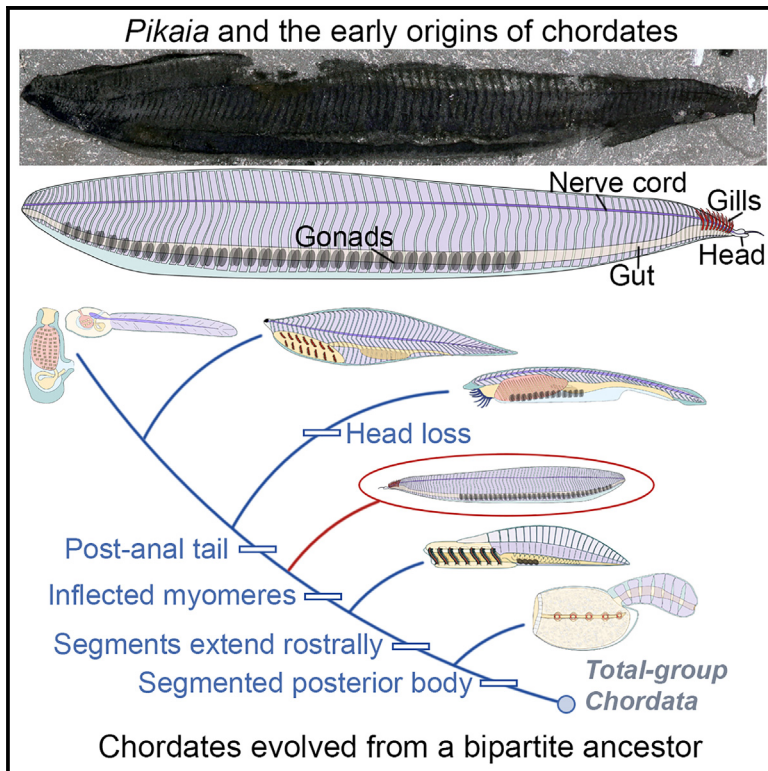


Current Biology

A new interpretation of *Pikaia* reveals the origins of the chordate body plan

Graphical abstract



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In brief

Mussini et al. identify a gut canal and a dorsal nerve chord in the Cambrian fossil *Pikaia* and resolve it as a stem-group chordate. Their morphological reinterpretation of *Pikaia* establishes phylogenetic links between vertebrates, amphioxus, and problematic Cambrian fossils with a bipartite body plan, unveiling a cryptic chapter in chordate history.

Highlights

- We describe a dorsal nerve cord and a gut canal in the Cambrian fossil *Pikaia*
- Its character combination identifies *Pikaia* as an unambiguous stem-group chordate
- *Pikaia*, *Yunnanozoon*, and vetulicolians record a chordate stem lineage
- Cambrian fossils support Romer's "somato-visceral" hypothesis of chordate origins

Report

A new interpretation of *Pikaia* reveals the origins of the chordate body plan

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SUMMARY

Our understanding of the evolutionary origin of Chordata, one of the most disparate and ecologically significant animal phyla, is hindered by a lack of unambiguous stem-group relatives. Problematic Cambrian fossils that have been considered as candidate chordates include vetulicolians,¹ *Yunnanzoon*,² and the iconic *Pikaia*.³ However, their phylogenetic placement has remained poorly constrained, impeding reconstructions of character evolution along the chordate stem lineage. Here we reinterpret the morphology of *Pikaia*, providing evidence for a gut canal and, crucially, a dorsal nerve cord—a robust chordate synapomorphy. The identification of these structures underpins a new anatomical model of *Pikaia* that shows that this fossil was previously interpreted upside down. We reveal a myomere configuration intermediate between amphioxus and vertebrates and establish morphological links between *Yunnanzoon*, *Pikaia*, and uncontroversial chordates. In this light, we perform a new phylogenetic analysis, using a revised, comprehensive deuterostome dataset, and establish a chordate stem lineage. We resolve vetulicolians as a paraphyletic group comprising the earliest diverging stem chordates, subtending a grade of more derived stem-group chordates comprising *Yunnanzoon* and *Pikaia*. Our phylogenetic results reveal the stepwise acquisition of characters diagnostic of the chordate crown group. In addition, they chart a phase in early chordate evolution defined by the gradual integration of the pharyngeal region with a segmented axial musculature, supporting classical evolutionary-developmental hypotheses of chordate origins⁴ and revealing a “lost chapter” in the history of the phylum.

RESULTS

Pikaia gracilens Walcott, 1911⁵ from the Burgess Shale of British Columbia (Wuliuan Stage, Miaolingian Series, c. 508 Ma) represents the first potential Cambrian chordate to be recognized.^{2,3,6} Nonetheless, the established model of *Pikaia*’s anatomy³ shows conspicuous discrepancies with living chordates^{3,7,8}: no clearly preserved dorsal nerve cord; no evident ventral digestive tract posterior to the foregut, despite the latter’s recurrent preservation among 114 described specimens^{3,7}; an internal, cuticularized, rod-like “dorsal organ”^{3,7,8}; a continuous “ventral blood vessel” unlike the branched main ventral vessels of amphioxus and vertebrates⁷; ventrally oriented “anterior appendages” lacking counterparts among chordate gills³; and myomere boundaries with apices pointing in the direction opposite those of amphioxus and vertebrates.^{3,7,8} Given these discrepancies and a lack of unambiguous chordate synapomorphies, several authors have suggested alternative placements among protostomes.^{9–11}

We reinterpret *Pikaia*’s morphology following the recognition of the ventral blood vessel³ as a nerve cord and of the dorsal

organ³ as a gut, justified below. This supports a dorsoventrally inverted *Pikaia* relative to the traditional interpretation, given that by comparison with all known bilaterians¹² the gut canal of *Pikaia* could not have run along the dorsal margin of the body.

Digestive tract

In their detailed study of *Pikaia*, Conway Morris and Caron³ identified a longitudinal tube-like structure running alongside one of the body edges, which they labeled a dorsal organ acting as an internalized support or storage structure.³ However, the dorsal organ has proved difficult to reconcile with the anatomy of any living or fossil groups.^{3,7,8}

Our reinterpretation of the dorsal organ as a digestive tract, speculatively suggested in Lacalli,⁸ is justified by the observation that the organ broadens and extends into the previously identified pharyngeal region³ (Figures 1E, 1F, 1J–1L, 2F, and 2J). Like the pharynx, parts of the organ are conspicuously three-dimensional³ (Figures 1E, 1F, 1H, and 1L) and characterized by calcium phosphate mineralization,³ revealed by energy-dispersive X-ray spectroscopy³ (Figure 3 in Conway Morris and Caron³). As with other

