

REVIEW

Pathogenic stromal cells as therapeutic targets in joint inflammation

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1 **Abstract**

2

3 Knowledge of how the joint functions as an integrated unit in health and disease
4 requires an understanding of the stromal cells populating the joint mesenchyme,
5 including fibroblasts, tissue resident macrophages and endothelial cells. Physiological
6 and pathological mechanisms in these mesenchymal cells that define the joint have
7 begun to cast new light on why joint inflammation persists. In this review, we highlight
8 how the shared embryological origins of fibroblasts and endothelial cells may shape
9 the behaviour of these cell types in diseased adult tissues. We review the molecular
10 mechanisms by which cells of mesenchymal origin sustain inflammation in the
11 synovial membrane and tendons, highlighting the importance of recently discovered
12 fibroblast subtypes and their associated cross talk with endothelial cells, tissue
13 resident macrophages and leukocytes. Finally, we discuss how this knowledge shapes
14 the future therapeutic landscape, emphasising the requirement for new strategies to
15 address the pathogenic stroma and associated cross talk of leukocytes with cells of
16 mesenchymal origin.

17

18 **Key points**

19

- 20 • Joint inflammation and tissue damage are mediated by stromal cells of
21 mesodermal origin
- 22 • Stromal activation and “memory” of previous inflammatory insults are shared
23 mechanisms exhibited by fibroblasts, tissue resident macrophages and
24 endothelial cells
- 25 • Recent advances characterising the phenotype and function of cells of
26 mesenchymal origin highlight the distinct fibroblast subtypes mediating joint
27 inflammation and tissue damage
- 28 • Mesenchymal stromal cell niches and their interactions with leukocytes are
29 implicated in the persistence of joint inflammation
- 30 • To be effective, strategies to treat residual joint disease should target
31 pathogenic stroma and associated immune cell cross talk

32

33

34 **Introduction**

35

36 Chronic inflammatory diseases affecting joint soft tissues include arthritis (synovium
37 and cartilage), enthesopathy and tendinopathy. Collectively, these diseases
38 comprise a significant global economic burden¹. Each are characterized by
39 inflammation of mesenchymal tissues that form the synovium, tendons, ligaments
40 and joint capsule and in some cases structural damage to bone and cartilage.
41 Inflammation of these tissues is broadly characterized by leuckocyte infiltration,
42 fibroblast accumulation and neovascularization supporting cell expansion. In this
43 article, we first review the pathophysiological basis of inflammation and tissue
44 damage with respect to the embryological origins of joint mesenchymal tissues. We
45 next discuss the stromal cell types populating joint mesenchymal tissues, including
46 fibroblasts, tissue resident macrophages (TRM) and endothelial cells (vascular and
47 lymphatic), highlighting their contribution and roles in chronic synovial inflammation
48 and tissue damage. Finally, we discuss potential future therapeutic strategies to
49 target inflammation across joint mesenchymal tissues that address the pathogenic
50 stroma and associated immune cell cross talk.

51

52 **1.0 Embryological origins of the tissues that mediate inflammation** 53 **and damage across the whole joint organ**

54

55 Inflammation and tissue damage are pivotal pathological processes affecting
56 structures across the whole joint organ. To further understand the mechanisms and
57 inter-relationships underpinning these fundamental disease processes, it is important
58 to consider the origins of joint tissues, given that an organ is best defined by its
59 embryological origin as well as function. This section discusses how the embryological
60 and anatomical origins of the tissues that comprise the joint might shape inflammation
61 and tissue damage, highlighting how this knowledge informs understanding of
62 'disease patterns' across the joint.

63

64 The anatomical basis of inflammation and tissue damage relative to their
65 embryological origin is summarized in Figure 1. Although parts of the axial skeleton
66 derive from neural crest, mesoderm is the precursor for mesenchymal tissues

67 comprising the appendicular skeleton, synovium, cartilage, tendons, ligaments, joint
68 capsule and their associated lymphatics and vasculature. These joint soft tissues are
69 predominantly composed of cells of mesenchymal origin, including fibroblasts,
70 vascular and lymphatic endothelial cells and TRM. The embryological origins of
71 stromal cells may shape the behaviour of these cell types in diseased adult tissues.
72 Notably, mesoderm derived fibroblast and endothelial cell populations both undergo
73 sustained phenotypic changes after exposure to inflammatory stimuli, exhibiting
74 stromal activation and a form of tissue ‘memory’^{2,3}. However, the distinct molecular
75 markers expressed by these cell types vary, as we later discuss. TRM also exhibit
76 complex activation states and “memory”⁴. The origins and renewal of TRM have been
77 extensively reviewed and will not be repeated here ⁵⁻⁸. The majority of TRM are
78 established during embryonic development and persist into adulthood, rather than
79 replacement from circulating adult monocytes ^{7,9-14}. During early gestation,
80 macrophages are first observed and expand in the extraembryonic yolk sac during
81 primitive hematopoiesis. Yolk sac derived hematopoietic stem cells (HSCs) emerge to
82 form bone marrow precursor cells, which subsequently give rise to all immune cell
83 lineages ^{7,15} (Figure 1). Importantly, yolk sac derived TRM are phenotypically distinct
84 from HSC derived progeny ¹⁰. The subspecialized adult tissue niches which TRM
85 occupy dictate heterogeneity in the phenotype and functions of these cells in health
86 and disease ¹⁶. We next review how these mesenchymal cell populations are
87 implicated in mediating inflammation and tissue damage in joint disease.

88

89 **2.0 Cells of mesenchymal origin in the joint**

90

91 In this section, we focus exclusively on cells of mesenchymal origin including
92 fibroblasts, endothelial cells and TRM rather than on haematopoietically derived cells
93 whose role in these processes (in particularly inflammation and damage) has been
94 well documented¹⁷⁻¹⁹. We discuss the roles of these cells in normal joint physiology
95 and their impact on inflammation and damage in joint disease. We highlight the
96 recently identified mechanisms implicated in sustaining synovial inflammation,
97 discussing the molecular features and pathological phenotypes of fibroblast subtypes,
98 endothelial cells and TRM.

