Decoding human fetal liver haematopoiesis

Dorin-Mirel Popescu[†1], Rachel A. Botting[†1], Emily Stephenson[†1], Kile Green[1], Simone Webb[1], Laura Jardine[1], Emily F. Calderbank[2], Krzysztof Polanski[3], Issac Goh[1], Mirjana Efremova[3], Meghan Acres[1], Daniel Maunder[1], Peter Vegh[1], Yorick Gitton[5], Jong-Eun Park[3], Roser Vento-Tormo[3], Zhichao Miao[3,4], David Dixon[1], Rachel Rowell[1], David McDonald[1], James Fletcher[1], Elizabeth Poyner[1,18], Gary Reynolds[1], Michael Mather[1], Corina Moldovan[6], Lira Mamanova[3], Frankie Greig[1], Matthew Young[3], Kerstin B. Meyer[3], Steven Lisgo[7], Jaume Bacardit[8], Andrew Fuller[1], Ben Millar[1], Barbara Innes[1], Susan Lindsay[7], Michael J. T. Stubbington[3], Monika S. Kowalczyk[10], Bo Li[10], Orr Ashenbrg[10], Marcin Tabaka[10], Danielle Dionne[10], Timothy L. Tickle[10,12], Michal Slyper[10], Orit Rozenblatt-Rosen[10], Andrew Filby[1], Peter Carey[11], Alexandra-Chloe Villani[9], Anindita Roy[13], Aviv Regev[10,14], Alain Chedotal[5], Irene Roberts[15], Berthold Göttgens[2], Sam Behjati[2,16*], Elisa Laurenti[2*], Sarah A. Teichmann[3,17*], Muzlifah Haniffa[1,3,18*]

Affiliations:

[1]Institute of Cellular Medicine, Newcastle University, Newcastle upon Tyne, NE2 4HH, UK

[2]Department of Haematology and Wellcome and MRC Cambridge Stem Cell Institute, University of Cambridge, Cambridge, CB2 2XY, UK

[3]Wellcome Sanger Institute, Wellcome Genome Campus, Hinxton, Cambridge CB10 1SA, UK

[4]European Molecular Biology Laboratory, European Bioinformatics Institute (EMBL-EBI), Wellcome Genome Campus, Cambridge, CB10 1SD UK

[5]Sorbonne Université, INSERM, CNRS, Institut de la Vision, 17 Rue Moreau, F-75012 Paris, France

[6]Department of Pathology, Newcastle Hospitals NHS Foundation Trust, Newcastle upon Tyne NE2 4LP, UK

[7]Institute of Genetic Medicine, Newcastle University, Newcastle upon Tyne, NE1 3BZ, UK

[8]School of Computing, Newcastle University, NE4 5TG, UK

[9]Broad Institute of Harvard and MIT, Cambridge, MA 02142, USA; Center for Immunology and Inflammatory Diseases, Massachusetts General Hospital, Boston, MA 02129, USA.

[10]Klarman Cell Observatory, Broad Institute of Harvard and MIT, Cambridge, MA, USA
[11]Haematology Department, Royal Victoria Infirmary, Newcastle-upon-Tyne Hospitals NHS Foundation Trust, Newcastle-upon-Tyne, UK
[12]Data Sciences Platform, Broad Institute of Harvard and MIT, Cambridge, MA, USA
[13]Department of Paediatrics, University of Oxford, Oxford OX3 9DS, UK
[14]Howard Hughes Medical Institute, Koch Institute of Integrative Cancer Research, Department of Biology, Massachusetts Institute of Technology, Cambridge, MA, USA
[15]MRC Molecular Haematology Unit and Department of Paediatrics, Weatherall Institute of Molecular Medicine, University of Oxford, and BRC Blood Theme, NIHR Oxford Biomedical Centre, Oxford OX3 9DS, UK
[16]Department of Paediatrics, University of Cambridge, Cambridge CB2 0SP, UK
[17]Theory of Condensed Matter Group, Cavendish Laboratory/Department of Physics, University of Cambridge, Cambridge CB3 0HE, UK
[18]Department of Dermatology and NIHR Newcastle Biomedical Research Centre, Newcastle Hospitals NHS Foundation Trust, Newcastle upon Tyne NE2 4LP, UK

[†]Equal contribution, [*]Corresponding authors

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Summary

Definitive haematopoiesis in the fetal liver supports self-renewal and differentiation of haematopoietic stem cells/multipotent progenitors (HSC/MPPs) but remains poorly defined in humans. Using single cell transcriptome profiling of ~140,000 liver and ~74,000 skin, kidney and yolk sac cells, we identify the repertoire of human blood and immune cells during development. We infer differentiation trajectories from HSC/MPPs and evaluate the impact of tissue microenvironment on blood and immune cell development. We reveal physiological erythropoiesis in fetal skin and the presence of mast cells, NK and ILC precursors in the yolk sac. We demonstrate a shift in fetal liver haematopoietic composition during gestation away from being erythroid-predominant, accompanied by a parallel change in HSC/MPP differentiation potential, which we functionally validate. Our integrated map of fetal liver

haematopoiesis provides a blueprint for the study of paediatric blood and immune disorders, and a valuable reference for harnessing the therapeutic potential of HSC/MPPs.

Introduction

The blood and immune systems develop during early embryogenesis. Our understanding of this process derives from murine and *in vitro* model systems as human fetal tissue is scarce. While haematopoietic development is conserved across vertebrates[1], important differences between mouse and human have been noted[2,3]. Comprehensive interrogation of human tissue to understand the molecular and cellular landscape of early hematopoiesis has implications beyond life *in utero,* providing a blueprint for understanding immunodeficiencies, childhood leukemias and anaemias and generating insights into HSC/MPP propagation to inform stem-cell technologies.

The earliest blood and immune cells originate outside the embryo, arising from the yolk-sac between 2-3 post-conception weeks (PCW). At 3-4 PCW, intra-embryonic progenitors from the aorta-gonad-mesonephros (AGM) develop[4]. Yolk-sac and AGM progenitors colonise fetal tissues such as the liver, which remains the major organ of haematopoiesis until the mid-second trimester. Fetal bone marrow (BM) is colonised around 11 PCW and becomes the dominant site of haematopoiesis after 20 PCW in human[4]. Yolk sac-, AGM-, fetal liver- and BM-derived immune cells seed peripheral tissues including non-lymphoid tissues (NLT), where they undergo specific maturation programs which are both intrinsically determined and extrinsically nurtured by the tissue microenvironment[5,6]. Systematic, comprehensive analysis of multiple blood and immune lineages during human development has not previously been attempted.

In this study, we used single cell transcriptomics to map the molecular states of human fetal liver cells between 7-17 PCW, when the liver represents the predominant site of human fetal haematopoiesis. We integrate imaging mass cytometry, flow cytometry and cellular morphology to validate the transcriptome-based cellular profiles. We construct the functional organisation of the developing immune network through comparative analysis of immune cells in fetal liver with those in yolk sac, and skin and kidney as representative NLT.

Results

Single cell transcriptome of fetal liver

To investigate blood and immune cell development in the fetal liver, we generated single cell suspensions from embryonic and fetal livers between 6 and 17 PCW. We FACS-isolated $CD45^+$ and $CD45^-$ cells using adjoining gates for comprehensive capture (Figure 1a and Extended Data 9a) for single cell RNA-sequencing (scRNA-seq) (both 10x Genomics platform Smart-seq2) (Figure 1, Extended Data 4d, and Supplementary Table 1). To allow parallel evaluation of blood and immune cell topography in NLT and the yolk sac during early development (Figure 1a) we profiled skin, kidney and yolk sac cells by FACS-isolation and 10x Genomics platform.

In total, 138,575 (n = 14) liver (an additional 1,206 cells were profiled using Smart-seq2), 54,690 (n = 7) skin, 9,643 kidney (n = 3) and 10,071 yolk sac (n = 3) cells passed quality control (QC) and doublet exclusion (Extended Data 1a-b, and Supplementary Table 2). We performed graph-based Louvain clustering and derived differentially expressed genes to annotate cell clusters. To minimize technical batch-effect while preserving biological variation due to gestational stage, we divided liver samples into four gestational stage categories and performed data integration between samples using Harmony (Extended Data 1c-e).

27 major cell states are present in the fetal liver (Figure 1b, Extended Data 1f). $VCAM1^+$ erythroblastic island (EI) macrophages were validated as a distinct cell state as a result of their interactions with erythroid cells (Extended Data 4a-f). We applied a descriptive nomenclature based on gene expression profiles. All cell states are found throughout the developmental period studied, but frequency varied by gestation stage (Figure 1c and Extended Data 1e). Neutrophils, basophils and eosinophils are not detected, consistent with reports of granulocytes emerging during fetal BM haematopoiesis[7]. Early stage samples show erythroid lineage bias, with later lymphoid and myeloid lineages representation, as previously shown[5] (Figure 1c and Extended Data 1g).

Our fetal liver dataset can be explored using an interactive web portal through the following weblink: https://developmentcellatlas.ncl.ac.uk//datasets/hca_liver/. We provide comprehensive expression profiles of genes known to cause primary immunodeficiencies[8] to aid future molecular phenotyping of these disorders (Extended Data 8).

Validation of genes and cell states

We manually selected 48 genes from the 4,471 differentially expressed genes between all clusters (log fold change >0.5) (Figure 2a). The predictive power of these 48 genes to determine cell states by the Random Forest classifier was on average 89% for precision and recall (Extended Data 2a). We designed a FACS panel for prospective cell isolation using genes encoding surface proteins (Figure 2a, Extended Data 9b), permitting validation of 19 cell types by mini-bulk transcriptome profiling (6 of the 19 cell types also by scRNA-seq) using Smart-seq2 (Extended Data 2b-d). Cytospins from FACS-isolated cells are morphologically consistent with their designated cell type, including hypogranularity of embryonic/fetal mast cells[9] and early erythroid cells resemblance to the previously reported 'early erythroid progenitors' (Figure 2b)[10].

Next, we evaluated the spatial distribution of erythroid, mast cell, myeloid and lymphoid lineages using imaging mass cytometry (Figure 2c). The liver architecture evolves considerably between 8 and 15 PCW. Organization of hepatocyte aggregates increases, though hepatic lobules around a central vein and portal triad are not clearly visible. Haematopoietic islands are present in sinusoids and surrounding hepatocyte aggregates. Sinusoidal $CD68^+$ macrophages are surrounded by GlycophorinA $(GYPA)^+$ erythroid cells (Figure 2c). $CD1c^+$ DCs and $CD79a/CD20^+$ cells from the B-lineage are sparsely distributed (Figure 2c). Cell proportions approximate our scRNA-seq profile for haematopoietic cells but not hepatocytes (Figure 2c and Figure 1b), in keeping with the fragility of hepatocytes following *ex vivo* isolation and their high expression of mitochondrial genes[11]. By validating our single cell transcriptome dataset with multiple modalities, we provide an integrated map of haematopoietic cells in the fetal liver.

Fetal liver and NLT haematopoiesis

Next, we inferred trajectories of haematopoietic development. By force directed graph (FDG), we identify three connections to a central HSC/MPP node featuring erythroid-megakaryocyte-mast cells, B cell and innate/T-lymphoid cells and myeloid cells (Figure 3a

and Supplementary Video 1). Partition based approximate graph abstraction (PAGA) also supports the presence of a shared megakaryocyte-erythroid-mast cell progenitor (MEMP) downstream of HSC/MPP (Extended Data 3a). Genes dynamically modulated in the specification of erythroid, megakaryocyte and mast cell lineages are distinct: *TAL1* and *KLF1* in erythroid; *F11R*, *PBX1* and *MEIS1* in megakaryocyte; and *HES1* in mast cell differentiation (Extended Data 3b)[12–15]. We explored supporting factors for fetal liver erythropoiesis using CellPhoneDB[16] to predict specific/enriched receptor-ligand interactions between erythroblasts and VCAM1$^+$ EI macrophages (Extended Data 4a). We identify statistically significant interactions for *VCAM1, ITGB1, ITGA4*, *SIGLEC1, ICAM4* and *SPN*, molecules known to be important in haematopoiesis (Extended Data 4a)[17,18]. The presence of VCAM1 on EI macrophages and ITGA4 on early/mid erythroid cells is confirmed by immunohistochemical analysis on serial fetal liver sections (Extended Data 4b). VCAM1$^+$ EI macrophage interaction with erythroblasts is also observed using Imaging Flow Cytometry ImageStream analysis (Extended Data 4d) and may explain their combined erythroblast and macrophage transcriptome (Figure 2a), which has also been described in mouse central EI macrophages (Extended Data 4g)[19].

Comparing across haematopoietic tissues, mast cells are also present in yolk sac (Extended Data 1b). Erythroblasts show expression of haemoglobin genes, and a temporal shift from Gower 1 and 2 subunit expression (*HBZ*, *HBE1)* to fetal haemoglobin subunit expression (*HBA1* and *HBG2)* between yolk sac and liver (Figure 3b).

Megakaryocytes, erythroid cells, mast cells and MEMP, are present in NLT, but HSC/MPPs are absent (Extended Data 1a, Extended Data 3c-d). We compared the highly expressed and differentially expressed genes of corresponding cell types in fetal liver, skin and kidney (Figure 3c). Mast cells, megakaryocytes and cells of the erythroid lineages show high connectivity (PAGA scores) across all four tissues (Extended Data 3d). Local maturation of progenitors in NLT or influx of cells at various differentiation stages are two possibilities. Erythroid cells are absent in kidney, suggesting restricted differentiation of the MEMP lineage in certain sites (Extended Data 1a, Figure 3c). Immunohistochemical analysis of serial skin sections show nucleated GYPA$^+$ cells inside and outside of CD34$^+$ blood vessels, in keeping with local differentiation of MEMPs (Figure 3d). Light sheet fluorescence microscopy supported this finding (Figure 3e, Supplementary Video 2). The proliferative capacity of MEMP in NLT is confirmed using *MKI67* and cell cycle gene expression

(Extended Data 3e). Skin MEMP express some early erythroblast genes including *MYL4* (Figure 3c)[20], suggesting that these may act as erythroid progenitors *in situ* in the skin. These findings demonstrate that during early development, the skin in physiological state can contribute to erythropoiesis and supplement fetal liver erythroid output.