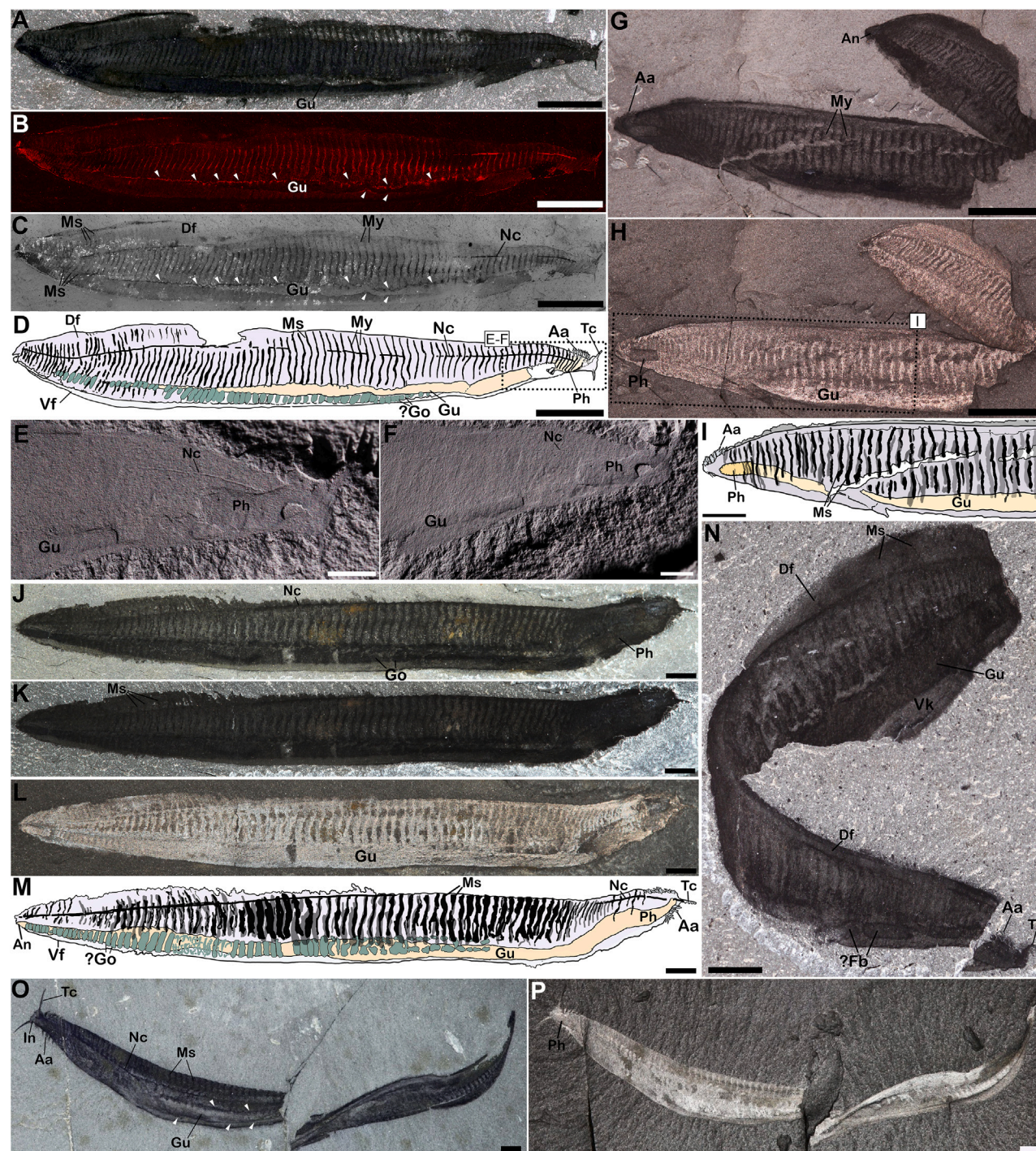


Figure 1. Anatomical reinterpretation of *Pikaia gracilens* Walcott, 1911: General body plan, axial musculature, and digestive tract

(A) USNM 198684, showing ventral gut canal, myosepta, and dorsal nerve cord. Note light-colored sediment infill of the gut canal, comparable to background matrix.

(B) Element map of (A) for carbon (C) obtained under energy-dispersive X-ray spectroscopy. Note dark, carbon-poor region of the gut canal, mapping onto the sediment infill in (A).

(C) View of (A) under backscattered scanning electron microscopy. Note light-colored sediment infill of the gut canal, comparable to background matrix.

(D) Interpretative drawing of (A)–(C).

(E and F) Photographs of regions corresponding to boxed area in (D) (part and counterpart), taken with specimen coated in ammonium chloride sublimate to highlight relief.

(G) USNM 198686.

(legend continued on next page)

Burgess Shale-type macrofossils, this three-dimensionality implies resistance to compaction, either through original biomineralization (inconsistent with the organ's trajectory and morphology³) or diagenetic permineralization.^{3,13,14} In Burgess Shale-type deposits, early diagenetic permineralization denotes labile, chemically reactive tissues, particularly gut and cecal systems.^{13,14} Additionally, anterior tracts of the organ are preserved with light-colored sediment infills (cf. Hughes¹⁵) similar to the surrounding matrix (Figures 1A–1C and 1O), suggesting that it was externally open rostrally. Overall, the organ's reflectivity, color, morphology, and relief³ match those of unambiguous gut tissues from other Burgess Shale-type macrofossils, including the chordate *Metaspriggina* (cf. Conway Morris and Caron¹⁶; Figures 1 and S1; cf. Butterfield, Hughes, and Briggs and Whittington^{13–15,17}).

Pikaia's newly identified digestive tract is delineated by diffuse carbonaceous margins (Figures 1A–1C, 1O, 2A, 2F, 2I, and 2J), also visible in *Metaspriggina*¹⁶ (Figure 1; Figure S5 in Conway Morris and Caron¹⁶) and consistent with a pigmented or lipid-rich peritoneum (cf., e.g., Heingård et al.²⁰). Previously uncharacterized, internal lensoid structures suggestive of food boluses (cf., e.g., Tian et al.²¹; Figures 1N³ and 2; Figure 12A in Conway Morris and Caron³) also support a digestive function. Furthermore, some specimens preserve repeated, organic-rich rounded structures with indistinct outlines, flanking the gut ventrally (Figures 1A–1D, 1J–1M, S1F, and S1G) and morphologically similar to the female gonads of cephalochordates (e.g., Yamaguchi and Henmi²²), *Mylokunmingia*,²³ and *Yunnanozoon*.^{18,24} The modest positive relief of these structures relative to the gut canal (Figure 1L) explains its apparently wrinkled or “creased”³ surface, previously interpreted as evidence for a cuticular wall.³

In summary, *Pikaia*'s digestive tract is identified by sediment infills, internal features interpreted as food boluses and phosphatized inclusions,³ and carbonaceous margins. The digestive tract comprises an expanded anterior pharynx and a tubular gut varying in width but tapering caudally. The pharynx comprises ~7%–10% of the total body length and constricts into the tubular gut slightly posterior to *Pikaia*'s anterior appendages³ (Figures 1A, 1E, 1F, 2H, and 2J). The digestive tract terminates at the body's caudal tip, indicating the absence of a post-anal tail (cf. Conway Morris and Caron¹⁶; Figures 1J–1M, 2I, and S1D).

Nervous system

The dark longitudinal strand running along *Pikaia*'s traditionally interpreted ventral margin was tentatively described as a ventral blood vessel,³ but this has been challenged^{7,25} due to

its morphological differences with the main ventral vessels of amphioxus and vertebrates, which branch into a subenteric hepatic portal vein and a ventral aorta separated by hepatic capillaries.⁸ The ventral blood vessel also lacks the adjoining vessels required to complete a circuit.²⁵ Accordingly, Conway Morris and Caron³ hypothetically suggested that the “vessel” may have been a nerve cord. We support and extend this hypothesis by reinterpreting this structure, using new observations and recent advances in the taphonomy of nervous systems,^{26–28} as a dorsal nerve cord (Figures 1A–1F, 1J–1M, 1O, 1P, 2A, S1A, S1C, and S1E).

The nerve cord is preserved as a highly organic-rich residue, consistent with nervous tissues from Burgess Shale-type localities.^{26–28} Occasionally, it is fragmented, indicating that it behaved as a relatively brittle structure during decay and early diagenesis (Figures 2A, 2B, and S1A). It shows modest three-dimensional preservation (Figures 1E and 1F), in contrast to putative cardiovascular systems in Cambrian organically preserved fossils²⁹ (Figure 3A; cf. Figure 3B): its thickness is reflected by occasional uneven splitting between part and counterpart, leaving an intermittent groove in place of original nerve cord tracts (cf. Ortega-Hernández et al.²⁶ and Figure 1) (e.g., Figure 2H; cf. Figure 3H). The nerve cord thickens rostrally into a “bulb”-like³ region (Figures 2B, 2C, and S1E), reminiscent of the tubular brain of cephalochordates and embryonic vertebrates,³⁰ before bifurcating into two carbonaceous strands extending into the innervations³ of *Pikaia*'s cephalic tentacles (Figures 2B, 2C, and 2H–2K; Figures 10B–10G, 10J, and 11A–11D in Conway Morris and Caron³). Together, the well-defined carbonaceous preservation, bilateral symmetry, and topology of *Pikaia*'s nervous system exclude alternative interpretations including cardiovascular or microbial origins²⁶ (cf. Liu et al.³¹).

We find no apparent notochord associated with the dorsal nerve cord in *Pikaia*, and no notochord was conclusively identified by previous studies^{3,7,8,25} (Data S1, character 168). However, this may not reflect a genuine absence given the frequent lack of visible notochords in unambiguous crown-chordate fossils (e.g., Gabbott et al.³²).

Myomeres

As shown by Conway Morris and Caron,³ the organic-rich transverse strands on the trunk of well-preserved *Pikaia* specimens are best interpreted as intercalary spaces between myomeres, coincident with the lipid-rich myosepta³ (cf. Briggs and Kear and Sansom et al.^{33,34}). This interpretation is supported by their

(H) Photograph of (G) coated in ammonium chloride sublimate, showing anterior pharynx in negative relief.

(I) Interpretative drawing of boxed area in (G) and (H); reflective myosepta are represented in dark gray; carbonaceous impressions of myosepta in black.

(J) ROMIP 61236, showing fully preserved digestive tract.

(K) (J) photographed under polarized light.

(L) (J) photographed with ammonium chloride sublimate coating to highlight relief of the gut canal.

(M) Interpretative drawing of (J)–(L); reflective myosepta are represented in dark gray; carbonaceous impressions of myosepta in black.

(N) ROMIP 951804, showing laterally compressed dorsal “fin”³ in profile and frontal view and digestive tract with potential food boluses.

(O) USNM 57628 (lectotype), showing the caudal section of the trunk rotated and preserved in oblique view. Note light-colored sediment infill of the gut canal.

(P) Specimen in (O) photographed with ammonium chloride sublimate coating to highlight relief of gut canal.

USNM, Department of Paleobiology, National Museum of Natural History, Smithsonian Institution; ROMIP, Royal Ontario Museum, Invertebrate Palaeontology; Df, dorsal fin; Vf, ventral fin; Ms, myosepta; My, myomeres; Nc, dorsal nerve cord; Aa, anterior appendages; ?Go, possible gonads; Gu, gut canal; Ph, pharyngeal cavity; An, anus; Tc, tentacles; In, innervation. White arrows in (B), (C), and (O) denote carbonaceous margins associated with the gut canal. Scale bars, 5 mm (A–D, G, and H); 1 mm (E, F, and I); 2 mm (other panels).