99 **2.1 Fibroblasts and the healthy joint**

100

101 The term 'stroma' was originally derived from the Greek word describing "a platform
102 on which to lie" and is used to describe the supporting substance of tissue. Its principle
103 role is to maintain the microenvironment required by the parenchyma; the important
104 functional elements of each body system. The stroma comprises connective tissue,
105 nerves, vessels and the extracellular matrices (ECM) and fluids which these cells
106 produce²⁰. Joint soft tissues including synovium, capsule, tendon and enthesis are
107 predominantly composed of mesenchymal stromal cells. Fibroblasts are the most
108 abundant cell type populating these joint connective tissues²¹ and synthesise the highly
109 organized collagen rich scaffold necessary for joint structure and movement.

110

111 Fibroblasts are defined by their spindle shaped morphology, the absence of specific
112 lineage markers of leukocytes, endothelium and epithelium and their ability to adhere
113 to tissue culture plastic *in vitro*²². They are believed to arise from 3 distinct cellular
114 origins: primary mesenchyme, local epithelial-mesenchymal transition (EMT) or bone
115 marrow derived precursors (circulating fibrocytes)^{23,24}. It is widely accepted that the
116 majority of fibroblasts originate from primary mesenchymal cells and that fibroblasts
117 can proliferate to generate new progeny^{25,26}. In physiological conditions, fibroblasts
118 provide mechanical strength to tissues by producing ECM components (type I, III and
119 V collagen and fibronectin) as well as factors that regulate ECM turnover, including
120 metalloproteinases (MMPs) and proteins involved in the formation of basement
121 membranes (type IV collagen and laminin)^{27,28}. Fibroblasts synthesise an array of
122 paracrine factors²⁹ and exhibit mechanosensitive properties³⁰ to effect functional
123 adaptation in normal joint physiology. The intimate relationship between fibroblasts
124 and mesenchymal stromal cells (MSC) and the clinical use of MSC to repair damaged
125 tissues has driven a renewed interest in fibroblasts as new therapeutic targets²¹.

126

127 **2.2 Mechanisms sustaining joint inflammation based on pathogenic stroma**

128

129 **2.2.1 Fibroblasts and the diseased joint**

130

131 Traditionally, the diversity of stromal cells and in particular fibroblasts and their roles
132 beyond those of space filling and ECM homeostasis have been underexplored in
133 inflammation. Mesenchymal tissues in the joint including the synovium, enthesis and

134 tendons undergo phenotypic changes as a consequence of inflammation³¹⁻³³. These
135 include molecular and structural changes to the ECM, impacting upon the functional
136 quality of the healed tissue³⁴. Whilst it remains challenging to discern which is the
137 initiating pathogenic cell type, it is clear that stromal cells populating these tissues
138 provide a niche conducive to sustaining chronic inflammation^{2,35,36}. Recent work
139 shows that fibroblasts vary phenotypically and functionally at different anatomical sites
140 and contribute significantly to the identity of individual tissues, providing a so-called
141 ‘stromal postcode’²⁶. Furthermore, it is known that, rather than acting as a bystander,
142 fibroblasts are capable of actively participating and indeed orchestrating inflammation
143 and immunity³⁶⁻³⁸. We next review how fibroblasts sustain inflammation, highlighting
144 the mechanisms underpinning their activation, “memory” and phenotypic diversity,
145 with particular focus on the synovial microenvironment.

146

147 ***Fibroblast activation and memory***

148

149 Fibroblast activation is a recognized feature of diseases affecting the joint, whereby
150 fibroblasts adopt a pro-inflammatory phenotype. This pathological feature has been
151 identified in cancer³⁹, rheumatoid synovium^{32,33} and tendon disease³¹. Fibroblast
152 activation and memory therefore span both innate and adaptive immune responses,
153 suggesting this is a highly conserved disease mechanism common to tissues of
154 mesenchymal origin. There is now a growing list of cell surface molecules and
155 secreted products which collectively provide a fibroblast activation marker “cassette”.
156 These include CD90 (Thy1), CD44, decay accelerating factor (CD55), VCAM-1
157 (CD106), uridine diphosphoglucose dehydrogenase, and prolyl-4-hydroxylase,
158 Podoplanin (PDPN/gp38), endosialin (CD248) and Fibroblast Activation Protein (FAP)
159^{31,36,37,40-42}. Fibroblast activation markers therefore represent important phenotypic
160 alterations implicated in effecting the switch from resolving to persistent inflammation
161⁴².

162

163 Epigenetic changes are implicated in fibroblast activation and memory. New insights
164 into the epigenetics of inflammatory rheumatic diseases have been recently reviewed
165 in detail elsewhere⁴³. Prolonged exposure of RA synovial fibroblasts to TNF α reduce
166 histone H4 levels and promote H4 acetylation⁴⁴. This study showed that TNF α

167 removed the chromatin barrier from the CXCL10 promoter, permitting abundant
168 binding of NF- κ B family transcription factors and recruitment of transcriptional
169 machinery⁴⁴. DNA methylation is another important epigenetic modification identified
170 in RA synovial fibroblasts occurring during the early stage of disease ⁴⁵. Further
171 studies are required to identify the mechanisms underpinning DNA methylation and
172 there appears to be important prognostic potential for differentially methylated genes
173 as disease biomarkers ⁴⁵. The activated and aggressive phenotype of RA synovial
174 fibroblasts is associated with global DNA hypomethylation⁴⁶. Gaur *et al.* investigated
175 if microRNAs moderate the methylation status of RA synovial fibroblasts, showing L-
176 methionine increased DNA methylation compared to betaine ⁴⁷. Collectively these
177 studies advance our understanding of how epigenetic changes are implicated in
178 fibroblast activation and memory, informing future strategies to selectively target
179 pathogenic fibroblasts.