Lymphoid lineages in fetal liver and NLT

Previous studies have reported the presence of T and B lymphocytes[21], NK cells[22], and ILCs[23] in the human fetal liver. We observe two lymphoid branches; an NK/T/ILC lineage and a B-lineage (Figure 4a and Extended Data 5a). The 'early lymphoid/T lymphocyte' cluster varies by gestational stage, with cells expressing *GATA3*, *KLRB1*, *CD3D, CD7* and *JCHAIN* at 7-8PCW before T cells emerge from the thymus (Figure 4a and Extended Data 5b-c). Early in gestation, this cluster may contain the fetal liver early thymocyte progenitor, which is capable of generating αβT cells upon co-culture with thymic epithelial cells[24,25]. At the 12-14 PCW stage, cells express *TRDC* and *TRAC,* but no *GZMB* or *PRF1*, the cytoplasmic granular products characteristic of mature $CD8^+$ T cells. *TRDC* expression is absent at the 15-17 PCW stage (Extended Data 5b-d). These findings are in keeping with the seeding of fetal liver by γδT cells and αβT cells sequentially following their exit from thymus after 12 PCW[26] and are consistent with previous reports of T cell identification only after 18 PCW[25,27].

NK cells (expressing *NCAM1*, *CD7*, *IL2RB* and *CD3E*) and ILC precursors (expressing *KIT*, *KLRB1*, *IL7R*, *RORC*) share a common origin in the lymphoid branch by PAGA and diffusion map analyses (Figure 2a, 4a, Extended Data 5a, 5h and Supplementary Table 3). This is in keeping with existing literature of a shared progenitor for NK and ILCs in human and
mouse[28,29].

The B-lineage cells are a continuum of differentiation states, from primitive 'pre pro-B' clusters expressing *CD34*, *SPINK2,* and *IGLL1* to 'pro-B' and 'pre-B' clusters with increasing expression of B cell transcripts *MS4A1*, *CD79B, DNTT* and *HLA-DRA* and reducing expression of *JCHAIN* and *LTB*[30] (Figure 2a, 4a and Supplementary Table 3). 'Pro/pre-B' cell clusters have high nuclear to cytoplasmic ratio, immature chromatin and nucleoli (Extended Data 5e). Differentially regulated genes in the HSC/MPPs to B cell

transition include *SPIB*, *SP100* and *CTSS* (Extended Data 5e). Pre-B cells are detected between 7-8 PCW, but mature B cells only after 9 PCW (Figure 1b-c and 4a). We observe a decline in HSC/MPP expression of *NFKBIA*, an inhibitor of NF-κB, and an increase in Kupffer cell *TNFSF13B* (BAFF) expression with gestation (Extended Data 5f). NF-κB and BAFF are known B cell survival and differentiation factors[31]. The cell-intrinsic *versus* tissue-microenvironment factors controlling B cell differentiation in the fetal liver require further investigation.

Comparing liver with yolk sac and NLT, pro-B, pre-B and B cells are present in NLT but HSC/MPPs and pre pro-B cells are absent (Figure 4b and Extended Data 5g). NK cell precursors, NK cells and ILC in NLT share a transcriptional signature with their liver counterparts, however tissue-specific expression of chemokine (*XCLI, CXCL8*) and cytotoxic granule genes (*GNLY*) suggest maturation and tissue adaptation in the skin and kidney (Figure 4c). ILC precursors in NLT lack the full characteristic markers and TFs of their mature progenies; ILC1, ILC2 and ILC3 (Extended Data 5h). NK cells and ILC precursors are present in yolk sac (Figure 4b). Together, these findings suggest that NLTs are seeded by NK and ILC precursors from fetal liver and potentially yolk sac, which differentiate *in situ* and acquire tissue-related gene expression profiles.

Tissue signatures in myeloid cells

In mice, fate-mapping studies have demonstrated that tissue macrophages are seeded by yolk sac and fetal liver progenitors[32,33], while dendritic cells (DCs) originate from BM-derived HSC/MPPs through a monocyte-independent lineage[34]. We observe myeloid progenitors, monocytes, macrophages, DC1 and DC2 in fetal liver and NLT as early as 7 PCW (Figure 1b, 5a-b, Extended Data 1a, 6a).

Myeloid lineages stem from HSC/MPP via three intermediates: a neutrophil-myeloid progenitor expressing *CD34*, *SPINK2*, *AZU1, PRTN3, ELANE*, *MPO* and *LYZ,* monocyte- and DC- precursors (Figure 2a, Figure 3a and Supplementary Table 3). DC1 differentiate from neutrophil-myeloid progenitors and DC-precursors link closely to DC2. DC and monocyte differentiation involve dynamic regulation of *CLEC11A*, *BATF3* and *ID2* while monocyte differentiation involves *S100A8/A9, FCGR1A/2A* and *S100A12* (Extended Data 6b). Plasmacytoid DC (pDC) precursors branch from both early myeloid precursors and pre

pro-B cells in keeping with recent reports of their mixed lymphoid and myeloid origin in mice (Extended Data 6a)[35,36]. We compared monocytes, macrophages and their putative precursors between placenta, yolk sac and fetal liver (Figure 5a-b and Extended Data 6c). The macrophage transcriptome profile is highly tissue specific, showing connectivity to other macrophage subtypes within their tissue of residence and obscuring potential ontogenic relationships (Figure 5a-b and Extended Data 6c-e).

Monocytes, macrophages, pDC, DC1 and DC2 are present in both skin and kidney (Extended Data 1a). NLT monocytes and DCs correlate strongly to fetal liver counterparts but macrophages are more tissue-specific, with *VCAM1* expression in liver and *F13A1* in skin, as is observed in adult tissue macrophages[37,38] (Extended Data 6e). Tissue specific gene expression patterns conferring functional specialization are discernible for DCs, for example *S100A4* in skin DC1 (a molecule involved in T cell activation)[39] and *AOAH* in liver DC2 (an enzyme involved in lipopolysaccharide response modulation)[40] (Extended Data 6e). DC activation despite the sterile fetal environment suggests an active role for fetal DCs in mediating tolerance as previously reported[41].

HSC/MPP differentiation potential by age

Our observation of an HSC/MPP cell state from which the earliest lineage-committed cells radiate is in keeping with recent observations from scRNA-seq analysis in post-natal mice and humans[35,42,43] (Figure 6a). At the base of this cluster by FDG visualization is a population expressing *CLEC9A, HLA-DRA* and highest levels of primitive genes including *MLLT3* consistent with a multipotent long-term repopulating (LT)-HSC (Figure 6a-b)[44]. Using differentially expressed genes and supervised analysis, HSC/MPP clusters with gene expression intermediate between LT-HSC and early progenitors similar to human lymphoid-primed short-term (ST)-HSCs[44], the mouse erythroid-biased MMP2, and myeloid-biased MMP3[45,46] are identifiable, demonstrating early transcriptome priming along all differentiation branches within the MPP pool (Figure 6a-b).

We hypothesized that the cellular composition of the developing fetal liver resulted from local modulation of HSC/MPP potential. To test this, we FACS-isolated single cells from the $CD34^+CD38^+$, $CD34^+CD38^-CD45RA^-$ and $CD34^+CD38^-CD45RA^+$ FACS gates and profiled them by both plate-based single cell transcriptomics (Smart-seq2) and single cell clonal differentiation assays[44,47] (Extended Data 7a). A support vector machine trained on the fetal

liver dataset identified enrichment of HSC/MPPs (~85%) in the $CD34^+CD38^-$ gate, with the majority of cells in the $CD34^+CD38^-CD45RA^+$ MLP gate also classified as HSC/MPP (Extended Data 7b). This is in agreement with reported transcriptional similarity between HSC/MPP and MLP[78] and our identification of lymphoid priming in the HSC/MPP compartment. Single cell culture from $CD34^+CD38^-CD45RA^-$ HSC/MPPs yield both uni- and multipotential colonies (Figure 6c-e and Extended Data 7c-g). There is a significant reduction in trilineage colonies with gestational age (Extended Data 7f). Colonies containing erythroid cells significantly decrease while those containing NK cells and B cells increase with gestational age (Figure 6c-e, Extended Data 7c-g). HSC/MPP from <9 PCW embryonic livers generate almost no B cells (Figure 6e), in keeping with the paucity of B cells at this stage (Figure 1c, 4a and Extended Data 1g). These findings support the hypothesis of differential HSC/MPP intrinsic potential by gestational stage and mirror our observation of early erythroid predominance and greater lymphoid representation at later stages (Figure 1c and Figure 4a).

Comparing HSC/MPPs and early progenitors across haematopoietic tissues during development, higher *MKI67* expression and cell cycle staining suggest enhanced proliferative potential of yolk sac and fetal liver progenitors/HSC/MPPs[48–50] (Extended Data 7h). The fraction of fetal liver HSC/MPP in G_0 increases with gestational age (Figure 6f, Extended Data 7i), indicating a progressive shift to quiescence during fetal life. Fetal liver HSC/MPP have higher expression of genes encoding a heat shock protein (*HSPA1A*), potentially for maintenance of genome and proteome integrity, and lower levels of MHC-I (*HLA-B*) suggesting reduced antigen presenting potential compared to cord blood and adult BM HSC/MPPs (Figure 6g).

Collectively, our findings demonstrate that intrinsic changes in HSC/MPP numbers, proliferation and differentiation potential occur over the first and second developmental trimesters. These changes are likely to be pivotal for fetal liver haematopoiesis to adapt to the needs of the developing fetus; first the establishment of an effective oxygen transport system and subsequently the development of a complete blood and immune system.

Discussion

Development of the human immune system *in utero* has remained poorly understood. Using single cell transcriptome profiling, we resolve the cellular heterogeneity and abstract dynamic

temporal information on blood and immune development in fetal liver, yolk sac, skin and kidney. Large scale scRNA-seq studies during human development must account for technical batch effects without compromising detection of biological variations over gestation. Our approach highlights key insights; physiological erythropoiesis in fetal skin, establishment of DC network as early as 7 PCW, potential dual myeloid and lymphoid origin of pDCs, seeding of mast cells, NK and ILCs from the yolk sac and tissue adaptation of NKs, ILCs, DCs and macrophages during development. Our findings reveal modulation of HSC/MPP intrinsic differentiation potential over gestation age suggesting this as an additional functional mechanism to regulate haematopoietic output of the fetal liver throughout the first and second trimesters.

In summary, our comprehensive fetal liver atlas provides a foundational resource for understanding fetal liver haematopoiesis and the developing immune system. Our reference dataset will be invaluable for studies on paediatric blood and immune disorders and exploiting HSC/MPPs for therapy. Our approach using single cell transcriptomics to study human development provides a framework that can be applied to study any temporal processes across the human lifespan.

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Figure Legends

Figure 1: Single cell transcriptome map of fetal liver. **a,** Schematic of tissue processing and cell isolation for scRNA-seq profiling of fetal liver, skin and kidney across four developmental stages (7-8, 9-11, 12-14, and 15-17 post conception weeks (PCW)), and yolk sac from 4-7 PCW. SS2, Smart-seq2. **b,** UMAP visualisation of fetal liver cells from 10x using 3' chemistry. Colours indicate cell state. HSC/MPP, haematopoietic stem cell/multipotent progenitor; ILC, innate lymphoid cell; NK, natural killer cell; Neut-myeloid, neutrophil-myeloid; DC, dendritic cell; pDC, plasmacytoid DC; Mono-mac, monocyte-macrophage; EI, erythroblastic island; Early L/TL, Early lymphoid/T lymphocyte; MEMP, megakaryocyte-erythroid-mast cell progenitor. Statistical significance of cell frequency change by stage shown in parentheses (negative binomial regression with bootstrap correction for sort gates; * $p < 0.05$, *** $p < 0.001$, and **** $p < 0.0001$ as per SI Table 8) with up/down arrows to indicate positive/negative coefficient of change, respectively. **c**, Liver composition by developmental stage as the mean percentage of each population per stage corrected by $CD45^+/CD45^-$ sort fraction. Colours indicate cell states as shown in **b.**

Figure 2: Multi-modal and spatial validation of cell types. **a,** Median scaled ln-normalised gene expression of 48 selected differentially expressed genes for the liver cell states from **1b** visualised by dot; asterisk (*) indicates markers used for FACS-isolation of cells. Gene expression frequency (% cells within cell type expressing) indicated by spot size and expression level by colour intensity. **b,** Representative Giemsa-stained cytospins showing morphology of populations isolated by FACS based on differentially expressed genes with * in **a**. Scale bar, 10μm. **c,** Overlay pseudo-colour Hyperion representative images for 8 PCW and 15 PCW fetal liver. Far left images are shown at 5x magnification with zoom of insets on right at 20x magnification (1μm/pixel). Bile ducts are marked with an *.

Figure 3: Fetal liver and NLT haematopoiesis. **a,** Force-directed graph (FDG) visualisation of all haematopoietic cells from **1b**. **b,** Dot plot showing the median scaled ln-normalised expression of globin genes encoding haemoglobin subunits; *HBZ* and *HBE1* (Gower 1), *HBE1* and *HBA1* (Gower 2) and *HBA1* and *HBG2* (fetal) in liver, skin, and yolk sac erythroid lineages (MEMP, early, mid and late erythroids). Gene expression frequency (% cells within cell type expressing) indicated by spot size and expression level by colour intensity. **c,** Heat map showing the scaled ln-expression of selected marker genes in fetal

liver, NLT and yolk sac subsets. **d,** Representative immunohistochemical staining of sequential sections of 8 PCW fetal skin for endothelium ($CD34^+$) and erythroblasts (nucleated and $GYPA^+$), nuclei stained with blue alkaline phosphatase. Zoom in of insets (right) bordered with black (top) indicate nucleated cells stained positive for GYPA within $CD34^+$ blood vessels, and those bordered with red (bottom) indicate nucleated $GYPA^+$ cells outside $CD34^+$ blood vessels. Scale bar, 100μm. **e,** Representative light sheet fluorescence microscopy of embryo (5 PCW) hand skin. Scale bar, 5μm; TO-PRO-3/nuclei = red, GYPA = green (see also Supplementary Video 2). < indicates extravascular nucleated erythroid cells.

Figure 4: Lymphoid lineages in fetal liver and NLT. a, FDG visualisation of fetal liver HSC/MPP and lymphoid cell types from **1b** showing changes over four developmental stages. **b,** FDG visualisation of fetal liver and corresponding skin, kidney and yolk sac lymphoid cells. **c,** ln-normalised median expression of selected known NK (left) and ILC precursor (right) marker genes and selected differentially expressed genes between liver (red), skin (blue) and kidney (green) visualised by violin plots (*** $p < 0.005$; **** $p < 0.001$).