See also Figure S1 and Data S1 and S2.

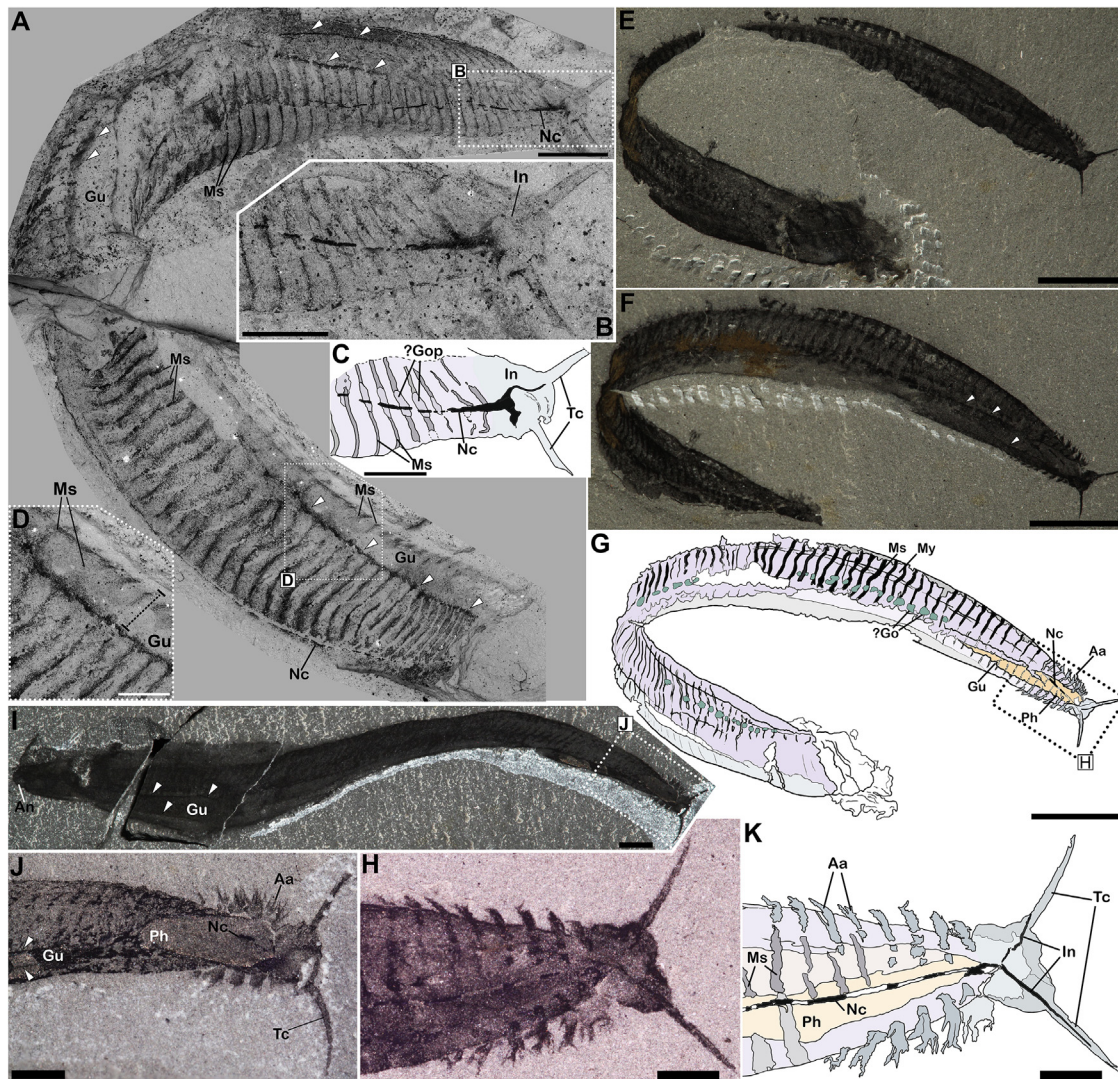


Figure 2. Anatomical reinterpretation of *Pikaia gracilens* Walcott, 1911: Nervous system and axial musculature

(A) Composite image of USNM 202220 under backscattered scanning electron microscopy, showing dark carbon-rich stains corresponding to the myosepta, nervous system remnants, and margins of the gut canal.

(B) Detail of boxed area in (A) showing thickened rostral portion of the dorsal nerve cord, partly preserved innervations of the tentacles, and apparent integumentary perforations recording possible gill openings.

(C) Interpretative drawing of (B).

(D) Detail of corresponding box in (A); dotted line denotes extension of myomeres spanning the width of the gut canal.

(E and F) USNM 198688 (part and counterpart).

(G) Interpretative drawing of USNM 198688 combining part and counterpart.

(H) Detail of boxed area in (F) highlighting rostral portion of the nerve cord, anterior appendages, and cephalic structures.

(I) ROMIP 951067.

(J) Detail of boxed area in (H) showing pharyngeal cavity, anterior-most midgut, and partly preserved nerve cord tract.

(K) Interpretative drawing of (G).

USNM, Department of Paleobiology, National Museum of Natural History, Smithsonian Institution; ROMIP, Royal Ontario Museum, Invertebrate Palaeontology; ?Gop, possible gill openings; Ms, myosepta; Nc, nerve cord; In, innervation; Tc, tentacle; My, myomeres; ?Go, possible gonads; Gu, gut canal; Ph, pharynx; An, anus; Aa, anterior appendages. Scale bars, 2 mm (A and I); 1 mm (B–D, H, J, and K); 5 mm (E–G). White arrows in (A), (F), (I), and (J) denote carbonaceous margins associated with the gut canal.

See also [Figure S1](#) and [Data S1](#), [S2](#), and [S3](#).

orderly arrangement and regular spacing ([Figures 1A–1D](#), [1J](#), [1K](#), [2A](#), [S1B](#), and [S1F](#)). Both contrast with the disorganized, offset configuration of similar organic-rich strands in decayed modern

chordates^{33,34} and *Metaspriggina*,¹⁶ which are often directly superimposed on the matrix following decomposition of surrounding tissues¹⁶ (cf. Briggs and Kear³³). This argues against

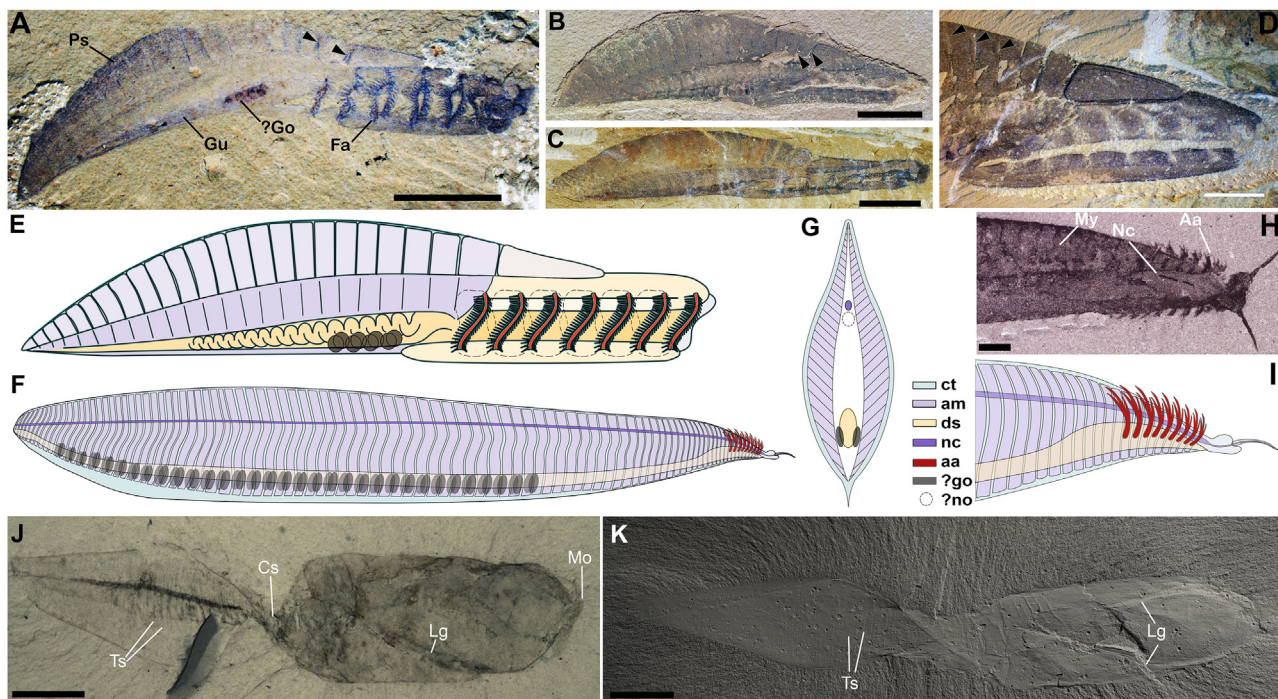


Figure 3. Representative stem-group chordates and comparisons with *Pikaia*

(A–E) *Yunnanozoon*.