180

181 Fibroblasts from different joint tissues maintain their phenotype, positional memory
182 and topographic differentiation despite culture *ex vivo*. Fibroblasts isolated from RA
183 synovium or diseased tendon exhibit stromal 'memory', whereby these cells show an
184 enhanced subsequent capacity to respond to an additional inflammatory stimulus
185 ^{2,31,44}. Therefore, sustained expression of activation markers by fibroblasts in the joint
186 reflects their 'primed' status after exposure to an inflammatory stimulus. The
187 processes underpinning innate memory have been extensively reported for leukocytes
188 ^{48,49} and are gaining acceptance in tissue resident cells of mesenchymal origin.
189 Engagement of TLR4 and downstream activation of the NF κ B pathway is a prominent
190 pathological feature of fibroblasts populating inflamed joint tissues ^{2,31,44}. These
191 studies suggest that fibroblast memory is associated with altered NF κ B
192 responsiveness to an inflammatory stimulus ⁵⁰. Given the longevity of fibroblasts as
193 tissue resident cells and the relatively low rates of cell turnover in the joint ⁵¹, the
194 effects of stromal memory in tissues such as synovium and tendon are likely to be
195 long lived. In contrast, dermal fibroblasts show higher rates of turnover and do not
196 exhibit memory, suggesting this process of stromal memory may vary according to
197 anatomical location ^{2,52,53}. Rheumatic diseases follow a characteristic anatomical
198 pattern of joint and organ involvement. Mechanisms regulating the predilection of
199 specific joints for developing particular forms of arthritis (for example osteoarthritis

200 (OA) compared to rheumatoid arthritis (RA)) have been reviewed in detail ⁵⁴. These
201 include site-specific local cell types driving disease, systemic triggers affecting local
202 cell types and site-specific exogenous factors activating cells locally. Therefore the
203 mechanisms underpinning activation of stromal cells depends on the local anatomical
204 tissue niche⁵⁴.

205

206 ***Fibroblast diversity***

207 Recent work shows that tissue resident fibroblasts help define the pattern of joints
208 involved in RA ^{55,56}. The concept of epigenetically-driven anatomical diversity of
209 synovial fibroblasts provides an attractive mechanism to explain the clinical
210 observations that different types of arthritis affect distinct types of joints. For example,
211 OA and psoriatic arthritis often involve the distal interphalangeal joints, whereas RA
212 is frequently symmetrical and more commonly affects the MCP joints. In contrast,
213 ankylosing spondylitis (AS) mainly targets spinal ligaments and enthesal tissue⁵⁷.
214 Such studies have prompted improved characterization of the phenotypes of fibroblast
215 subsets and their different proposed roles. In RA, synovial fibroblasts undergo distinct
216 changes in function, including loss of immunosuppressive response in early disease,
217 followed by later acquisition of an immuno-stimulatory phenotype⁵⁸. Highly conserved
218 homeobox (HOX) transcription factors specify regional identities of cells and tissues
219 throughout the body ^{59,60} and adult fibroblasts retain key features of embryonic
220 positional HOX gene expression ⁵⁶. Fibroblasts also vary according to their
221 anatomical location in relation to tissue structures at an individual site and the
222 exogenous stimuli which they receive ^{54,56,61}. Whether variability can be attributed to
223 the plasticity of individual fibroblasts necessary for responding to different
224 environmental cues and whether phenotypic variation can be used to define distinct
225 subsets of fibroblasts specialized for different niches remains unclear.

226

227 The synovium is composed of lining and sub-lining layers of fibroblasts which vary in
228 terms of phenotype and function. Single cell RNA sequencing and
229 immunohistochemistry have revealed that RA synovial fibroblasts can be broadly
230 characterized into 3 subsets, highlighted in Figure 2. Synovial lining fibroblasts are
231 CD34⁻CD90⁻CD55⁺ and Cadherin 11⁺. This lining subset synthesizes MMP-1 and
232 MMP-3 which mediate tissue damage in the inflamed joint ⁶². Fibroblasts populating
233 the synovial sublining are predominantly comprised of 2 populations. CD34⁺CD90⁻

234 fibroblasts release CXCL12, CCL2 and IL-6 and drive fibroblast accumulation and
235 invasion. A second population of CD34⁺CD90⁺ fibroblasts with a pro-inflammatory
236 phenotype express markers of fibroblast activation^{62,63}. These fibroblast subsets
237 between them degrade articular cartilage, mediate stromal memory, sense tissue
238 damage via TLR4 activation and have altered responsiveness to signalling pathways
239 converging on NFκB responsiveness^{26,33,50,62} (Figure 2). Having highlighted the
240 complexity of discrete synovial fibroblast subtypes, we next discuss the phenotypes
241 and functions of other mesenchymal cell types including endothelial cells and TRM
242 and their respective roles in joint disease.

243

244 ***2.2.2 The endolymphatic niche in the diseased joint***

245

246 Other mesenchymal stromal tissues including the vasculature and lymphatics
247 contribute to sustaining inflammation across the joint organ. Neo-angiogenesis is a
248 prominent feature of disease of mesenchymal joint tissues and impacts upon changes
249 in tissue architecture and pain perception⁶⁴. In health, vascular endothelial cells
250 regulate blood flow, vessel wall permeability and leukocyte extravasation into tissues,
251 regulating the inflammatory process⁶⁵⁻⁶⁸. In lymph nodes and tertiary lymphoid
252 tissues, high endothelial vessels (HEVs) provide specialized microenvironments for
253 efficient entry of lymphocytes into tissues in an L-selectin dependent process⁶⁹. The
254 phenotypes of endothelial cells change as inflammation transitions from acute to
255 chronic and also between activation of innate and adaptive immune systems⁶⁷.
256 Endothelial cell phenotypes are poorly characterized in tendon and enthesal tissues.
257 However, in RA synovium, these cells have been described as activated, angiogenic,
258 apoptotic and leaky, a process found in many tumour microenvironments⁷⁰. During
259 prolonged exposure to inflammatory stimuli endothelial cells become activated, exhibit
260 memory and express adhesion molecules including ICAM, VCAM-1 and CD31
261 (PECAM-1)^{3,71-73} (Figure 2). These activated endothelial cells also present
262 chemokines and initiate leukocyte migration from blood to local tissues⁷⁰. Endothelial
263 activation is a cause and consequence of endothelial dysfunction^{74,75}, culminating in
264 increased microvascular permeability, extravasation of plasma and joint oedema.
265 Release of angiogenic factors including VEGF triggers angiogenesis, provide
266 necessary nutrients and oxygen to meet the metabolic demands of the inflamed tissue.