Figure 5: Tissue signatures in developing myeloid cells. a, FDG visualisation of HSC/MPP, myeloid progenitors, monocytes and macrophages from fetal liver, decidua/placenta and yolk sac. Mac, Macrophage; Monocyte prec., Monocyte precursor; Neut-myeloid prog., Neutrophil-myeloid progenitor. **b,** PAGA connectivity scores of the populations shown in **a.**

Figure 6: HSC/MPP differentiation potential by gestation. a, FDG visualisation of liver HSC/MPP and early haematopoietic progenitor populations from **Figure 1b**. **b,** Violin plots showing ln-normalised median gene expression of statistically significant, dynamically variable genes that are up or downregulated during HSC/MPP transition to neutrophil-myeloid progenitors, MEMP and pre pro-B cells from fetal liver. Bar and 'ns' indicate not significant. H/M, HSC/MPP. **c,** Stacked barplot of all different types of colonies generated by single 'HSC pool' gate cells in an assay with MS5 stroma. * $p < 0.05$, *** $p < 0.001$, individual samples shown in **Extended Data 7c**. The colour of the stat bar corresponds to the type of colony tested vs all others. My, Myeloid; Ery, Erythroid, Meg, Megakaryocyte. **d**, Percentage of colonies generated by single 'HSC pool' gate cells containing erythroid cells (defined as the sum of Ery, Ery/Meg, Ery/Meg/My, Ery/My, Ery/NK and Ery/NK/My

colonies shown in **c**), *** $p < 0.001$. My, Myeloid; Ery, Erythroid; Meg, Megakaryocyte. **e,** Percentage of colonies containing B cells following culture in B/NK optimized conditions from 10 cells from 'HSC pool' gate (** $p < 0.01$). **f**, Mean +/- s.d. percentage of cells in the G_0 phase of the cell cycle assessed using Ki67 and DAPI flow cytometry analysis (* $p = 0.0136$). **g,** ln-normalised median expression of selected genes in yolk sac progenitors, cord blood HSC and adult bone marrow HSC with significant differential expression compared to fetal liver HSC/MPP, visualised by violin plots (**** $p < 0.0001$).

Extended Data 1: Single cell transcriptome map of fetal liver. **a**, Fetal skin and kidney haematopoietic cells visualised by UMAP. Colours indicate cell state. Inset: colours indicate tissue type. **b**, UMAP visualisation of yolk sac haematopoietic cells. Colours indicate cell state. Inset: colours indicate location within yolk sac. **c**, UMAP visualisation of 3' liver 10x cells post batch correction, coloured by sample. **d**, UMAP visualisation (top) of 3' 10x liver sample sex mixing grouped by developmental stage, and violin plots (bottom) showing ln-normalised median expression of *XIST* (green) and *RSP4Y1* (purple), which marks female and male samples respectively. **e**, UMAP visualisation of fetal liver composition by developmental stage. Colours indicate cell state. **f,** UMAP visualisation of fetal liver cells profiled using Smart-seq2. Colours indicate cell states as shown in **e**. **g**, Frequency (mean +/- s.e.m.) of B cells in the $CD34^-$ cells detected in 6-19 PCW fetal livers by flow cytometry (* $p < 0.05$; *** $p = 0.003$; **** $p < 0.001$).

Extended Data 2: Transcriptome validation of fetal liver cells. a, Assessment of 48 genes from the 4,471 highly variable genes by using a Random Forest classifier to assign cell labels, where 'true cell label' indicates the manual annotation based on the full list of variable genes. **b,** Comparison of representative mini bulk RNAseq data (in coloured triangles) and liver erythroblastic island (EI) populations (early, mid and late erythroids, $VCAM1^+$ EI macrophages), Kupffer cells and endothelium validated by SS2 (in colour) overlaid on whole liver SS2 populations (grey). **c,** Dot plot showing representative median scaled ln-normalised gene expression of 100 FACS-isolated liver cells based on marker gene expression in **Figure 2a**. Gene expression indicated by spot size and colour intensity. **d,** Dot plot showing median scaled ln-normalised gene expression of FACS sorted single cells from liver erythroblastic island (EI) populations (early, mid and late erythroids, $VCAM1^+$ EI macrophages), Kupffer cells and endothelium shown as coloured dots in **b** based on marker gene expression in

Figure 2a. Gene expression frequency (% cells within cell type expressing) indicated by spot size and expression level by colour intensity.

Extended Data 3: Fetal liver and NLT haematopoiesis. a, PAGA analysis of fetal liver HSC/MPP, erythroid, megakaryocyte and mast cell lineages from **Figure 3a.** Lines symbolise connection; line thickness corresponds to the level of connectivity (thin to thick = low to high PAGA connectivity). **b,** Heat map showing min-max normalised expression of statistically significant ($p < 0.001$), dynamically variable genes from pseudotime analysis for erythroid, megakaryocyte and mast cell inferred trajectories. Transcription factors in bold, asterisk (*) mark genes not previously implicated for the respective lineages. **c,** FDG visualisation of fetal liver, skin and kidney HSC/MPP, MEMP, erythroid, megakaryocyte and mast cell lineages. **d,** PAGA connectivity scores of HSC/MPP, erythroid, megakaryocyte and mast cell lineages between fetal liver, skin, kidney (K) and yolk sac. **e,** Stacked barplots (right) of the mean +/- s.d. percent of fetal liver (red), skin (blue) and kidney (green) HSC/MPP, MEMP, Erythroid, Megakaryocyte and Mast cells in each stage of the cell cycle (G_1 (navy), G_2M (blue), and S (white) phase), and ln-normalised median expression of *MKI67* transcript (right) in corresponding liver vs NLT cell types (total percent of *MKI67* expressing cells stated above plots; each dot represents a single cell). * $p < 0.05$; ** $p < 0.01$; *** $p < 0.005$.

Extended Data 4: Investigation of interactions between fetal liver macrophages and erythroid cells. a, Representative immunohistochemical staining of fetal liver for erythroblasts and macrophages with GYPA and CD68, respectively. Scale bar, 50μm. Statistically significantly ($p < 0.05$) enriched receptor-ligand interactions from CellPhoneDB between $VCAM1^+$ EI macrophages (purple) and two erythroid populations (early and mid; red) ($n = 14$ biologically independent samples). Asterisk (*) indicate protein complexes. Violin plots show ln-normalised median gene expression value of VCAM1 and ITGA4 in cells analysed by CellPhoneDB (marked by # in dot plot). **b,** Representative immunohistochemical staining of sequential sections of 8 PCW fetal liver for $VCAM1^+$ EI macrophages ($VCAM1^+$) and $CD49d^+GYPA^+$ cells with nuclei stained using blue alkaline phosphatase. Zoom in of insets (right) with coloured arrows indicating erythroblast (yellow) and $VCAM1^+$ EI macrophage (purple). Scale bar, 100μm. **c,** Representative gating strategy used to visualise fetal liver erythroid cells, $VCAM1^+$ EI macrophages, Kupffer cells, Mono-macs, and mast cells. **d**, Bright field, VCAM1 (CD106), CD34, CD45, KIT (CD117), GYPA,

CD14, and HLA-DR images for each cell type within gates shown in **c**. **e,** Representative bright field images of cells found within the single cell and doublet gates. **f,** Barplots showing the mean +/- s.d. proportion of each cell type within the single cell gate (white) or doublet gate (grey); * $p = 0.0194$. **g,** Comparison of macrophage and erythroid gene expression in mouse macrophages (red) and EI macrophages (blue), $n = 3$ from Li *et al*.[19].

Extended Data 5: Lymphoid lineages in fetal liver and NLT. a, PAGA analysis of fetal liver HSC/MPP and lymphoid cell types from **Figure 1b** showing changes over four developmental stages. Lines symbolise connection; line thickness corresponds to the level of connectivity (thin to thick = low to high PAGA connectivity). **b,** Feature plots and **c,** violin plots showing ln-normalised median expression of selected known NK, ILC and T cell genes over gestation for early lymphoid/T lymphocyte cluster; ** $p < 0.001$; *** $p < 0.005$; **** $p < 0.0001$. **d,** Dot plot showing median scaled ln-normalised median expression of V(D)J transcripts in fetal liver lymphoid cell types. Gene expression indicated by spot size and colour intensity. **e,** Heat map showing normalised expression of statistically significant, dynamically variable genes from pseudotime analysis for B cell lineage inferred trajectory (likelihood ratio test). Transcription factors are in bold. Morphology of liver Pro/Pre B cells and B cells by Giemsa stain after cytospin. **f**, ln-normalised expression (mean +/- s.e.m.) of *TNFSF13B* in Kupffer cells and *NFKBIA* in HSC/MPPs and cells in the B cell lineage across 4 developmental stages spanning 6-17 PCW; trend lines showing linear regression. **g,** PAGA connectivity scores of HSC/MPP and lymphoid cells from fetal liver, skin, kidney and yolk sac. **h,** Violin plots showing ln-normalised median expression of selected known ILC and NK cell genes expressed in ILC precursors from fetal liver, skin, and kidney.

Extended Data 6: Tissue signatures in developing myeloid cells. a, Diffusion map of fetal liver HSC/MPP, progenitors and precursors from **1b**. **b,** Heat map showing min-max normalised expression ($p < 0.001$) of dynamically variable genes from pseudotime analysis for monocyte, DC1 and DC2 inferred trajectories. Transcription factors in bold, * mark genes not previously implicated for the respective lineages. **c,** Heat map visualisation comparing scaled expression of the top marker genes of decidua/placenta (red), fetal liver (black) and yolk sac (purple) progenitor and myeloid populations. **d,** PAGA connectivity scores of HSC/MPP and myeloid cells from fetal liver, skin and kidney. **e,** ln-normalised median expression of 3 known marker genes & 3 differentially expressed genes in corresponding

myeloid populations across fetal liver, skin and kidney visualised by violin plots (* $p < 0.05$; *** $p < 0.005$; **** $p < 0.0001$).

Extended Data 7: HSC/MPP differentiation potential by gestation. a, Experimental design for single cell transcriptome and culture of fetal liver cells from representative FACS gates illustrated. **b**, Alignment of 349 scRNA-seq profiled cells from FACS gates in **a** with 10x profiled HSC/MPPs and early progenitors visualised using FDG, point shape corresponds to sequencing type (triangle = SS2 plate data, circle = 10X data). **c,** Stacked barplot of all different types of colonies generated by single 'HSC pool' gate cells (gate defined in **a**). **d,** Stacked bar plot of all different types of colonies generated by single 'HSC pool' gate cells without MS5 stroma layer (gate defined in **a**) by stage (left) and in individual samples (right), * $p < 0.05$. **e,** Percentage of colonies generated by single 'HSC pool' cells without MS5 stroma layer containing erythroid cells (sum of Ery, Ery/Meg, Ery/Meg/My, and Ery/My colonies shown in **c**), ** $p < 0.01$. **f**, Percentage of colonies from single cell culture (shown in **6c**) that differentiated along 3 lineages (defined as sum of Ery/NK/My and Ery/Meg/My colonies) branches (*** $p < 0.005$). **g,** Percentage of colonies containing NK cells following B/NK optimised culture of 10 cells from 'HSC pool' gate (* $p < 0.05$, ** $p < 0.01$). **h,** Percentage (Mean +/- s.e.m.) of HSC/MPP and early progenitors in fetal liver, yolk sac, cord blood and adult bone marrow expressing *MKI67* (* $p < 0.05$, ** $p < 0.01$, **** $p < 0.001$). **i**, Mean +/- s.d. percentage of $CD34^+CD38^-$ and $CD34^+CD38^+$cells in the indicated cell cycle phases (right) as determined by flow cytometry analysis (left, representative plot of $n = 8$ biologically independent samples) (G_0: $Ki67^-DAPI^-$, G_1: $Ki67^+DAPI^-$, S-G_2-M: $Ki67^+DAPI^+$ (left)).

Extended Data 8: Expression of known Primary Immunodeficiency (PID)-linked genes in fetal liver. Dot plots showing relative expression of genes known to be associated with major PID disease categories in fetal liver cell types from **Figure 1b**. Early L/T L, Early lymphoid/T lymphocyte. Gene expression frequency (% cells within cell type expressing) indicated by spot size and expression level by colour intensity.

Extended Data 9: FACS gating strategy for scRNA-seq analysis. a, Gating strategy used to FACS-isolate cells for droplet-(10x) and plate-based scRNA-seq (Smart-seq2) for samples F2-F17. **b,** Gating strategy used to FACS-isolate cells for cytospins, scRNA-seq (Smart-seq2) and 100 cell RNA-seq. **c,** Flow cytometry gating strategy used to identify the colonies

cultured *in vitro* from single cells as shown in **Figure 6c. d,** Flow cytometry gating strategy used to identify B and NK colonies cultured *in vitro* from 10 cells as shown in **Figure 6e.**

Materials and methods

Tissue Acquisition

Human fetal tissues were obtained from the MRC/Wellcome Trust-funded Human Developmental Biology Resource (HDBR; http://www.hdbr.org)[51] with appropriate written consent and approval from the Newcastle and North Tyneside NHS Health Authority Joint Ethics Committee (08/H0906/21+5). HDBR is regulated by the UK Human Tissue Authority (HTA; www.hta.gov.uk) and operates in accordance with the relevant HTA Codes of Practice.

Embryos and fetal specimens used for light sheet fluorescence microscopy were obtained with written informed consent from the parents (Gynecology Hospital Jeanne de Flandres, Lille, France) with the approval of the local ethics committee (protocol N°PFS16-002). Tissues were made available in accordance with the French bylaw (Good practice concerning the conservation, transformation and transportation of human tissue to be used therapeutically, published on December 29, 1998). Permission to utilize human tissues was obtained from the French agency for biomedical research (Agence de la Biome´decine, Saint-Denis La Plaine, France).

Tissue Processing

All tissues were processed immediately after isolation using the same protocol. Tissue was transferred to a sterile $10mm^2$ tissue culture dish and cut into $<1mm^3$ segments before being transferred to a 50mL conical tube. Yolk sac content was aspirated for analysis prior to yolk sac digestion. Tissue was digested with 1.6mg/mL collagenase type IV (Worthington) in RPMI (Sigma-Aldrich) supplemented with 10%(v/v) heat-inactivated fetal bovine serum (Gibco), 100U/mL penicillin (Sigma-Aldrich), 0.1mg/mL streptomycin (Sigma-Aldrich), and 2mM L-Glutamine (Sigma-Aldrich) for 30 minutes at 37°C with intermittent shaking. Digested tissue was passed through a 100µm filter, and cells collected by centrifugation (500g for 5 minutes at 4°C). Cells were treated with 1X RBC lysis buffer (eBioscience) for 5 minutes at room temperature and washed once with flow buffer (PBS containing 5%(v/v) FBS and 2mM EDTA) prior to counting.

Fetal developmental stage assignment and chromosomal assessment

Embryos up to 8 post conception weeks (PCW) were staged using the Carnegie staging method[52]. After 8 PCW, developmental age was estimated from measurements of foot length and heel to knee length and compared against a standard growth chart[53]. A piece of skin, or where this was not possible, chorionic villi tissue was collected from every sample for Quantitative Fluorescence-Polymerase Chain Reaction analysis using markers for the sex chromosomes and the following autosomes 13, 15, 16, 18, 21, 22, which are the most commonly seen chromosomal abnormalities. All samples were karyotypically normal.

