiras <i>et al.</i> (2011)	Inver.	19	10.1	3500	5113.9	30
Flaspohler <i>et al.</i> 2010	Ver.	18	0.07	55.7	168	10
Ford (1987)	Ver.	20	0.14	18	48.2	48
Galle (2008)	Inver.	15	0.009	0.5	1.2	51
Galli <i>et al.</i> (1976)	Ver.	10	0.01	24	52.7	35
Ganzhorn <i>et al.</i> (1999)	Ver.	7	3	600	1261	10
Gaublomme <i>et al.</i> (2008)	Inver.	10	5.3	88.2	276.7	91
Gavish <i>et al.</i> (2012)	Inver.	12	0.11	3.9	12.8	114
Gavish <i>et al.</i> (2012)	Inver.	12	0.06	2.8	10.6	115
Gillespie & Walter (2001)	Ver.	7	420	16804	51144	69
Haila <i>et al.</i> (1993)	Ver.	13	0.7	4.4	30.3	29
Hatt (1948)	Ver.	15	1.01	15125.5	24516.2	10
Hatt (1948)	Ver.	15	1.01	15125.5	24516.2	152
Hatt (1948)	Ver.	15	1.01	15125.5	24516.2	31

Hattori & Ishida (2000)	Pla.	38	0.003	7.8	17.7	177
Holbech (2005)	Ver.	15	2360	58790	258020	147
Hu <i>et al.</i> (2012)	Pla.	152	0.02	130.8	839.5	383
Ishida <i>et al.</i> (1998)	Pla.	29	0.025	16.4	33.5	119
Johnson (1975)	Ver.	20	3367	211085	1480444	74
Kelt (2000)	Ver.	14	1.89	124.9	246.8	9
Kitchener <i>et al.</i> (1980)	Ver.	23	34	5119	22841	69
Kitchener <i>et al.</i> (1980)	Ver.	23	34	5119	22841	24
Kratter (1992)	Ver.	20	1	106830	247791	48
Langrand (1995)	Ver.	8	0.64	1250	1512.6	51
Lomolino & Davis (1997)	Ver.	24	600	2116500	3683700	23
Lomolino & Perault (2001)	Ver.	20	0.93	58.9	470.5	18
Lomolino <i>et al.</i> (1989)	Ver.	27	689	1113433	1913140	23
Lumaret <i>et al.</i> (1997)	Pla.	8	0.005	0.2	0.7	193
Maldonado-Coelho & Marini (2003)	Ver.	9	3.6	384.5	865.2	72
Marini (2001)	Ver.	6	7.5	230	479.5	145
Matthiae & Stearns (1981)	Ver.	22	0.4	40	103.5	13
McCollin (1993)	Ver.	16	0.74	14.5	76.8	49
Meynard & Quinn (2008)	Ver.	9	180	60830	169040	51
Miyashita <i>et al.</i> (1998)	Inver.	7	0.2	27	57.4	34
Miyashita <i>et al.</i> (1998)	Inver.	9	0.4	15.1	46.4	52
Mohd-Azlan & Lawes (2011)	Ver.	13	0.67	594.5	1720.1	70
Newmark (1991)	Ver.	10	0.1	521	566.9	31
Nores (1995)	Ver.	9	1300	360000	1085100	54
Nufio <i>et al.</i> (2011)	Inver.	13	0.85	37.3	149.8	38
Nyeko (2009)	Inver.	6	10	150	452	45
Peltzer <i>et al.</i> (2003)	Ver.	8	0.001	0.7	1.7	20
Pineda & Halfpeter (2004)	Ver.	5	16	72	222	21