267 Importantly, neo-angiogenesis further promotes the retention and survival of immune
268 cells at inflamed sites, thereby sustaining chronic inflammation ³⁸. These angiogenic
269 processes occur during normal inflammatory immune responses (i.e vaccination)⁷⁶,
270 however whether angiogenesis that occurs in joint disease is a cause or effect of
271 pathology remains unclear.

272

273 Stromal lymphatic vessels form a one-way conduit for tissue fluid and leukocytes in
274 health and disease ⁷⁷. During adaptive immune responses, antigen presenting cells
275 travel to lymph nodes via lymphatic vessels, which highly express PDPN, implicated
276 in fibroblast activation⁷⁸. The permeability of lymphatic vessels is a tightly regulated
277 dynamic process that alters during health and disease ⁷⁹. Lymphatic vessel growth
278 (lymphangiogenesis) is a primary response during acute inflammation, which
279 becomes dysregulated in chronically inflamed adult tissues⁸⁰. In experimental murine
280 models of inflammatory arthritis, lymphatic vessels and nodes draining the diseased
281 joint undergo an initial expansion phase to expedite lymphatic clearance. This
282 expansion phase is followed by a collapsed phase, characterized by structural
283 damage to lymphatic vessels and reduced lymphatic clearance ^{79,81}. Studies
284 demonstrate alteration in lymphatic vessel function and lymph node volume also occur
285 in patients with RA flare ⁸². Therapies targeting aberrant lymphatic function have
286 shown promise in preclinical models of inflammatory arthritis and may prove
287 efficacious in RA ⁷⁹.

288

289 ***2.2.3 Tissue Resident Macrophages in the diseased joint***

290

291 TRM mediate a diverse range of biological actions. They are appropriately positioned
292 and transcriptionally primed to respond to local environmental challenges, maintaining
293 tissue homeostasis. TRM direct immune surveillance, induce inflammation and
294 promote subsequent resolution, reviewed in detail elsewhere ^{34,83}. Given the biological
295 complexity of these roles, TRM are highly heterogeneous and exhibit diverse
296 phenotypic and functionally distinct subtypes within a single tissue type ^{5,84}.

297

298 In inflamed synovium, TRM mediate immune surveillance through expression of a
299 variety of pattern recognition receptors, notably Toll-like receptors (TLR) TLR2 and
300 TLR4 and facilitate the recruitment of infiltrating leukocytes, including monocyte

301 derived macrophages⁸⁵⁻⁸⁷. TRM induce joint inflammation through release of TNF α ,
302 IL-1 β IL-6, GM-CSF and PGE₂, driving fibroblast accumulation, angiogenesis,
303 leukocyte recruitment and tissue damage via protease secretion (Figure 2). The
304 essential role of non-classical Ly6C-monocytes has been reported in murine arthritis
305 models⁸⁸. This study highlights the phenotypic heterogeneity of synovial TRM,
306 demonstrating how macrophage activation status regulates disease progression and
307 resolution. In support of this, human RA synovial macrophages exhibit distinct
308 transcriptional profiles associated with disease activity and therapy⁸⁹. However,
309 distinction between TRM and infiltrating macrophages is currently hampered by a lack
310 of specific markers that distinguish between these populations in diseased human
311 tissues.

312

313 The pro-inflammatory milieu in the inflamed synovium triggers an active process of
314 lipid mediator class switching and the subsequent release of families of specialized
315 proresolving mediators (SPM). These include lipoxins, resolvins, protectins and
316 maresins, that are generated via transcellular biosynthesis and are concerned with
317 mediating resolution of inflammation⁹⁰⁻⁹⁴. These bioactive lipid mediators initiate
318 programmes which halt neutrophil infiltration, potentiate monocyte recruitment,
319 moderate vascular permeability and promote phagocytosis and drainage of apoptotic
320 cells⁹⁵. The mechanisms mediating resolution in inflammatory arthritis have been
321 reviewed in detail and are not covered here⁹⁶. TRM are key regulators of repair and
322 fibrosis across all tissue types³⁴ and are also implicated in mediating resolution of
323 inflammation. Distinct populations of resolution phase macrophages have been
324 identified in systemic murine inflammation models that express Alox15, Timd4 and
325 Tgfb2, which terminate leukocyte recruitment and promote clearance⁹⁷. However, the
326 precise phenotypes of TRM mediating resolution in human joint disease requires
327 further investigation.

328

329

330

331 ***2.2.4 Cross talk between cells of mesenchymal origin***

332

333 Having highlighted the molecular features and phenotypes of mesenchymal cells and
334 their roles in mediating joint pathology, we next discuss how cross talk between these
335 cell populations sustains inflammation. Damage sensing mechanisms, cytokine and
336 chemokine gradients are pivotal pathological processes involving cross talk between
337 fibroblast, endothelial cell, TRM and leukocyte populations that sustain inflammation
338 in the diseased joint ^{26,98,99}.

339
340 RA synovial fibroblasts act as sentinel cells that can “sense” tissue damage. This
341 occurs via the binding of damage associated molecular patterns (DAMPs) including
342 HMGB1, heat shock and S100 proteins^{100,101}. Tenascin-C a matrix protein induced
343 upon tissue damage also activates TLR4 mediated sterile inflammation ¹⁰². Binding of
344 these ligands to TLR4 induces a high alert state, favouring the development of chronic
345 inflammation ^{50,103}. Engagement of TLR4 activates Myd88 signalling pathways,
346 inducing pro-inflammatory cytokine release via NFκB activation ⁴⁸. Consequently,
347 activated synovial fibroblasts are primed to release a broad range of pro-inflammatory
348 mediators. These localised cytokine and chemokine gradients promote the migration,
349 retention and survival of leukocytes and TRM,^{42,104} creating a complex functional
350 syncytium conducive to sustaining inflammation, highlighted in Figure 3. The
351 processes mediating leukocyte trafficking between stromal compartments in RA are
352 recently reviewed in detail elsewhere ¹⁰⁵.