(A) RCCBYU 10310, showing gut tube, posterior segments, and filamentous arches.

(B) YKLP 13005.

(C) YKLP 13032.

(D) YKLP 13016. Black arrows denote separation between dorsal “segments.”

(E) Schematic anatomical reconstruction after Cong et al. and Tian et al.^{18,19}

(F) *Pikaia*, schematic reconstruction illustrating dorsoventrally inverted anatomical reinterpretation.

(G) Hypothetical cross-sections of trunk segments in *Pikaia*³ under the new dorsoventrally inverted interpretation, showing dorsal and ventral “fins.”³ Color legend on the right applies to (E)–(I).

(H) USNM 198688, detail of the anterior region showing bilobed head, anterior appendages, pharyngeal cavity, and dorsal nerve cord.

(I) Schematic reconstruction of the anterior region of *Pikaia* under the new anatomical interpretation, showing the anterior appendages as dorsally directed gills.

(J and K) *Banffia constricta*.

(J) ROMIP 49914.

(K) ROMIP 49898, photographed under ammonium chloride sublimate to highlight relief of lateral grooves and segmental boundaries.

RCCBYU, Research Center of Chengjiang Biota, Yunnan University, Kunming; YKLP, Yunnan Key Laboratory for Palaeobiology, Kunming; USNM, Department of Paleobiology, National Museum of Natural History, Smithsonian Institution; ROMIP, Royal Ontario Museum, Invertebrate Palaeontology; Ps, posterior segments; Gu, gut; ?Go, possible gonads; Fa, filamentous arches; ct, connective tissue, integument; am, axial musculature; ds, digestive system; nc, dorsal nerve cord; aa, anterior appendages; ?no, hypothetical notochord; My, myomeres; Ph, pharynx; Lg, lateral grooves; Cs, constriction; Ts, tail segments; Mo, mouth opening. Scale bars, 5 mm (A, C, and D); 10 mm (B, J, and K); 1 mm (H).

See also [Data S1](#) and [S2](#).

interpreting *Pikaia*’s strands as decaying myomeres shrunk away from myosepta^{3,33} and provides a counterpart to probable myosepta described in fossil vertebrates.³³

Our reinterpretation establishes a new orientation for *Pikaia*’s myomeres (cf. Conway Morris and Caron³). The myomeres, defined by the spaces between the observed myosepta, are sigmoidal along their dorsoventral trajectory and kink or inflect anteriorly in the dorsal portion of the body (Figures 1A–1D, 1J–1M, S1B, S1F, and S1G), consistent with myomeres in crown chordates, including amphioxus, *Mylokunmingia*, and *Metaspriggina*.^{16,35,36} Additionally, they sometimes show weaker, rostrally convex ventral inflections (Figures 1J, 1K, and 2D). The myomeres extend along *Pikaia*’s entire trunk, from behind the bilobed “head”³ to the tip of the tail. Caudally, myosepta

can be traced into the dorsal “fin”³ as fainter impressions (Figures 1A–1C, 1J–1N, and S1H), consistent with them occupying a reduced volume within this laterally compressed extension of the trunk³ (Figure 1N).

External features—fins and gills

Conway Morris and Caron identified laterally compressed dorsal and ventral median “fins” in *Pikaia*,³ inverted in our reconstruction. In the reoriented configuration, a thin, keel-like ventral fin extends for about 80% of the total body length, whereas a more prominent posterior dorsal “fin”³ extends for 30%–40% of total body length. Both fins terminate at the caudal tip of the body. Unlike in crown chordates, the myomeres extend into the dorsal “fin”³ (Figures 1A–1D, 1J–1N, and S1H) with no

evidence for supportive fin rays, suggesting that the fin was more robust and less clearly demarcated from the trunk than in extant chordates.

Pikaia's external, filamentous "anterior appendages"³ (Figures 2H, 2J, 2K, 3H, and 3I) match the configuration of free-hanging gills in aquatic vertebrates and bilaterians more generally, oriented dorsally and away from the substrate (e.g., Nokhbatolfighahai and Downie and Stundl et al.^{37,38}). Their branching morphology and similar preservation of these structures as organic-rich stains in both vertebrate and invertebrate fossils (cf. Gao and Shubin and Caron et al.^{39,40}) further support an interpretation as gills.³

Phylogenetic analysis

Our novel interpretation of *Pikaia* informed a new deuterostome-focused phylogenetic analysis, including 102 taxa and 625 characters (Data S2) and sampling extensively from Li et al. and Tian et al.^{41,19} (Data S1). Twenty-five new characters in our analyses (Data S1) were subject to different scoring criteria testing the robustness of our topology to alternative anatomical interpretations (Data S2 and S3). We incorporated a number of other relevant fossil taxa exhibiting potential shared features with living chordates, including vetulicolians, *Yunnanozoon*, and the stem-group vertebrates *Metaspriggina*¹⁶ and *Mylokunmingia*^{23,35} (Data S2 and S3).

Vetulicolians (Figures 3J and 3K) and *Yunnanozoon* (Figures 3A–3D) show a bipartite body plan with an "anterior body" occupied by a pharynx lined by lateral arches or pores and a "posterior body" partitioned into metameric units with a recalcitrant body wall.^{18,19,42,43–46} Both taxa lack apparent myomeres or myosepta,^{18,42,47,45,46} which preserve consistently in other fossils from the same deposits.^{23,35} The anterior body of both taxa comprises two sclerotized "halves" joined along the lateral sides to form a groove-like region, clearly opened to the exterior in vetulicolids⁴³ and lined by the pharyngeal openings.^{18,42–47} The digestive tract extends through the caudal region: in vetulicolians, it extends medially and opens into a terminal anus; in *Yunnanozoon* it is displaced ventrally as in *Pikaia*.^{18,19,42,43–46}

Tree searches were performed in MrBayes 3.2.7,⁴⁸ using Bayesian Inference under an MK + gamma model.⁴⁹ We recover *Pikaia* as a stem chordate, subtending the crown node with high posterior probability ($p = 98$; Figure 4A). Together with *Yunnanozoon* and *Cathaymyrus*,⁶ *Pikaia* forms a grade of stem chordates crownward of vetulicolians, resolved as a grade of earlier diverging chordates (Figure 4A). The topology is robust to alternative character coding, taxon sampling, and model choices (Data S1; Figures S2–S4). The alternative scenario of vetulicolian monophyly is recovered as unlikely in our steppingstone sampling analysis ($2 \times \log_e B_{10} = -7.20$; Table S1).

DISCUSSION

Our reinterpretation of *Pikaia* provides new evidence for diagnostic chordate synapomorphies: a dorsal nerve cord and unambiguous myomeres.^{3,51} By contrast, *Pikaia*'s digestive tract has a terminal anus, unlike in crown chordates,^{16,35,41} which possess a post-anal tail.^{16,35,41} We resolve *Pikaia* as a stem chordate (Figures 4 and S2–S4), elucidating the sequence of character acquisition in the group.

The sigmoidal myosepta of *Pikaia*³ show conspicuous dorsal inflections pointing cranially, as in vertebrates^{7,16,35} and amphioxus,^{7,25,35} but their weakly recurved ventral sections confirm a more similar shape to vertebrates.⁷ *Pikaia*'s weakly bent myomeres are intermediate between the steeply angled crown-chordate muscle blocks^{7,16,25,35,36} and the subrectangular axial units of *Yunnanozoon*,^{18,42,43,47,52} recovered immediately stemward of *Pikaia* (Figures 4A and 4C). This topology suggests a transition toward increasingly inflected segmental musculature along the chordate stem²⁵ (Figure 4C), likely accompanying the evolution of faster, more powerful swimming.²⁵ The anatomical correspondence between myomeres and yunnanozoan axial units is independently supported by the extension of myomeres into *Pikaia*'s dorsal fin. Metameric muscle blocks extending into a dorsal fin³ are unknown in crown chordates^{7,16,25,35,36} but find plausible counterparts in *Yunnanozoon*'s fin-like dorsal region,^{18,42,43,47} partitioned into segmental blocks (Figures 3A–3E). Therefore, homology between myomeres and *Yunnanozoon*'s axial units is upheld by positional and morphological similarity, as well as congruence with other homologies⁵³: *Pikaia*'s filamentous "anterior appendages,"³ suggestive of external free-hanging gills, further support an affinity with *Yunnanozoon* by providing a counterpart to its similarly exposed and filamentous arches^{3,19} (Figures 3A and 3E^{18,42,43,47}).