Flow cytometry and FACS for scRNA-seq

Antibody panels were designed to allow enrichment of cell fractions for sequencing and cell type validation. Antibodies used for FACS isolation are listed in Supplementary Table 14. An antibody cocktail was prepared fresh by adding 3µL of each antibody in 50µL Brilliant Stain Buffer (BD) per tissue. Cells ($<10x10^6$) were resuspended in 50-100µL flow buffer and an equal volume of antibody mix was added to cells from each tissue. Cells were stained for 30 minutes on ice, washed with flow buffer and resuspended at $10x10^6$ cells/mL. DAPI (Sigma-Aldrich) was added to a final concentration of 3µM immediately prior to sorting. Flow sorting was performed on a BD FACSAria™ Fusion instrument using DIVAv8, and data analysed using FlowJo (v10.4.1, BD). Cells were gated to exclude dead cells and doublets, and then isolated for scRNA-seq analysis (10x or Smart-seq2). For 10x, cells were sorted into chilled FACS tubes coated with FBS and prefilled with 500µL sterile PBS. For Smart-seq2, single cells were index-sorted into 96-well lo-bind plates (Eppendorf) containing 10µL lysis buffer (TCL (Qiagen) + 1% (v/v) β-mercaptoethanol) per well. B cells were also investigated by flow cytometry as per Roy *et. al.*[54].

Cytospins and mini bulk RNA-seq validation

Fetal liver cells were immunostained with two separate panels (see Supplementary Table 15 for antibody details). Cells were stained for 30 minutes on ice followed by DAPI staining. FACS was performed on a BD FACSAria™ Fusion instrument, and data analysed using FlowJo (v10.4.1). Cells were isolated into chilled FACS tubes coated with FBS and prefilled with 500µL sterile PBS for cytospin (500 – 2000 cells), or into 1.5mL microfuge tubes containing 20µL lysis buffer (100 cells). Giemsa staining (Sigma-Aldrich) was used to determine the morphology of sorted cells on cytospins. Slides were viewed using a Zeiss AxioImager microscope, images taken of 4 fields from $n = 3$ samples using the 100x objective, and viewed using Zen (v2.3).

HSC/MPP Culture

MS5 in log-phase growth (DSMZ, Germany, passage 6-10, or imported from Prof. Katsuhiko Itoh at Kyoto University; all tested mycoplasma-free, not authenticated) were seeded into 96-well flat-bottom plates (Nunclon delta surface; Thermo) at a density of 3000 cells per well 24 hours prior to sorting. Medium was Myelocult H5100 (Stem Cell Technologies) supplemented with 100U/mL Penicillin and 0.1mg/mL Streptomycin (Sigma-Aldrich). On the day of sorting, media were replaced with Stem Pro-34 SFM media (Life Technologies) supplemented with 100U/mL Penicillin and 0.1mg/mL Streptomycin, 2mM L-glutamine (Sigma-Aldrich), stem cell factor 100ng/ml (Miltenyi), Flt3 20ng/ml (Miltenyi), TPO 100ng/ml (Miltenyi), EPO 3ng/ml (Eprex), IL-6 50ng/ml (Miltenyi), IL-3 10ng/ml (Miltenyi), IL-11 50ng/ml (Miltenyi), GM-CSF 20ng/ml (Miltenyi), IL-2 10ng/ml (Miltenyi), IL-7 20ng/ml (Miltenyi) and Lipids (hLDL) 50ng/ml (Life Technologies)[44].

Frozen fetal liver cells were thawed and stained with 10µl/10^6 cells of antibody cocktail (see Supplementary Table 16-17 for antibody details) for 30 minutes on ice. Three populations of HSC/MPPs and progenitors were isolated from fetal liver suspension. Populations were identified from the $DAPI^-$, doublet-excluded gate as $CD3/CD16/CD11c/CD14/CD19/CD56^-$, $CD34^+$ cells (see Supplementary Table 16 for antibody details). The HSC/MPP pool and MLP were found within the 20% of cells with lowest CD38 expression: HSC/MPP pool were $CD90^{+/-}$ and $CD45RA^-$ whilst MLP were $CD90^-CD45RA^+$. Progenitors with the highest 20% of CD38 expression were sorted for comparison. Single cells were sorted using a BD FACSAria™ Fusion, and sorted directly onto MS5 or medium for culture, or into 96-well lo-bind plates containing 10µl/well lysis buffer for Smart-seq2 scRNAseq (Supplementary Table 12). Single-cell-derived colonies analysis was performed as described by the Laurenti Lab[44]. In brief, colonies were harvested into 96 U-bottom plates using a plate filter to prevent the carryover of MS5 cells. Cells were stained with 50µl/well of antibody cocktail (Supplementary Table 16-17 for antibody details), incubated for 20 minutes in the dark at room temperature and then washed with 100µl/well of PBS + 3% FBS. The type (lineage composition) and the size of the colonies formed were assessed by high-throughput flow cytometry (BD FACS Symphony). Colony output was determined using the gating strategy shown in Extended Data 9c. A single cell was defined as giving rise to a colony if the sum of cells detected in the $CD45^+$ 5^+ and $GYPA^+$ gates was ≥ 30 cells. Erythroid colonies were identified as $CD45^-GYPA^+ \geq 30$ cells, Megakaryocyte colonies as $CD41^+ \geq 30$ cells, Myeloid

colonies as [($CD45^+CD14^+$) + ($CD45^+CD15^+$)] ≥ 30 cells, NK colonies as $CD45^+CD56^+$ ≥ 30 cells. All high-throughput screening flow cytometry data was recorded in a blinded way, and correlation between the colony phenotype and originating population was only performed at the final stage. Two-tailed Fisher's exact test, performed in Prism (v8.1.2, GraphPad Software), were applied to the numbers of colonies of each type by stage to determine statistical significance in lineage differentiation potential with development. For differentiation of B cells from HSC/MPP pool, 10 cells were sorted directly on MS5 stroma in the same conditions as Doulatov *et al.*[47] (see Supplementary Table 17 for antibody details). An example of gating strategy is shown in Extended Figure 8d. For Ki67 staining, cells were stained using antibody panel in Supplementary Table 18, $CD34^+$ cells were sorted, fixed/permeabilised using the BD Cytofix/Cytoperm kit according to the manufacturer's instructions and then stained overnight for Ki67-FITC followed by DAPI as a DNA dye.

ImageStream analysis of fetal liver cell suspensions

Frozen fetal liver cells were thawed and stained with the antibody cocktail (see Supplementary Table 19 for antibody details) for 30 minutes on ice. Cells were washed with flow buffer and resuspended at the same cell density employed for cell sorting experiments ($10x10^6$ cells/mL). DAPI (Sigma-Aldrich) was added to a final concentration of 3μM immediately prior to acquisition in order to identify and exclude dead cells from the experiment. Samples were acquired on a fully calibrated ImageStream X MKII system (Luminex Corporation, USA) using 488nm, 561nm, 405nm and 642nm excitation lasers and the 60x magnification collection optic. Laser powers were set in order to maximise signal resolution but minimise any saturation of the CCD camera with bright-field (BF) images collected in channels 1 and 9. A minimum of 50,000 cell events were collected per sample. In order to calculate spectral compensation, single stained antibody capture beads were acquired with the bright-field illumination turned off. Spectral compensation and data analysis were performed using the IDEAS analysis software (v6.2.64, Luminex Corp). Briefly, dead cells were first excluded based on DAPI positivity. Overt doublets and debris were excluded from the live cell population using the aspect ratio and area of the BF image in channel 1. Single cells had an intermediate area value and an aspect ratio between 1 and 0.6 and were gated as such. Overt doublets present as having an increased area and a lower aspect ratio value and were also gated as such. In all cases the position and boundaries of a gate was checked for appropriateness using the underlying imagery. The key cell types within the sample were then identified and gated using the total integrated (post-

compensation) fluorescence signals from each labelled antibody in the panel in an analogous fashion to conventional flow cytometry data. Each major cell type was then interrogated using the associated multi-spectral images for true single cell identity or for the presence of "pernicious doublets" (cells with either significant debris attached or large cells with much smaller cells attached). The same phenotypic analysis was extended to the cells in the overt doublet gate. Further analysis was performed in FlowJo (v10.4.1). Kruskal-Wallis with Dunn's post hoc, performed in Prism (v8.1.2, GraphPad Software), were applied to the proportion of each cell type found within the doublet gate to determine statistical significance in doublet rates across fetal liver cell types.

Library Preparation and Sequencing

For the droplet-encapsulation scRNA-seq experiments, 7,000 live, single, $CD45^+$ or $CD45^-$ FACS-isolated cells were loaded onto each channel of a Chromium chip before encapsulation on the Chromium Controller (10x Genomics, Pleasanton, CA, USA). Single cell sequencing libraries were generated using the Single Cell 3' v2 and V(D)J Reagent Kits (for T cell receptor repertoire analysis) as per the manufacturer's protocol. Libraries were sequenced using an Illumina HiSeq 4000 using v4 SBS chemistry to achieve a minimum depth of 50,000 raw reads per cell. The libraries were sequenced using the following parameters: Read1: 26 cycles, i7: 8 cycles, i5: 0 cycles; Read2: 98 cycles to generate 75bp paired end reads.

For the plate-based scRNA-seq experiments, a slightly modified Smart-seq2 protocol was used as previously described[55]. After cDNA generation, libraries were prepared (384 cells per library) using the Illumina Nextera XT kit (Illumina Inc, San Diego, CA, USA). Index v2 sets A, B, C and D were used per library to barcode each cell before multiplexing. Each library was sequenced to achieve a minimum depth of 1-2 million raw reads per cell using an Illumina HiSeq 4000 using v4 SBS chemistry to generate 75bp paired end reads.

For the mini bulk RNA-seq experiments, each cell lysate was transferred into a 96-well lo-bind plate (Eppendorf) then processed using the same modified Smart-seq2 protocol as described above. After cDNA generation, libraries were prepared using the Illumina NexteraXT kit with Index v2 set A to barcode each mini bulk library before multiplexing. All libraries were sequenced on one lane of an Illumina HiSeq 4000 using v4 SBS chemistry to generate 75bp paired end reads and aiming to achieve a minimum depth of 10 million reads per library.

841

842 **Immunohistochemistry**

843 Formalin fixed, paraffin embedded blocks of fetal livers aged 6 PCW, 8 PCW, 10 PCW and
844 13 PCW were obtained from the HDBR. Each was sectioned at 4μm thickness onto APES-
845 coated slides. Sections were dewaxed for 5 minutes in Xylene (Fisher Chemical) then
846 rehydrated through graded ethanol (99%, 95% and 70%; Fisher Chemical) and washed in
847 running water. Sections were treated with hydrogen peroxide block (1%v/v in water; Sigma)
848 for 10 minutes and rinsed in tap water prior to antigen retrieval. Citrate antigen retrieval was
849 used for all sections. Citrate buffer, pH6 was used with pressure heating for antigen retrieval,
850 and then slides placed in TBS, pH7.6 for 5 minutes prior to staining. Staining was done using
851 the Vector Immpress Kit (Vector Laboratories). Sections were blot dried and blocked
852 sequentially with 2.5% normal horse serum, avidin (Vector Laboratories) and then biotin
853 (Vector Laboratories) for 10 minutes each and blot dried in between. Sections were incubated
854 for 60 minutes with primary antibody diluted in TBS pH7.6 (see Supplementary Table 20 for
855 antibody details). Slides were washed twice in TBS pH7.6 for 5 minutes each before
856 incubation for 30 minutes with the secondary antibody supplied with the kit. Slides were
857 washed twice in TBS pH7.6 for 5 minutes each, and developed using peroxidase chromogen
858 DAB. Sections were counterstained in Mayer's Haematoxylin for 30 seconds, washed and
859 put in scots tap water for 30 seconds. Slides were dehydrated through graded ethanol (70% to
860 99%) and then placed in Xylene prior to mounting with DPX (Sigma-Aldrich). Sections
861 were imaged on a Nikon Eclipse 80i microscope using NIS-Elements Fv4.

862

863 **Alignment, quantification and quality control of scRNA-seq data**

864 Droplet-based (10x) sequencing data was aligned and quantified using the Cell Ranger
865 Single-Cell Software Suite (version 2.0.2, 10x Genomics Inc) using the GRCh38 human
866 reference genome (official Cell Ranger reference, version 1.2.0). Smart-seq2 sequencing
867 data was aligned with *STAR* (version 2.5.1b), using the STAR index and annotation from the
868 same reference as the 10x data. Gene-specific read counts were calculated using *htseq-count*
869 (version 0.10.0). Cells with fewer than 200 detected genes and for which the total
870 mitochondrial gene expression exceeded 20% were removed. Genes that were expressed in
871 fewer than 3 cells were also removed. We detected on average ~3,000 genes per cell with the
872 10x Genomics platform and ~6,000 genes with the Smart-seq2 protocol.

873

874 **Doublet detection**

Doublets were detected with an approach adapted from Pijuan-Sala *et al.*[56]. In the first step of the process, each 10X lane was processed independently. Scrublet[57] was run, obtaining per-cell doublet scores. The standard Seurat-inspired *Scanpy* processing pipeline was performed up to the clustering stage, using default parameters. Each cluster was subsequently separately clustered again, yielding an over clustered manifold, and each of the resulting clusters had its *Scrublet* scores replaced by the median of the observed values. The resulting scores were assessed for statistical significance, with *p*-values computed using a right-tailed test from a normal distribution centred on the score median and a MAD-derived standard deviation estimate. The MAD was computed from above-median values to circumvent zero-truncation. The *p*-values were FDR-corrected with the Benjamini-Hochberg procedure, and a significance threshold of 0.1 was imposed. In the second step of the process, all 10X lanes for a single tissue were pooled together and the Seurat-inspired *Scanpy* processing was repeated, with the addition of *Harmony*[58] with a theta of 3 for batch correction between the lanes before the neighbour graph identification step. The joint manifold was clustered, and the frequency of identified doublets was computed. The same statistical framework as in the first step was used to identify clusters significantly enriched in doublets, which were subsequently flagged as doublets in their entirety and removed.

Clustering and annotation

Downstream analysis included data normalisation (*NormalizeData*, LogNormalize method, scaling factor 10000), data feature scaling (*ScaleData*), variable gene detection (*FindVariableGenes*), PCA (*RunPCA*, from variable genes) and Louvain graph-based clustering (*FindClusters*, data dimensionality reduction using PCA, clustering resolution (res.30)) performed using the R package *Seurat* (version 2.3.4). Cluster cell identity was assigned by manual annotation using known marker genes and computed differentially expressed genes (DEGs) using *FindAllMarkers* function in *Seurat* package (one-tailed Wilcoxon rank sum test, *p*-values adjusted for multiple testing using the Bonferroni correction; Supplementary Table 3). For computing DEGs all genes were probed provided they were expressed in at least 25% of cells in either of the two populations compared and the expression difference on a natural log scale was at least 0.25. Manual annotation was performed iteratively, which included validating proposed cell labels with known markers and further investigating clusters whose gene signatures indicated additional diversity.