353

354 *Fibroblast – immune cell cross talk*

355

356 RA synovial fibroblasts promote leukocyte retention via release of cytokines and
357 chemokines and via contact with other cells of mesenchymal origin. Pro-inflammatory
358 cytokines released by retained monocytes, T cells and TRM including IFN γ , TNF α and
359 IL-1 β induce activated synovial fibroblasts to release high levels of PGE₂, GM-CSF,
360 IL-6. These cytokines exert differing effects on leukocyte activation. PGE₂ moderates
361 chemokine production and promotes Th2, Th17 and Treg responses ¹⁰⁶. IL-6 drives
362 CD4+ T cells towards Th17 activation ¹⁰⁷, whereas GM-CSF promotes neutrophil
363 survival and monocyte differentiation in the inflamed synovium ^{26,108}. Nguyen *et al.*
364 demonstrated that IL-6 and other inflammatory cytokines and chemokines are
365 regulated by a positive feedback loop that selectively operates in fibroblasts involving

366 leukemia inhibitory factor (LIF), LIF receptor and STAT4¹⁰⁹. TGF β , also found at high
367 levels in RA synovium induces persistent expression of CXCR4 on synovial T cells,
368 leading to their active CXCL12 mediated retention, providing an additional mechanism
369 for immune cell retention¹¹⁰. RA synovial fibroblasts also release a repertoire of
370 chemokines, generating a gradient consisting of CCL2, CCL4, CCL5, CCL8, CXCL8,
371 CXCL12 and IFN β ^{26,111,112}. This chemokine gradient actively promotes the
372 recruitment, retention and survival of monocytes and CD4+ T cells at the inflamed
373 synovial site (Figure 3). CXCL12, VCAM-1 (CD106) and IL-6 therefore constitute part
374 of a 'stromal address code', critical for leukocyte survival and differentiation²⁶.

375

376 *Endothelial cell cross talk*

377

378 Resident stromal cells populating inflamed synovium modulate the ability of
379 endothelial cells to recruit leukocytes via release of soluble mediators or direct cell-
380 cell contact. Fibroblasts isolated from healthy patients are known to regulate the
381 cytokine-sensitivity of vascular endothelium, while fibroblasts associated with chronic
382 inflammation adopt a pro-inflammatory phenotype^{29,113}. Cytokine and chemokine
383 gradients mediate and sustain cross talk between endothelial cell, synovial fibroblast
384 and TRM populations. IL-6, TGF β 1 and VEGF released from TRM provide the
385 necessary cues to promote an angiogenic environment required to sustain endothelial
386 cell activation and dysfunction (Figure 3). This is supported by antibody neutralisation
387 of IL-6, which diminished the ability of endothelial cells to bind lymphocytes in co-
388 cultures with RA fibroblasts²⁹.

389

390 The RA synovial fibroblast milieu further sustains an angiogenic environment through
391 chemokine gradients comprising CXCL1-5 and CXCL8²⁶ (Figure 3). RA fibroblasts
392 regulate expression of endothelial cell adhesion molecules, potentiate leukocyte
393 extravasation⁵⁸ and induce unstimulated HUVEC to bind flowing lymphocytes via a
394 CXCR4-CXCL12 dependent manner²⁹. Consequently, the interactions between cells
395 of mesenchymal origin create and sustain an inflammatory milieu, whereby synovial
396 inflammation persists and potentially becomes independent of its inciting cause. We
397 next consider how persistent inflammation culminates in tissue damage across soft
398 tissues that comprise the joint.

399

400 **2.3 Mesenchymal cells and their role in joint damage**

401

402 In health, early damage repair mechanisms maintain the integrity of joint soft tissues.
403 In joint disease, sustained inflammation, tissue remodeling and fibrosis ensue,
404 resulting in irreversible tissue damage. We next discuss how cells of mesenchymal
405 origin mediate fetal scarless healing and highlight the mechanisms by which they
406 induce damage across adult joint tissues.

407 In contrast to normal adult tissues, early human and murine fetal wounds and wounds
408 in Nude (FoxN1 deficient) mice heal without scar formation¹¹⁴. Fetal wounds show
409 diminished numbers of immune cells and lower levels of cytokines compared to adult
410 tissues¹¹⁵⁻¹¹⁸. Differences between embryonic and adult tissue healing are also
411 attributed to the milieu of pro-fibrotic growth factors released by TRM, including those
412 of the TGF β family. TGF β 1 levels are reduced and this growth factor shows
413 accelerated clearance in embryonic compared to adult tissue repair¹¹⁹⁻¹²¹. Collectively
414 these studies indicate a role for immune cell derived cytokines including TNF α and
415 TGF β in tissue scarring and healing¹²². Other studies highlight differences between
416 fetal and adult fibroblasts and localized production of MMP-9 and MMP-13 in the
417 scarring process¹¹⁴. Fetal fibroblasts show enhanced synthetic function, increased
418 rate of turnover of collagen, hyaluronic acid, ECM components and increased
419 migration velocity compared to adult fibroblasts, suggesting rapid healing may also
420 play a role in scarless tissue repair¹²³⁻¹²⁵.

421 In adult tissues, fibroblasts and TRM directly contribute to joint destruction, bony
422 erosions and remodeling through expression of enzymes such as MMPs¹²⁶. MMP-2,
423 MMP-9 and MMP-13 have been specifically implicated in the pathogenesis of RA and
424 OA¹²⁷. MMP-9 is also upregulated by CXCL12 (SDF-1) a key chemokine secreted by
425 synovial fibroblasts¹²⁸. FAP is highly expressed within RA synovium and co-localises
426 with MMP-13, where it appears to play a role in tissue degradation¹²⁹. Cathepsins, a
427 major group of proteases involved in joint remodeling are also upregulated in the
428 diseased joint¹³⁰. Additionally fibroblasts can indirectly contribute through cross talk
429 with TRM and lymphocytes, further amplifying processes driving tissue damage
430 (Figure 3), whilst also presenting antigen to tissue infiltrating lymphocytes¹³¹.