Yunnanozoon, in turn, shares characters with the vetulicolians, recovered as stem-group chordates.^{41,19,52} These include a bipartite body plan with a voluminous anterior pharynx and a segmented tail, a medial anterior cleft reminiscent of the vetulicolian "lateral groove," and a robust cuticle-like integument^{18,42–47,52,54} (Figures 3A–3E). The homology of these characters is supported by our phylogenetic results (Figures 4A and S2–S4). A chordate placement for vetulicolians^{41,19,52} fits with the recognition of a terminal anus in stem-group ambulacrarians,⁴¹ supporting independent origins of post-anal structures in ambulacrarians and chordates⁴¹ and undermining the main rationale for an alternative, stem-deuterostome placement.⁴⁶ Serialized lateral openings, segments of probable mesodermal⁴⁵ origin, a voluminous anterior pharynx, and laterally compressed bodies in most vetulicolians also support a chordate affinity.^{1,41} However, unlike other vetulicolians, banffozoans (Figures 3J and 3K)—recovered for the first time as the earliest diverging stem chordates (Figure 4A)—lack a laterally compressed body plan and visible lateral pouches or other serialized pharyngeal structures,^{44,46,54} suggesting that these traits were acquired in a stepwise fashion in a paraphyletic vetulicolian stem group (Figure 4A). If so, pharyngeal openings in the deuterostome last common ancestor might have been non-serialized pores,^{41,55} as in the stem-group ambulacrarian *Herpetogaster*.^{41,50}

The grade of "bipartite animals" comprising vetulicolians and *Yunnanozoon* on the chordate stem prompts us to propose a new evolutionary scenario reminiscent of Romer's "somato-visceral" hypothesis of chordate origins⁴ (Figure 4B). Drawing on vertebrate neuroanatomy and metamerism, and inspired by Garstang's⁵⁶ tunicate-based model of the chordate ancestor, Romer proposed that the archetypal chordate bauplan was subdivided into a "somatic"⁴ posterior body comprising the central nervous system, axial skeleton, and somites and a "visceral"⁴ anterior, encompassing the pharynx and digestive organs. Romer's hypothesis addressed the distinct embryonic origins and

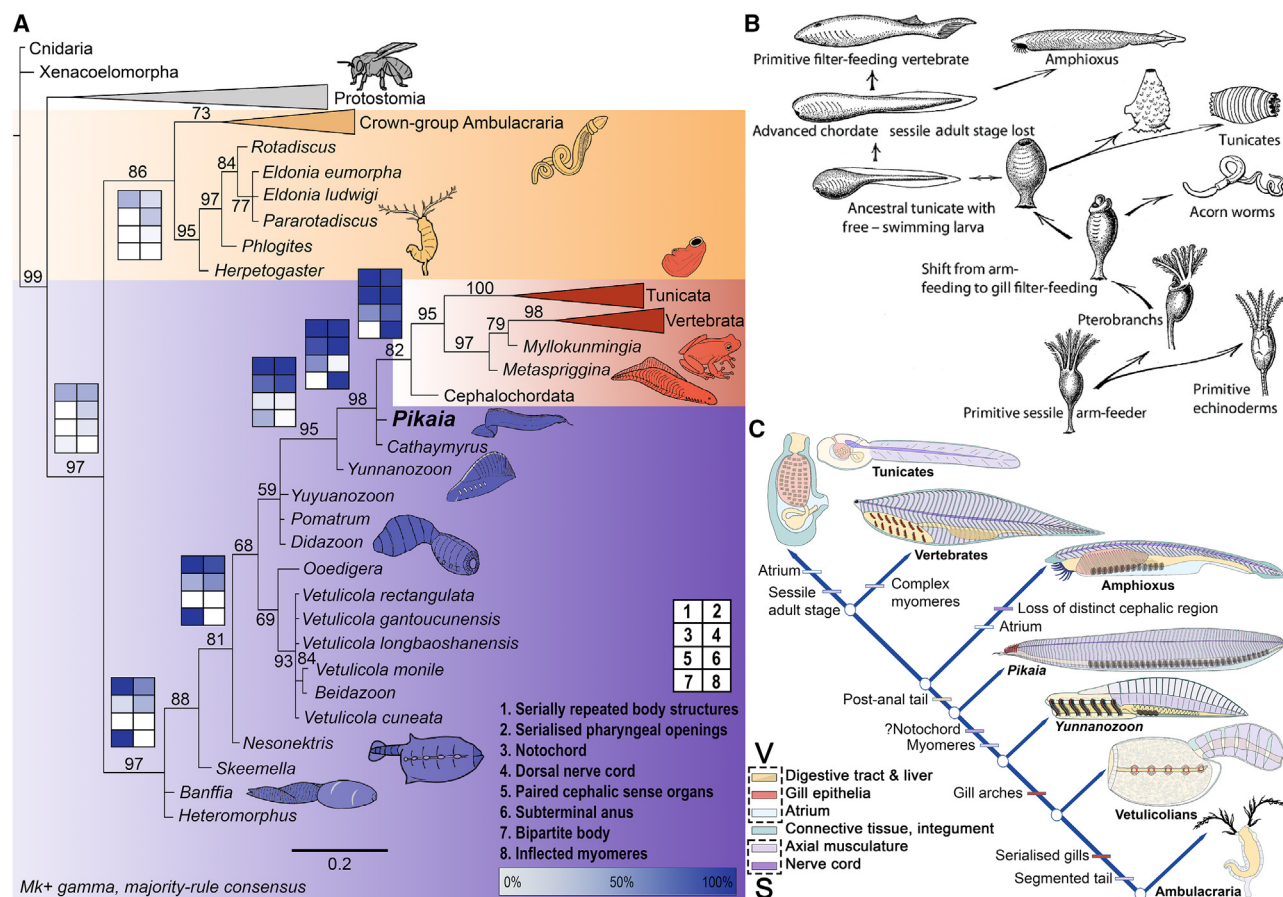


Figure 4. The phylogenetic relationships and evolutionary implications of stem-group chordates

(A) Phylogenetic tree of Bilateria based on Bayesian inference (BI), showing the position of *Pikaia* and other Cambrian deuterostomes. Branch lengths are proportional to the number of character state changes (scale bar on the bottom left). Node numbers indicate posterior probability values (BI). Tie plots show reconstructed ancestral state probabilities (0% to 100%; legend on the bottom right) at their corresponding nodes. Yellow background shading denotes ambulacrarians; purple, stem-group chordates; red, crown-group chordates.

(B) Romer's hypothetical "somato-visceral" scenario of chordate evolution, adapted from Romer,⁴ for comparison with our updated hypothesis in (C).

(C) Scenario of chordate evolution based on (A), charting the integration of somatic (S) and visceral (V) components (legend at the bottom left) and the acquisition of key chordate characters (colored bars). Stem-group vertebrate (*Metaspriggina*) after Morris and Caron,¹⁶ *Yunnanozoon* after Cong et al. and Tian et al.,^{18,19} and stem-group ambulacrarian (*Herpetogaster*) after Caron et al.⁵⁰

See also Data S1, S2, and S3 and Figures S2–S4.

anatomical offset of chordate somites and gill arches and their manifestations in associated muscular, nervous, and skeletal structures.⁴ It envisaged a gradual integration of the two "bodies," with the "visceral" encapsulated by the anterodorsal extension of the "somatic."⁴

Gee¹ speculated that the bipartite vetulicolian body plan embodied Romer's⁴ hypothetical ancestral chordate. Under this interpretation, the "somatic" posterior unit would correspond to the segmented tail of vetulicolians, and the "visceral" unit to the anterior body.¹ This hypothesis is supported by recovering a grade of vetulicolians stemward of all other chordates and reconstructing their bipartite condition as plesiomorphic for the phylum (Figures 4A and S2–S4). Moreover, the recovery of *Yunnanozoon* as a phylogenetic intermediate between vetulicolians and more derived chordates lacking a bipartite body plan (Figure 4C) supports a trend toward greater integration of the two "bodies"⁴ along the chordate stem.

Our revised "somato-visceral"⁴ model fits evolutionary-developmental data revealing highly conserved gene-regulatory elements among deuterostomes, including modular gene clusters underpinning gill-slit development,⁵⁷ cardiopharyngeal regulatory networks,⁵⁸ and core modules for notochord and neural tube development.⁵⁹ These discrete gene-regulatory elements, showing conserved and functionally important syntenic linkages,⁵⁷ suggest ancient origins as distinct developmental and functional modules for the defining "visceral" and "somatic" organs⁴: the pharyngeal apparatus,⁵⁷ and the central nervous system and axial skeleton.⁵⁹ Moreover, our hypothesis reconciles stem-chordate morphologies with the conserved organization and Hox gene expression patterns of the deuterostome trunk (cf. Lacalli⁸): the vetulicolian and yunnanozoan "posterior bodies" would comprise somite homologs, whereas the gill-bearing anterior would embody the condition of the trunk prior to the evolution and rostral extension of somites,^{1,4}

as exemplified by hemichordates⁸ and possibly early-diverging echinoderms.⁶⁰ By contrast, both the bilobed cephalic unit of *Pikaia* and the lobate circumoral rim of *Yunnanozoon*^{3,19} lie anterior to the trunk and therefore the anterior-most boundary of Hox gene expression,⁸ supporting a “head” identity.⁸ *Pikaia*’s distinct head with paired, cerebrally innervated sense organs suggests simplification of the anterior central nervous system in amphioxus (Figures 4A and S2B), consistent with secondary losses of complex neuroectodermal signaling centers in cephalochordates.⁶¹

This scenario restores *Pikaia* from specialized and phylogenetically “peripheral”^{7,35} to a keystone evolutionary intermediate, illuminating the morphology of the chordate LCA (Figure 4). *Pikaia* and other Cambrian problematica exhibit a mosaic of overlapping plesiomorphic features and nested subsets of crown-group traits, as expected for stem-group relatives of any clade.⁶² Our scenario also restores Romer’s “dual animal”⁴ from useful heuristic device⁶³ to genuine evolutionary model, supplying a historical explanation for the evolutionary-developmental evidence in its favor^{1,57–59,64} and elucidating the assembly of an ecologically revolutionary body plan.^{65,66}

STAR★METHODS

Detailed methods are provided in the online version of this paper and include the following:

- KEY RESOURCES TABLE
- RESOURCE AVAILABILITY
 - Lead contact
 - Materials availability
 - Data and code availability
- EXPERIMENTAL MODEL AND SUBJECT DETAILS
- METHOD DETAILS

SUPPLEMENTAL INFORMATION

Supplemental information can be found online at <https://doi.org/10.1016/j.cub.2024.05.026>.