Number of each cell type per sample, annotations per cell, and nGene and nUMI per cell type are reported in Supplementary Tables 4, 5, 8 and 9, respectively.

Clustering and cell type assignment for fetal liver data was assessed using two additional clustering methods (not shown): Agglomerative clustering (with Ward linkage and Euclidean affinity) and Gaussian mixture (*AgglomerativeClustering* class from *cluster* module and *GaussianMixture* from *mixture* module in *sklearn* version 0.19.1 Python 3.6.3). Consensus agreement between the 3 clustering methods was measured by Rand index and adjusted mutual information implemented in the *metrics* module in *sklearn* package. The Rand Index scores were 0.89 and 0.85 for Agglomerative and Gaussian Mixture clustering methods respectively.

After annotation was completed, a cell type classifier was built by training an SVM on labelled fetal liver scRNA-seq data with grid search for parameter optimization based on training data. 70% of the data was used for training and the other 30% for test. The SVM was previously compared in terms of accuracy and recall with a random forest and logistic regression classifiers trained on the same data. Out of the 3 classifiers the SVM was chosen due to showing a mean accuracy and weighted mean recall of 95%. Random forest showed 89% for both precision and recall (Supplementary Table 11). The SVM classifier was used for automatic annotation of the Smart-seq2 and mini bulk RNA sequencing data sets to allow identification of biologically meaningful clusters and DEG computation.

Data generated from fetal skin, kidney and yolk sac was pre-processed, normalised, clustered and manually annotated, in parallel with, and using the same pipeline as, the liver data. Annotation by cell type for skin and kidney, and yolk sac are reported in Supplementary Tables 6 and 7, respectively. Skin and kidney data were combined using the *MergeSeurat* function. Clusters characterised by differentially expressed immune gene markers were extracted from the NLT dataset for subsequent comparative analysis with liver-derived immune populations. Human cord blood and adult bone marrow datasets were downloaded from Human Cell Atlas data portal (https://preview.data.humancellatlas.org/). These were processed using the same approach as described above, followed by manual annotation. Decidua and placental data from Vento-Tormo *et al.*[16], were downloaded from ArrayExpress record E-MTAB-6701 (https://www.ebi.ac.uk/arrayexpress/experiments/E-MTAB-6701/).

Data integration

We used *Harmony* data integration[58] to correct for batch effect between sample identities. The average kBET rejection rate statistically significantly improved from 0.735 to 0.471 (Supplementary Table 13) following *Harmony* data integration (*p*-value 3.83e-3 in Kolmogorov–Smirnov test and *p*-value 8.8e-6 in Wilcoxon signed-rank test). The manifold was subjected to re-clustering using *Harmony* adjusted PCs with parameters as mentioned above in "Clustering and annotation". Cell type classifications were then ascertained through re-annotation of the clusters derived from *Harmony* adjusted PCs to produce the final annotation.

Changes in cell proportions over development

Comparison of cell proportions across gestational stages was assessed by modelling cell number data with negative binomial regression based on Poisson-gamma mixture distribution. Cell numbers were corrected for $CD45^-/CD45^+$ FACS sorted ratio (Supplementary Table 1) prior to applying negative binomial regression modelling. Modelling was achieved using the *glm.nb* function in the R *MASS* package. Modelled cell number data were studied for regression coefficient significance (variable coefficient *p*-value ≤ 0.05) to the response variable of gestational age with the corresponding *z*-score and *p*-values taken (Supplementary Table 10)

Dimensionality reduction and trajectory analysis

Dimensionality reduction methods included tSNE (*Seurat*, computed from the first 20 PCs, Barnes-Hut fast computation), UMAP (Python *UMAP* package, 5 nearest neighbours, correlation metric, minimum distance 0.3, computed from the first 20 PCs), FDG (*ForceAtlas2* class from *fa2* Python Package, Barnes-Hut implementation for faster computation with theta 0.8, 2000 iterations) and partition-based approximate graph abstraction (PAGA) (*paga* in *scanpy* Python package version 1.2.2). Development trajectories were inferred by comparing FDG, PAGA and diffusion map plots. Inferred trajectory analysis included computing diffusion map (*scanpy tl.diffmap* with 20 components), pseudotime (*scanpy tl.dpt* setting the earliest known cell type as root) and variable genes across pseudotime. Order of cells in pseudotime was statistically significant using Kruskal-Wallis test ($p < 1x10^{-7}$).

Comparisons of trajectories across stage were performed by subsetting liver dataset by stage using *SubsetData* function, computing dimensional reduction coordinates, batch correcting by sample using Harmony, and plotting PAGA and FDG by stage. Cell type comparisons across tissue involved subsetting for cell types of interest using *SubsetData* function, merging cross-tissue datasets using *MergeSeurat* function, and processing data using the same approach as for the liver and NLT datasets. *Harmony* batch correction was then performed by tissue type, with results presented as combined UMAPs, FDGs and PAGA score heatmaps.

Dynamically expressed genes across pseudotime

Genes that vary across pseudotime were calculated using *DifferentialGeneTest* function in *Monocle* in R (version 2.6.4) and a cut-off of adjusted *p*-value < 0.001 applied. This was applied on the entire pseudotime range and also on the pseudotime intervals specific to each cell type in order to avoid limitation to the genes characterised by monotonic changes across the inferred trajectory. Expression of pseudotime variable genes were min-max normalised prior to visualization and annotated based on each gene's involvement in relevant cell-specific functional modules or hallmark functional pathways from MSigDB v6.2, a curated molecular signature database[59]. Peak expression for each gene over pseudotime was calculated and grouped into 'Early', 'Mid' or 'Late' categories. For visualisation purposes, the resulting gene lists were minimised by ordering them from those present in the most selected functional pathways to least, as well as ensuring coverage across pseudotime. These genes were manually compared against current literature to determine if they have known functional or cell type associations. The top 20-25 genes in each list were displayed using the *ggplot2* package. Transcription factors were marked within the dataset based on AnimalTFDB transcription factor prediction database[60]. The full pseudotime gene list is available in the interactive files accompanying diffusion maps.

Visualisation by animated force-directed graph representation

The FDG animation was created using an in-house modified version of the *ForceAtlas2* class in *fa2* Python package by saving all the intermediate states (published version only outputs the final state and discards all intermediates). The FDG coordinates at each iteration were plotted and the resulting graphs were assembled in a mp4 video format using *VideoWriter* in *cv2* (version 3.3.1) Python package.

Differential gene extraction and validation

Differential gene validation was done using a random forest classifier (*RandomForestClassifier* class in *ensemble* module of *sklearn* Python package v0.19.1, with 500 estimators, *min_sample_split of 5, class weights set to the "balanced" policy and all other parameters set to default*). The Random Forest algorithm was chosen as it resembled the FACS gating hierarchy. 70% of the data was used for training and 30% for test. Parameter tuning was performed on training data using grid search. To determine whether tissue-related transcriptome variations were present in equivalent immune populations between liver, skin and kidney, each equivalent population was taken in turn and grouped according to its tissue of origin. *Seurat FindMarkers* function was then applied in a pair-wise manner between each tissue subset to produce a cell type-specific list of genes marking each tissue subset. These were investigated in turn for biological relevance, with representative genes displayed using *VlnPlot* function of *Seurat*.

DEGs from B cell pseudotime were studied for significant expression change across stage and differentiation state using a one-way ANOVA with Tukey's multiple comparison test. DEGs displaying significant variance in ln-normalised expression were further studied for correlation to DEGs identified within all other cell-types across stage. B cell pseudotime DEGs with significantly correlated trends of expression to DEGs within other cell-types across stage (p-value <0.05, Two-tailed Pearson's R at 95% CI) were plotted in Prism (v8.1.2, GraphPad Software). All graphs presented in the manuscript were plotted using *ggplot2* R package, *Seurat* implementation of *ggplot2, matplotlib* Python package, Prism (v.8.1.2, GraphPad Software) or FlowJo (v10.4.1). Spot plots are shown throughout the manuscript, displaying scaled expression of ln-normalised counts.

Primary immunodeficiency (PID) gene list curation

Disease and genetic deficiency information was extracted from Picard *et al.*[8] and manually annotated to include HGNC symbol names for each disease-associated genetic defect for subsequent correlation with the liver dataset. Diseases implicated in PID were divided according to the International Union of Immunological Societies (IUIS) major categories and screened across the liver scRNA-seq dataset. 315 unique genes were identified in the dataset from the 354 inborn errors of immunity highlighted in the article. For each disease category a dot plot was generated using Seurat *DotPlot* function and ordered by highest expression

across each gene and across each cell type, highlighting those cell types in each disease category which express the highest number of genes associated with a genetic defect.

CellPhoneDB analysis

CellPhoneDB v2.0 (www.cellphonedb.org)[61] was used for the receptor-ligand analysis in Figure 3d. Significant ($p < 0.05$) receptor-ligand interactions between $VCAM1^+$ Erythroblastic Island macrophages and the two erythroid (early and mid) populations were displayed.

Whole genome sequencing and fetal cell identification

To identify maternal cells present in our data we combined the information from fetal whole genome DNA sequencing with the single cell RNA-seq data. For each sample we measured the allele frequency in the fetal DNA of SNPs from the 1000 genomes project[62] falling within exons with a population allele frequency in excess of 1%. We then consider only those SNPs which are homozygous in the fetal DNA for follow up in the scRNA-seq data. A SNP was considered to be homozygous if its allele frequency in the fetal DNA was less than 0.2 or greater than 0.8 and had an FDR adjusted *p*-value of less than 0.01 under a binomial test for the null hypothesis that the allele frequency in the DNA was in the range [0.3,0.7].

The allele frequency of each of these SNPs with population allele frequency > 1% that are known to be homozygous in the fetal DNA was then measured in each cell in the scRNA-seq data. Any deviations from homozygosity in the RNA-seq data must be a consequence of either sequencing errors, RNA editing, or the genotype of the cell differing from the fetal DNA. For each cell, we calculated the total fraction of reads at the SNPs (selected as described above) that differ from the fetal genotype. We then assume that the genome-wide rate of deviations due to sequencing errors and RNA editing is less than or equal to 2%. For maternal cells, the expected genome wide rate of deviation at these SNPs is equal to half the mean of the population allele frequency at the interrogated SNPs. Finally, for each cell we calculated the posterior probability of the cell being fetal or maternal assuming a binomial distribution with rate 2% for a fetal cell and half the mean of the population allele frequency for the maternal cell and assign a cell as: maternal/fetal if either posterior probability exceeds 99%, ambiguous otherwise. We validated this method using samples for which both the fetal and maternal DNA were available.

'Hyperion' Imaging mass cytometry (IMC)

Antibodies were conjugated to metals using the Fluidigm MaxPar conjugation kits and the associated method with following modifications; the lanthanides were used at 1.5 mM and washed for a shorter duration (4x 5 minutes) in W buffer in prior to elution. Ultrapure MilliQ water was used throughout for any dilutions and washes. 4 µm Formalin-fixed paraffin-embedded sections obtained from 8 and 15 PCW fetal liver tissue blocks were incubated at 60 °C for 1 hour then dewaxed in Xylene (Fisher). After rehydration through graded alcohols (Fisher) and a 5 minute wash in water, the sections were subjected to Heat-Induced Epitope Retrieval with Citrate buffer (pH- 6.0). Sections were then washed in water and PBS (Gibco) and blocked with 3% BSA (Sigma-Aldrich) for 45 minutes. A mixture of 8 metal-conjugated antibodies diluted in 0.5% BSA (see Supplementary Table 21 for antibody details, was added to the sections for overnight incubation at 4 °C in a humidified chamber. Slides were washed twice in 0.2% Triton X-100 diluted in PBS for 8 minutes and then twice in PBS for 8 minutes.

To counterstain nucleated cells, sections were incubated with 312.5nm (193 Ir) Intercalator-Ir (Fluidigm) for 30 minutes at room temperature. Slides were then washed in water for 5 minutes, and allowed to air-dry at room temperature prior to imaging on the Hyperion imaging mass cytometer. Using expected target cell frequencies from previous fluorescence flow cytometry data, Region of Interest (ROI) size was set to 2.8mm by 3.8mm. The ablation energy was set at 2 db with a laser frequency of 200Hz. Each session of ablation generated a .MCD image file containing information for every panorama and ROI measured whereby each 1µm piece of tissue liberated by the laser was analysed for ionic content on a per channel basis by Time of Flight. Single cell segmentation and feature extraction was performed using CellProfiler (v3.1.5). Nuclei were identified using the "IdentifyPrimaryObjects" module where the input images were the sum of the DNA stained Iridium channels (191 and 193) constructed by the "ImageAfterMath" module. The diameter range set for Nuclei identification was 4-15 pixel units. The "ExpandOrShrink" module was used to grow the nuclear segmentation area by 3 pixels to define the cellular area and the "MeasureObjectIntensity" module was used to determine the mean intensity for each cell object identified.

Light sheet fluorescence microscopy

Male embryos at 5, 7 and 11 PCW deemed devoid of morphological anomalies were dissected after overnight fixation in 4% PFA. Whole-mount and cryosection immunostaining were performed as described in Belle *et al.*[63], with the following conditions: tissue was incubated with primary antibodies (see Supplementary Table 22 for antibody details) for 9 days at 37°C, with secondary antibody for 16 hours at 37°C using dedicated host species antibodies and reagent combination. TO-PRO-3 647 was used at 1:100 in whole embryos and 1:5000 on cryosections. Whole-mount specimens were solvent-cleared as described[63], and imaged in dibenzylether with a Miltenyi Lavision Biotech ultramicroscope (Olympus MXV10 stereomicroscope and PCO Edge SCMOS CCD camera using the dedicated Imspector pro acquisition software. Four lasers (at 488, 561, 647 and 790nm wavelengths) were used to generate light sheets. IMaris (v9.2, BitPlane) was used for image conversion and processing. Photoshop (Adobe) was used to create panels. All raw files are being made available on demand through our dedicated KeenEye Technologies-hosted Platform (www.transparent-human-embryo.com, <<request database access>>).

Statistics and reproducibility

For all analyses of fetal liver 3' 10x data, n = 14 biologically independent samples were included. This includes Figure 1b, Figure 2a, Figure 3a-c, Figure 4b-c, Figure 5a-b, Figure 6a-b, Figure 6g, Extended Data 1c, Extended Data 2a, Extended Data 3a-e, Extended Data 4a, Extended Data 5e, Extended Data 5g-h, Extended Data 6a-e, Extended Data 7b and h, and Extended Data 8.

For all analyses of fetal liver 3' 10x data by developmental stage, n = 4 7-8 PCW, n = 4 9-11 PCW, n = 3 12-14 PCW, and n = 3 15-17 PCW biologically independent samples were used. This includes Figure 1c, Figure 4a, Extended data 1d-e, Extended Data 5a-c, and Extended Data 5f.