431

432 Pathological conditions in which cells of mesenchymal origin play a role include
433 chronic inflammation (e.g. RA, chronic skin wound healing), tissue fibrosis (e.g.
434 COPD) and cancer (e.g. breast cancer). Interestingly, while these diseases differ
435 dramatically in aetiology and genetic predispositions, they converge in terms of
436 phenotype and function of the stromal component. Fibroblasts expand in the RA
437 synovial tissue and in the tumor parenchyma, while fibrosis is characterized by
438 profound changes in myofibroblast phenotype and function across different organs
439 such as the lungs and kidneys ¹³². Whether these fibroblast properties are intrinsic
440 phenotypic changes acquired as a consequence of exposure to chronic inflammation,
441 or are derived from the conditioning of the pathogenic infiltrating cells is still under
442 investigation and seems to differ in the different conditions³⁷. Lafevre *et al* reported
443 epigenetically programmed aggressive cells may “spread” arthritis from inflamed to
444 uninfamed joints in the early stages of disease, ¹³³. PDPN expressing lining synovial
445 fibroblasts are migratory and mediate release of cartilage destructive MMPs ^{33,62}.
446 Collectively, these data raise the possibility of distinct mesenchymal cell subsets
447 implicated in mediating the effects of tissue damage in the diseased joint. We next
448 discuss how the possibility of selectively targeting pathogenic stromal subpopulations
449 mediating inflammation and tissue damage informs the development of future
450 strategies to successfully treat joint disease.

451

452 **3.0 Shaping the future landscape: therapeutic targeting of** 453 **mesenchymal cells**

454

455 Cells of mesenchymal origin including fibroblasts, TRM and endothelial cells constitute
456 the major cell types populating joint soft tissues. We have discussed the roles and
457 mechanisms by which these cells mediate joint inflammation, highlighting their ability
458 to act as immune sentinel cells, their capacity for activation, positional memory and
459 their altered phenotypes comprising multiple cellular sub-populations. Multidirectional
460 cross talk between stromal cell populations further fuels the development of persistent
461 inflammation. Given these important roles and associated biological complexities, it
462 is likely that residual disease activity in patients treated with immune therapies may
463 be attributable to stromal mediated inflammatory responses, which are refractory to

464 current therapies that target immune cell populations ¹³⁴. New therapeutic approaches
465 are therefore required to 'break the cycle and reset the system', particularly in
466 scenarios where inflammation becomes independent of the inciting stimulus. Given
467 the limited capacity of joint tissues to regenerate once damaged, there are significant
468 challenges associated with curbing tissue damage, which might be accomplished
469 through moderating persistent inflammation as a driver of fibrosis. We next discuss
470 the requirement for future strategies to address the pathobiology concerned with the
471 stromal microenvironment, targeting cells of mesenchymal origin. We review the drug
472 classes in current clinical use, those in early phase clinical trials and strategies with
473 pre-clinical potential to target stromal mediated joint disease. The cellular and
474 molecular targets and the mechanism of action through which these drug classes
475 function are summarized in Table 1.

476

477 *Existing licensed therapies*

478

479 Nonsteroidal anti-inflammatory drugs (NSAIDs) and corticosteroids provide
480 symptomatic relief for a broad array of conditions targeting inflammation and pain. Their
481 clinical use in the management of a multitude of diseases affecting the joint is well
482 established ¹³⁵⁻¹³⁸. These therapies target fibroblasts, TRM and endothelial cells via
483 differing biological modes of action. Inhibition of COX activity by NSAIDs dampens
484 release of prostaglandins, leukotrienes and thromboxane A₂. Corticosteroids act via
485 the glucocorticoid receptor to inhibit cPLA₂, regulate expression of NFκB / MAPK
486 target genes and dampen release of inflammation initiating eicosanoids. Whilst
487 NSAIDs and corticosteroids continue to provide background anti-inflammatory therapy
488 for many rheumatic diseases, they are both associated with well documented adverse
489 systemic effects. Importantly, COX-2 selective NSAIDs also dampen protective
490 endogenous resolution responses ^{139,140}, which may paradoxically impede the
491 capacity of inflamed joint tissues to heal.

492

493 Monoclonal antibodies enable precise molecular targeting of cytokines mediating joint
494 inflammation. The biological modes of action and efficacy of therapeutic inhibitors of
495 IL-1, IL-6, TNFα and IL-17 in current clinical use are well reported and listed in Table
496 1. One disadvantage associated with selective cytokine inhibition is the failure of this

497 approach to fully target stromal mediated inflammatory responses and address the
498 complex multidirectional cross talk between mesenchymal cell populations. Similarly
499 targeting chemokine gradients is an attractive strategy to moderate leukocyte retention
500 ¹⁴¹. However chemokine antagonists including AMD3100 targeting CXCR4 are
501 associated with adverse systemic effects ¹⁴² and the plethora of chemokines mediating
502 stromal inflammatory responses presents a further therapeutic challenge.

503

504 *Therapies in early phase clinical trials*

505

506 GM-CSF, predominantly produced by activated T cells, monocytes and macrophages
507 is also released by tissue resident cells of mesenchymal origin ¹⁴³. Humanised IgG1
508 monoclonal antibodies to GM-CSF prevent interaction of this cytokine with its receptor,
509 reducing downstream signalling pathways converging on NF κ B. GM-CSF has shown
510 potential as a therapeutic target in autoimmune and inflammatory disorders, including
511 RA. Therapies targeting GM-CSF or its receptor have shown encouraging results in
512 more recent pre-clinical studies and are reviewed in detail elsewhere ¹⁴³. Recent
513 phase IIb studies have demonstrated that long term mavrilimumab treatment
514 maintained clinical responses and was well tolerated in RA patients with inadequate
515 response to DMARD's¹⁴⁴. Further investigation is required to determine the efficacy of
516 GM-CSF targeted therapies to modulate stromal mediated inflammatory responses in
517 the joint.

518 Kinase inhibitors targeting JAK and SYK signalling pathways have been investigated
519 for their therapeutic utility to reduce cytokine release through JAK STAT ^{145,146} or
520 MAPK / PKC ^{147,148} blockade respectively (Table 1). Baricitinib, an oral reversible
521 inhibitor of JAK1 and JAK2 has shown therapeutic value in RA patients. This treatment
522 was associated with significant clinical improvements in patients with an inadequate
523 response to methotrexate compared with placebo and adalimumab treated groups ¹⁴⁹.
524 Protein kinase inhibitors target a broad range of cells types with reported off target
525 effects, highlighting the importance of understanding the pharmacology of these drugs
526 beyond the kinome ¹⁵⁰.