Ramanamanjato (2000)	Ver.	11	10	457	1925	21
Ramanamanjato (2000)	Ver.	11	10	457	1925	41
Ribas <i>et al.</i> (2005)	Inver.	18	3	299	637	136
Rosenblatt <i>et al.</i> (1999)	Ver.	10	1.8	600	817.6	16
Ruiz-Gutiérrez <i>et al.</i> (2008)	Ver.	7	1.5	262	369.5	184
Shreeve & Mason (1980)	Inver.	22	2	175	938	26
Silva (2001)	Ver.	14	0.07	5.8	38	11
Silva & Pôrto (2009)	Pla.	10	22.976	2628.6	5091.1	99
Simberloff & Martin (1991)	Ver.	35	0.4	101	351.9	45
Smith <i>et al.</i> (1996)	Ver.	26	0.5	174	411.3	19
Suarez <i>et al.</i> (1998)	Inver.	40	0.4	101.6	851.1	45
Summerville <i>et al.</i> (2002)	Inver.	25	0.004	0	0.3	30
Matthews <i>et al.</i> (2014)	Ver.	40	0.11	150	525.5	60
Matthews <i>et al.</i> (2014)	Ver.	42	0.06	110	557.5	47
Matthews <i>et al.</i> (2014)	Ver.	41	0.2	150	926.1	55
Matthews <i>et al.</i> (2014)	Ver.	37	0.5	172	757.8	43
Tonn & Magnuson (1982)	Ver.	18	2.4	89.8	522.3	23
Usher & Keiller (1998)	Inver.	18	0.5	30.6	115.4	214
Vallan (2000)	Ver.	7	0.16	1250	1433	28
Viveiros de Castro & Fernandez (2004)	Ver.	8	1.2	13.3	70.7	11
Wang <i>et al.</i> (2010)	Ver.	14	0.97	1289.2	1565.1	11
Wang et al (2010)	Ver.	42	0.3	1289.2	1761.3	93
Watson (2003)	Ver.	17	2	159246	650075	129
Weaver & Kellman (1981)	Pla.	10	0.82	8.1	36	30
Willson <i>et al.</i> (1994)	Ver.	10	0.5	350	721.5	25
Yong <i>et al.</i> (2011)	Ver.	6	1.1	184.7	403.2	75
Zimmerman & Bierregaard (1986)	Ver.	7	1	500	722	40

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Appendix S3 Expanded study methods

SAR model comparison

We fit a set of 20 SAR models to both the ISAR and SAC forms of each dataset. After unsatisfactory fits (e.g. fits with non-normal residuals, or fits in which the model fitting algorithm did not converge) were removed, we considered the model with the lowest Akaike's information criterion corrected for small sample size (AIC_c), as providing the best fit (Burnham & Anderson, 2002). The 20 models are listed as having specific shapes (i.e. either of linear, convex or sigmoidal). However, the actual observed shape of the model fits can vary from these listed shapes. For example, sigmoidal models can produce observed convex or linear fits depending on the parameter values. Thus, we used the algorithm of Triantis *et al.* (2012) to determine the observed shape of the best fitting model. This algorithm works by first determining whether the observed shape of a model fit was linear. This is achieved by comparing the shape of the model fit with a straight line which joins the fitted values for the smallest and largest island area in the dataset. One hundred equally spaced increments between the smallest and largest island area are created, and the shape of the fit is classified as linear if all of the differences between the number of species calculated from the straight line and from the model fit are less than 0.001 species. If the shape of the fit was not classified as linear, the second derivative of the model fit was examined for an inflexion point to distinguish between a convex and sigmoid shape. If the second derivative changed sign, the model fit displayed an inflection point in the observed range of the data, and thus the fit was classified as sigmoid; if not, convex.

Determining the degree of deviation of the archipelagic point

We plotted the ISAR data in log–log space and fitted the power (log–log; base 10) model to these data. We then calculated the archipelagic point as the cumulative total area of all habitat islands in the dataset plotted against the cumulative species richness total; and predicted the number of species in the archipelagic point ('SApred') using the ISAR model of the constituent islands. To determine if SAobs deviated substantially from SApred we used one of the simple (rule of thumb) indices employed by Santos *et al.* (2010), first calculating the residuals of the ISAR model (observed minus predicted species richness) and then selecting the maximum absolute value (i.e. MaxRes). MaxRes was then expressed as a proportion of the predicted log(species richness) of the island from which MaxRes was generated from (PropMaxRes). SAobs was said to deviate from SApred if SAobs was greater than $SApred \pm (SApred \times PropMaxRes)$. We also followed Santos *et al.* (2010) and calculated the archipelagic residual ('ArcRes') as the difference between log(SAobs) and SApred, standardised by log(SAobs).

Appendix S4 Supplementary results

Table S3 Model parameters (c and z) for the power model (non-linear) fitted to the ISAR (IS), Large-Small SAC (LS), Small-Large SAC (SL), Rich-Poor SAC (RP), Poor-Rich SAC (PR), and Ran SAC (R) versions of each dataset (see Table S1 in Appendix S1 for descriptions of each SAC type; full references given in Appendix S1). The ArcRes (AR) and SApred (SA) values for each dataset are also provided (for definitions, see Appendix S3). The dataset numbers correspond to the order in Table S2 in Appendix S2. NA relates to situations where it was not possible to fit the power model to certain SAC forms due to the fact that either the largest or most species rich island in a dataset contained the full set of species in that dataset.