For analyses including 10x sequencing data of skin, kidney, yolk sac, decidua and placenta, n = 7, 3, 3, 11 and 5 biologically independent samples were used, respectively. These analyses are shown in Figure 3b-c, Figure 4b-c, Figure 5a-b, Figure 6g, Extended Data 1a-b, Extended Data 3c-e, Extended Data 5g-h, and Extended Data 6c-e.

For analysis including sequencing data of cord blood and adult bone marrow, n = 8 biologically independent samples of each were used. These analyses are shown in Figure 6g and Extended Data 7h.

For all scRNA-seq data shown, all cells of a given label from indicated tissues are shown, no down-sampling or sub-setting was performed. The following cells numbers generated using scRNA-seq are displayed in each of the listed figures:

Figure 1b-c	113,063 fetal liver cells
Figure 2a	113,063 fetal liver cells
Figure 3a	104,515 fetal liver cells
Figure 3b	43,507 fetal liver cells, 2,455 yolk sac cells, and 243 skin cells
Figure 3c	52,327 fetal liver cells, 362 skin cells, 28 kidney cells, and 2,793 yolk sac cells were analysed, and a maximum of 20 cells displayed of each cell type per tissue
Figure 4a	16,919 fetal liver cells
Figure 4b	16,919 fetal liver cells, 2,757 skin cells, 213 kidney cells, and 259 yolk sac cells
Figure 4c	6,706 NK cells and 1,726 ILC precursors from fetal liver, 1,479 NK cells and 1,142 ILC precursors from skin, and 155 NK cells and 36 ILC precursors from kidney
Figure 5a-b	38,464 fetal liver cells, 6,887 yolk sac cells, and 10,008 cells from decidua and placenta
Figure 6a-b	5,673 fetal liver cells
Figure 6g	3,439 fetal liver HSC/MPP, 205 yolk sac progenitors, 1,082 cord blood HSCs, and 3,668 adult bone marrow HSCs
Ext Data 1a	10,258 skin cells, and 17, 95 kidney cells
Ext Data 1b	10,071 yolk sac cells
Ext Data 1c-e	113,063 fetal liver cells
Ext Data 1f	1,206 fetal liver cells from n = 2 biologically independent samples profiled using Smart-seq2 (also displayed in Extended Data 2b)
Ext Data 2a	113,063 fetal liver cells
Ext Data 2b	1,206 fetal liver cells from n = 2 biologically independent samples profiled using Smart-seq2; and 381 fetal liver erythroid liver erythroblastic island (EI) populations (early, mid and late erythroids, VCAM1$^+$ EI macrophages), Kupffer cells and endothelium validated by SS2 from n = 2 biologically independent fetal liver samples

1175	Ext Data 2d	381 fetal liver erythroid liver erythroblastic island (EI) populations (early, mid
1176		and late erythroids, VCAM1^{+} EI macrophages), Kupffer cells and endothelium
1177		validated by SS2 from n = 2 biologically independent fetal liver samples
1178	Ext Data 3a-b	52,237 fetal liver cells
1179	Ext Data 3c	52,237 fetal liver cells, 362 skin cells, and 28 kidney cells
1180	Ext Data 3d	52,237 fetal liver cells, 362 skin cells, 28 kidney cells, and 2,588 yolk sac cells
1181	Ext Data 3e	3,439 HSC/MPP, 1,342 MEMP, 11,985 Early Erythroid, 27,000 Mid
1182		Erythroid, 3,180 Late Erythroid, 3,983 Megakaryocytes, and 1,308 Mast cells
1183		from fetal liver; 55 MEMP, 51 Mid Erythroid, 137 Late Erythroid, 11
1184		Megakaryocytes, and 108 Mast cells from skin; and 2 MEMP and 26
1185		Megakaryocytes from kidney.
1186	Ext Data 5a	16,919 fetal liver cells
1187	Ext Data 5b-c	767 fetal liver Early lymphoid/T lymphocytes
1188	Ext Data 5d	16,666 fetal liver cells from n = 7 biologically independent samples.
1189	Ext Data 5e	7,467 fetal liver cells
1190	Ext Data 5f	32,308 fetal liver cells
1191	Ext Data 5g	16,919 fetal liver cells, 2,775 skin cells, 213 kidney cells, and 464 yolk sac
1192		cells
1193	Ext Data 5h	1,726 fetal liver ILC precursors, 1,142 skin ILC precursors, and 36 kidney
1194		ILC precursors
1195	Ext Data 6a	6,606 fetal liver cells
1196	Ext Data 6b	11,653 fetal liver cells
1197	Ext Data 6c	38,646 fetal liver cells, 6,887 yolk sac cells, and 10,008 cells from decidua
1198		and placenta were analysed, and a maximum of 50 cells displayed of each cell
1199		type per tissue
1200	Ext Data 6d	43,498 fetal liver cells, 8,350 skin cells, and 1,514 kidney cells
1201	Ext Data 6e	24,841 Kupffer cells, 2,586 monocytes, 253 pDC precursors, 336 DC1, and
1202		3,954 DC2 from fetal liver; 5,474 macrophages, 704 monocytes, 36 pDCs, 99
1203		DC1, and 527 DC2 from skin; and 1,075 macrophages, 82 monocytes, 1 pDC,
1204		1 DC2, and 271 DC2 from kidney
1205	Ext Data 7b	6,606 fetal liver cells and 161 of 349 scRNA-seq profiled cells
1206		(Supplementary Table 12) from n = 3 biologically independent samples,
1207		sorted as per FACS gates in Extended Data 7a

Ext Data 7h	3,439 HSC/MPPs, 1,341 MEMPs, 234 Pre pro B cells, 658 Neutrophil-myeloid progenitors, 350 Monocyte precursors, 253 pDC precursors, and 330 DC precursors from fetal liver; 205 yolk sac progenitors; 1,082 cord blood HSCs; and 3,668 adult bone marrow HSCs
Ext Data 8	113,063 fetal liver cells

Representative mini bulk RNA-seq data of 100 cells per cell state from 1 of $n = 3$ biologically independent fetal liver samples is shown in Extended Data 2b-c.

Cytospin images shown in Figure 2b and Extended Data 5e are representative from 1 of $n = 3$ biologically independent samples.

Hyperion images shown in Figure 2c are representative from 1 of $n = 4$ biologically independent 8 PCW fetal livers and 1 of $n = 4$ biologically independent 15 PCW fetal livers.

Immunohistochemical (IHC) staining of 8 PCW fetal skin in Figure 3d is representative from 1 of $n = 3$ biologically independent samples. IHC staining of fetal liver in Extended Data 4-b are from independent samples and both are representative from 1 of $n = 3$ biologically independent samples.

Light sheet fluorescence microscopy of embryo (5PCW) hand skin shown in Figure 3e is a representative image from 1 of $n = 3$ biologically independent samples.

Statistical analysis of differential gene expression was done using one-tailed Wilcoxon rank sum test with Bonferroni correction, including those shown in heatmaps and violin plots.
Statistically significant gene expression shown in Figure 4c comparing expression in NK cells and ILC precursors across tissues was $p < 0.001$ where **** was shown, and $p = 0.00236$ for *TXNIP* expression in ILC precursors in liver compared to those in kidney.
In Figure 6b, expression of genes in HSC/MPP 1 compared to each other cell cluster, and MEMP/neutrophil-myeloid progenitor/pre pro B cell compared to each other cell cluster was statistically significant unless specified as not significant (ns). All $p < 0.001$, except expression of *KLF1* between HSC/MPP1 vs HSC/MPP2 ($p = 0.00934$), *IL1RL1* between HSC/MPP 1 vs HSC/MPP 2 ($p = 0.00148$), *GATA2* between MEMP vs HSC/MPP2 ($p =$

0.00802), *BCL11A* between HSC/MPP 1 vs HSC/MPP 7 (p = 0.00114), *LYZ* between HSC/MPP 1 vs HSC/MPP 5 (p = 0.00675), *AZU1* between HSC/MPP 1 vs HSC/MPP 8 (p = 0.00494), *CSF1R* between Neutrophil-myeloid progenitors vs HSC/MPP 8 (p = 0.213). In Figure 6g, **** indicates $p < 0.0001$ when comparing expression of *ANXA1*, *DUSP1*, *HLA-B*, and *HSPAIA* between samples. Expression of genes in fetal liver Early lymphoid/T lymphocytes was compared across developmental stages and displayed in Extended Data 5b-c. No significant difference across stage was observed in the genes displayed in Extended 5b. The following comparisons shown in Extended Data 5c were significant: *CD2* expression at 7-8 PCW vs 9-17 PCW ($p < 0.0001$) and vs 12-17 PCW ($p < 0.0001$); *TRDC* expression at 7-8 PCW vs 9-17 PCW ($p < 0.0001$) and 9-11 PCW vs 12-17 PCW ($p < 0.0001$); *CD8A* expression at 7-8 PCW vs 9-17 PCW (p = 0.00714); *CD27* expression at 7-8 PCW vs 9-17 PCW ($p < 0.0001$) and 9-11 PCW vs 12-17 PCW ($p < 0.0001$); *IL7R* expression at 7-8 PCW vs 9-17 PCW ($p < 0.0001$), 9-11 PCW vs 12-17 PCW (p = 0.00168), and 12-14 PCW vs 15-17 PCW (p = 0.00216); *JCHAIN* expression at 7-8 PCW vs 9-17 PCW ($p < 0.0001$); *CD3D* expression at 7-8 PCW vs 9-17 PCW ($p < 0.0001$); *KLRB1* expression at 9-11 PCW vs 12-17 PCW ($p < 0.0001$); *TRAC* expression at 7-8 PCW vs 9-17 PCW ($p < 0.0001$) and 9-11 PCW vs 12-17 PCW ($p < 0.0001$); and *PRF1* expression at 9-11 PCW vs 12-17 PCW ($p < 0.0001$). Expression of genes in ILC precursors in fetal liver, skin and kidney, as shown in Extended Data 5h, was compared between tissues and no significant difference was observed. Extended Data 6e shows comparisons between fetal liver, skin and kidney for macrophages, monocytes, pDCs, DC1, and DC2. The following comparisons were statistically significant ($p < 0.0001$) between macrophages from different tissues: *CD14* expression in kidney vs liver and vs skin; *CD68* expression in liver vs skin and vs kidney; *CD163*, *RNASE1* and *F13A1* expression between all tissues; and, *VCAM1* expression in liver vs skin and vs kidney. The following comparisons were statistically significant between monocytes from different tissues: *CD14* expression in liver vs skin ($p < 0.0001$); *CD68* expression in liver vs skin ($p < 0.0001$); *S100A9* expression in liver vs kidney ($p < 0.0001$), and skin vs kidney (p = 0.0245); *FCGR3A.* expression in liver vs skin (p = 0.0004); *POSTN* expression in liver vs skin ($p < 0.0001$) and vs kidney ($p < 0.0001$), and skin vs kidney (p = 0.0411). No significant difference was observed when comparing genes in pDCs across tissues. *CLEC9A* expression was statistically significantly different in liver vs skin DC1 ($p < 0.0001$). The following comparisons were statistically significant between DC2 from different tissues: *CD1C* expression in skin vs liver ($p < 0.0001$) and vs kidney ($p < 0.0001$); *CLEC10A* expression in kidney vs liver ($p < 0.0001$). and vs skin ($p < 0.0001$); *S100B* expression in skin vs liver ($p <$

0.0001) and vs kidney ($p = 0.0162$); and *FCER1A* and *CD83* expression between each tissue ($p < 0.0001$).

Statistical analysis of HSC colony assays shown in Figure 6c-e, and Extended Data 7 was done by applying two-tailed Fisher's exact tests to colony counts. Figure 6c and d, and Extended Data 7f shows 125, 217 and 124 colonies from 7-8 PCW, 12-14 PCW and 15-17 PCW fetal liver samples, respectively from $n = 2$ biologically independent samples per development stage. The number of colonies per sample is 93, 32, 93, 124, 84, 40, which are shown in Extended Data 7c. The number of colonies between the following stages in Figure 6c were statistically significant: Erythroid colonies in 7-8 PCW vs. 15-17 PCW ($p = 0.0238$), Erythroid/Megakaryocyte/Myeloid colonies in 7-8 PCW vs. 15-17 PCW ($p = 0.0294$), NK colonies in 7-8 PCW vs. 15-17 PCW ($p = 0.0357$), and Erythroid/Myeloid colonies in 7-8 PCW vs 12-14 PCW ($p = 0.0188$) and 15-17 PCW ($p < 0.001$), and 12-14 PCW vs 15-17 PCW ($p = 0.0232$). The number of Erythroid-containing colonies was significant between each stage shown in Figure 6d ($p < 0.001$). The number of colonies that differentiated along 3 lineages was significant between 7-8 PCW vs 12-14 PCW ($p = 0.0041$), and 7-8 PCW vs 15-17 PCW ($p = 0.0027$). Figure 6e shows 141, 74 and 124 colonies from 7-8 PCW, 12-14 PCW and 15-17 PCW fetal liver samples, respectively from $n = 2$ biologically independent samples per development stage. The number of B cell-forming colonies in Figure 6e was significant between 7-8 PCW vs 12-14 PCW ($p = 0.0014$) and 15-17 PCW ($p = 0.0044$). Extended Data 7d-e shows 163, 196, 182 colonies from $n = 3$ 7-8 PCW, $n = 2$ 12-14 PCW, and $n = 2$ 15-17 PCW biologically independent fetal liver samples, respectively on the left, and 42, 74, 47, 97, 99, 59, 123 colonies by individual sample on the right. The number of Erythroid colonies compared to all other types shown in Extended Data 7d was statistically significant between 12-14 PCW and 15-17 PCW ($p = 0.0307$). The number of Erythroid-containing colonies was significant between 7-8 PCW vs 15-17 PCW ($p = 0.0013$), and 12-14 PCW vs 15-17 PCW ($p = 0.0497$), as shown in Extended Data 7e. Extended Data 7f shows 125, 217 and 124 colonies from 7-8 PCW, 12-14 PCW and 15-17 PCW fetal liver samples, respectively from $n = 2$ biologically independent samples per development stage. Extended Data 7g shows 141, 74 and 124 colonies from 7-8 PCW, 12-14 PCW and 15-17 PCW fetal liver samples, respectively from $n = 2$ biologically independent samples per development stage. The percentage of NK-containing colonies was statistically significant in 7-8 PCW vs 15-17PCW ($p = 0.0032$), and 12-4 PCW vs 15-17 PCW ($p = 0.0115$).

Flow cytometric analysis of cell cycle phases, as shown in Figure 6g and Extended Data 7i was performed on cells from n = 3 7-8 PCW and n = 3 12-16 PCW biologically independent fetal liver samples, and n = 2 biologically independent cord blood samples. One-way ANOVA with Tukey's multiple comparison test was used to determine statistical significance between stages (7-8 PCW and 12-14 PCW) and samples (fetal liver and cord blood). The percent of $CD34^{+}CD38^{-}$ cells in G_0 was significantly higher in 12-14 PCW livers compared to 7-8 PCW livers ($p = 0.0136$).