527

528 *Potential future strategies to target pathogenic stroma*

529

530 Developments in cancer medicine targeting cancer associated fibroblasts populating
531 tumour stroma have informed potential future strategies to target pathogenic stroma
532 in rheumatic disease ^{151,152}. Targeting pathogenic stroma presents a considerable
533 therapeutic challenge due to the biological complexity underpinning activation,
534 memory and phenotypic diversity exhibited by these mesenchymal cell populations.
535 Potential future strategies to treat residual rheumatic disease might include targeting
536 activated fibroblast subtypes, use of epigenetic modifiers or resolution agonists to
537 target stromal mediated inflammation. Pre-clinical evidence supporting these
538 approaches are discussed below.

539

540 Selective targeting of distinct fibroblast subtypes mediating joint inflammation and
541 tissue damage is a potential therapeutic strategy to target pathogenic stroma.
542 Cadherin-11 is known to regulate synovial fibroblast inflammation, synergizing with
543 IL-1 β and TNF α to regulate IL-6 release ¹⁵³. This study showed that cad-11 deficient
544 mice or anti-cad-11 mAb therapies reduced inflammation in arthritic mice, suggesting
545 that cadherin expression regulates the inflammatory capacity of synovial fibroblasts.
546 Cyclin dependent kinases regulate cell proliferation and survival via specific inhibitors
547 (CDKi) and are potential therapies to target fibroblast accumulation in RA synovium
548 (Table 1). CDK pathways become dysregulated in cancer, leading to the development
549 of anti-cancer drugs including the CDKi Roscovitine ¹⁵⁴. In synovial fibroblasts, IL-6
550 and MMP-1 are known to be regulated by CDKi p21 ¹⁵⁵. Given that CD34⁺CD90⁻
551 'immunoregulatory' fibroblasts are highly proliferative, invasive and produce IL-6⁶²,
552 CDKi therapies are a potential strategy to target this fibroblast subset mediating joint
553 disease.

554

555 We previously discussed how epigenetic changes are implicated in mediating
556 fibroblast activation and memory. Epigenetic alterations in RA synovial fibroblasts are
557 listed in Table 1, identifying DNA methylation, histone modification and miRNA as
558 potential processes to therapeutically target ^{43,45-47,156}. Moderating the epigenetic
559 landscape is likely to have broad ranging effects on a variety of cell types, with off
560 target effects. Hence improved understanding of the pharmacology of these drugs
561 beyond the epigenome is essential before we can appreciate their potential utility to
562 treat joint disease.

563

564 The roles of proresolving mediators in joint health and disease are increasingly
565 understood, identifying resolution agonists as potential therapies to moderate joint
566 inflammation and promote tissue repair ⁹⁶. The biological modes of action of
567 proresolving mediators or ‘immunoresolvents’ are well established from *in vitro* and
568 *in vivo* studies and include limiting PMN infiltration, stimulating efferocytosis and
569 activation of endogenous tissue protective mechanisms ^{90-93,157,158}. Whilst
570 immunoresolvents target leukocytes, their biological actions are not associated with
571 immunosuppression ^{83,159}. Importantly, proresolving mediators also target fibroblasts,
572 TRM and endothelial cells types ¹⁶⁰⁻¹⁶² and therefore possess the capacity to modulate
573 stromal mediated inflammatory responses across joint tissues. Approaches to
574 potentiate resolution processes include dietary supplementation with proresolving
575 precursors, blocking catabolism of proresolving mediators or local delivery of stable
576 analogues binding proresolving receptors ⁹⁶. The pro-resolving mediator RvD3 was
577 found to limit leukocyte infiltration and paw joint eicosanoid levels in murine
578 inflammatory arthritis ¹⁶³. The stable epimer 17R-RvD1 significantly attenuated arthritis
579 severity, cachexia, paw oedema, leukocyte infiltration and shortened the remission
580 interval, showing cartilage protective actions in murine models of acute inflammatory
581 arthritis¹⁶⁴. *In vitro* studies also highlight the capacity of 15-epi-LXA₄ and MaR1 stable
582 epimers to regulate PDPN, STAT-1 and IL-6 in IL-1 β stimulated diseased human
583 tendon stromal cells ^{35,165}. Collectively these studies suggest resolution pharmacology
584 may be an important future therapeutic tool to address stromal pathobiology in the
585 joint.

586

587 **Conclusions**

588

589 Stromal cells of mesenchymal origin including fibroblasts, tissue resident
590 macrophages and endothelial cells are pivotal populations regulating health and
591 disease in musculoskeletal tissues. New insights are beginning to reveal the
592 mechanisms underpinning the activation and dysfunction of mesenchymal stromal
593 cells and their contribution to sustaining chronic joint inflammation. The discovery that
594 distinct synovial fibroblast subsets mediate joint inflammation and damage will inform
595 precision therapeutic targeting of pathogenic stromal cell populations. These

596 discoveries shape the future therapeutic landscape, presenting exciting new
597 approaches to address the pathogenic stromal microenvironment. Harnessing the
598 capacity to modulate cross talk between leukocyte and pathogenic stromal cell
599 populations is a critical barrier to overcome in our quest to advance therapeutic
600 strategies for patients with refractory joint disease.

601

602 **Glossary of terms**

603

604 **Mesoderm:** Middle embryonic primary germ layer residing between ectoderm and
605 endoderm

606

607 **Mesenchymal:** Embryonic connective tissue derived from the mesoderm

608

609 **Mesenchymal tissue:** Tissue of the musculoskeletal, circulatory and lymphatic
610 systems

611

612 **Stromal cell:** Non-haematopoietic, tissue resident cells.

613

614 **Stromal cell activation:** Process whereby stromal cells including fibroblasts, tissue
615 resident macrophages and endothelial cells adopt a pro-inflammatory phenotype and
616 express distinct molecular markers after exposure to an inflammatory stimulus.