No.	IS c	IS z	LS c	LS z	SL c	SL z	RP c	RP z	PR c	PR z	R c	R z	AR	SA
1	9.4	0.17	3.8	0.51	18.6	0.14	5.6	0.43	13.5	0.21	8.7	0.33	0.2	18
2	10.2	0.15	1.6	0.3	12.6	0.15	29.6	0.07	2.5	0.26	7.9	0.18	<0.1	71
3	18.0	0.06	<0.1	1.42	15.2	0.14	<0.1	0.75	11.5	0.16	0.3	0.45	0.2	38
4	5.3	0.25	6.2	0.21	4.5	0.27	6.2	0.21	4.6	0.27	4.4	0.27	-0.1	36
5	9.4	0.17	3.4	0.41	16.2	0.16	6.9	0.29	12.4	0.20	10.4	0.22	0.1	26
6	0.1	0.37	0.4	0.26	0.4	0.26	9.9	0.02	0.1	0.33	0.6	0.22	-0.2	21
7	1.1	0.16	0.5	0.23	1.8	0.14	2.8	0.10	1.1	0.17	0.8	0.20	0.1	10
8	0.3	0.29	0.2	0.3	1.2	0.19	5.9	0.07	0.9	0.21	0.7	0.22	-0.1	20
9	9.4	0.32	4.3	0.58	7.2	0.44	3.7	0.62	7.2	0.44	7.4	0.43	<0.1	36
10	38.8	0.13	0.4	0.79	41.9	0.16	33.6	0.18	41.1	0.15	12.0	0.33	0.1	104
11	2.2	0.37	0.6	0.63	1.8	0.42	2.4	0.38	1.2	0.50	1.0	0.55	0.3	8
12	9.6	0.50	28.2	0.12	12.8	0.31	27.3	0.13	11.0	0.34	16.1	0.25	-0.4	248
13	4.7	0.03	1.2	0.29	5.4	0.05	NA	NA	1.2	0.30	4.2	0.09	0.2	5
14	64.7	0.28	69.1	0.55	97.5	0.32	102	0.20	78.0	0.46	90.2	0.33	<0.1	106
15	6.7	0.22	<0.1	1.77	12	0.22	11.5	0.20	4.9	0.35	2.0	0.52	0.1	24
16	4.0	0.38	9.2	0.21	7.0	0.29	15.2	0.11	3.1	0.43	6.6	0.27	-0.3	78
17	12.4	0.24	23.2	0.94	28	0.24	24.8	0.46	28.1	0.52	24.6	0.38	-0.1	36
18	43.5	0.00	0.3	1.11	72.5	0.07	68.1	0.07	18.1	0.31	21.3	0.30	0.2	39
19	0.1	0.31	0.2	0.27	0.2	0.28	0.3	0.26	0.1	0.36	<0.1	0.40	-0.7	75
20	9.0	0.11	6.6	0.4	11.8	0.20	13.7	0.05	8.5	0.31	9.9	0.23	<0.1	17
21	10.2	0.17	8.9	0.31	12.5	0.21	17.4	0.05	6.8	0.44	11.2	0.23	0.1	13
22	2.0	0.10	1.3	0.16	3.1	0.10	6.2	0.03	0.1	0.39	1.2	0.17	0.2	6