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AUTHOR CONTRIBUTIONS

J.V. first suggested a dorsoventrally inverted reinterpretation of *Pikaia*. G.M., F.S.D., M.P.S., and J.V. conceptualized the study. G.M. conducted specimen descriptions and interpretations with input from J.V., F.S.D., and M.P.S.; G.M., F.S.D., M.P.S., I.A.R., J.V., and D.J.E.M. contributed to assembling the phylogenetic dataset. G.M. ran the phylogenetic analyses, made the figures, and wrote the original draft of the manuscript. All authors contributed to review and editing.

DECLARATION OF INTERESTS

The authors declare no competing interests.

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STAR★METHODS

KEY RESOURCES TABLE

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Deposited data		
Morphological character matrices in NEXUS format and as readable Mesquite files	This study	Data S1 , S2 , and S3
Software and algorithms		
MrBayes 3.2.6	Ronquist et al. ⁴⁸	http://nbisweden.github.io/MrBayes/
Other		
<i>Pikaia</i> specimens	National Museum of Natural History, Smithsonian Institution; Royal Ontario Museum, Invertebrate Paleontology	USNM, ROIMP

RESOURCE AVAILABILITY

Lead contact

Further information and requests should be directed to and will be fulfilled by the lead contact, Giovanni Mussini (gm726@cam.ac.uk).

Materials availability

All specimens of *Pikaia* illustrated and analyzed in this study are deposited at the Royal Ontario Museum (ROMIP) or at the National Museum of Natural History, Smithsonian Institution (USNM), with accession numbers provided in the captions of [Figures 1](#), [2](#), and [S1](#). All data analyzed in this study are available as part of the Report, [Figures 1](#), [2](#), [3](#), [4](#), and [S1–S4](#), or in the [Data S1](#).

Data and code availability

Our main phylogenetic dataset is available as a readable Mesquite file in [Data S2](#), with character definitions listed in [Data S1](#). Our alternative phylogenetic dataset is provided as a Mesquite file in [Data S3](#). The parameters used for the phylogenetic analysis are described in the [STAR Methods](#) section below. Details of the MrBayes code used in our phylogenetic analyses are available in our [Data S1](#).

EXPERIMENTAL MODEL AND SUBJECT DETAILS

Photographs of previously described specimens of *Pikaia*³ were made and provided by Jean-Bernard Caron. All illustrated specimens are deposited at the Royal Ontario Museum (ROM) or at the National Museum of Natural History, Smithsonian Institution (USNM). Accession numbers are provided in the captions of [Figures 1](#), [2](#), [3](#), and [S1](#).

METHOD DETAILS

Phylogenetic analyses were performed using Bayesian Inference (BI), recently shown to be more robust to missing morphological data than alternative parsimony-based techniques^{67–70} and generally outperform them in topological accuracy.^{70–72} Markov chain Monte Carlo (MCMC) tree searches were performed in MrBayes 3.2.7⁴⁸ under a MK + gamma model.⁴⁹ Two parallel runs of 100,000,000 generations were requested, with 25% of samples discarded as burn-in. Our analyses only included characters which are parsimony informative ('informative' using the `lset` function) to account for the ascertainment bias. Convergence was checked using the `sump` command (standard deviation of split frequencies < 0.01, ESS scores > 200, PSRF ~ 1.0 across all parameters) and the results visualized as a majority-rule consensus tree ([Figure 4A](#)). The MCMC search was repeated using alternative models, topological constraints, character coding, and taxon sampling (see [Data S1](#) and [S3](#)).

To test the sensitivity of our topology, we reran our analyses using two datasets with alternative scorings of key characters (see [Data S1](#) and [S3](#) for alternative phylogenetic dataset). To estimate the marginal likelihood of our original tree ([Figure 4A](#)) against a scenario of vetulicolian monophyly, the MCMC search was also repeated after constraining vetulicolians as a clade ([Data S1](#); [Figure S4A](#)). Then, a stepping-stone sampling analysis⁷³ (SSA) was performed for both resulting topologies by applying hard constraints to all nodes and using two independent runs of sampling, with 40,000,000 generations (~10 times the highest number required for

convergence) requested. Both analyses used 50 steps. A Bayes factor comparison⁷⁴ was used to test support for the original and alternative, monophyletic topologies ([Data S1](#)). Bayes factors for hypothesis testing were interpreted after Bergsten et al.⁷⁵ ([Table S1](#)).

An ancestral states reconstruction (ASR) was performed on the unconstrained Bayesian tree ([Figure 4A](#); [Data S1](#)) using the method outlined in Huelsenbeck and Bollback⁷⁶ to integrate phylogenetic uncertainty across all parameters, including topology. Hard constraints were applied across the entire tree, with model settings matching those of the MCMC search. Ancestral state probabilities were recovered using the report function and visualized through Tieplots ([Figure 4A](#)).

Current Biology, Volume 34

Supplemental Information

A new interpretation of *Pikaia* reveals the origins of the chordate body plan

Giovanni Mussini, M. Paul Smith, Jakob Vinther, Imran A. Rahman, Duncan J.E. Murdock, David A.T. Harper, and Frances S. Dunn