617

618 **Stromal cell memory:** A change in the capacity of stromal cells to respond to
619 inflammatory stimuli

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Table 1: Drugs to target the pathogenic stroma and associated immune cell cross talk in joint disease

Drug Class	Target Mesenchymal Cell	Molecular Target	Mechanism of Action	References
NSAIDs	Fibroblast (F) Tissue Resident Macrophage (TRM) Endothelial Cell (EC)	COX-1 COX-2	Selective / non-selective inhibition of COX to reduce release of prostaglandins, leukotrienes, thromboxane	135,136
Corticosteroids	F, TRM, EC	glucocorticoid receptor	cPLA2 inhibition regulate NFκB / MAPK target genes reduce prostaglandins, leukotrienes, thromboxane	137,138
Monoclonal Ab				
IL-1	TRM (F)	IL-1R	Reduce effects of inflammasome and caspase activation	166,167
IL-6	TRM, F	IL-6R	Reduce STAT-3 signalling	168-172
TNF	TRM (F)	TNFR 1/2	Reduce NFκB / MAPK signalling	173-178
GM-CSF	TRM, F, EC	GM-CSFR	Reduce JAK STAT, PI3K, MAPK and NFκB signalling	179,180
IL-17	TRM	IL-17R family	Reduce TRAF6, MAPK, TAK1 & NFκB signalling	181-183
Kinase Inhibitors				
JAK inhibitors	F, TRM	JAK1 JAK2 JAK3 TYK2	Blockade of cytokine signalling via JAK STAT	145,146,149
SYK inhibitors	F, TRM	Fcγ receptor	Reduce IL-6 via MAPK / PKC	147,148
Fibroblast activation				
Cadherin-11 mAb	F	Cadherin-11	Reduce MAPK, NFκB, IL-6	153
Cyclin dependent kinase inhibitors (CDKi)	F	CDK1,2,4,6	Inhibit cell proliferation & survival, induce apoptosis	142,154,155
Epigenetic Modifier	F	DNA methylation Histone modification miRNA	Hypomethylation Increase H4 acetylation CXCL10 promoter Increase H4 acetylation IL-6 promoter Reduce miR-22 Reduce miR-20a Reduce miR203	45,47,156 44 184 185 186 187
Pro-resolving				
17-R RvD1	F, TRM, EC	ALX, DRV1	Chondroprotective	164
Annexin A1		ALX	Chondroprotective, increased TGFβ, prevent apoptosis	188
RvD3		ALX	Reduce leukocyte infiltration, prostaglandins, leukotrienes and thromboxane	163
15-epi-LXA ₄		ALX	Reduced STAT-1, IL-6, Podoplanin	35,165

Figures:

Figure 1. Embryological origins of mesenchymal tissues in the whole joint organ.

To further understand the mechanisms and inter-relationships underpinning inflammation and tissue damage across the joint, it is important to consider the embryonic origins of joint tissues, which may shape the behaviour of these cell types in diseased adult tissues. Mesoderm is the precursor for mesenchymal tissues comprising the appendicular skeleton, synovium, cartilage, tendons, ligaments, joint capsule and their associated lymphatics and vasculature. Adult joint soft tissues are predominantly composed of cells of mesenchymal origin, including fibroblasts, endothelial cells and tissue resident macrophages (TRM). Yolk sac derived TRM are phenotypically distinct from HSC derived lineages. TRM occupy subspecialized niches which dictate their heterogeneity and phenotype in adult tissues.

Figure 2. Molecular features of cells of mesenchymal origin in Rheumatoid synovium.

Inset shows topographical location of cell types comprising RA synovium, consisting of lining and sublining layers. Synovial lining fibroblasts (blue) are CD34⁻CD90⁻, express PDPN, CD55 and release MMP-1 and MMP-13 implicated in tissue destruction. Fibroblast subsets concerned with proliferation, accumulation and inflammation occupy the synovial sublining. Immunoregulatory fibroblasts (green) promote fibroblast accumulation and invasion. These cells express CD34 and release chemokines and cytokines generating gradients that promote leukocyte retention. Pathogenic fibroblasts (red) are a CD34⁻CD90⁺ subpopulation that highly express markers of fibroblast activation and exhibit inflammation memory. Pathogenic fibroblasts express TLR4 which mediates the damage sensing properties of these cells and downstream activation of the NF κ B pathway via MAPK, JNK and JAK-STAT signalling pathways. These phenotypic features sustain the pro-inflammatory pathogenic phenotype of this fibroblast subset. Fibroblasts in the synovial sublining are in close proximity to activated endothelial cells, expressing CD31, VCAM-1 and ICAM-1 and CD68⁺ tissue resident macrophages (TRM) which release pro-inflammatory mediators and proteases.

Figure 3: Mechanisms sustaining synovial inflammation, highlighting cross talk between cells of mesenchymal origin and leukocytes.

Cells of mesenchymal origin including fibroblast subsets, endothelial cells and tissue resident macrophages (TRM) are engaged in multidirectional cross talk, which sustains synovial inflammation. RA synovial fibroblasts promote leukocyte retention via release of cytokines and chemokine gradients and via contact with other cells of mesenchymal origin. Pro-inflammatory cytokines released by retained monocytes, T cells and TRM including $\text{IFN}\gamma$, $\text{TNF}\alpha$ and $\text{IL-1}\beta$ induce activated synovial fibroblasts to release high levels of PGE_2 , GM-CSF and IL-6. $\text{TGF}\beta$ released by TRM induces persistent expression of CXCR4 on synovial T cells, leading to their active CXCL12 mediated retention. RA synovial fibroblasts also release chemokines including CCL2, CCL4, CCL5, CCL8, CXCL8, CXCL12 and $\text{IFN}\beta$ that promotes the recruitment, retention and survival of monocytes and CD4^+ T cells. IL-6, $\text{TGF}\beta 1$ and VEGF released from TRM provide the necessary cues to promote an angiogenic environment required to sustain endothelial cell activation and dysfunction. The RA synovial fibroblast milieu further sustains an angiogenic environment through chemokine gradients comprising CXCL1-5 and CXCL8.