Cell cycle phases determined by transcriptome analysis of fetal liver cells, fetal skin cells, and fetal kidney cells is shown in Extended Data 3e. Statistical significance of the proportion of MEMP and Megakaryocytes in each cell cycle phase was compared between fetal liver, skin and kidney using Kruskal-Wallis with Dunn's post hoc test. Statistical significance of the proportion of Mid and Late Erythroids, and Mast cells in each cell cycle phase was compared between fetal liver and skin using two-tailed Mann-Whitney tests. The following comparisons were statistically significant: Megakaycoytes in fetal liver vs fetal kidney in G_1 ($p = 0.0317$), G_2M ($p = 0.0317$) and S ($p = 0.0139$); Megakaryocytes in fetal liver vs fetal skin in S ($p = 0.0039$); Mid Erythroids in fetal liver vs skin in G_1 ($p = 0.0031$) and G_2M ($p < 0.0001$); Late Late Erythroids in fetal liver vs fetal skin in G_1 ($p = 0.0021$) and S ($p < 0.0001$); and, Mast cells in fetal liver vs fetal skin in G_1 ($p = 0.0248$) and S ($p = 0.0337$).

Statistical comparison of the percent of MEMP, Mid and Late Erythroids, Megaryocytes and Mast cells expressing *MKI67* in fetal liver vs NLT (skin and kidney) was performed using two-tailed Mann-Whitney tests. This is displayed in Extended Data 3e, where the following comparisons were statistically significant: Megakaryocytes in fetal liver vs fetal NLT ($p = 0.007$), Mid Erythroid in fetal liver vs fetal skin ($p = 0.0305$), and Late Erythroid in fetal liver vs fetal skin ($p = 0.0368$).

Statistical comparison of the percent of fetal liver HSC/MPP expressing *MKI67* compared to the percent of fetal liver MEMP, Pre pro B cells, Neutrophil-myeloid progenitors, Monocyte precursors, pDC precursors and DC precursors, Yolk Sac progenitors, and HSC from cord blood and adult bone marrow expressing *MKI67* was performed using Kruskal-Wallis with Dunn's post hoc test. This is displayed in Extended Data 7h, where the percent of *MKI67*-expressing fetal liver HSC/MPPs was statistically significant when compared to the following populations: fetal liver MEMP ($p = 0.0180$), Monocyte precursors ($p < 0.0001$), DC

precursors ($p < 0.0001$), cord blood HSC ($p < 0.0001$), and adult bone marrow HSC ($p = 0.0076$)

Extended Data 1g displays flow cytometric analysis of the frequency of B cells in the CD34- cells from $n = 13$ 6-9 PCW, $n = 13$ 9-12 PCW, $n = 14$ 12-15 PCW and $n = 9$ 15-19 PCW biologically independent fetal liver samples. Statistical significance across the developmental stages was compared using Kruskal-Wallis with Dunn's multiple comparison post-test, for which the following comparisons were statistically significant: 6-9 PCW vs 12-15 PCW ($p < 0.0001$), 6-9 PCW vs 15-19 PCW ($p = 0.0003$), 9-12 PCW vs 12-15 PCW ($p = 0.0157$), and 9-12 PCW vs 15-19 PCW ($p = 0.0287$).

Statistically significant, dynamically variable genes from pseudotime were determined by Likelihood of ratio test applied in monocle (see Methods). Select genes were displayed for Erythroid, Mast cell and Megakaryocyte lineages shown in Extended Data 3b, B cell lineage shown in Extended Data 5e, and DC1, DC2 and Monocyte lineage shown in Extended Data 6b.

Investigation of potential receptor:ligand interactions between all fetal liver VCAM1$^+$ EI macrophages (161 cells) and Early and Mid Erythroids (11,985 and 27,000 cells respectively), as shown in Extended Data 4a, was performed using CellPhoneDB (see Methods). A permutation test was applied to determine statistical significance, which is indicated by the colour of the dots. ln-normalised median expression of *ITGA4* and *VCAM1* in the same cells is also displayed in a violin plot (right panel).

ImageStream analysis of fetal liver cells shown in Extended Data 4c-f was performed on $n = 3$ biologically independent samples. Extended Data 4c-d display representative data from one sample. Extended Data 4e shows representative images of cells from 38,576 single cells and 1,945 doublets from one sample. The percent of each cell type that was observed within the doublet of singlet gate, as per Extended Data 4c, was compared using Kruskal-Wallis with Dunn's post hoc test. The percent of doublets in VCAM1$^+$ EI macrophages was significantly different the percent of doublets in Erythroids ($p = 0.0194$).

Statistical significance of *TNFSF13B* expression in Kupffer cells over time compared to *NFKBIA* in HSC/MPPs ($p = 0.0245$), Pre pro B cells ($p = 0.0008$), Pro B cells ($p = 0.0004$),

Pre B cells (p = 0.0197) and B cells (p = 0.0343) across 4 developmental stages spanning 6-17 PCW was determined using two-tailed Pearson's R test.

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Author contributions

M.H.; S.A.T. and E.L. conceived and directed the study. M.H.; S.A.T.; E.L. and R.A,B. designed the experiments. Samples were collected by S.L. and S.L., isolated by R.A.B. and libraries prepared by E.S.; L.M.; D.M.P.; R.V-T; J.P. and J.F. Flow cytometry and FACS experiments were performed by R.A.B.; E.F.C.; L.J.; D.M. and A.F. Imaging mass cytometry experiments were performed by M.A.; B.M.; B.I.; D.M. and A.F. Cytospins were performed by D.D.; J.F.; and *in vitro* culture differentiation experiments were performed by L.J.; D.M. and E.F.C. Immunohistochemistry was performed by B.I.; M.A.; F.G. and C.M.; P.C. and M.A. interpreted immunohistochemistry and developmental pathology sections. Y.G. and A.C. performed and interpreted light sheet fluorescence microscopy experiments. M.S.K.; B.L.; O.A.; M.T.; D.D.; T.L.T.; M.S.; O.R-R. and A.R. generated adult and cord blood scRNA-seq datasets. D.M.P.; K.G.; K.P.; S.W.; I.G.; M.E.; P.V.; M.Y.; Z.M. and J.B performed the computational analysis. M.H.; D.M.P.; R.A.B.; B.G.; E.L.; I.R.; A.R.; E.F.C; L.J.; A-C.V.; R.R.; E.P.; M.M.; J.P.; G.R.; K.B.M.; M.J.T.S.; A.F.; K.G.; S.W.; I.G.; S.B. and J.B. interpreted the data. M.H.; L.J.; R.A.B.; E.S.; D.M.P; B.G.; E.L.; I.R; K.R.; S.W.; I.G.; A-C.V. and A.R wrote the manuscript. All authors read and accepted the manuscript.

Competing Interests

None declared

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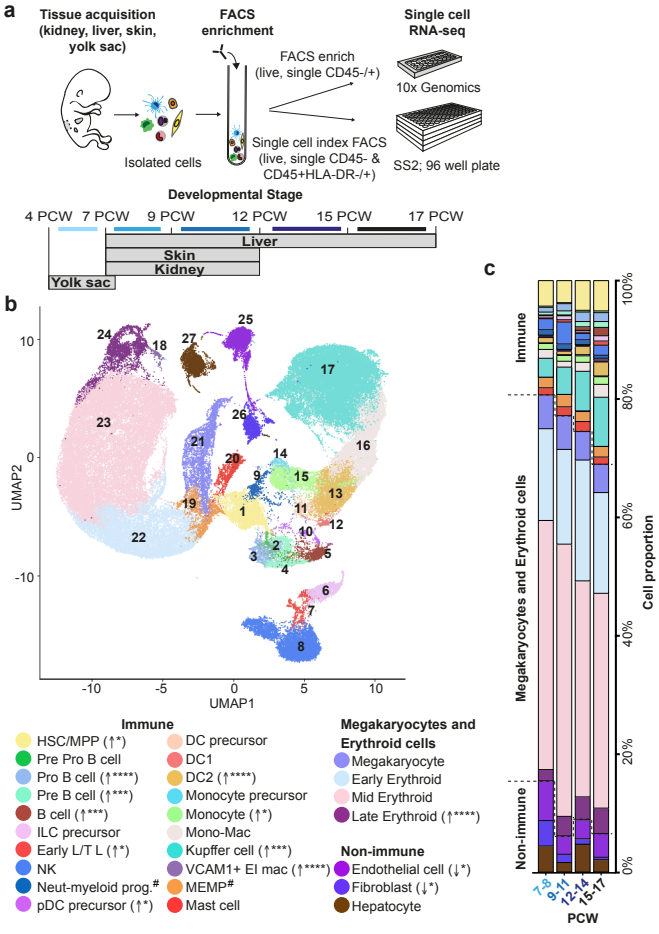
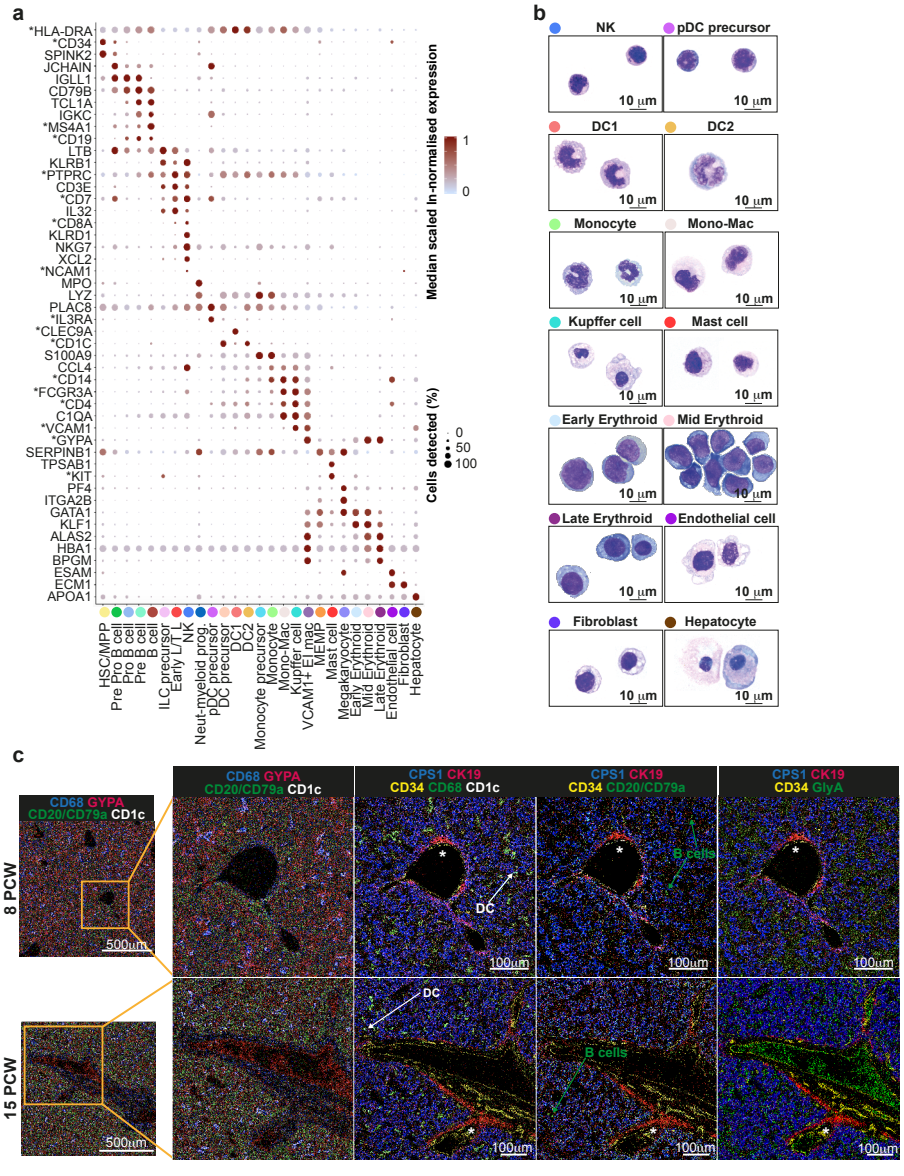
Data and materials availability

The raw sequencing data, expression count data with cell classifications are deposited at ArrayExpress: https://www.ebi.ac.uk/arrayexpress/experiments/E-MTAB-7407/