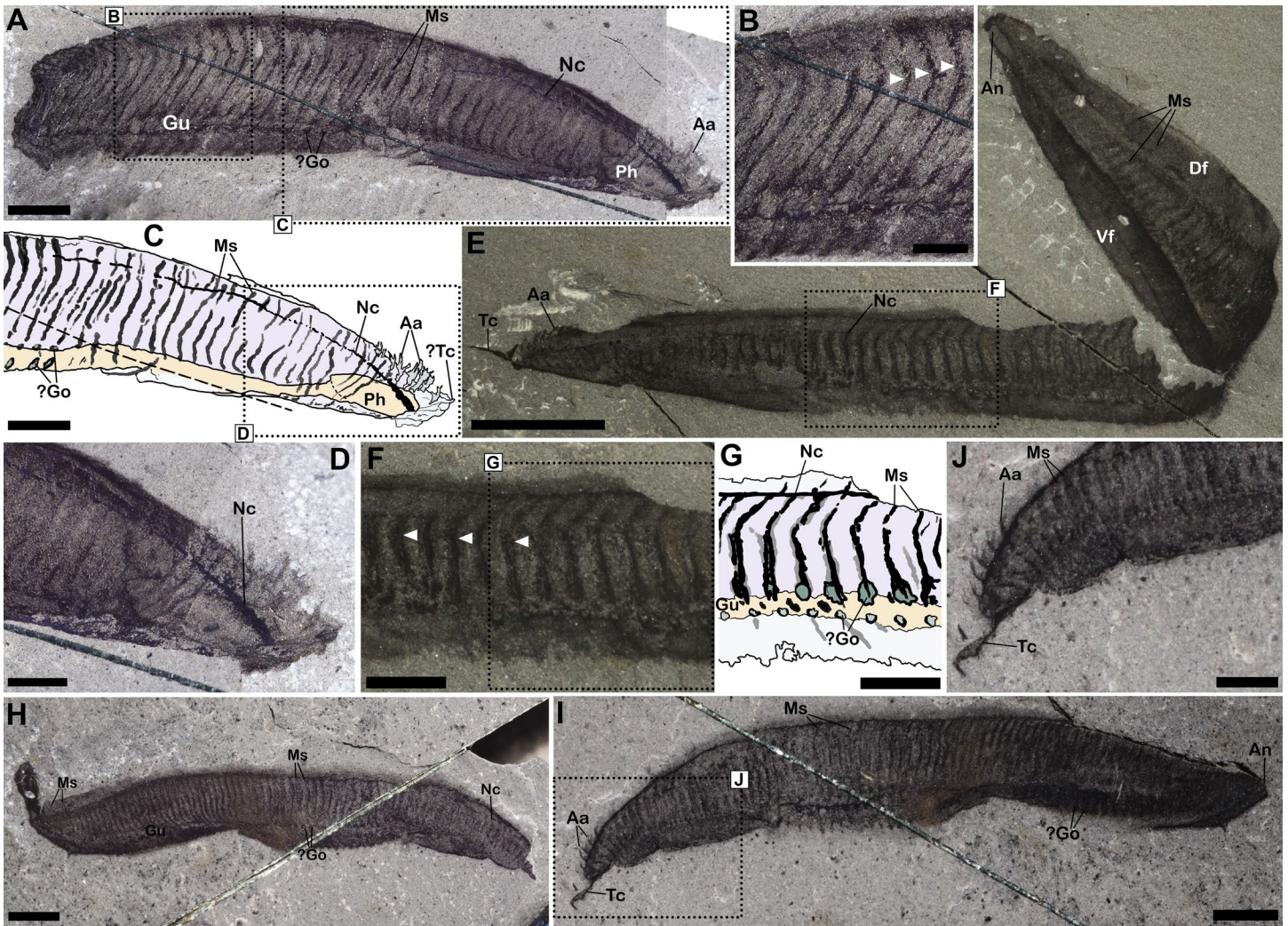


Figure S1. *Pikaia gracilens* Walcott 1911, additional specimens. Related to Figures 1-2. **A**, ROMIP 61191, showing the rostral portion of the body. Merged image of part and counterpart (using the function 'Apply' and the 'Darken' blending mode in Photoshop®). **B**, detail of boxed area in **A**, showing rostral inflections in the dorsal portion of myomeres. **C**, interpretative drawing of boxed area in **A**, with annotated morphological structures. **D**, detail of boxed area in **C**, showing rostral expansion of dorsal nerve cord. **E**, USNM 200530, a specimen showing rostrally inflected myomeres, anterior appendages, dorsal and ventral fins, and terminal anus. **F**, detail of boxed area in **E** showing myomere inflections dorsal to the digestive tract. **G**, interpretative drawing of boxed area in **F**. **H-I**, ROM 61188 (part and counterpart). **J**, detail of boxed area in **I**, showing cephalic tentacles and dorsally pointing anterior appendages. Abbreviations: USNM, Department of Paleobiology, National Museum of Natural History, Smithsonian Institution; ROMIP, Royal Ontario Museum, Invertebrate Palaeontology. Df, dorsal fin; Vf, ventral fin; Ms, myosepta; Nc, dorsal nerve cord; Aa, anterior appendages; ?Go, possible gonads; Gu, gut canal; Ph, pharyngeal cavity; An, anus; Tc, tentacles. White arrows indicate direction of myomere dorsal inflections. Scale bars: 2 mm except for **B**, **D**, **J** (1 mm) and **E** (5 mm).

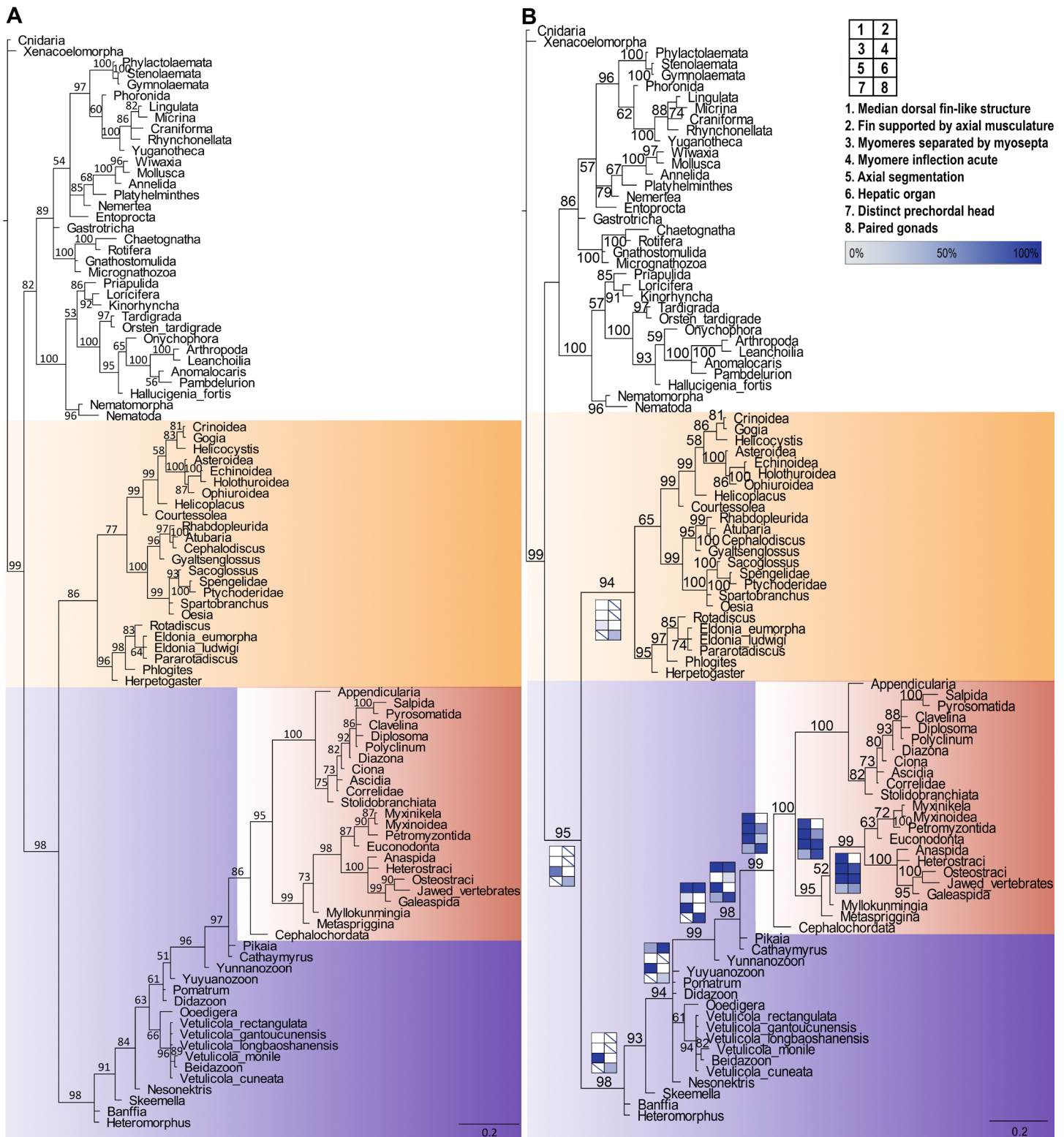


Figure S2. Supplemental phylogenetic trees. Related to Figure 4 and the STAR methods. **A**, Majority rule consensus tree using BI under the Mk model, with dataset (Data S2) analysed under the log normal (instead of gamma; cf. Figure 4a) distribution, using symmetrical transition frequencies. **B**, Majority rule consensus tree using BI under the Mk model, based on our alternatively scored dataset (Data S3) analysed under the gamma distribution, and using symmetrical transition frequencies (the default option in MrBayes). Node numbers indicate posterior probability support values. Branch lengths are proportional to the number of character state changes (scale bars at the bottom of the image); node numbers indicate posterior probability support values; yellow background shading denotes ambulacrarians, purple stem-group chordates, red

crown-group chordates. Tieplots show reconstructed ancestral states probabilities for characters of interest (0 to 100%; legend on the bottom right) at the corresponding nodes; transverse blue bars denote inapplicable character reconstructions.