23	52.9	0.15	<0.1	2.13	63.4	0.17	<0.1	2.13	59.0	0.18	21.7	0.32	<0.1	159
24	89.8	0.01	0.6	0.86	104.5	0.14	97.7	0.14	22.2	0.34	26.5	0.33	0.2	96
25	1.1	0.50	4.5	0.25	1.4	0.43	10.5	0.12	0.6	0.55	1.9	0.38	-0.1	36
26	13.1	0.15	2.8	0.62	19.6	0.16	16.0	0.18	14.3	0.24	16.2	0.19	0.1	27
27	5.1	0.39	37.6	0.02	10.2	0.25	31.0	0.06	4.5	0.41	5.3	0.37	<0.1	41
28	7.9	0.18	2.3	0.42	8.6	0.23	18.6	0.09	6.4	0.25	6.4	0.26	0.1	24
29	1.7	0.31	0.2	0.57	8.0	0.15	0.8	0.43	1.0	0.40	0.8	0.43	0.2	13
30	4.2	0.17	6.9	0.07	4.5	0.13	6.9	0.07	4.2	0.14	3.2	0.23	<0.1	10
31	21.4	0.18	14.2	0.31	27.9	0.17	23.7	0.18	24.6	0.19	22.3	0.21	0.0	47
32	19.1	0.15	38.2	1.14	51.3	0.25	48.1	0.23	48.7	0.37	48.2	0.45	0.3	19
33	16.3	0.25	34.5	<0.1	19.3	0.18	34.8	<0.1	18.7	0.19	17.2	0.19	-0.1	52
34	2.7	0.17	3.6	0.14	3.2	0.17	3.5	0.15	2.2	0.21	0.9	0.34	0.1	9
35	34.2	0.03	0.5	0.93	31.1	0.2	26.4	0.23	0.2	1.07	12.9	0.35	0.2	35
36	36.2	0.25	20.9	0.65	59.9	0.28	24.4	0.59	52.1	0.35	35.7	0.46	0.1	74
37	42.6	0.21	22.6	0.68	64.2	0.27	38.7	0.46	52.8	0.34	44.6	0.41	0.1	71
38	7.3	0.17	0.2	0.55	4.9	0.25	6.8	0.21	4.9	0.24	1.2	0.38	0.1	47
39	9.4	0.31	14.5	0.19	11.9	0.25	16.6	0.17	8.0	0.36	10.0	0.32	0.1	23
40	0.2	0.39	1.5	0.18	1.0	0.23	1.1	0.22	0.2	0.42	0.1	0.47	-0.4	28
41	8.3	0.28	19.9	0.20	17.7	0.21	19.3	0.20	8.2	0.29	2.3	0.42	<0.1	160
42	1.3	0.26	<0.1	2.14	1.7	0.30	<0.1	0.69	1.4	0.32	0.1	0.62	-0.3	98
43	84.9	0.16	27.4	0.65	131.1	0.15	49.6	0.44	127.7	0.15	88.8	0.25	<0.1	164
44	40.3	0.09	8.6	0.23	38.0	0.11	45.3	0.09	34.3	0.12	23.4	0.15	<0.1	127
45	58.4	0.10	0.1	1.29	163.9	0.15	31.0	0.37	92.0	0.21	57.5	0.28	0.2	116
46	47.9	0.21	16.6	0.56	64.0	0.21	29.1	0.40	57.9	0.23	47.4	0.27	<0.1	107
47	2.7	0.23	0.7	0.33	6.7	0.17	21.0	0.09	1.5	0.27	2.9	0.23	0.1	61
48	4.2	0.01	<0.1	1.14	3.1	0.22	5.3	0.10	1.5	0.30	1.9	0.29	0.4	4
49	5.8	0.17	<0.1	0.80	4.5	0.29	9.7	0.20	2.3	0.34	1.7	0.37	0.1	40
50	2.2	0.20	0.1	0.52	1.5	0.29	2.0	0.25	0.7	0.35	1.2	0.30	<0.1	21
51	2.1	0.27	43.1	0.01	9.1	0.14	40.7	0.01	8.6	0.14	3.1	0.22	0.1	36
52	21.1	0.13	NA	NA	25.1	0.11	NA	NA	24.3	0.12	14	0.18	<0.1	60
53	0.2	0.30	<0.1	0.53	0.8	0.23	<0.1	0.65	0.1	0.36	0.1	0.38	-0.1	29
54	4.7	0.14	2.7	0.31	7.9	0.15	9.5	0.10	2.5	0.32	4.1	0.25	0.2	11