Code availability

All scripts are available at https://github.com/haniffalab/FCA_liver


a
Tissue acquisition (kidney, liver, skin, yolk sac)
FACS enrichment
Single cell RNA-seq
FACS enrich (live, single CD45-/+)
10x Genomics
Isolated cells
Single cell index FACS (live, single CD45- & CD45+HLA-DR-/+)
SS2; 96 well plate
Developmental Stage
4 PCW
7 PCW
9 PCW
12 PCW
15 PCW
17 PCW
Liver
Skin
Kidney
Yolk sac
b
UMAP2
UMAP1
Immune
HSC/MPP (↑*)
Pre Pro B cell
Pro B cell (↑****)
Pre B cell (↑***)
B cell (↑***)
ILC precursor
Early L/T L (↑*)
NK
Neut-myeloid prog.#
pDC precursor (↑*)
DC precursor
DC1
DC2 (↑****)
Monocyte precursor
Monocyte (↑*)
Mono-Mac
Kupffer cell (↑***)
VCAM1+ EI mac (↑****)
MEMP#
Mast cell
Megakaryocytes and Erythroid cells
Megakaryocyte
Early Erythroid
Mid Erythroid
Late Erythroid (↑****)
Non-immune
Endothelial cell (↓*)
Fibroblast (↓*)
Hepatocyte
c
Immune
Megakaryocytes and Erythroid cells
Non-immune
Cell proportion
PCW
7-8
9-11
12-14
15-17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LYZ
PLAC8
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S100A9
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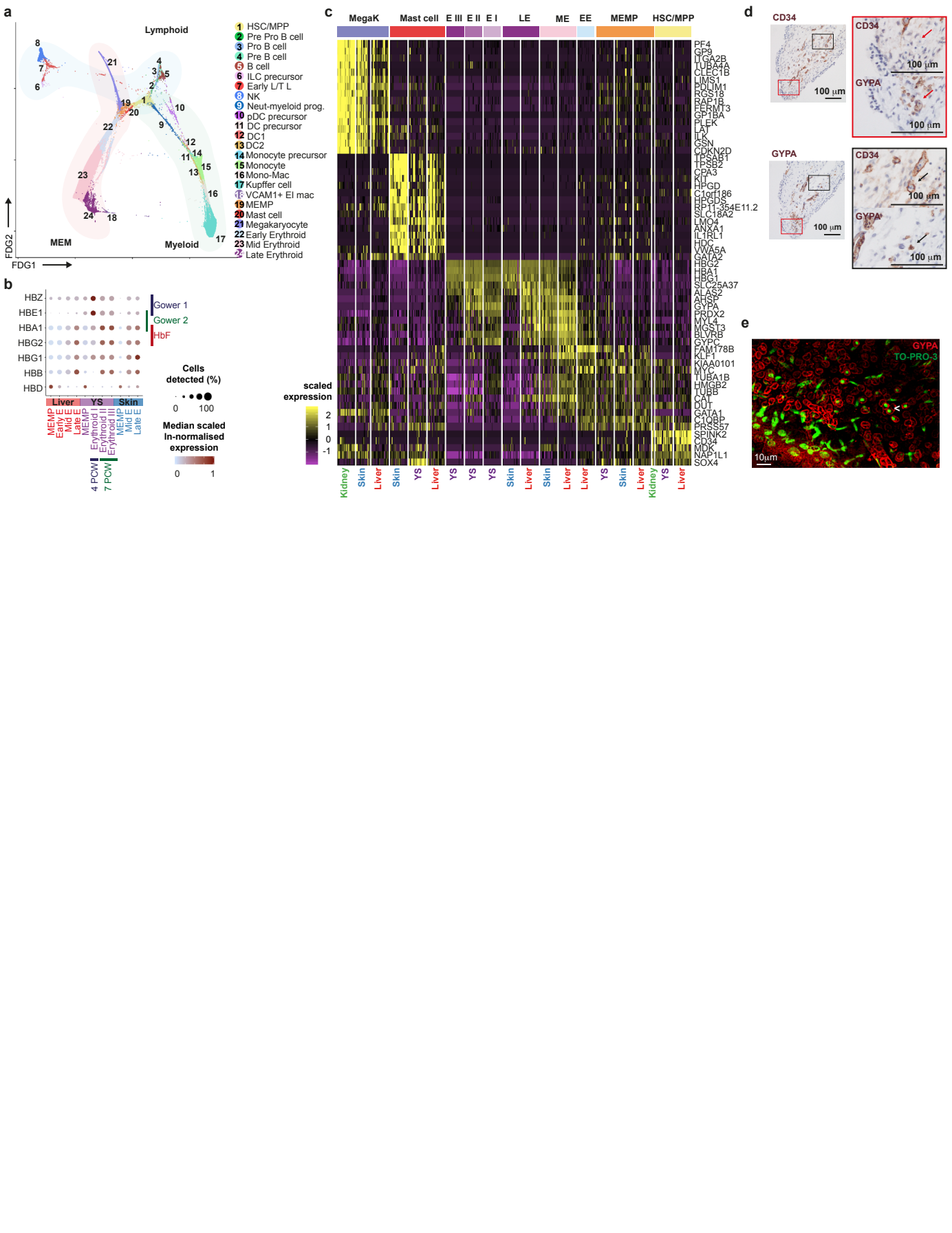
ALAS2
HBA1
BPGM
ESAM
ECM1
APOA1
HSC/MPP
Pre Pro B cell
Pro B cell
Pre B cell
ILC precursor
Early LMK
NK
Neut-myeloid prog
pDC precursor
DC precursor
DC1
DC2
Monocyte precursor
Monocyte
Mono-Mac
Kupffer cell
VCAM1+ EI mac
MEMP
Mast cell
Megakaryocyte
Early Erythroid
Mid Erythroid
Late Erythroid
Endothelial cell
Fibroblast
Hepatocyte
Median scaled ln-normalised expression
1
0
Cells detected (%)
0
50
100
b
NK
pDC precursor
DC1
DC2
Monocyte
Mono-Mac
Kupffer cell
Mast cell
Early Erythroid
Mid Erythroid
Late Erythroid
Endothelial cell
Fibroblast
Hepatocyte
10 μm

c
8 PCW
15 PCW
CD68 GYPA
CD20/CD79a CD1c
CPS1 CK19
CD34 CD68 CD1c
CD34 CD20/CD79a
CD34 GlyA
DC
B cells
500μm
100μm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Lymphoid
MEM
Myeloid
FDG1
FDG2
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2 Pre Pro B cell
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5 B cell
6 ILC precursor
7 Early L/T L
8 NK
9 Neut-myeloid prog.
10 pDC precursor
11 DC precursor
12 DC1
13 DC2
14 Monocyte precursor
15 Monocyte
16 Mono-Mac
17 Kupffer cell
18 VCAM1+ EI mac
19 MEMP
20 Megakaryocyte
21 Early Erythroid
22 Mid Erythroid
23 Late Erythroid
b
HBZ
HBE1
HBA1
HBG2
HBG1
HBB
HBD
Gower 1
Gower 2
HbF
Liver
YS
Skin
MEMP
Early E
Mid E
Late E
MEMP
Erythroid I
Erythroid II
Erythroid III
MEMP
Mid E
Late E
4 PCW
7 PCW
Cells detected (%)
0
100
Median scaled ln-normalised expression
0
1
c
MegaK
Mast cell
E III
E II
E I
LE
ME
EE
MEMP
HSC/MPP
scaled expression
2
1
0
-1
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ITGA2B
TUBA4A
CLEC1B
LIMS1
PDLIM1
RGS18
RAP1B
FERMT3
GP1BA
PLEK
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TPSB2
CPA3
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DUT
GATA1
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PRSS57
SPINK2
CD34
MDK
NAP1L1
SOX4
Kidney
Skin
Liver
Skin
YS
Liver
YS
YS
YS
Skin
Liver
Skin
Liver
Liver
YS
Skin
Liver
Kidney
YS
Liver
d
CD34
100 μm
GYPA
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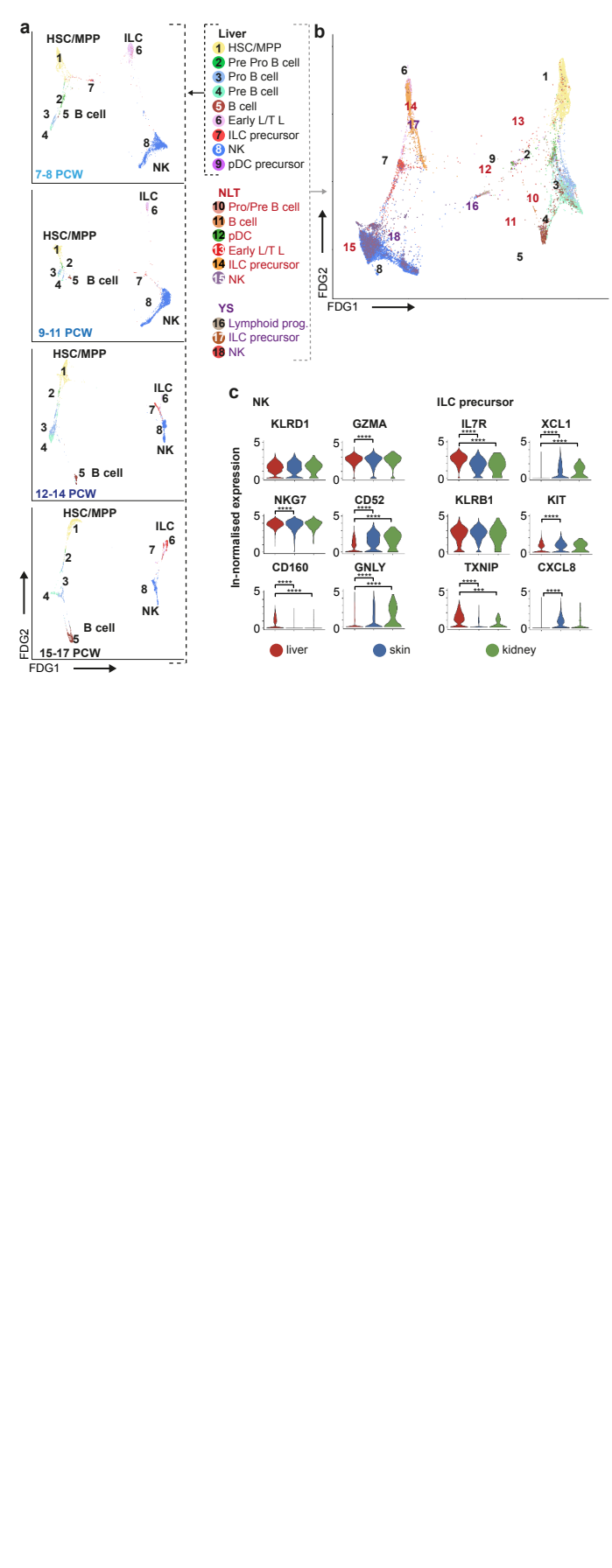
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HSC/MPP
ILC
B cell
NK
7-8 PCW
9-11 PCW
12-14 PCW
15-17 PCW
FDG1
FDG2
Liver
1 HSC/MPP
2 Pre Pro B cell
3 Pro B cell
4 Pre B cell
5 B cell
6 Early L/T L
7 ILC precursor
8 NK
9 pDC precursor
NLT
10 Pro/Pre B cell
11 B cell
12 pDC
13 Early L/T L
14 ILC precursor
15 NK
YS
16 Lymphoid prog.
17 ILC precursor
18 NK
b
c
NK
ILC precursor
KLRD1
GZMA
IL7R
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KIT
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GNLY
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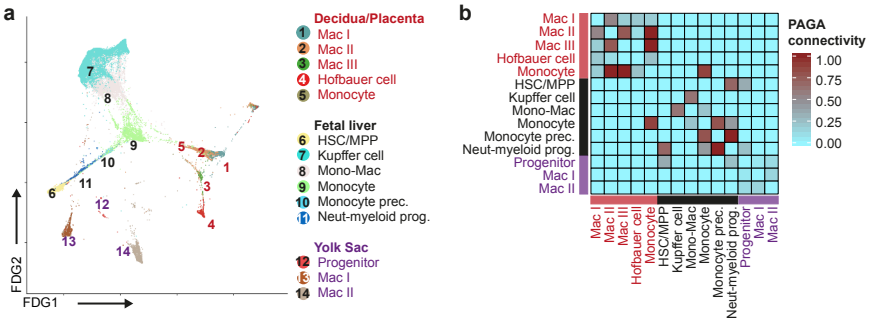
ln-normalised expression
liver
skin
kidney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FDG2
FDG1
Decidua/Placenta
1 Mac I
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3 Mac III
4 Hofbauer cell
5 Monocyte
Fetal liver
6 HSC/MPP
7 Kupffer cell
8 Mono-Mac
9 Monocyte
10 Monocyte prec.
11 Neut-myeloid prog.
Yolk Sac
12 Progenitor
13 Mac I
14 Mac II

b
Mac I
Mac II
Mac III
Hofbauer cell
Monocyte
HSC/MPP
Kupffer cell
Mono-Mac
Monocyte prec.
Neut-myeloid prog.
Progenitor
Mac I
Mac II
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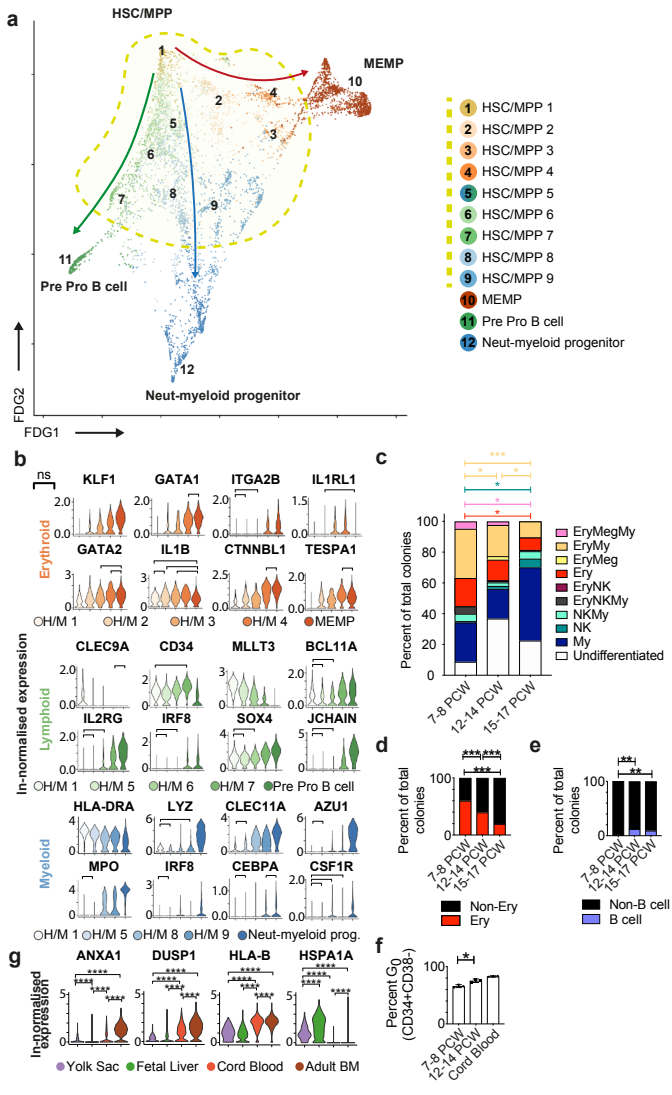
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HSC/MPP
MEMP
Pre Pro B cell
Neut-myeloid progenitor
FDG2
FDG1
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2 HSC/MPP 2
3 HSC/MPP 3
4 HSC/MPP 4
5 HSC/MPP 5
6 HSC/MPP 6
7 HSC/MPP 7
8 HSC/MPP 8
9 HSC/MPP 9
10 MEMP
11 Pre Pro B cell
12 Neut-myeloid progenitor
b
ns
KLF1
GATA1
ITGA2B
IL1RL1
GATA2
IL1B
CTNNBL1
TESPA1
Erythroid
H/M 1
H/M 2
H/M 3
H/M 4
MEMP
CLEC9A
CD34
MLLT3
BCL11A
IL2RG
IRF8
SOX4
JCHAIN
Lymphoid
H/M 5
H/M 6
H/M 7
Pre Pro B cell
HLA-DRA
LYZ
CLEC11A
AZU1
MPO
IRF8
CEBPA
CSF1R
Myeloid
H/M 8
H/M 9
Neut-myeloid prog.
ln-normalised expression
c
Percent of total colonies
7-8 PCW
12-14 PCW
15-17 PCW
EryMegMy
EryMy
EryMeg
Ery
EryNK
EryNKMy
NKMy
NK
My
Undifferentiated
d
Percent of total colonies
Non-Ery
Ery
e
Percent of total colonies
Non-B cell
B cell
f
Percent G0 (CD34+CD38-)
Cord Blood
g
ANXA1
DUSP1
HLA-B
HSPA1A
ln-normalised expression
Yolk Sac
Fetal Liver
Cord Blood
Adult BM