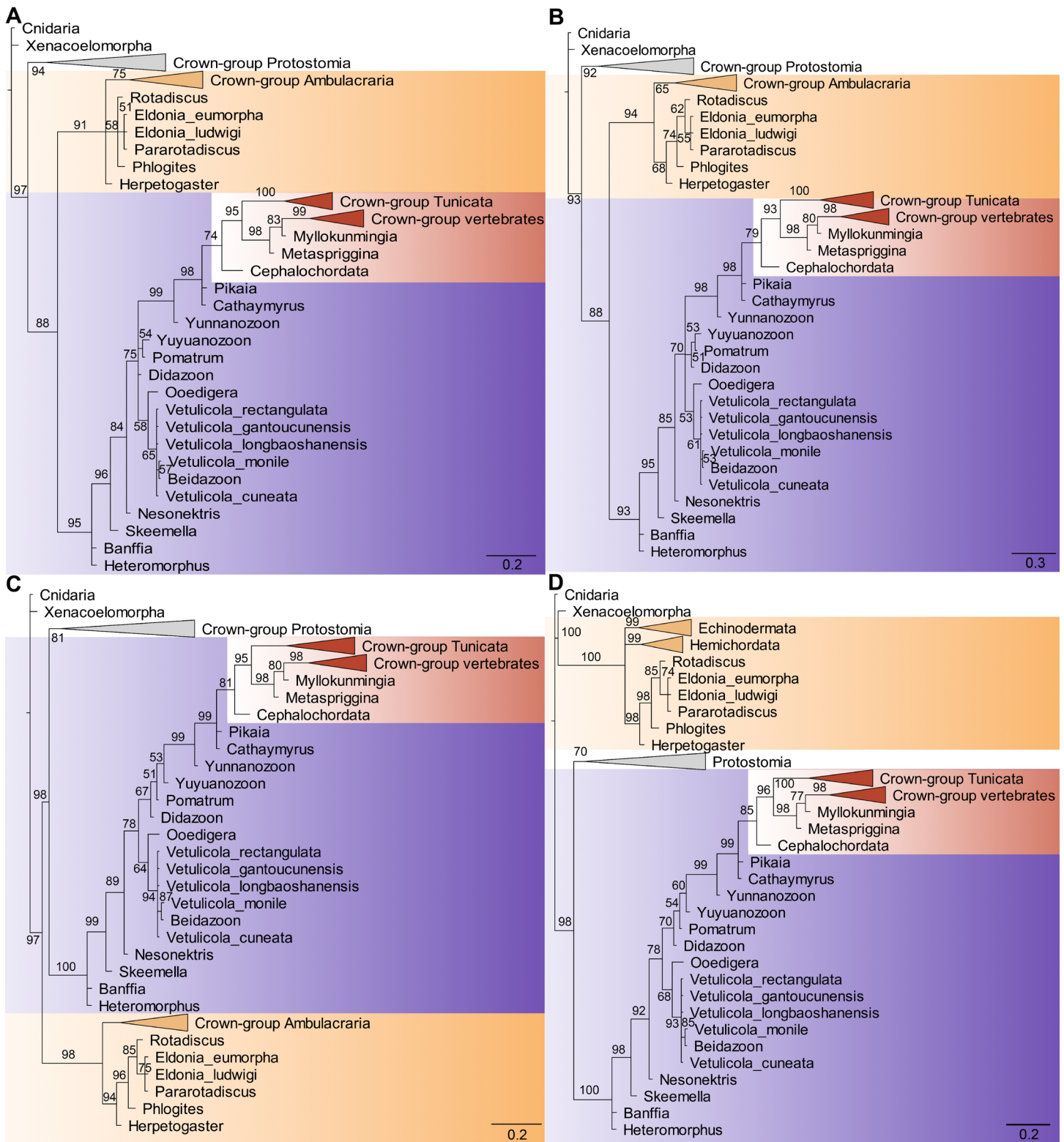


Figure S3. Additional supplemental phylogenetic trees. Related to Figure 4 and the STAR methods. A, Majority rule consensus tree based on our main dataset (Data S2) analysed using the Mk model under the gamma distribution using asymmetrical transition frequencies, corresponding to a uniform prior on the proportions of state frequencies. **B,** Majority

rule consensus tree based on our main dataset (Data S2) analysed using the Mk model under the log normal distribution, using asymmetrical transition frequencies. **C**, Majority rule consensus tree based on Data S2 and using the Mk + gamma model (symmetrical transition frequencies), with Chordata and Protostomia constrained as sister-taxa. **D**, Majority rule consensus tree based on Data S2 and using the Mk + gamma model (symmetrical transition frequencies), with Xenacoelomorpha constrained as the sister-group to the ambulacrarians. Note that by imposing this constraint deuterostomes are recovered as paraphyletic. Branch lengths are proportional to the number of character state changes (scale bars at the bottom of the image); yellow background shading denotes ambulacrarians, purple stem-group chordates, red crown-group chordates. Node numbers indicate posterior probability support values.

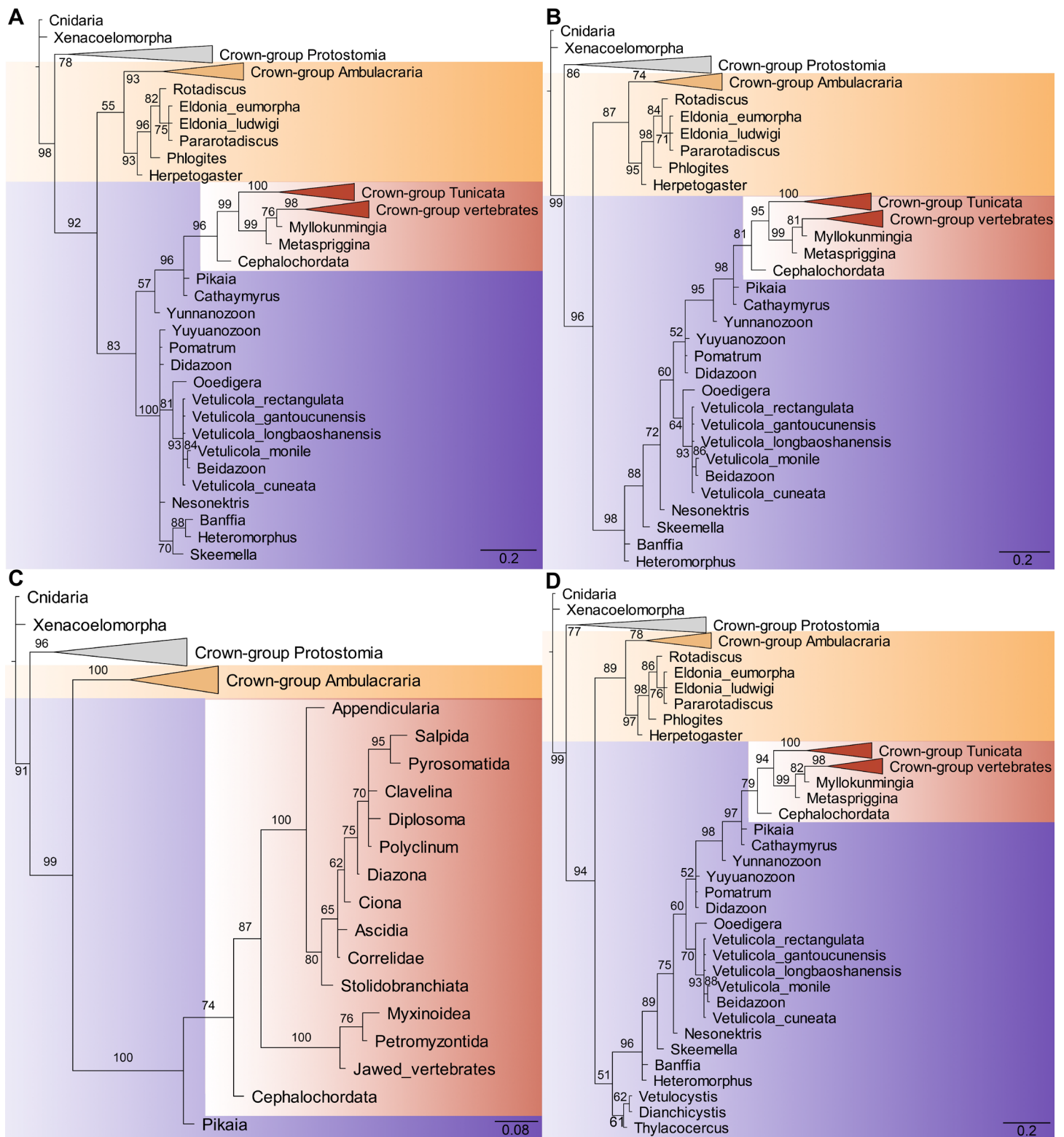


Figure S4. Additional supplemental phylogenetic trees. Related to Figure 4 and the STAR methods. **A**, Majority rule consensus trees yielded by BI based on Data S2 under the Mk + gamma model (symmetrical transition frequencies) with vetulicolians constrained as monophyletic. **B**, Majority rule consensus trees based on Data S2 under the Mk + gamma model (symmetrical transition frequencies) with a notochord scored as present in *Nesonektris*. **C**, Majority rule consensus trees based on Data S2, yielded by the Mk + gamma model (symmetrical transition frequencies), with all fossil taxa excluded except for *Pikaia*. **D**, Majority rule consensus trees based on Data S2, yielded by the Mk + gamma model (symmetrical transition frequencies), including vetulocystids, tentatively recovered as a clade of total-group chordates stemward of

Banffia. Branch lengths are proportional to the number of character state changes (scale bars at the bottom of the image); yellow background shading denotes ambulacrarians, purple stem-group chordates, red crown-group chordates. Node numbers indicate posterior probability support values.

SS run	Marg. Log Lik. H0 = Vetulicolian paraphyly (Figure 4A)	Marg. Log Lik. H1 = Vetulicolian monophyly (Figure S4A)	$2 \times \log_e B_{10}$ (H0 vs H1)	Interpretation
1	- 3432.42	- 3439.63	-7,21	Vetulicolian paraphyly strongly supported
2	- 3432.42	- 3439.61	-7,19	
Mean	- 3432.42	- 3439.62	- 7,20	

Table S1. Steppingstone sampling analysis results. Related to Figure 4 and the STAR methods. Results of steppingstone sampling analysis showing Bayes factor comparison of mean marginal likelihoods with accompanying interpretation. Marginal likelihood values of vetulicolian paraphyly (H0) and monophyly (H1) are estimated using two independent steppingstone sampling runs (see STAR Methods).

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