55	0.4	0.27	<0.1	0.68	0.9	0.24	0.2	0.32	0.3	0.31	0.2	0.33	-0.1	30
56	74.0	<0.1	237.4	0.70	209.7	0.15	214.3	0.23	245.3	0.62	234.3	0.42	0.2	70
57	30.0	0.09	4.4	0.41	38.1	0.10	3.7	0.43	31.7	0.13	16.0	0.23	0.1	53
58	30.6	0.21	8.1	0.46	34.0	0.23	7.9	0.47	31.0	0.25	12.2	0.40	0.1	107
59	5.4	0.23	13.0	<0.1	8.0	0.10	13.0	<0.1	5.7	0.19	6.5	0.16	<0.1	14
60	16.5	0.18	7.4	0.44	19.8	0.23	24.1	0.16	16.7	0.25	16.3	0.26	0.1	38
61	18.1	0.04	0.1	0.52	10.2	0.14	5.9	0.18	7.4	0.16	1.9	0.28	0.1	29
62	11.8	0.21	1.5	0.77	12.9	0.25	16.2	0.18	11.9	0.24	7.0	0.39	<0.1	29
63	20.1	0.18	13.4	0.34	24.7	0.20	15.1	0.32	22.8	0.22	15.9	0.31	0.1	41
64	36.3	0.03	0.5	0.66	36.0	0.10	36.1	0.09	23.8	0.13	17.2	0.19	0.1	46
65	7.4	0.21	<0.1	2.27	9.5	0.20	<0.1	2.16	9.2	0.20	1.9	0.44	<0.1	32
66	0.1	0.46	14.7	0.09	0.7	0.32	18.8	0.08	0.1	0.44	0.1	0.45	<0.1	45
67	9.9	0.25	5.7	0.39	9.6	0.28	18.2	0.15	8.4	0.29	8.6	0.30	<0.1	33
68	15.2	0.14	13.7	0.19	14.3	0.19	14.2	0.19	9.2	0.25	7.2	0.30	0.1	34
69	22.4	0.59	18.6	0.12	16.6	0.38	18.7	0.12	16.6	0.38	16.8	0.39	<0.1	21
70	5.2	0.19	2.9	0.34	6.2	0.24	1.4	0.50	4.3	0.31	2.0	0.44	0.1	14
71	0.4	0.65	17.7	0.02	0.7	0.46	17.7	0.02	0.7	0.45	3.0	0.26	-0.3	59
72	4.4	0.30	14.5	0.14	3.9	0.30	13.8	0.15	3.4	0.32	4.7	0.29	<0.1	48
73	12.3	0.34	0.9	0.78	23.4	0.28	0.8	0.80	13.3	0.38	10.6	0.40	<0.1	133
74	7.3	0.11	1.2	0.39	7.3	0.13	11.6	0.04	6.8	0.14	4.3	0.20	<0.1	15
75	55.1	0.17	2.2	0.75	59.4	0.20	2.9	0.70	62.1	0.19	27.3	0.33	<0.1	146
76	5.1	0.29	11.4	0.12	8.3	0.18	18.4	0.05	3.7	0.30	8.4	0.17	-0.3	65
77	5.9	0.03	3.5	0.30	7.9	0.08	7.6	0.11	2.7	0.41	4.9	0.23	0.3	6
78	15.6	0.11	<0.1	1.41	12.5	0.25	17.8	0.20	2.5	0.42	1.4	0.50	0.2	42
79	6.5	0.38	8.5	0.29	6.4	0.36	13.2	0.21	4.7	0.41	9.2	0.28	-0.4	211
80	2.3	0.36	8.9	0.13	3.2	0.27	9.3	0.12	1.5	0.41	1.5	0.43	0.1	14
81	2.0	0.45	1.4	0.51	2.5	0.44	4.1	0.36	0.1	0.89	2.4	0.44	0.1	31
82	565	1.00	37.2	0.17	49.8	0.46	37.4	0.17	58.7	0.65	44.9	0.35	-0.6	220
83	10.0	0.29	9.1	0.30	20.0	0.19	9.5	0.29	14.5	0.25	14.0	0.24	0.1	50
84	9.5	0.24	10.3	0.23	22.9	0.12	10.9	0.23	11.1	0.24	12.2	0.22	0.1	35
85	6.6	0.31	6.0	0.32	11.6	0.25	9.5	0.26	8.5	0.29	9.6	0.26	-0.1	68
86	8.8	0.25	10.3	0.21	14.6	0.17	13.4	0.18	10.7	0.22	11.4	0.20	-0.1	52

87	3.2	0.24	0.3	0.71	5.6	0.24	9.6	0.14	3.3	0.32	2.3	0.37	0.1	19
88	88.5	0.05	17.1	0.53	126	0.12	91.2	0.18	76.8	0.22	69.8	0.24	0.1	113
89	10.6	0.12	3.8	0.27	13.7	0.10	5.3	0.22	12.7	0.11	3.4	0.30	0.1	24
90	5.1	0.20	6.5	0.13	5.6	0.17	11.0	<0.1	4.3	0.22	5.8	0.16	<0.1	12
91	2.8	0.13	<0.1	0.80	5.2	0.11	2.3	0.2	2.8	0.20	1.0	0.33	0.2	6
92	23.9	0.15	0.1	0.89	38.2	0.13	0.1	0.89	32.0	0.16	10.1	0.30	0.1	70
93	7.3	0.20	0.4	0.44	21.5	0.14	14.9	0.16	14.8	0.17	2.5	0.30	0.1	101
94	12.1	0.11	3.9	0.58	11.5	0.28	14.8	0.20	5.0	0.49	9.2	0.33	0.2	18
95	9.0	0.13	4.9	0.24	13.9	0.10	4.5	0.26	13.2	0.11	3.9	0.29	0.1	21
96	10.5	0.29	1.7	0.63	10.9	0.33	3.0	0.54	8.4	0.37	3.8	0.50	0.1	58
97	7.1	0.28	13.4	0.17	7.0	0.27	18.2	0.12	7.0	0.27	3.5	0.38	<0.1	45